Fish skin acellular dermal matrix: potential in the treatment of chronic wounds

Munir Patel
John C Lantis II
Department of Vascular Surgery, Mount Sinai St. Luke’s-West Hospital Center, New York, NY, USA

Abstract: The role of cellular and tissue-based therapies (CTPs) in the treatment of chronic wounds continues to grow. However, the utility of these is only partially investigated and very few head-to-head studies exist. In general, the ideal CTP would provide a structural matrix which would allow for dermal and epidermal regeneration with a single application. At present, this Holy Grail is very far from where we are. However, new CTPs continue to be designed with the goal of wound closure in mind. Acellular fish skin (AFS) is one such therapy. The histologic features make it amenable to cellular modulation, and its ability to provide omega-3 fatty acids to the local tissue makes it an unprecedented anti-inflammatory, antibacterial, antiviral, and analgesic component. There is burgeoning research on AFS that continues to validate its efficacy, although, much more substantial data will be required before its general acceptance. In this review, we will go through the current literature to evaluate the strengths and gaps in our current knowledge with the goal of providing a context for use in modern clinical practice.

Keywords: fish skin, chronic wounds, dermal matrix

Introduction

The role of cellular and tissue-based therapies (CTPs) in the treatment of chronic wounds continues to grow. However, the utility of these is only partially investigated and very few head-to-head studies exist. In general, the ideal CTP would provide a structural matrix which would allow for dermal and epidermal regeneration with a single application. At present, this Holy Grail is very far from where we are. Currently, there are broadly three categories of CTPs; they are the ones that combine living cells and a collagen matrix, extracellular matrices (ECMs), and amniotic tissues. The living cellular therapies functionally allow for the deposition of growth factors and beneficial cytokines. Early on, these showed great promise; they were used in a host of injuries— including acute wounds. Unfortunately, few large-scale studies were done and the ones that were done successfully directed these therapies to treat venous leg ulcers and diabetic foot ulcers.1–3 The amniotic tissues fundamentally call in mesenchymal precursor cells, and generally require frequent applications.

However, functional skin cannot be created without a functional ECM. There are three methods to enhance the ECM function in a wound: protect against the degradation of ECM, enhance its deposition, and replace it out of the box. Of the three methods, to replace it out of the box is the most clinically useful. There are broadly three categories of ECMs that one can apply right off the shelf. They are made up of xenografts (from

Correspondence: Munir Patel
Department of Vascular Surgery, Mount Sinai St. Luke’s-West Hospital Center, 1090 Amsterdam Avenue, 12th Floor, New York, NY 10025, USA
Tel +1 212 523 4797
Fax +1 212 523 4720
Email Munir.patel@mountsinai.org
nonhuman sources), allografts, and bioengineered products. Within the subset of xenografts, there is a continuum from modulatory ECMs to scaffold ECMs. One can think of these as products that require frequent application (weekly) and modulate the chronic wound environment to ones that only require one application and become highly incorporated into the wound bed. No ECM is completely one or the other, but ECMs may lean one way or the other. An example of a modulatory ECM, possibly with unique properties, is a xenograft composed of North Atlantic cod fish skin which is naturally rich in omega-3 fatty acids (acellular fish skin, AFS) (Figure 1).

**AFS origin and properties**

AFS is remarkably similar to human skin, yet fundamentally different from mammalian-derived matrices, because of the preservation of structure, lipids, and other soluble components. Mammalian scaffolds require harsh chemical processing to reduce viral and prion transmission risk; such risk from the Atlantic cod (*Gadus morhua*), found mainly in the salt waters of the Atlantic ocean, is nonexistent. Fish skin grafts are therefore subjected to gentle processing where structure and bioactive composition, including omega-3 polyunsaturated fatty acids, are preserved. Studies have shown that omega-3 fatty acids possess antiviral/antibacterial properties and also act as regulators of inflammation.

Acellular mammalian-derived products possess prolonged shelf life compared to cellular skin substitutes, but, per US Food and Drug Administration (FDA) requirements, have to undergo “viral inactivation”. Viral inactivation is performed with detergents that remove all soluble components from the tissue leaving behind an inert matrix of collagenous structure. The harsh viral inactivation processing removes lipids, glycans, elastins, hyaluronic acid, soluble collagen, and other important biologic components from the tissue that are potentially beneficial to wound healing. The fish skin graft has been used to treat a large number of wounds of various etiologies, both acute and chronic.

For the purpose of this paper, we will review the literature related to AFS, while trying to place it the context of this study. When possible, we will make connections between the clinical and basic science, while relaying some practical pearls.

**Prospective double-blind controlled trials in an acute forearm punch biopsy model**

Like most ECMs, AFS had to go through the 510 (k) process which is an FDA regulatory pathway that states that a device is “substantially equivalent” to a predicate device (one that has been cleared by the FDA or marketed before 1976). Most ECMs demonstrate similarities to porcine submucosa. In total, there have been two blinded randomized controlled...
clinical trials showing that treatment of acute forearm wounds with AFS allows for significantly faster wound closure than porcine small intestine submucosa (SIS)-derived matrices and dehydrated human amnion/chorion membrane (dHACM).

In 2014, the manufacturer of AFS needed to show the FDA that AFS was similar or superior to a presently commercially available ECM product in its ability to close wounds. As in almost all xenograft ECM product evaluations, they selected SIS as a predicate product. They then undertook a study with 81 healthy volunteers in which 162 acute 4 mm forearm punch model wounds were treated with SIS or AFS. There was significantly ($P=0.041$) faster wound closure in the AFS group (Figure 2), and no autoimmune reaction, as evidenced by ELISA tests on rheumatoid factor, antinuclear antibodies, antinuclear cytoplasmic antibody, extractable nuclear antibody, anti-double stranded DNA, anti-cyclic citrullinated peptide, and anticollagen I and II. These positive data allowed the entry of the AFS to the US market through the 510 (k) pathway.

In late 2017 and early 2018, in response to the massive if somewhat unexplainable interest and use of amniotic/chorionic products in the chronic wound marketplace, the makers of AFS sponsored another head-to-head acute wound closure study of dHACM vs AFS in the same 4 mm forearm punch model with 170 wounds. Eight-five wounds were treated with dHACM, and 85 were treated with AFS. Wounds treated with fish skin healed significantly faster over a 28-day period, with an HR of 2.37 (95% CI: 1.75–3.21) at a $P$-value of 0.001, compared to dHACM allograft-treated wounds (Figure 3).

While the results of acute wound models are difficult to apply to the chronic wound patients, one can postulate that after complete surgical excision of a wound the local milieu may be similar. However, the patients are not similar. In both these studies, the patients were healthy volunteers in their third decade (on average) of life, which is usually not the case in the chronic wound population. Though, in both cases, the AFS promoted faster wound closure than comparative therapies, which would support the thought that at minimum it is a superior biologic dressing.

**Histologic features of Atlantic cod fish skin**

When studying the mechanism of action of an ECM, it may be “low hanging fruit” to simply look at the histologic properties of the acellular ECM and compare them to those of the native acellular ECM of human skin. While there may indeed be a sweet spot for this ECM anatomic profile, it is somewhat hard to support though many commercial companies do. In the case of AFS, this is no different. Histologic analysis demonstrates that after grafting, the AFS is incorporated into the damaged area and infiltrated by autologous cells that convert the graft into functional, living tissue while the graft slowly breaks down.

Investigation into the properties of the AFS that are important for tissue repair and regeneration has been undertaken employing microscopy-based techniques. Grossly, AFS
has been compared to dHACM allograft, with respect to structure and ability to sustain fibroblast ingrowth in culture. In addition, the ability of the AFS to resist bacterial invasion has been studied.

Porosity and thickness
The porosity of the naturally occurring ECM was counted as the number of pores per 100 µm of sample, and then software was implemented to measure the diameter of the pores. Pores with a diameter of 1 µm or less were not imaged to calculate the diameter of pores under the scanning electron microscope. The microstructure of the biologic materials was evaluated, and it was found that the fish skin graft is highly porous compared to human amnion/chorion membrane. The fish skin contains 16.7 holes (n=6) per 100 µm on average, whereas the dHACM contains 1.7 holes (n=6) per 100 µm on average. The average diameter of the pores in the fish skin graft is 16.1 µm (n=5) and that of pores in the dHACM is 1.3 µm (n=3). The thickness of the dHACM is 20.1 µm (n=6) on average, and the thickness of the fish skin graft is 450 µm (n=6) on average.14

Cell ingrowth assessment and quantification
To assess cell ingrowth capability of the biologic materials, the scaffolds (AFS and dHACM) were seeded with cells. The cell cultures were then incubated. For further assessment, fluorescent staining was carried out with a hydrophobic barrier pen to surround the fixed tissue on the slides.14 Cell ingrowth was assayed and quantitated by counting fluorescently labeled cells within the biologic materials. Cells were only counted when observed within the material. Cells were only counted when observed within the biologic materials, but not if they were on top of the material. Fibroblasts were seen to have infiltrated and remodeled the fish skin graft but did not interact with human amnion/chorion membrane. AFS graft showed superior cell ingrowth when compared to human allograft. After evaluating the porosity of the biologic materials, it was hypothesized that the fish skin grafts are well suited to support cell ingrowth since the pore size is within the range of typical cell size. H&E staining indicated that the fibroblasts remodeled the fish skin graft as they migrated and proliferated into the graft. The fibroblasts, however, formed a layer on top of dHACM allograft material and did not interact with the graft to the same extent. In this specific model, the AFS grafts showed significantly (P<0.0001) more three-dimensional ingrowth of cells when compared to the dHACM. While none of these images alone could provide any specific evidence of how the material behaves, they, in general, support that the ECM is easily penetrated by precursor and circulating blood components and at a 100 µm pore size, which appears to be in line with other ideal ECM structures.14

The antimicrobial properties of fish skin
It also appears from preclinical, small-mammal, and clinical data that the AFS grafts can withstand bacterial invasion for up to 48–72 hours. The original work chronologically started when Dr Baldursson was looking at the closure rate of a mouse burn model in mice treated with AFS. In this model, mice were burned for 30 seconds with brass blocks to create a third degree burn. The wounds were infected 5 minutes post-burn with 400–480×10⁶ colony-forming units.
(cfu) of *Proteus mirabilis* strain ATCC51393. The animals in which AFS was placed on the burn wound before infection with *P. mirabilis* showed 100% survival rate and significantly (*P*<0.05) more survival compared to control (50%) mice and mice treated with collagen matrix product indicated for burns.

In parallel to this work, the barrier function of AFS was assessed. A dilute *Staphylococcus aureus* culture was prepared. Once fully incubated, colonies were counted on the plates and the number of cfu was counted in the initial suspension. Then, 200 µL of precultured bacteria corresponding to 7,000 cfu was inoculated into one of the two chambers of a two-chamber unit in which a membrane made of fish skin to be tested was placed to cover the insert. The chambers were incubated at 37.0°C ±1.0°C. The AFS graft was tested by constraining the material between the two chambers and adding medium with bacteria on the other side and medium only in the second chamber. In this two-chamber assay, the fish skin graft preformed as a bacterial barrier for close to 60 hours compared to 5–45 hours observed for human amnion/chorion membrane products and several collagen matrix products. In addition to this, spiking the fish skin with more omega-3 fatty acids (>24%) enhanced its bacterial barrier function by roughly 80% (Figure 4).14

To follow this up, a very interesting observation was noted in a retrospective review of 54 nonhealing wounds which were treated with AFS for a period of 4 weeks and two applications on average. Improvement was defined as a >20% reduction of wound area. Data were extracted from the patient records database at the Landspitali University Hospital in Reykjavik, Iceland, which was approved by the National Bioethics Committee of Iceland (VSN-15-137). Antibiotics therapy was not a predetermined end point since this was a retrospective study. However, it was noted that after treatment with AFS there was a 38% reduction in antibiotics use (Figure 5). Overall, 87% of nonhealing wounds treated with fish skin grafts either improved or healed after 4 weeks of treatment. Interestingly, while it was only a retrospective observational study, no wound became worse in the course of the 4-week treatment with the AFS.15 These benchtop, small-animal data, supported by clinical observations, create the beginning of a story that supports the possibility that this naturally occurring ECM may have intrinsic antimicrobial properties, which may be secondary to the omega-3 fatty acids that it naturally contains.

**Clinical experience with fish skin**

If one looks at the peer-reviewed literature, other than the 166 patients in the two acute wound randomized controlled trials, there are only 59 patients treated with AFS (Table 1). There are two case series that are referred to in the US on specific wound types, including one by our group evaluating “hard-to-heal” venous leg ulcers, and as part of a very large cohort (yet to be published), and a small subset analysis of diabetic foot ulcers treated in Dr Chris Winters podiatric practice in Indiana, USA.

---

**Figure 4** Efficacy of AFS as a bacterial barrier and the increased efficacy seen with the addition of greater omega-3 fatty acids.

**Notes:** Magnusson S, Baldursson BT, Kjartansson H, Rolfsön O, Sigurjonsson GF, Regenerative and antibacterial properties of acellular fish skin grafts and human Amnion/Chorion membrane: implications for tissue preservation in combat casualty care. Mil Med, 2017, 182, S1, 383–388, by permission of Oxford University Press.14

**Abbreviation:** AFS, acellular fish skin.

**Figure 5** Clinical use of antibiotics before and after the application of AFS in a cohort of chronic wound patients.

**Notes:** Reproduced from Lantis J, Baldursson B, Magnusson S, Kjartansson SG. Acellular fish skin graft for surgical, trauma, venous and arterial and diabetic wounds: a retrospective clinical study. Paper presented at: 17th Congress of the Asian Society for Vascular Surgery; October 20–23, 2016; Singapore.15

**Abbreviation:** AFS, acellular fish skin.
<table>
<thead>
<tr>
<th>Study</th>
<th>Title</th>
<th>Study design</th>
<th>Objective</th>
<th>N</th>
<th>Population</th>
<th>Results</th>
<th>Conclusion</th>
<th>Dosage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fish skin as a treatment for autograft donor sites</td>
<td>Alam and Jeffery</td>
<td>Case study</td>
<td>To test fish skin as a donor-site treatment in patients undergoing skin autografting</td>
<td>10</td>
<td>Patients undergoing split-thickness skin grafting for burn injury</td>
<td>Time to 90% epithelialization was an average of 8.5 days (range 7–13). Time to 100% epithelialization was an average of 11.5 days (range 10–16)</td>
<td>Both the analgesic effect noted and the relatively short average times until 100% reepithelialization are promising and need to be studied further</td>
<td>2–4 applications</td>
</tr>
<tr>
<td>Fish skin for complicated wounds</td>
<td>Dorweiler et al</td>
<td>Case series</td>
<td>To report the cumulative experience of three centers for vascular surgery regarding use of fish skin in wound therapy</td>
<td>23</td>
<td>Patients with vascular disease- and/or diabetes mellitus-associated complicated wounds and partially exposed bony segment wounds</td>
<td>The time to healing varied between 9 and 41 weeks. A reduction of analgesics uptake was noted following fish skin treatment</td>
<td>Fish skin is beneficial to healing with added benefits such as reduced nociception/analgesic consumption, as well as being a possible treatment option in an outpatient setting</td>
<td>9 applications on average</td>
</tr>
<tr>
<td>Improved granulation in wounds with poor circulation</td>
<td>Clasen et al</td>
<td>Case series</td>
<td>To assess fish skin as a remedy for poor granulation in chronic wounds</td>
<td>3</td>
<td>The three cases were all caused by different pathologies: chronic infection, diabetes mellitus with atherosclerosis, and critical ischemia</td>
<td>Granulation tissue formation was induced in all cases. A complete pain relief was seen in morning VAS after 1 week for patient with chronically painful ulcer</td>
<td>Fish skin grafts successfully kickstarted granulation in all of the wounds</td>
<td>1–5 applications</td>
</tr>
<tr>
<td>Fish skin in painful, chronic wound</td>
<td>Cyrek et al</td>
<td>Case study</td>
<td>To treat a painful, chronic, and recurrent ulceration which had not responded to several months of traditional wound therapy</td>
<td>1</td>
<td>86-Year-old female with venous insufficiency</td>
<td>Local wound healing with fish skin resulted in a rapid progressive wound healing with complete healing of the ulcerations within 14 weeks</td>
<td>Fish skin achieved a progressive wound reduction, robust granulation, and epithelialization resulting in complete long-term wound healing. A potential beneficial analgesic effect of the fish skin was noted and resulted in reduction of analgesia over time</td>
<td>14 applications</td>
</tr>
<tr>
<td>Fish skin for the treatment of complicated wounds: experiences associated with amputations in the lower limb in diabetic patients</td>
<td>Trihn et al</td>
<td>Case series</td>
<td>Last-resort treatment of seven wounds at the level of the thigh (n=2) and the forefoot (n=5) in order to avoid further amputation</td>
<td>5</td>
<td>Patients with diabetes mellitus and complicated wounds in the lower limb with exposed bony segments</td>
<td>In all wounds, a reduction of wound size of 50% could be achieved within the first third of treatment. Thigh wounds took 26 weeks to achieve wound closure, and forefoot wounds healed at between 13 and 41 weeks. Analgesics intake was reduced for all patients after the fish skin treatment was initiated</td>
<td>Fish skin represents a viable treatment option for complicated wounds in the lower limb of diabetic patients to circumvent any otherwise-necessary proximalization of amputation level</td>
<td>Weekly application for 13–41 weeks</td>
</tr>
<tr>
<td>Study Title</td>
<td>Study Design</td>
<td>Population</td>
<td>Results</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>---------------------------------------------------------------------------</td>
<td>--------------------------------</td>
<td>--------------------------------------------------</td>
<td>-------------------------------------------------------------------------------------------------------------------</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Efficacy of intact fish skin graft compared to dHACM</strong></td>
<td>Blinded, prospective, randomized, clinical trial on 170 acute wounds shows significantly faster healing rate with intact fish skin compared to dHACM (MiMedx): comparison of time to heal of whole-thickness biopsy wounds</td>
<td>Healthy adults</td>
<td>Wounds in the fish skin-treated group healed significantly faster (HR 2.37; 95% CI: 1.75–3.22; P=0.001) compared to amnion/chorion allograft-treated wounds</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Effectiveness of fish skin in a high risk patient with hemophilia and other comorbidities</strong></td>
<td>Double-blinded prospective RCT</td>
<td>To treat a unique case of dehiscence in a person with diabetes and hemophilia and dehisced chronic wound</td>
<td>Wound healed within 14 weeks after six applications of fish skin</td>
<td>The fish skin was effectively used to treat a chronic wound unresponsive to previous treatments</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Chronic wounds unresponsive to NPWT</strong></td>
<td>Case series</td>
<td>To treat chronic wounds in a real-world environment</td>
<td>75% of the wounds healed within the study period</td>
<td>There was efficient and rapid granulation tissue formation seen in many of the wounds and improved esthetic outcomes — reduced scar formation from what would be expected</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Prospective evaluation in chronic hard-to-heal lower-extremity ulcers</strong></td>
<td>Case series</td>
<td>A prospective, single-center, non-blinded, non-comparative, post-market compassionate clinical evaluation of a novel acellular fish skin graft which contains omega 3 fatty acids, for the closure of hard to heal lower extremity chronic ulcers</td>
<td>A 40% decrease in wound surface area (P&lt;0.05) and a 48% decrease in wound depths (P&lt;0.05) were observed. Complete closure was seen in three of 18 patients by the end of the study phase</td>
<td>Fish skin is a promising wound-closing adjunctive ECM in this compassionate setting, where many other products may fail</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Testing of safety and efficacy of fish skin for wound care</strong></td>
<td>Parallel-group, double-blinded RCT</td>
<td>Fish skin vs porcine SIS ECM (Oasis): comparison of time to heal of whole-thickness biopsy wounds</td>
<td>Fish skin showed accelerated wound healing with significantly (P&lt;0.05) more wounds fully healed on days 14, 21, and 25 when compared to pig allograft</td>
<td>Healing time of wounds dressed with fish skin was noninferior to the healing time of wounds dressed with porcine SIS ECM</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Abbreviations:** AFS, acellular fish skin; dHACM, dehydrated human amnion/chorion membrane; ECM, extracellular matrix; NA, not available; NPWT, negative-pressure wound therapy; RCT, randomized controlled trial; SIS, small intestine submucosa.
We prospectively evaluated the percentage of wound closure area from baseline after five weekly AFS applications in 18 patients with at least one “hard-to-heal” criterion (area >10 cm² or duration of wound >52 weeks). The AFS was placed weekly for 5 weeks, followed by 3 weeks of standard care; secondary wound dressings were applied per the underlying etiology. Wound area reduction and signs of infection were noted and reviewed weekly. There was a 40% decrease in wound surface area with five weekly applications (P<0.05). In addition, the patients reported a notable reduction in pain and drainage. interestingly, the wound area reduction ceased when the product application was halted (Figure 6).11

Dr Chris Winters in the US used data from his practice and created a very interesting cost simulation model in which data from 27 diabetic wounds treated with AFS were inserted into the prognostic model, and compared with the actual outcome of the treatment. The prognostic model was built on Dr Margolis’ widely publicized work, in which a model was generated using data from 27,630 patients with diabetic foot ulcers. The model predicts the likelihood of a wound not healing after 20 weeks based on some of the following parameters being met: size >2 cm², previous duration of wound >2 months, and Wagner ulcer grade ≥3 (Table 2).16 The model was then used to check the predicted healing of the 27 patients vs the actual healing when these patients were treated with AFS. At 20 weeks, according to the predictive model, in 11 of the 27 patients, the wounds should have been closed with standard care, but, with AFS treatment, 24 patients had healed.17

Various case series and case reports have been published, including, but not limited to, those focusing on traumatic wounds, skin graft donor sites, and diabetic foot wounds with exposed bone or tendon (Table 1). At present, clearly more prospective evaluation of clinical series and controlled studies is necessary to support the application timing and to generate information on the expected outcomes. There needs to be a much more aggressive attempt by practitioners and the makers of the product to develop a registry of patients by which one can better understand the application schedule and expected outcomes of the product.

Based on the authors’ personal use of the product and the proctoring of other physicians, the application of this product requires a wound bed that can range from a surgically prepped one (application in the operating room [OR]) to the one that fundamentally needs a biologic dressing, such as a venous leg ulcer. It appears that this modulatory ECM needs to be reapplied at 4–7 days after its first application, and at present, weekly application is the standard. However, an application interval window of 2 weeks certainly seems to occur frequently in practice and in our experience. The ideal “dosing frequency” per wound type has yet to be defined. However, no other CTP has documented appropriate dosing guidelines to date (with the possible exception of the dermal (re)generative templates), most likely due to an eye toward sales, and not clinical efficacy (Figure 7).

### Translational research and the omega-3 fatty acids question

The primary question that exists around AFS is whether the intrinsic omega-3 fatty acids in the product allow for any special properties for this CTP. Answering the primary question requires answers to a couple of other questions: the first, does AFS contain relevant amounts of omega-3 fatty acids that matter in vitro, and the second, does this amount of omega-3 fatty acids make a difference in vivo. The AFS

![Figure 6 Mean wound surface area as noted during the 5-week AFS treatment and 3-week follow-up of venous leg ulcers.](Image)


**Abbreviation:** AFS, acellular fish skin.

![Figure 7](Image)

**Table 2 Likelihood of healing by prognostic factors**

<table>
<thead>
<tr>
<th>Size &gt;2 cm²</th>
<th>Ulcer grade ≥3</th>
<th>Duration &gt;2 months</th>
<th>Modeled expected likelihood of healing</th>
</tr>
</thead>
<tbody>
<tr>
<td>✗</td>
<td>✗</td>
<td>✗</td>
<td>0.66</td>
</tr>
<tr>
<td>✗</td>
<td>✗</td>
<td>✗</td>
<td>0.52</td>
</tr>
<tr>
<td>✗</td>
<td>✗</td>
<td>✗</td>
<td>0.53</td>
</tr>
<tr>
<td>✗</td>
<td>✗</td>
<td>✗</td>
<td>0.47</td>
</tr>
<tr>
<td>✗</td>
<td>✗</td>
<td>✗</td>
<td>0.33</td>
</tr>
<tr>
<td>✗</td>
<td>✗</td>
<td>✗</td>
<td>0.31</td>
</tr>
<tr>
<td>✗</td>
<td>✗</td>
<td>✗</td>
<td>0.28</td>
</tr>
<tr>
<td>✗</td>
<td>✗</td>
<td>✗</td>
<td>0.22</td>
</tr>
</tbody>
</table>

**Notes:** The table shows the likelihood that a wound with a given prognostic factor or combination of prognostic factors will heal after 20 weeks. ✗ denotes the presence of one of the prognostic factors.
less inflammation, the same downstream effects observed with NSAIDs.\textsuperscript{18}

Interestingly, this NSAID effect has been seen by our group clinically with significant reduction in pain in patients with venous leg ulcer (n=18), as well as in 23 patients with vascular disease and diabetes mellitus who had foot wounds. The clinical significance has been great enough to support an application to the US Department of Defense (DOD) for the use of AFS to help reduce narcotic dependence in patients with severe wound pain. AFS may also have a role in atypical wounds such as vasculitis. To date, there is really only one CTP, a dermal regenerative template, with a documented role in atypical wounds.\textsuperscript{19}

In an attempt to further assay the effect of omega-3 fatty acids on wound closure, an in vitro keratinocyte wound model was created in a study. The constituent fatty acids from the AFS were delivered to the keratinocyte injury model, while the control received no fatty acids. The group treated with the lipid extract closed 20\% faster with the lipid extract increasing the migration of cells to the wound bed. The mechanism of action by which this was promoted may include omega-3 fatty acids-induced activation of specialized pro-resolvin pathways, which lead to reduction of prostaglandins and

Figure 8 Levels of omega-3 and omega-6 fatty acids in various products with y-axis being the percent of fatty acid and x-axis being the type of fatty acid.

Note: Reproduced from Lantis J, Magnusson S, Baldursson B, Kjartansson H, Rolfsson O, Sigurjonsson G. Fish skin omega-3 PUFAs induce cell migration and transcription of ALOX15, a specialized pro-resolving mediator forming lipoxygenase. Paper presented at: SAWC Fall; October 20–22, 2017; Las Vegas, NV.\textsuperscript{18}

Figure 7 (A) Post-debrided foot with exposed bone. (B) Completion of transmetatarsal amputation. (C) Placement of AFS. (D) Post-op day 4 with no negative-pressure wound therapy. (E) Three weeks after grafting.

Abbreviation: AFS, acellular fish skin.
includes patients with vasculitis, which if enrolled will be the first CTP trial to actively recruit vasculitic patients prospectively.

**Using fish skin in burns**

The burn market remains one of the targets for skin substitutes, despite the fact that the number of severely burned individuals in most of the world is decreasing due to public health initiatives. In addition to this, an immediate post-trauma/modern battlefield dressing is attractive if it has intrinsic hemostatic and antimicrobial properties. Also, an agent that can enhance the speed of closure gains attention, particularly a dressing that can enhance the quality of wound bed and reduce the time to closure.

Recently, the US Army Institute of Surgical Research Combat Trauma and Burn Injuries Research Group, under the direction of Randolph Stone II, PhD, have reported on their assessment of AFS in an established porcine burn model. The objective of their project was to evaluate decellularized fish skin graft on deep partial-thickness (DPT) and full-thickness (FT) burn wounds in a preclinical porcine model. More specifically, the project aimed to evaluate fish skin graft as an acellular dermal matrix for its ability to integrate and improve healing of DPT burn wounds, to assess the use of fish skin graft as a temporary dressing prior to grafting, and to look at its ability to improve healing of highly meshed autografts on FT burn wounds. The animals were wounded and treated as shown in Table 3. There were six animals in each group, and 12 wounds were created per animal. Wound closure, wound perfusion, hydration, and contracture were assayed.

### Table 3 Porcine burn model treatment groups

<table>
<thead>
<tr>
<th>Group number</th>
<th>Burn depth (day 1)</th>
<th>Primary treatment after excision of dead tissue (day 0)</th>
<th>mSTSG application (day 7) (meshing ratio)</th>
<th>Second application of Kerecis (day 7)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Full thickness</td>
<td>Kerecis</td>
<td>Yes (1:5:1)</td>
<td>No</td>
</tr>
<tr>
<td>2</td>
<td>Full thickness</td>
<td>Kerecis</td>
<td>Yes (3:1)</td>
<td>Yes</td>
</tr>
<tr>
<td>3</td>
<td>Full thickness</td>
<td>Cadaver skin</td>
<td>Yes (1:5:1)</td>
<td>No</td>
</tr>
<tr>
<td>4</td>
<td>Deep partial thickness</td>
<td>PriMatrix</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td>5</td>
<td>Deep partial thickness</td>
<td>Kerecis</td>
<td>No</td>
<td>Yes</td>
</tr>
</tbody>
</table>

**Conclusion**

We are just starting to understand AFS, an ECM which is new to the market. The unique omega-3 fatty acid content in addition to the relatively biologically ideal three-dimensional ECM structure makes it an intriguing solution to the clinical needs of the chronic wound-treating physicians. So far clinically, it has appeared to act as a very efficient modulatory agent that can enhance the speed of closure gains attention, particularly a dressing that can enhance the quality of wound bed and reduce the time to closure.
ECM that is resistant to bacteria, and the anti-inflammatory omega-3 fatty acids content makes it very attractive for their potential added pain reduction. The fact that in some cases it can be skin grafted early at 7 days is also a benefit. At present, it can be used immediately after burn excision or wide soft tissue resection in the OR, in negative-pressure wound therapy, and also in the clinic after wound debridement with appropriate supportive dressing.

Figure 9 Progression of porcine wound closure when treated with AFS with and without skin grafting.

Abbreviations: AFS, acellular fish skin; DPT, deep partial thickness; FBC, fetal bovine collagen.
Disclosure
John C Lantis II is a paid consultant for Kerecis. Munir Patel reports no conflicts of interest in this work.

References
13. Lantis JI, Petursdottir K, Baldursson B, Kirsner RS, Double-Bline WD. Prospective, randomized clinical trial on 170 acute wounds shows significantly faster healing rate with intact fish skin compared to human amniotic membrane. Presentation at a conference. American Professional Wound Care Association (APWCA) sept 6–8, 2018 in Baltimore, MD, USA.
18. Lantis J, Magnusson S, Baldursson B, Kjartansson H, Rolffso O, Sigurjonsso G. Fish skin omega-3 PUFA induces cell migration and transcription of ALOX15, a specialized pro-resolving mediator forming lipoxygenase. Paper presented at: SAWC Fall; October 20–22, 2017; Las Vegas, NV.