Androgen upregulates the palmitoylation of eIF3L in human prostate LNCaP cells

Luwei Cui1,2
Ming Liu2
Shicong Lai2,3
Huimin Hou2,3
Tongxiang Diao1,2
Dalei Zhang2
Miao Wang2,3
Yaoguang Zhang1,2
Jianye Wang1,2

1Peking University Fifth School of Clinical Medicine, Beijing, People’s Republic of China; 2Department of Urology, Beijing Hospital, National Center of Gerontology, Beijing, People’s Republic of China; 3Graduate School of Peking Union Medical College and Chinese Academy of Medical Sciences, Beijing, People’s Republic of China

Introduction

Updated statistics in 2018 indicate that prostate cancer remains the most commonly diagnosed malignant tumors in men. A total of 164,690 new cases of prostate cancer have been diagnosed and 29,430 related-deaths have been reported in the United States.1 An estimated 450,000 males have been diagnosed with prostate cancer in Europe, comprising a large proportion of the overall burden of cancers worldwide.2 Prostate cancer is the most common noncutaneous malignancy diagnosed in American
men and androgen-dependent malignant tumor among males.\textsuperscript{3,4} The interaction between androgen and androgen receptor (AR) is the cornerstone that leads to progression, thus drugs targeting the AR signaling axis remain the principal therapy for advanced prostate cancer.\textsuperscript{5} Androgen deprivation therapy (ADT) is initially effective, leading to a marked decrease in PSA. However, the majority of the patients with advanced prostate cancer can progress and become refractory to ADT owing to aberrant AR activation, which is castration-resistant prostate cancer. The median survival at this stage is historically less than 2 years.\textsuperscript{6,7} The associated mechanisms include the occurrence of AR splice variants,\textsuperscript{8} inducing 20–40\% of the patients to respond poorly to enzalutamide or abiraterone, both of which are large breakthroughs in the treatment of metastatic castration-resistant prostate cancer (CRPC). Secondary resistance to these drugs is regarded as an adaptive change in prostate cancer cells. Thus, therapeutic methods targeting the novel mechanism need to be developed to meet the clinical need of patients with metastatic CRPC.

Posttranslational modification helps provide protein with additional functions, among which protein fatty acylation is rather common.\textsuperscript{9} Protein fatty acylation is the covalent attachment of lipids onto proteins, controlling protein–protein and protein–membrane interactions, which play a pivotal role in modulating the biological functions of proteins and mediating their targeting for activation.\textsuperscript{10,11} Thus, this modification is essential for cells to maintain homeostasis and to respond instantly to extracellular signals! Several forms of protein fatty acylation exist in eukaryocytes, which primarily include N-myristoylation and S-palmitoylation. S-palmitoylation is the addition of a 16-carbon saturated fatty acid, the so-called palmitate, to cysteine residues via a labile thioester linkage\textsuperscript{12} (Figure 1A). Among these modifications, palmitoylation is a common form of protein fatty acylation, by which protein hydrophobicity is efficiently increased to facilitate membrane association. In addition, palmitoylation is closely associated with the regulation of protein trafficking, stability and activity.\textsuperscript{13,14} Several studies have demonstrated that protein palmitoylation is associated with the development of disease, tumorigenesis and prognosis. Studies have proved the presence of aberrant palmitoylation in Huntington disease, caused by a CAG expansion in the huntingtin (HTT) gene. The palmitoyl acyltransferases for HTT are disturbed, resulting in reduced palmitoylation of HTT and exhibiting itself with motor, cognitive and psychiatric deficits.\textsuperscript{15} Moreover, Ras tumorigenesis is also regulated by palmitoylation. The Ras family of proteins widely consists of cancer drivers and plays

![Figure 1](https://www.dovepress.com/)

**Figure 1** (A) Structure of common protein fatty acylation. S-palmitoylation is the addition of the 16-carbon palmitate to a cysteine residue via a reversible thioester linkage. (B) Reversible S-palmitoylation. Palmitate is transferred from palmitoyl-CoA, which is produced by acyl-CoA synthetase, to a protein by protein acyl transferase. By contrast, palmitate on proteins is cleaved by palmitoyl-protein thioesterase.
a role in signaling, cell proliferation, differentiation and survival. Two reports in 2005 indicated that the presence of Ras was controlled by a transport cycle via palmitoylation and depalmitoylation thus determining its location to membrane and subsequent oncogenic signaling (Figure 1B). Therefore, targeting and disrupting membrane interactions of specific Ras isoforms represent a significant therapeutic strategy for regulating Ras-driven tumorigenesis. Nature in 2017 indicated that palmitoylation-mediated activation of MC1R could prevent melanogenesis. From the aforementioned discussion, palmitoylation plays a key role in membrane location, subsequent signaling pathway and tumorigenesis. Considering the limitations of the current drug therapy for CRPC, we explore palmitoylated proteins in androgen-treated LNCaP cells and non-treated LNCaP cells, to reveal the mechanism of prostate cancer progression by protein palmitoylation and identify a new drug target!

Click-chemistry-based chemical probes for the detection of protein palmitoylation hasten the discovery of novel palmitoylated proteins and elucidate their biological functions. The combination of mass-spectrometry-based quantitative proteomics with click chemistry probes can be used to explore the dynamics of protein palmitoylation under different physiological or pathological conditions. In the present study, we used the chemical tool and compared the palmitoylated proteins between androgen-treated LNCaP cells and non-treated LNCaP cells, to reveal the mechanism of prostate cancer progression by protein palmitoylation and identify a new drug target.

Materials and methods

S-palmitoylation assay
Prior to the experiments, LNCaP cells were seeded with complete media onto 6-cm dishes (5×10⁵ cells/dish) and incubated for 48 hrs. We purchased LNCaP cells from American Type Culture Collection, which were grown in RPMI-1640 media supplemented with 10% FBS and incubated in a 5% CO2 humidified chamber at 37°C for 48 hrs before any experiment. The cells were then treated with R1881 (10 nM) or DMSO in RPMI-1640 media supplement with 0.5% FBS. After 24 hrs, the cells were harvested for S-palmitoylation assay.

The S-palmitoylation assay was performed in accordance with the protocols described previously and slightly modified. The following are the inclusive reagents needed in this experiment: Nethylemaleimide (NEM, Sigma), Protein (A/G) UltraLink Resin (Thermo), Rabbit anti-eIF3L (1:2000, Protein tech), hydroxylamine (Sigma), tris (2-carboxyethyl) phosphate hydrochloride (TEP, Sigma), tris ([1-benzyl-1H-1, 2, 3-triazol-4-yl] methyl) amine (TBTA, Sigma), Thiopropyl Sepharose 6B (Sigma) and azide-agarose beads (Nanocs). LNCaP cells were homogenized in lysis buffer (10 mM sodium phosphate, 2 mM Na2EDTA, 0.32 M sucrose, 1% Triton X-100, 50 mM NEM and Pierce protease and phosphatase inhibitor cocktail, pH 7.4) for 30 mins. The lysates were then immunoprecipitated using protein A/G resin preloaded with eIF3L antibody overnight. The following day, the protein A/G resin was washed three times and incubated with elution buffer (1% SDS, 10 mM sodium phosphate, 2 mM Na2EDTA, 0.32 M sucrose) at 50°C for 5 mins to release eIF3L proteins. Eluted samples were divided into two equal portions—one treated with 1 M hydroxylamine and the other with 1 M Tris-HCl (pH 7.4) (as a control) with the presence of activated thiol-Sepharose 4B (Sigma). After incubation for 2 hrs at room temperature, the protein A/G resin was washed with a wash buffer (10 mM sodium phosphate, 2 mM Na2EDTA, 0.32 M sucrose, 1% Triton X-100, 500 mM NaCl and 0.2% SDS) three times. Western blot analyses were performed to determine the presence of eIF3L protein.

Metabolic labeling and click chemistry accumulation
Before the experiments, the LNCaP cells were seeded with complete media onto 6-cm dishes (5×10⁵ cells/dish) and then incubated for 48 hrs. Metabolic labeling and click chemistry were performed in accordance with the protocols previously described and then modified slightly. The ω-alkynyl fatty acid analog, Alk-C16, was dissolved in DMSO to generate 50 mM stock solutions and stored at −80°C. Prior to cell treatment, Alk-C16 was dissolved in RPMI-1640 serum-free media supplement with 5% BSA (fatty acid free) at a final concentration of 100 μM. The
fatty acid media solutions were sonicated for 15 mins at room temperature. The fatty acid media were then divided into two equal portions—one added with R1881 (10 nM) and the other added with DMSO as a control. The seeded LNCaP cells were washed with PBS once and added the two fatty acid media, respectively, for 24 hrs at 37°C/5% CO₂. After incubation for 24 hrs, the cells were washed three times and homogenized in 500 μL lysis buffer (1% Nonidet P-40/150 mM NaCl/Pierce protease and phosphatase inhibitor cocktail/100 mM sodium phosphate, pH 7.5) for 30 mins at 4°C. The protein extracts were then subjected to the probe labeling reaction for 1 hr at room temperature, at final concentrations of the following reagents: 1 mM azide-agarose beads, 1 mM TCEP dissolved in water, 0.2 mM TBTDA dissolved in DMSO/tert-butanol (20%/80%) and 1 mM CuSO₄ in PBS. The order of addition of the reagents to the protein extracts is important for the reaction. With click chemistry, the Alk-C16-conjugated proteins were accumulated in the azide-agarose beads. The agarose beads were washed three times with the lysis buffer at room temperature. The proteins accumulated in the beads were subsequently digested for mass spectrometry.

**nLC-MS/MS analysis**
Proteomic profiling was performed on an EASY nLC1000 (Thermo Fisher) system with an LTQ-Orbitrap-Elite (Thermo Fisher) mass spectrometer. Tryptic peptides were desalted using 0.1% fluoroacetic acid and loaded on a trap column (PepMap C18, 300 μm×5 mm). The peptides were then eluted to a 3 μm fused silica capillary column (75 μm×500 mm). Finally, the peptides were separated at two gradients: the gradient from 100% mobile phase A (97.9% H₂O, 2% acetonitrile, 0.1% formic acid) to 30% mobile phase B (80% acetonitrile, 19.02% H₂O, 0.08% formic acid) in the first 80 mins and the mobile phase B from 30% to 100% in the next 10 mins. The eluted peptides were directly sprayed with a voltage of 1.6 kV into the on-line coupled LTQ-Orbitrap-Elite MS using a nano electro-spray ionization source equipped with a metal-coated nano-scale emitter. Mass spectra were obtained over a mass-to-charge ratio (m/z) range of 300–1800 Th at a resolving power of 30,000 at 400 m/z.

**Database search**
The recorded MS spectra were analyzed using MaxQuant Software (version 1.5.5.1). The MS/MS peak list file was performed by searching against a forward and reverse version of the UniProtKB/Swiss-Prot human database (generated from version 2017_05, human taxonomy, 20,316 entries). The cutoff of the false discovery rate for peptide and protein identification was set to 0.01, and only peptides with ≥7 amino acid residues were allowed for identification. Label-free quantitation (LFQ) was conducted using MaxQuant on identified razor and unique peptides to properly quantify identified proteins. Gene ontology (GO) analysis was performed using DAVID (NIH) configured to assign the identified proteins with cellular function GO terms.

**Data processing and statistical analysis**
Protein abundances normalized using the LFQ algorithm integrated in MaxQuant were Log2 transformed for further analyses. Filtering was conducted in Microsoft Excel 2010. DanteR (version 1.0.1.1) and Perseus (version 1.5.5.3) were used to perform different types of statistical analysis including Log2 transformation, correlation plot, statistical tests, imputation, P-value adjustment and volcano plot.

**Results**

**Design and synthesis of clickable palmitate probe (Alk-C16)**
Both alkyne and azide analogs of palmitic acid were developed for click chemistry applications. In the current study, we chose the alkyne analog because it exhibited physicochemical properties similar to those of the wild type fatty acid carbon chain and maintained the hydrophobicity for membrane affinity. (Figure 2) We added the alkyne group at the ω-position of a fatty acid. The position is the terminal end farthest from the carboxyl functionality. Our ω-Alkynyl palmitate analog (Alk-C16) was synthesized from alcohols containing internal alkyynes via a zipper reaction. In the reaction, an internal alkyne was isomerized to a terminal alkyne followed by Jones oxidation to yield the ω-alkynyl palmitate analog with high yield and purity.

**Proteomic experiments using clickable palmitate probe (Alk-C16)**
To compare the palmitoylated proteins between androgen-treated LNCaP cells and non-treated LNCaP cells in the palmitoylome profiles, Alk-C16, a palmitate fatty acid analog, was metabolically incorporated onto cellular proteins and the proteins modified with Alk-C16 were analyzed using MS-based proteomics and an informatics-assisted
label-free strategy. First, the LNCaP cells were added with Alk-C16 and then incubated for 24 hrs. When the cells were metabolically incorporated with Alk-C16, the cells were divided into two parts to check the androgen-induced palmitoylated proteins: one treated with androgen (R1881) and the other with DMSO as the control. After 24 hrs, the cells were harvested and lysated. Alk-C16 incorporated onto acylated proteins was chemoselectively ligated to azide agarose beads by Cu1-catalyzed alkyne-azide [3+2] cycloaddition reaction (click chemistry). The conjugated proteins, which were theoretically palmitoylated proteins, were accumulated in azide agarose beads by ligation. The conjugated proteins were then digested on the agarose beads for MS and label-free quantitation.

Androgen promotion of eIF3L palmitoylation

We supplied three individual pairs of androgen-treated and non-treated LNCaP cells to screen the androgen-induced palmitoylated proteins. As a result, we identified 4351 unique peptides corresponding to 835 proteins (mascot score>2, P<0.05). Meanwhile, we filtered the outlier proteins and contaminant proteins. The identified proteins were then analyzed in LFQ by MaxQuant version. A number of these identified proteins were palmitoylated proteins, particularly the eIF3 initiation factors, including its subunits eIF3j, eIF3g, eIF3c, eIF3b, eIF3a and eIF3L. We have also compared the amino acid sequences of eIF3L of Homo sapiens (human) with the mass spectrum results of eIF3L, and they are consistent (Figures 3 and 4). The protein with the relatively higher LFQ activity was eIF3L, one subunit of eIF3, both in androgen-treated and in non-treated LNCaP cells. Among these 6 subunits of eIF3, the androgen-induced candidates were identified as their LFQ activities were significantly upregulated (fold changes>1.5, P<0.05) in triplicate samples of androgen-treated vs non-treated LNCaP cells. Notably, both eIF3a and eIF3L were identified as the candidates (Figure 5).

To strengthen the evidence, thiopropyl captivation of S-palmitoylated protein assay was further performed to explore whether androgen promotes the eIF3a and eIF3L palmitoylation levels in LNCaP cells, when the cells were treated with R1881 (androgen). The eIF3L palmitoylation level was significantly upregulated. This result was consistent with that of MS. We concluded from the aforementioned results that androgen upregulates the palmitoylation of eIF3L in human prostate LNCaP cells (Figure 6).

To further elucidate the regulatory effect of androgen on eIF3L palmitoylation, androgen (R1881) and abiraterone were added in the cultured LNCaP cells at the same time, but the concentration of abiraterone is varied from 0.1 µmol/L to 0.5 µmol/L; as the result shown in Figure 7, abiraterone could downregulate the eIF3L palmitoylation level by inhibiting androgen (R1881). From the above results, it can be seen that eIF3L palmitoylation is clearly regulated by androgen.

Discussion

Protein fatty acylation is critical in maintaining cellular homeostasis and regulating signaling pathway. We have shown that the clickable palmitate probe (Alk-C16) can be used for biochemical detection and cellular imaging of lipid-modified proteins. It facilitates our study of the dynamic behavior of lipid-modified proteins in various diseases and the determination of the pathological mechanism. It also has several advantages in cell culture for the detection of proteins that are palmitoylated under metabolic conditions such as drug treatment.22,23 Moreover, combined with quantitative proteomics, the clickable palmitate probe can quantify the palmitoylated proteins as well.

Protein synthesis is increasingly recognized as an important component in tumorigenesis and progression. Enhanced translation rates are closely associated with
Genome-wide analyses of mRNA translation by ribosome profiling indicates that translational control contributes significantly to overall gene expression. The initiation phase of protein synthesis is rate-limiting for most mRNAs. The translational components, particularly some initiation factors, play a pivotal role in the translational regulation. elf3 is the largest of the initiation factors, comprising 13 non-identical protein subunits (elf3a to m). It binds stably to the 40S ribosomal subunit and promotes the binding of methionyl-tRNAi and mRNA to form the 40S initiation complex. The individual over-expression of 6 different elf3 subunits is observed in various tumors, by enhancing the translation of mRNAs that encode growth-promoting proteins.

Figure 3. The mass spectrometry results of elf3 palmitoylation.
junction of the 60S subunit. The 80S initiation complex formed can then enter the elongation phase of protein synthesis. The ribosomal complex, often called the 43S preinitiation complex or (43S PIC), comprises the 40S ribosome, eIF1, eIF1A, eIF3, and the ternary complex, eIF2-GTP-Met-tRNAi.

In our recent study, we detected the androgen-induced palmitoylated proteins by comparing the palmitoylome profile of androgen-treated LNCaP cells with that of non-treated cells. The results demonstrated that androgen significantly increased the level of palmitoylated eIF3L in androgen-treated LNCaP cells. This rapid response to

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**Figure 4** The amino acid sequences of eIF3L of *Homo sapiens* (human). The partial highlighted red sequences of eIF3L are consistent with the mass spectrum results by comparing Figure 3 with Figure 4.

**Figure 5** Regulation of androgen on the palmitoylation level of eIF3 subunits in LNCaP cells. The LNCaP cells labeled by Alk-C16 were treated with R1881 or DMSO as a control. The palmitoylated proteins were accumulated by click chemistry. They were identified by MS and quantified by label-free strategy. Among the six eIF3 subunits, the androgen-induced candidates were identified as LFQ activities were significantly up-regulated (fold changes >1.5, \( P < 0.05 \)) in triplicate samples of androgen-treated vs non-treated LNCaP cells. The \( P \)-values were analyzed using two-tailed Student’s \( t \)-distribution function. \( * P < 0.05 \).
extracellular signal: the androgen is closely associated with the dynamic palmitoylated level of eIF3L. Prostate cancer is an androgen-driven solid tumor; thus, many therapeutic strategies target the AR signaling axis. However, when patients progress to the CRPC stage, the administration of drug therapy remains a challenge for urologists.

This preliminary study identified that androgen can induce the palmitoylation of eIF3L, which serves as the initiation factor and regulates protein expression and cell growth. Over-expression of eIF3 subunits is observed in many tumors by enhancing the translation of mRNAs. Further studies should be performed to confirm the correlation of palmitoylated eIF3L and cell proliferation. In addition, the correlation between the palmitoylated eIF3L and the prognosis of PCs patients was also worthy of further study. It is hoped that all the questions will be resolved with our proposed methods.

### Conclusion

The results in this study indicate that other mechanisms may exist in the development and progression of prostate cancer. Androgen-induced protein palmitoylation may provide a new direction for the treatment of prostate cancer and the level of palmitoylated eIF3L may be a biomarker to detect early prostate cancer.

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### Disclosure

The authors report no conflicts of interest in this work.

### References


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### Table

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**Figure 6** Androgen promotion of eIF3L palmitoylation by S-palmitoylation assay. The LNCaP cells were treated with R1881 or DMSO for 24 hrs and then the cells were harvested and performed by S-palmitoylated protein assay to test the palmitoylation level of eIF3L.

**Abbreviations:** HAM, hydroxylamine; Palm, palmitoylation; IB, immunoblot; IP, immunoprecipitation.

**Figure 7** The palmitoylation level of eIF3L was downregulated with the increase of the concentration of abiraterone. The cultured LNCaP cells were treated with androgen R1881 and different concentrations of abiraterone (0.1 µmol/L or 0.5 µmol/L), then the S-palmitoylated protein assay revealed that abiraterone could downregulate eIF3L palmitoylation level by inhibiting androgen.


