Anti-nociceptive and anti-inflammatory activities of crude root extract and solvent fractions of *Cucumis ficifolius* in mice model

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**Background:** Societies in developing countries use traditional medicine as alternatives for management of pain and inflammation. The plant *Cucumis ficifolius* has been used in Ethiopia to treat many ailments including inflammation and pain. The objective of this study was to evaluate the antinociceptive and anti-inflammatory activities of the crude root extract and solvent fractions of *C. ficifolius*.

**Methods:** The analgesic activity of crude extract and solvent fractions of *C. ficifolius* was evaluated with acetic acid-induced writhing, hot plate, and formalin-induced paw licking tests. The anti-inflammatory effect of crude methanolic root extract and solvent fractions of *C. ficifolius* was evaluated using carrageenan-induced paw edema. The crude extract was given at 200, 400 and 800 mg/kg. Butanol and aqueous fractions were given at 100 and 200 mg/kg doses. The negative control groups were treated with distilled water (10 mL/kg). Standard drugs used were acetylsalicylic acid (ASA) in acetic acid, formalin tests and carrageenan-induced paw edema and morphine (20 mg/kg) in hot plate test.

**Results:** The crude extract, at its maximum dose, produced comparable analgesic activity (72.5%) to ASA in acetic acid writhing test. In the hot plate test, both the crude extract and solvent fractions exhibited a significant prolongation of nociception reaction time. Formalin test result indicated a significant reduction of mean lick time with maximal protection of 64% (early phase) and 83% (late phase). Aqueous and butanol fractions showed good analgesic activity in the three models. Inflammation was decreased by 69% with butanol (200 mg/kg); 71% (800 mg/kg) of crude extract and by 41% and 56% with the use of aqueous fraction at 100 and 200 mg/kg, respectively ($p<0.001$).

**Conclusion:** The present study indicates that the crude methanolic root extract, as well as butanol and aqueous solvent fractions, showed anti-nociceptive and anti-inflammatory activities.

**Keywords:** hot plate test, writhing test, paw edema, formalin test, carrageenan, 80% methanol

**Introduction**

Pain and inflammation impose an enormous economic and health problem globally. They are associated with a number of adverse health consequences including the inability to carry out daily activities, impaired patients’ quality of life and risk of all-cause mortality.¹

Currently, a number of drug classes are available to manage inflammation and pain. However, the clinical uses of anti-inflammatory and analgesic agents are...
limited by their affordability, accessibility, and adverse drug reactions, and many medicines are not effective as expected in all patients. Tolerance and dependence is another challenge, particularly with the chronic use of opioids.

Pharmaceutical development has led to a great number of medicines, however many of them have limited efficacy; patients can encounter escalating health-care costs; and adverse drug reactions are reported in some patients.

As a result, there is an increased tendency to use traditional medicine, with an extent not less than 80% worldwide.

Plants have been used since ancient times to treat diseases and infections. Medicinal plants are cheap, easily available and affordable. Besides traditional use, scientific study of medicinal plants has found herbal medicines to be a medicinal resource for drug discovery.

A variety of medicinal plants have been used to treat pain and inflammation. These plants include Malva verticillata, Otostea integrifolia, croton macrostachyus, Ocimum suave, Cucumis ficifolius, Arisaema schimperianum, Euclera racemosa, Malva verticillata, Impatiens tinctoria, Balanites, Ehretia cymosa, and a host of others.

Cucurbit plants (cucurbitaceae) are cultivated widely in the subtropical and tropical countries. They have demonstrated anti-inflammatory, antitumor, hepato-protective, cardiovascular, immunoregulatory, anti-fungal, antibacterial, anti-viral, anti-diabetic, anti-tumor and anti-HIV/AIDS activities. The diverse biological actions of the Cucurbitaceae family is believed to be due to the presence of different bioactive constituents such as cucurbitacins, triterpenes, sterols, alkaloid, saponins, tannins, flavonoids, and phenolic compounds. Cucumis melo extract demonstrated antioxidant and anti-inflammatory properties. Methanolic extract of Cucumis colosso exhibited significant analgesic anti-inflammatory activity.

In this study, the roots of Cucumis ficifolius (Cucurbitaceae) were collected from Kilte Awlaelo district in Ethiopia in 2017. This pre-clinical study was performed 1) to predict the safety and efficacy data from animal models to support further research in human beings; and 2) to check the relationship between traditionally claimed use and scientific research. The plant is known locally in Amharic as “yemdir enbuay”; and in Tigriigna as “Rambo-ambo”. It is found widely distributed in East Africa, especially in Ethiopia and Kenya between altitudes of 1,000 and 2,700 m. It is perennial, usually prostrate herb that stems up to 1 m long, deeply lobed leaves and small, yellow flowers followed by oblong, dark yellow fruits to about 5 cm long that are covered in bristly pustules.

The root of C. ficifolius is chewed with the diseased teeth to treat toothache; crushed, macerated with water, filtered and the fluid is drunk to treat joint pain; mixed with bark of Croton macrostachyus, the dried paste is mixed with butter and drunk or the product is chewed and then the fluid is drunk to treat stomach ache. The plant is also used to treat bloody diarrhea, evil eye, liffe (wound) and expel ear-mites. An 80% methanolic extract of C. ficifolius showed significant anti-oxidant and hepatoprotective activities. Phytochemical screening demonstrated that C. ficifolius constituted secondary metabolites such as, phenols, flavonoids, terpenoids, steroids and saponins, which might confer anti-oxidant and hepatoprotective effects.

Thus, the aim of this study is to investigate the anti-inflammatory and anti-nociceptive activities of C. ficifolius.

Materials and methods

Chemicals and drugs

Chemicals and solvents of laboratory and analytical grade were used throughout this project: carrageenan (Tokyo Chemical Industries, Tokyo, Japan), methanol (Alpha Chemika, Mumbai, India), n-butanol (Carlo Erba Reagents SAS, Val de Reuil, France), chloroform (Labort Fine Chem Pvt Ltd, Surat, India), acetylsalicylic acid (ASA) (Addis Pharmaceutical Factory, Adigrat, Ethiopia/APF; Batch no., CA1704016), acetic acid (Thermo Fisher Scientific, Waltham, MA, USA), morphine (Amino Ltd, Neuenhof, Switzerland; Batch no., 6910/10) and formalin (Albert Rose Chemicals IP Ltd, Ahmedabad, India) were used.

Collection, identification, and preparation of plant materials

The roots of C. ficifolius were collected from Kilte Awlaelo district of Tigray Regional State, 47.7 km from Mekelle and 829 km from Addis Ababa. Identification and authentication of the plant were carried out by Dr Getnet Maresha, and sample specimen was deposited at Herbarium unit of Department of Biology, College of Computational and Natural Science, University of Gondar for future reference with voucher number of DG0024/210.

Preparation of extracts and fractions of plant material

The roots were cleaned of dust and debris and washed gently with water. The root was reduced to an appropriate size and air dried under shade for two weeks. The dried
root material was then pulverized with a grinder. A total amount of 1.335 kg dried root powder was macerated in methanol (80%) (1:6) with occasional shaking using orbital shaker at 120 rotations per minute for 72 h. The mixture was first filtered using muslin cloth followed by Whatman filter paper no.1. To maximize yield, remaceration was done for another 72 h twice. The filtrates were then evaporated with an oven dryer at 40°C until dried. The 80% methanolic crude extract of C. ficifolius was gummy and yellowish in color with 5.69% (w/w) yield. Among the fractions, the highest yield was found from the aqueous fraction. The dried powder of the methanolic crude extract was kept in a refrigerator at 4°C in airtight containers until use.

The liquid–liquid extraction was performed three times starting from chloroform then n-butanol. After collecting the chloroform and n-butanol fractions, the remaining residue was considered as an aqueous fraction. An oven dryer at 40°C was used to concentrate the fractions. The dried matter was stored in a refrigerator at 4°C until use.

**Experimental animals**

The experiments were performed on an equal number of male and female in-house bred Swiss albino mice age of 6–8 weeks and weight of 25–35 g. The animals were housed in polypropylene cage and maintained under standard animal housing condition (at ambient temperature, and with a 12 h light-dark cycle) and allowed free access to the standard pellet diet and water ad libitum. The animals were acclimatized to laboratory conditions for seven days prior to initiation of the experiment. All procedures have been undertaken as per the guidelines for the care and use of laboratory animals. The protocol was approved by the Health Research Ethics Review Committee HRERC, College of Health Sciences, Mekelle University with protocol number (ERC1206/2018). Animals were sacrificed under anesthesia after completion of the experiments.

**Acute oral toxicity test**

The acute oral toxicity test was performed on five female Swiss albino mice using the limit test recommendations of the Organization for Economic Co-operation and Development (OECD) 425 Guideline. For this test, we used healthy, nulliparous and non-pregnant female Swiss albino mice (age of 8–12 weeks) weighing 20–30 g. On day one, a four-hour fasted mouse was given 2,000 mg/kg (1 mL/100 g) of the extract orally in distilled water. The mouse was observed for 24 h for possible behavioral or physical changes. More emphasis was given to the first 4 h. After 24 h, four other mice were given the same dose and observed for 14 days for manifestations such as tremors, convulsions, salivation, diarrrhea, lethargy, sleep and coma.

**Anti-nociceptive activity**

**Acetic acid-induced writhing test**

As described by Birhane et al, the test was carried out to assess the peripheral analgesic action of C. ficifolius. Mice of either sex (26–35 g) were used. Animals fasted overnight (for 12 h) were randomly assigned into five groups for crude extract and six groups for solvent fractions. In each group we used six mice.

The first group of mice was given distilled water (DW) 10 mL/kg as a negative control. Groups II, III and IV were administered the crude methanolic root extract at 200, 400, and 800 mg/kg, respectively, and the fifth group received 150 mg/kg of ASA. In solvent fractions, six groups of animals were used: Group I received DW (10 mL/kg); Group II were challenged with 100 mg/kg and Group III with 200 mg/kg of aqueous fraction. Mice under group IV and V received 100 mg/kg, and 200 mg/kg of butanol fraction, respectively. The positive control group (Group VI) were challenged with ASA 150 mg/kg. Thirty minutes after administration of vehicle, standard drug and test substance; the animals were subjected to intraperitoneal (i.p.) injection of acetic acid solution (0.6%, 10 mL/kg).

**Nociception response was quantified after 5 min of latency for 20 min. The observation for the response includes stretching of the hind limbs, contraction of abdominal muscles and arching of the back. Percent protection was calculated by applying the formula:**

$$\text{% Analgesic activity} = \frac{\text{Mean no. writhes (control)} - \text{mean no. of writhes (treated)}}{\text{Mean no. of writhes (control)}} \times 100\%$$

**Hot plate test**

The hot plate test was used to evaluate the central anti-nociceptive action of the plant material. The hot plate apparatus was maintained at 55±0.1°C. Overnight fasted mice were randomly assigned into different groups. The positive control group received morphine 20 mg/kg orally. The animals were treated with the vehicle, standard and test substances as described in the acetic acid-induced writhing test.

The pre-drug nociception latencies were measured twice at 15 min intervals. The first reading was discarded.
to avoid the effect of stress on the nociception latency values, and the second reading was used as a pre-drug latency.28 Then post-drug latencies were quantified at 30, 60, 90 and 120 min after administering standard drug, test substance or vehicle.29 A cutoff time of 20 s was considered by taking three times the mean pre-drug latency to minimize tissue damage. The nociceptive latency in seconds was quantified by taking the time interval between the animal reaching the hot plate and licking its paw or jumping off the hot plate.

The post-drug latency: T1 was estimated according to the reaction time of each mouse at 30, 60, 90 and 120 min after treatment. T0 represented the mean pre-drug latencies. For each group, the percentage of protection against thermal stimulus was determined by using the formula:30

\[
\text{% Protection against thermal stimulus} = \frac{T0 - T1}{T0} \times 100
\]

**Formalin test**

As described by Hunskaar et al, a formalin-induced lick test was performed.31 Mice fasted overnight with the provision of water were used. Then the overnight fasted mice were randomly selected and assigned into groups of five, each group with six mice. Group I receiving DW (10 mL/kg) was assigned as the negative control while mice receiving ASA at a dose of 200 mg/kg served as positive control (group II). The remaining groups (III to V) were given the test extract at a dose of 200, 400 and 800 mg/kg, and butanol and aqueous fractions at a dose of 100 and 200 mg/kg. Just before induction of inflammation, the leg of each mouse was marked on the skin over the lateral maleous.36 The basal volume of the right hind paw of individual mice was measured with a digital plethysmometer (PLM 02). Then, a 0.05 mL of 1% carrageenan in normal saline was injected into the dorsal surface of the right hind paw. The volume of injected paw was measured at 1, 2, 3 and 4 h after carrageenan injection.

Paw diameter before carrageenan injection was compared with the same paw diameter after administration of carrageenan by calculating the percentage inhibition applying the following formula:5

\[
\text{% Inflammation inhibition} = \frac{(Vt - Vo) \text{ control} - (Vt - Vo) \text{ treated}}{(Vt - Vo) \text{ control}} \times 100
\]

Where: \( Vt = \text{post-carrageenan injection mean paw volume in treated and negative control groups at time t} \) and \( Vo = \text{pre-carrageenan injection mean paw volume in treated and negative control groups} \).

In addition, the increase in paw volume, ie, inflammation expressed in percentage was calculated according to the formula given by:5

\[
\text{% Inflammation (\%) = } \frac{Vf - Vi}{Vi} \times 100
\]

Where: \( Vi \) is the initial mean paw volume before carrageenan injection and \( Vf \) is the final volume (after carrageenan injection).

**Anti-inflammatory activity**

Swiss albino mice fasted overnight were randomly assigned into nine groups each with six mice. Thirty minutes before injection of carrageenan, DW (10 mL/kg), the test substances and ASA (200 mg/kg) were administered orally. The rest of the seven groups were treated with crude extract at doses of 200, 400 and 800 mg/kg, and butanol and aqueous fractions of carrageenan in normal saline was injected into the dorsal surface of the right hind paw. The volume of injected paw was measured 30 min after formalin injection.

The percentage inhibition of nociception for the two phases was calculated using the following formula:32

\[
\text{% Inhibition} = \frac{\text{Control mean} - \text{Test mean}}{\text{Control mean}} \times 100
\]

**Statistical analysis**

Data were entered and analyzed with the IBM statistical package for social sciences (SPSS) version 21 (IBM Corporation, Armonk, NY, USA). The data obtained in the study were tabulated and expressed as mean ± standard errors of the mean (SEM). The statistical analysis was carried out
using one-way analysis of variance (ANOVA) followed by Tukey post-hoc test to compare variations among groups. The result was considered significant when \( P<0.05 \).

**Results**

**Oral acute toxicity test**

Acute toxicity studies revealed that administration of the crude extract of *C. ficifolius* (at a dose of 2,000 mg/kg) did not cause manifestations such as drowsiness, salivation, tremor, restlessness, convulsion, piloerection, diarrhea, nor caused mortality in the first 24 h observation as well as during the two-week follow up period.

**Analgesic activity**

**Writhing test**

The reduction in the mean number of writhes produced by all dose levels of crude methanolic extract, aqueous and butanol fractions of *C. ficifolius* were significant compared to negative control. The percentage pain protection produced by the use of the crude at three doses (200, 400 and 800 mg/kg), and 200 mg/kg of ASA was 24.9% \( (P<0.05) \), 48.7% \( (P<0.001) \), 72.5% \( (P<0.001) \) and 80.4% \( (P<0.001) \), respectively, as compared to the vehicle received group. As illustrated in Figure 1, 800 mg dose of crude methanolic extract of *C. ficifolius* revealed a comparable reduction in the mean number of writhes to ASA.

The butanol fraction resulted in greater reduction of the frequency of writhing response with the protection of 35% \( (P<0.01) \) and 68% \( (P<0.001) \) in the lower and higher dose levels, respectively. And also, a significant reduction in the mean number of writhing was observed at the dose of 100 mg/kg (33%, \( P<0.01 \)) and 200 mg/kg (62%, \( P<0.001 \)) of aqueous fraction (Figure 2).

**Hot plate test**

The crude root extract and solvent fractions of *C. ficifolius* increased the latency of pain response at all dose levels. Treatment of mice with 200 and 400 mg/kg crude extract significantly increased the latency to the maximum record of 10.12 (46.62%) and 11.16 (52.88%) seconds at 120 min of observation compared to control, respectively. The highest dose level (800 mg/kg) showed greater protection at all times of observation in relation to the vehicle group with maximum pain protection (82%, \( P<0.001 \)) at 90 min (Table 1). The mean difference between the middle dose and lowest dose levels was not significant throughout the observation. However, 800 mg/kg of the crude root extract showed a significant mean difference compared to post-drug values of the lowest dose while a significant difference was obtained at 90 min \( (P<0.01) \) compared to the middle dose.

With regard to the aqueous and butanol fractions, the delay in nociception reaction was significant with the exception of 100 mg/kg of the aqueous fraction at 30 min, compared to the negative control. Nevertheless, effects attributed to the use were within a considered level of significance in time intervals beyond 30 min (Table 2). The remaining doses of the two fractions significantly prolong the nociception latency in a dose-dependent manner. The aqueous fraction, 100 mg/kg (60%, \( P<0.01 \)), 200 mg/kg (86.8%, \( P<0.001 \)) and butanol 100 mg/kg (76%, \( P<0.001 \)) resulted in greater

**Figure 1** Anti-nociceptive effect of the crude methanolic extract of *Cucumis ficifolius* on acetic acid-induced writhing test. The results are expressed as mean ± SEM, \( n=6 \). *Compared with vehicle group, DW10, \( \* \) compared with CM200, \( \^ \) compared with CM400, \( \# \) compared with CM800, and \( \$ \) compared with ASA150. \( P<0.05 \), \( \hat{P}<0.001 \), \( \hat{P}<0.001 \).

**Abbreviations:** ASA150, aspirin 150 mg/kg; CM200, *C. ficifolius* extract 200 mg/kg; CM400, *C. ficifolius* extract 400 mg/kg; CM800: *C. ficifolius* extract 800 mg/kg; DW10, distilled water 10 mL/kg; SEM, standard error of the mean.

**Figure 2** Anti-nociceptive effect of oral aqueous and butanol fractions of *Cucumis ficifolius* on acetic acid writhing test. The results are expressed as mean ± SEM, \( n=6 \). *Compared with vehicle group, DW10, \( \* \) compared with AQ100g, \( \^ \) compared with AQ200, \( \# \) compared with B100, \( \$ \) compared with B200, and \( \$ \) compared with ASA150. \( \hat{P}<0.05 \), \( \hat{P}<0.001 \), \( \hat{P}<0.001 \).

**Abbreviations:** AQ100, aqueous fraction 100 mg/kg; AQ200, aqueous fraction 200 mg/kg; ASA150, acetysalicylic acid 150 mg/kg; B100, butanol fraction 100 mg/kg; B200, butanol fraction 200 mg/kg; SEM, standard error of the mean.
The results are expressed as mean ± SEM, n=6.

Notes: *Compared with DW10, **Compared with CM200, †Compared with CM400, ‡Compared with CM800, and §Compared with MO20. *P < 0.05, †P < 0.01, **P < 0.001.

Abbreviations: CM200, C. ficifolius 200 mg/kg; CM400, C. ficifolius 400 mg/kg; CM800, C. ficifolius 800 mg/kg; DW10, distilled water 10 mL/kg; MO20, morphine 20 mg/kg; SEM, standard error of the mean.

Table 1 Anti-nociceptive effect of Cucumis ficifolius methanolic crude root extract in hot plate test

<table>
<thead>
<tr>
<th>Group</th>
<th>Dose (mg/kg)</th>
<th>Mean latency (s) ± SEM (% protection)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>30 min</td>
<td>60 min</td>
</tr>
<tr>
<td>CM400</td>
<td>7.12±0.31 (6.22)</td>
<td>9.95±0.49 (44.2)</td>
</tr>
<tr>
<td>CM800</td>
<td>10.86±0.60 (48.77)</td>
<td>10.92±0.34 (49.59)</td>
</tr>
<tr>
<td>MO20</td>
<td>14.12±0.76 (90.77)</td>
<td>14.35±0.58 (93.92)</td>
</tr>
<tr>
<td></td>
<td>90 min</td>
<td></td>
</tr>
<tr>
<td>CM400</td>
<td>6.80±0.27 (1.49)</td>
<td>9.31±0.51 (36.47)</td>
</tr>
<tr>
<td>CM800</td>
<td>10.04±0.54 (37.53)</td>
<td>10.86±0.60 (48.77)</td>
</tr>
<tr>
<td>MO20</td>
<td>13.17±0.56 (77.93)</td>
<td>14.12±0.76 (90.77)</td>
</tr>
<tr>
<td></td>
<td>120 min</td>
<td></td>
</tr>
<tr>
<td>CM400</td>
<td>7.12±0.31 (6.22)</td>
<td>9.95±0.49 (44.2)</td>
</tr>
<tr>
<td>CM800</td>
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</tr>
</tbody>
</table>

Notes: The results are expressed as mean ± SEM, n=6. *Compared with vehicle group, †Compared with AQ100, ‡Compared with AQ200, †Compared with B100, §Compared with DW10, and ¶Compared with MO20. *P < 0.05, †P < 0.01, ‡P < 0.001.

Abbreviations: AQ100, aqueous fraction 100 mg/kg; AQ200, aqueous fraction 200 mg/kg; B100, butanol fraction 100 mg/kg; B200, butanol fraction 200 mg/kg; DW10 mL/kg, distilled water; MO20, morphine 20 mg/kg; SEM, standard error of the mean.

Table 2 Anti-nociceptive effect of aqueous and butanol fractions of Cucumis ficifolius on hot plate test

Protection at 90 min, whereas 200 mg/kg butanol fraction provided greater protection at 60 min (89.8%, p < 0.001).

Formalin test

The crude methanolic root extract of C. ficifolius significantly diminished the mean time of animals spent licking the injected paw in the two phases. In the early phase, the 200, 400 and 800 mg/kg doses of the test extract shortened the time of licking by 26%, 53%, and 64%, respectively. In the late phase, percentage protection of 46%, 67%, and 83% were recorded from the lower to the higher dose levels. The 400 (53%) and 800 (64%) mg/kg crude extract exhibited better activity in the early phase of the test as compared to the standard drug, ASA (31%). Although statistically not significant, 800 mg/kg methanolic extract of C. ficifolius exhibited greater pain protection (83%) compared to ASA (72%) in the late phase (Table 3). The analgesic gap between ASA and different doses of the test extract was wider, especially in the initial phase.

The aqueous fraction at 100 mg/kg and 200 mg/kg decreased the time of licking by 28% and 56%, respectively, in the early phase while butanol fraction showed higher protection action as compared to the aqueous fraction. In the early phase, 37% and 62% of pain protection were attained with the lower and higher doses of butanol fraction (Figure 3). In the late phase, the aqueous fraction decreased the time by 29% and 57% while butanol fraction reduced by 41% and 69% at the respective lower and higher dose levels (Figure 4).
Anti-inflammatory activity test

As indicated in Table 4, all the tested doses of the crude extract significantly decreased paw volume compared to negative control. The lower dose of methanolic extract achieved a minimum (22%) anti-inflammatory activity at the first hour and maximum value (40%) at the fourth hour while the middle and higher doses inhibit inflammation by 56% (P<0.001) and 71% (P<0.001), respectively, at the fourth hour.

The aqueous and butanol fractions produced an inhibitory action in all times of observations. At the first hour, the 100 mg/kg of aqueous and butanol fractions inhibited inflammation by 24% and 34%, respectively. On the other hand, maximum activity was recorded with the same doses at the fourth hour with the aqueous fraction (41%, P<0.001) and butanol fraction (53%, P<0.001). The 200 mg/kg of the two fractions attained greater anti-inflammatory action as compared to the lower dose levels and were statistically significant as compared to the negative control group. The aqueous fraction produced 56% (P<0.001) and butanol fraction resulted in 69% (P<0.001) anti-inflammatory activity at the fourth hour (Table 4).

Discussion

In this study, the potential anti-nociceptive and anti-inflammatory effects of crude methanolic extract and solvent fractions of *C. cifolius* were investigated using different animal models. In the acetic acid, hot plate and formalin nociception models the test substances significantly provided pain protection. In addition, carrageenan-induced paw edema model was used to evaluate the anti-inflammatory activity of the test substances.

The abdominal constrictions response induced by acetic acid is a sensitive procedure to evaluate peripherally acting analgesics, and a primary tool for screening analgesic potential of test compounds. Intraperitoneal injection of acetic acid causes irritation in the peritoneal cavity where various endogenous inflammatory mediators such as histamine, serotonin, bradykinin substance P, and prostaglandins are released. These inflammatory mediators sensitize C fibers in acetic acid-induced visceral pain. The stimulation of the nerve endings of the primary afferent nerves produces pain characterized by
The plate was and C-

In agreement with this, morphine produced

observed in the present study could be due to

AQ100, aqueous fraction 100 mg/kg; AQ200, aqueous fraction

Percent in

**

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The second pain model used was the hot plate test which is a

thermic stimulus involving stimulation of Aδ fibers. In this
test two behavioral responses, namely paw licking and jumping,
are produced. The two responses are supraspinally inte-
grated, and sensitive to opioid analgesics.38,40 The plate
was maintained at 55°C, and at this temperature only opioid-like
agents are active.28 In agreement with this, morphine produced
significant analgesia at all times of observation. It was selected
as a reference drug by considering its advantages like provi-
sion of longer analgesia and less variability of response among
animals.41

The highest dose (800 mg/kg) of methanol extract
achieved maximum prolongation of reaction time similar to
the standard drug (morphine). However, the lowest and
middle doses (200 and 400 mg/kg) produced maximum
protection later in the course of observation (120 min)
(Table 1). The 200 mg/kg butanol fraction exhibited
more protection at 60 min. Variation in effect may be
attributed to the plasma concentration of the test
substances.42. Overall, methanol extract and solvent
fractions elicited a considerable nociception latency compar-
ing to the reaction time values to thermal stimulus in the
negative control group. Presumably, the effectiveness of
the methanolic extract and solvent fractions of the root of
C. ficifolius observed in the present study could be due to
an opioid-like action via Aδ and C-fibers.

The third model (formalin test) is known for its valid-
ity, reliability, and sensitive for various classes of analge-
sic drugs as compared to other models of nociception.43
Unlike other pain models, it provides a number of advan-
tages, such as little or no restraint, unhindered observation
of the complete range of behavioral responses, and greater
resemblance to clinical pain.31,44,45

As the nociception response has a biphasic nature with
different pain pathways, two phases of behavioral
responses were observed after formalin injection. The early
phase (neurogenic phase) involves stimulation of nociceptors
directly by a chemical and detected by Aδ fibers of the central nociceptive primary afferent terminals. The late phase is due to direct activation of nociceptors by
different chemical mediators that resulted in an increased
input from C fibers.33,38,39,45–47

The late phase of formalin-induced paw licking behav-
ior can be inhibited by 1) NSAIDs via their action against
prostaglandins; 2) steroids, and 3) centrally acting
analgesics.43 The first phase response is mediated by

Table 4 Anti-inflammatory activity of 80% methanolic extract
and solvent fractions of Cucumis ficifolius on carrageenan-induced
paw edema

<table>
<thead>
<tr>
<th>Group</th>
<th>Percent inflammation ± SEM (% inflammation inhibition)</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>1 hr</td>
</tr>
<tr>
<td>NC</td>
<td>54 ±0.043</td>
</tr>
<tr>
<td>CM200</td>
<td>48 ±0.06 (24)*</td>
</tr>
<tr>
<td>CM400</td>
<td>43 ±0.08 (44)</td>
</tr>
<tr>
<td>CM800</td>
<td>36 ±0.035 (51)***</td>
</tr>
<tr>
<td>AQ100</td>
<td>49 ±0.089 (24) **</td>
</tr>
<tr>
<td>AQ200</td>
<td>45 ±0.045 (37)***</td>
</tr>
<tr>
<td>B100</td>
<td>48 ±0.059 (34)***</td>
</tr>
<tr>
<td>B200</td>
<td>44 ±0.063 (48)***</td>
</tr>
<tr>
<td>ASA200</td>
<td>50 ±0.06 (29) ***</td>
</tr>
</tbody>
</table>

Note: *p<0.01, **p<0.001.

Abbreviations: AQ100, aqueous fraction 100 mg/kg; AQ200, aqueous fraction 200 mg/kg; ASA200, acetylsalicylic acid 200 mg/kg; B100, butanol fraction 100 mg/kg; B200, butanol 200 mg/kg; CM200, crude extract 200 mg/kg; CM 400, crude extract 400 mg/kg; CM800, crude extract 800 mg/kg; NC, negative control; SEM, standard error of the mean.

constriction of abdominal muscle with the extension of the
forelimbs and elongation of the body.37,39

The percentage pain protection produced by the use of
the crude extract at three doses (200, 400 and 800 mg/kg)
was 24.9% (P<0.05), 48.7% (P<0.001) and 72.5%
(P<0.001), respectively, as compared to the vehicle
group (Figure 1). Butanol and aqueous solvent fractions also
exhibited a greater reduction of the frequency of writhing
response (Figure 2). Analgesic effect of the crude extract
and solvent fractions in this study probably linked to

C fibers mediated via inhibition of prostaglandins histag-
mine, serotonin bradykinin, and substance P.
Substance P, whereas serotonin, histamine, bradykinin, and prostaglandins are reported to involve in the second phase. The crude methanolic root extract of *C. ficifolius* significantly diminished the mean time animals spent licking the injected paw in the early and late phases formalin-induced pain (Table 3). The aqueous fraction at 100 mg/kg and 200 mg/kg decreased the time of licking by 28% and 56%, respectively, in the early phase while butanol fraction showed higher protection action as compared to the aqueous fraction (Figure 3). In the late phase, the aqueous fraction decreased the time by 29% and 57% while the butanol fraction reduced by 41% and 69% at the respective lower and higher dose levels (Figure 4). In formalin-induced pain the mechanism of test substances may involve inhibition of nociception transmission via central pain pathways in the early phase and inhibitory of inflammatory mediators, such as serotonin, histamine, bradykinin, prostaglandins, in the second phase.

The presence of edema is one of the prime signs of inflammation. Carrageenan-induced paw edema is a well-defined model of acute inflammation and a variety of inflammatory mediators participate in its development. The evolution of carrageenan-induced acute inflammation indicates the presence of two phases, namely initial and late phases.

Following injection of carrageenan, inflammatory mediators such as bradykinin, serotonin and histamine contribute the initial phase occurring from 0 to 2.5 h. As indicated in Table 4, the maximum peak of edema was observed at 180 min, which is thought to be due to the release of kinin-like substances, particularly of bradykinin. The second phase of edema is a result of overproduction of prostaglandins in tissues and may occur from 2.5 to 6 h post-carrageenan injection.

The treatments achieved maximum anti-inflammatory activity at the fourth hour. This is supported by reports that the second phase is proven to be sensitive to the commonly used anti-inflammatory drugs. COX mediated production of different prostaglandins accounted for the emergence of the late phase of induced inflammation but not to lipoxygenase inhibitors.

Based on the result of the present study, a methanolic crude extract, aqueous and butanol fractions of *C. ficifolius* significantly decreased paw edema in both phases of carrageenan-induced acute inflammation. This suggests that bioactive constituents in the crude extract and solvent fractions may suppress both phases of acute inflammation by interfering with the release and/or activity of the chemical mediators, such as histamine, bradykinin, and serotonin in the first phase. In the late phase, a reduction in edema may be attributed to COX inhibitory action of 80% methanol extract and solvent fractions.

Previous phytochemical screening on methanol extract has revealed the presence of phenols, tannins, saponins, terpenoids, and flavonoids. The butanolic extract of *Cucumis sativus* demonstrated the presence of flavonoids, saponins, and steroids. In addition, carbohydrates, tannins, alkaloids, saponins, flavonoids, glycosides steroids were found in aqueous extract of *Cucumis melo*. The anti-nociceptive and anti-inflammatory effect of many plants has been attributed to their flavonoid, terpenoid, steroid, tannin, phenol, alkaloid and saponin constituents. Flavonoids exhibited anti-inflammatory through their free radical scavenging activity, for example, reactive oxygen species (ROS), and interfere with the action of pro-inflammatory cytokines, such as interleukin-6 (IL-6), TNF-α, IL-1β, and nuclear factor-kappa B. Hence the analgesic and anti-inflammatory activities of *C. ficifolius* might be due to the presence of the aforementioned phytoconstituents.

It can be concluded from this study that the crude methanolic extract, aqueous and butanol fractions of *C. ficifolius* proved to have analgesic properties against thermal and chemical noxious stimuli pain models; and anti-inflammatory activity in carrageenan-induced paw edema. The study justified the local use of *C. ficifolius* roots in the management of inflammatory and pain conditions.

**Conclusion and recommendation**

In this study crude methanolic extract, aqueous and butanol fractions of *C. ficifolius* proved to have analgesic properties against thermal and chemical noxious stimuli pain models; and anti-inflammatory activity in carrageenan-induced paw edema. The crude extract and solvent fractions possess peripheral and central analgesic activity. The mechanism of anti-inflammatory actions may involve a multitude of inflammatory mediators which needs further investigation.

Further constituent isolation, binding studies, and electrophysiological procedures may be useful to fully elucidate the anti-nociceptive and anti-inflammatory effects and mechanism of *C. ficifolius*.

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Author contributions
All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure
The authors report no conflicts of interest in this work.

References


