

# Wnt5a induces ROR1 and ROR2 to activate RhoA in esophageal squamous cell carcinoma cells

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**Background:** Wnt5a is a nontransforming Wnt family member and identified as an oncogenic role on cell motility of breast cancer and glioblastoma. However, Wnt5a signaling in esophageal squamous cell carcinoma (ESCC) progression remains poorly defined.

**Materials and methods:** Immunohistochemistry assays were used to measure the Wnt5a expression in ESCC sections. We evaluated the role of receptor tyrosine kinase-like orphan receptor (ROR)1/2 and RhoA on the invasion of ESCC cells by using cell invasion assay, immunoprecipitation, immunofluorescence, and Rho activation assay.

**Results:** Wnt5a was highly expressed in invasive ESCC tissues compared with that in noninvasive and nonmalignant tissues. In vitro assay showed that sfrp2 (Wnt5a antagonist) largely blocked the invasion but not the colony formation of KYSE410 and KYSE520 ESCC cells. Anti-ROR1 mAb and ROR2-shRNA markedly inhibited the disheveled-associated activator of morphogenesis 1 (DAAM1) activity, RhoA activity, microfilament formation and the invasion of ESCC cells. Fluorescent phalloidin staining experiment showed ROR1/ROR2, receptors of Wnt5a signaling, and regulated the reassembly of actin filaments in ESCC cells. Further experiments showed that ROR1 was strongly associated with ROR2 in KYSE410 cells. The activation of RhoA, not Rac1 or Rac2, was involved in ROR1/ROR2 signaling pathway. By using DAAM1 shRNA, we found that RhoA was downstream of DAAM1, which could be rescued by the overexpression of wild-type DAAM1. This could be further proved by a RhoA inhibitor CCG-1423 which could inhibit the invasion of ESCC cells but not DAAM1 activity.

**Conclusions:** Wnt5a promotes ESCC cell invasion via ROR1 and ROR2 receptors and DAAM1/RhoA signaling pathway.

**Keywords:** ESCC, invasion, Wnt5a, DAAM1, ROR

## Introduction

Esophageal squamous cell carcinoma (ESCC) is a common malignant tumor of digestive system and its mortality rate ranks sixth all around the world.<sup>1</sup> Surgical resection is the first choice for patients with early ESCC. However, patients mainly suffer from advanced ESCC which is often accompanied by lymph node metastasis or other complications.<sup>2</sup> The stage of ESCC invasion based on TNM staging system helps guide clinical decision and the subsequent therapeutic regimen making.<sup>3-5</sup> Cancer cells invading the para-carcinoma tissues causes cancer cells spreading to lymph nodes or distant organs, resulting in the deterioration of this disease. However, the underlying mechanism of ESCC cell invasion is still largely unknown.

According to the comprehensive genomic analysis of 158 ESCC cases, the International Cancer Genome Consortium research project reveals that somatic

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variations are mainly involved in Wnt and Notch pathways.<sup>6</sup> The Wnt signaling pathways begin with proteins passing signals into the cell through cell surface receptors, including Frizzled family receptors, lipoprotein receptor-related proteins, and receptor tyrosine kinase-like orphan receptors (RORs).<sup>7,8</sup> Wnt signaling pathways have been characterized: the canonical Wnt pathway, the noncanonical planar cell polarity pathway, and the noncanonical Wnt/calcium pathway.<sup>9</sup> Aberrant expression and location of beta-catenin, a member of the Wnt signaling pathway, regulates the progression of oesophageal cancer.<sup>10</sup> Wnt2 secreted by tumor fibroblasts enhances the motility and the invasiveness of ESCC cells by inducing epithelial–mesenchymal transition.<sup>11</sup> Treatment with galangin (an active pharmacological ingredient from propolis and *Alpinia officinarum* Hance) and berberine (a major component of *Berberis vulgaris* extract) causes the decreased expression of Wnt3a/ $\beta$ -catenin and induces apoptosis in ESCC cells.<sup>12</sup>

Wnt5a, as a nontransforming Wnt family member, mediates cancer initiation and metastasis.<sup>13,14</sup> Previous studies found that Wnt5a triggers disheveled 2/disheveled-associated activator of morphogenesis 1 (DAAM1) signaling pathway and activates RhoA, resulting in the elevation of the migratory capacity of breast cancer cells and the invasion of glioblastoma cells.<sup>15,16</sup> DAAM1, an element of cellular actin cytoskeleton, interacts with disheveled and RhoA and can polymerize actin filaments at the barbed end.<sup>13,15,17</sup> The active DAAM1 is elevated by the treatment of Wnt5a or type IV collagen and DAAM1 participates in the breast cancer cell migration and haptotaxis.<sup>15,18</sup> However, the role of Wnt5a in the invasion of ESCC cells is still largely unknown.

In this study, we first demonstrate that Wnt5a is upregulated in invasive ESCC tissues and promotes the invasion of ESCC cells. We also describe the mechanism underlying the Wnt5a/ROR1/2/DAAM1/RhoA signaling pathways that regulate ESCC cell invasion. Overall, these data identify ROR1/2 as the novel therapeutic targets in limiting esophageal cancer invasion.

## Materials and methods

### Clinical samples

Twenty-two ESCC patients were recruited by The Second Hospital of Nanjing from 2014 to 2018. The areas of higher tumor cell density for immunohistochemistry (IHC) were histopathologically confirmed by a pathologist. Formalin-fixed and paraffin-embedded tumor samples were obtained for IHC analysis of Wnt5a. Pathologic staging was

performed in accordance with the Union for International Cancer Control. The study was conducted in accordance with the Declaration of Helsinki and reviewed and approved by the research ethics committee, The Second Hospital of Nanjing (No. 2018-LY-KY068). Written informed consent was taken from each participant.

### Cell culture and transfections

The human ESCC cell lines KYSE410 and KYSE520 were from the Cell Bank of Chinese Academy of Sciences (Shanghai, China). These cells were grown in RPMI-1640 medium (Cat. KGM31800-500, KeyGEN BioTECH, Nanjing, China) supplemented with 10% (v/v) FBS (Cat. SH30068.03, Hyclone, Logan, UT, USA) and 0.5  $\mu$ g/mL penicillin/streptomycin in a humidified incubator at 37°C with 5% CO<sub>2</sub>.

Short hairpin RNAs (shRNAs) specific for DAAM1 (5'-GCCACTTTGTATCCTATCAGG-3'), ROR2 shRNA (Cat. sc-72390-SH, Santa Cruz Biotechnology, Dallas, TX, USA) and/or wild-type (WT) ROR2 constructs were transfected into KYSE410 and KYSE520 cells using Lipofectamine 2000 reagent (Cat. 11668-019, Invitrogen, Carlsbad, CA, USA). The cells were switched to fresh medium containing 10% FBS 6 hrs after transfection and cultured for 48 hrs. All cells were cultured in a 37°C incubator with 5% CO<sub>2</sub>.

### Immunohistochemistry (IHC)

ESCC pathological sections were deparaffinized at 55°C for 30 mins. Then, the sections were washed with xylene for three 5-min washes. The sections were rehydrated by 5-min successive washes in 100%, 90%, and 70% ethanol. Antigens were retrieved by microwaving the samples for 4 mins in 250 mL of 10 mmol/L sodium citrate (pH 6.0). Furthermore, endogenous peroxidase activity was blocked by incubation with 0.3% hydrogen peroxidase (ZSGB-Bio, Beijing, China) for 20 mins. The primary anti-Wnt5a (1:100 dilution, Cat. sc-365370, Santa Cruz Biotechnology) antibody was used. Diaminobenzidine (DAB) and hematoxylin counterstain (ZSGB-Bio) were used to visualize the antibody staining.

The percentage of positively stained cells was scored from 0 to 4: 0 (<10%), 1 (10–25%), 2 (26–50%), 3 (51–75%), and 4 (76–100%). The staining intensity was scored on a scale of 0 to 3: 0 (negative), 1 (weak), 2 (moderate), and 3 (strong). The immunoreactivity score (IRS) for each case was multiplied by the percentages of positive cells and staining intensities. The IRS ranged from 0 to 12 (0, 1, 2, 3, 4, 6, 8, 9, and 12) and was performed without prior clinical

records. All these results come from the same block in which both tumor and nonmalignant cells were counted. The invasive ESCC is judged by an experienced pathologist who showed that tumor cells penetrate the mucosa of esophagus.

## Immunoblotting

KYSE410 and KYSE520 cells treated with CCG-1423 (Cat. S7719, Selleck Chemicals, Houston, TX, USA) or transfected with indicated constructs were washed with PBS and lysed with RIPA lysis buffer.<sup>19</sup> The lysates were then clarified by centrifugation at 12,000×g for 20 mins at 4°C. The protein extracts were separated by 8% or 10% SDS-PAGE. The following antibodies were used: anti-Wnt5a (1:1,000 dilution, Cat. sc-365370, Santa Cruz Biotechnology), anti-β-actin (1:5,000 dilution, Cat. 60008-1-Ig, ProteinTech Group, Wuhan, China), anti-ROR1 (1:1,000 dilution, Cat. 20629-1-AP, ProteinTech), anti-ROR2 (1:1,000 dilution, Cat. 88639, Cell Signalling Technology, Danvers, MA, USA), anti-flag (1:1,000 dilution, Cat. 14793, Cell Signalling Technology), anti-GST (1:1,000 dilution, Cat. 2624, Cell Signalling Technology), anti-RhoA (1:1,000 dilution, Cat. 10749-1-AP, ProteinTech), and anti-DAAM1 (1:1,000 dilution, Cat. sc-100942, Santa Cruz Biotechnology) antibodies. Horseradish peroxidase-conjugated antibodies and Pierce™ ECL western blotting substrate (Cat. 32209, Thermo Fisher Scientific, Waltham, MA, USA) were used to exposure protein bands. The pictures were taken by Tanon 5200 Multi Imaging Workstation (Tanon, Shanghai, China) and the densitometry of immunoblots was analyzed by ImageCal software (Tanon).

## Pulldown assays

GST-RhoA beads were used to detect active DAAM1 and first incubated with 0.1 mmol/L GTPγS (Sigma-Aldrich, St. Louis, MO, USA) at 30°C for 15 mins. Cells were washed with PBS and lysed with lysis buffer.<sup>18</sup> The equal amount of cellular lysates was incubated with GTPγS-incubated GST-RhoA beads captured on MagneGST glutathione particles (Cat. V8611, Promega, Fitchburg, WI, USA) for 90 mins at 4°C with constant rotation. The beads were washed three times with washing buffer.<sup>18</sup> After that, beads were gathered by a magnet in a magnetic stand. The SDS-PAGE and the measurement of the immunoblot bands density were undergone according to standard protocol.

The bacterially expressed recombinant GST-ROR1 and flag-ROR2 were purified by using GST-Sepharose 4B (Cat. 17-0756-01, GE Healthcare, Uppsala, Sweden) and Pierce Anti-DYKD4K (FLAG) Magnetic Agarose Beads (Cat. A36797, Thermo Fisher Scientific), respectively.

GST-ROR1 and flag-ROR2 beads were stored for 90 mins at 4°C with constant rotation. After rinsing with washing buffer three times, the beads were enriched by centrifugation at 12,000 × g for 5 mins.

## Enzyme-linked immunosorbent assay (ELISA)

KYSE410 and KYSE520 cells were grown in normal cell cultural condition. Equal volumes of cell culture media were diluted in ice-cold PBS buffer. The experiments were then performed in accordance with the protocol of the Wnt5a ELISA kit (Cat. CSB-EL026138HU, CusaBio, Wuhan, China). The concentration was calculated according to the concentration curve for Wnt5a standard samples.

## Boyden chamber assays

A modified Boyden chamber system (8.0 μm pore diameter, Cat. 3422, Costar, Corning, NY, USA) were used in this study. The membranes of Boyden chambers were coated with Matrigel™ Basement Membrane Matrix (Cat. 356234, BD Biosciences, Bedford, MA, USA) for 30 mins at 37°C. KYSE410 and KYSE520 cells were treated with ROR1 rabbit mAb (1:1,000 dilution, Cat. 16540, Cell Signalling Technology) or vehicle and then suspended in serum-free culture medium supplemented with 5 μg/mL BSA. The suspensions containing 1×10<sup>5</sup> cells were added into the upper sides of Boyden chambers. The cells were allowed to invade at 37°C. After that, stationary cells were removed with a cotton-tipped applicator, and the membranes were stained with 0.5% crystal violet. The number of cells passing through the membrane was evaluated and counted in a microscope (Mshot MF53, Micro-shot Technology Co., Guangzhou, China).

## Clonogenic assay

KYSE410 and KYSE520 cells were placed into 6-well plates (5,000 cells/well) and incubated with 1 μg/mL secreted Frizzled-related protein 2 (sfrp2) at 37°C for 15 days, fixed and stained with crystal violet. The pictures of tumor colonies were taken by a commercial digital camera (PowerShot S110, Canon, Japan). The number of colonies was counted using an Adobe Photoshop CS2 software from five independent replicates.

## Immunoprecipitation (IP)

To test the interaction between ROR1 and ROR2, KYSE410 cells were lysed with lysis buffer.<sup>18</sup> Cellular lysates were centrifuged at 10,000×g for 10 mins. Clarified lysates were

incubated with ROR2 or IgG antibodies overnight and then treated with protein A/G conjugated agarose beads (Pierce, Rockford, IL, USA). After incubation for 1.5 hrs with agitation, beads were washed with lysis buffer for three times. After boiling in 60  $\mu$ L of 2 $\times$ SDS sample buffer for 5 mins, samples were subjected to SDS-PAGE and blotted with anti-ROR1 (1:1,000 dilution) or anti-ROR2 antibodies (1:1,000 dilution).

### Rho GTPase activation assays (G-LISA small GTPase activation assays)

Rho GTPase activation assays (Cat. BK121 for RhoA, Cat. BK127 for Cdc42, Cat. BK125 for Rac1,2,3, Cytoskeleton Inc., Denver, CO, USA) were used to measure the Rho activity. Briefly, cells were plated onto 24-well plates (Costar) and grown to the desired confluency. Cell extracts were equalized with ice-cold lysis buffer containing protease inhibitor cocktail. Ten microliters of each lysate or lysis buffer and 290  $\mu$ L of Precision Red Advanced Protein Assay Reagent were added into a 1.5 mL tube. The mixture was incubated for 1 min at room temperature. The Precision Red Advanced Protein Assay Reagent results in a red to purple/blue color change which can be characterized by an increase in absorbance at 600 nm. The absorbance of each sample was read in a blank spectrophotometer (AOE Instruments, Shanghai, China).

### Immunofluorescence and actin cytoskeleton staining

Cells adhering on glass slides were fixed in 4% paraformaldehyde/PBS for 20 mins and then permeabilized in 0.2% Triton X-100. Cells were subsequently blocked in PBS containing 1% BSA for 60 mins at room temperature. Next, cells were stained with tetramethylrhodamine (TRITC)-labelled phalloidin (5  $\mu$ g/mL, Sigma-Aldrich) for 40 mins in room temperature. After washing with PBS, DAPI Fluoromount G (Southern Biotech, Birmingham, AL, USA) was added to the glass slides. The images were snapshoted by a fluorescence microscope (Zeiss LSM710, Oberkochen, Germany).

### Statistical analysis

All statistical analyses were calculated by SPSS 23.0 software (Chicago, IL, USA). The data of ELISA assays (Figure 1E) were analyzed by Student's *t*-test, the data of correlation analysis (Table 1) were analyzed by  $\chi^2$  test, and the other data were analyzed by one-way ANOVA. The number of replications was five if not noted, which were acquired in five independent

experiments. For all analyses, bar charts show means  $\pm$  SD and a two-sided *p*-value  $<0.05$  was defined as statistically significant.

## Results

### Wnt5a is highly expressed in invasive ESCC tissues

To investigate whether Wnt5a signaling regulated the pathogenesis of human ESCC, the expression patterns of Wnt5a were examined in invasive ESCC tissues. According to the analysis of 22 ESCC cases, we found that the Wnt5a was highly expressed in invasive tumor tissues compared with that in noninvasive tumor tissues (Figure 1A and B). There is no expression in nonmalignant esophageal tissues (Figure 1A). However, we did not find the differential Wnt5a expression between in small tumor tissues and in large tumor tissues (Figure 1C). Table 1 shows the clinical characteristics of 22 esophageal cancer patients. Wnt5a expression seems highly associated with lymph node metastasis (Table 1), although more patients' samples are needed for further analysis.

We previously investigated EGFR expression and telomere in ESCC cells KYSE410 and KYSE520 and found KYSE410 and KYSE520 cells had the most difference in all nine KYSE cell lines.<sup>20–22</sup> In this study, we found the high expression and secretion levels of Wnt5a in KYSE410 and KYSE520 ESCC cells (Figure 1D and E), which were subjected to the measure of the ability of Wnt5a-induced cell invasion. The quantity of secretory endogenous Wnt5a was  $236.2 \pm 40.96$  ng/mL and  $243.6 \pm 35.91$  ng/mL in KYSE410 and KYSE520 cells, respectively (Figure 1E).

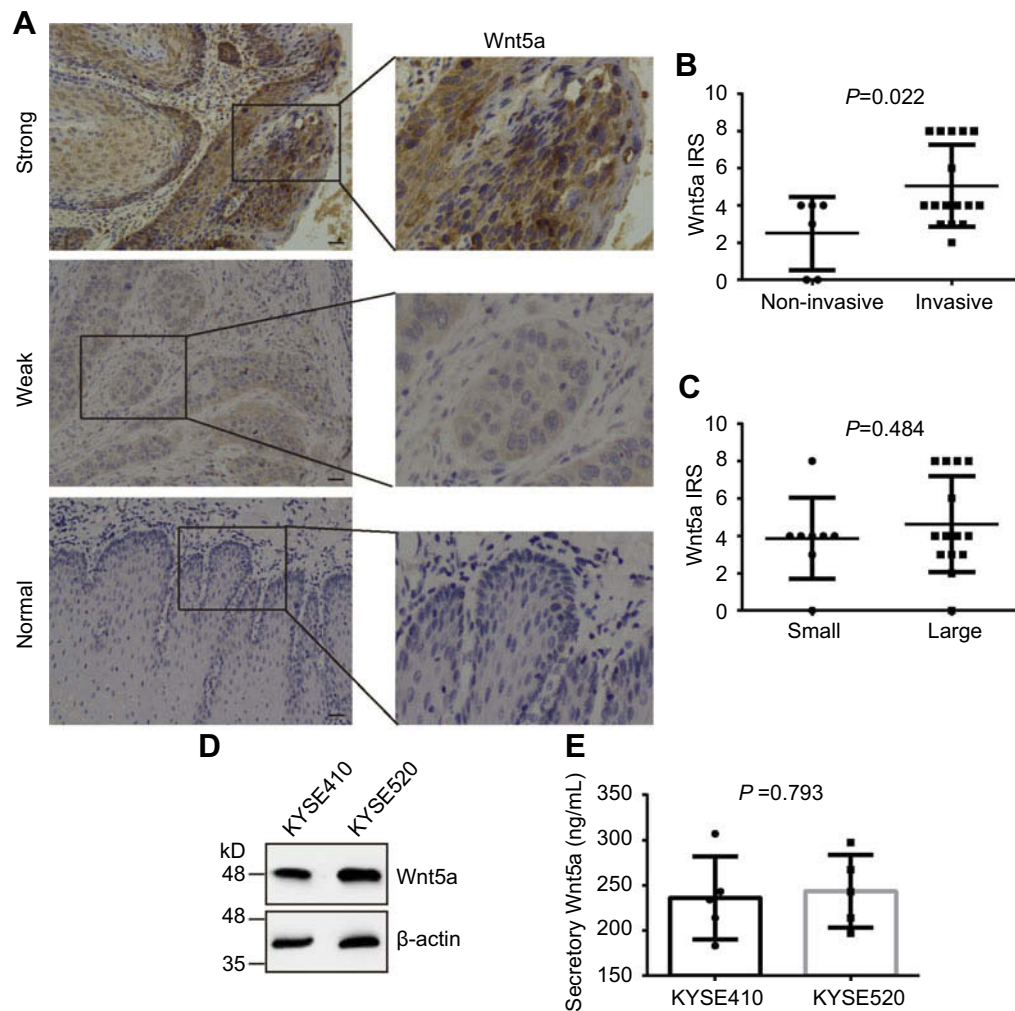
### Wnt5a induces the invasion of ESCC cells

To determine whether Wnt5a induced the invasion of ESCC cells, we examined the invasion of KYSE410 and KYSE520 cells through 8.0- $\mu$ m pores in membranes coated with matrigel. An approximately four-fold decrease was shown in the number of invasive cells treated with 1  $\mu$ g/mL secreted Frizzled-related protein 2 (sfrp2), a Wnt5a antagonist (Figure 2A and B). Moreover, sfrp2 did not alter the ability of colony formation of KYSE410 and KYSE520 ESCC cells (Figure 2C and D). Thus, Wnt5a signaling participates in the regulation of ESCC cell invasion.

### ROR1 and ROR2 participate in the invasion of ESCC cells

To investigate the effect of the receptors of Wnt5a signaling on the invasion of ESCC cells, we blocked the ROR1





**Figure 1** Wnt5a is highly expressed in the invasive ESCC tissues. **(A)** The representative images of Wnt5a expression in the pathological sections of ESCC and nonmalignant esophageal tissues. Wnt5a is highly expressed in invasive ESCC tissue. DAB substrate solution was applied to reveal the brown color of Wnt5a antibody staining. Hematoxylin staining showed the blue color of nucleus. Bar = 100  $\mu$ m. Objective lens, magnification,  $\times 20$ ; numerical aperture, 0.75. **(B)** Wnt5a was highly expressed in the invasive ESCC tissues ( $n=16$ ) than that in the noninvasive ESCC tissues ( $n=6$ ). Wnt5a immunostaining was analyzed by the evaluation of the percentage of tumor-stained cells and staining intensity, allowing assessment of an IRS. **(C)** Wnt5a expression was not shown correlation with the clinical and pathological characteristics of tumor size in ESCC. Tumour size, small,  $\leq 2$  cm. Large,  $>2$  cm. **(D)** Western blotting revealed the endogenous Wnt5a expression in KYSE410 and KYSE520 ESCC cells.  $\beta$ -actin as the loading control. **(E)** The Wnt5a secretion was measured and the culture media of KYSE410 and KYSE520 cells by ELISA.

**Abbreviations:** ESCC, esophageal squamous cell carcinoma; IRC, immunoreactive score.

and/or ROR2 signaling of ESCC cells and examined the cell invasion by Boyden chamber assays. ROR2-shRNA knocked down ROR2 expression by  $\sim 70\%$  in KYSE410 and KYSE520 cells, which can be rescued by WT ROR2 overexpression (Figure 3A). Silence of ROR2 gave rise to a significant reduction of cell invasion, which also can be rescued by WT ROR2 overexpression (Figure 3B and C).

Next, we tested whether ROR1 physically interacted with ROR2 in human ESCC cells. Interestingly, ROR1 was strongly associated with ROR2 in KYSE410 cells (Figure 3D). An in vitro binding assay demonstrated that purified bacterially expressed recombinant ROR1 was directly binding to purified ROR2 (Figure 3E).

Thus, ROR1/ROR2 receptors participate in the invasion of ESCC cells.

## RhoA functions as a downstream target of ROR1/ROR2

The significantly decreased activity of Rac1, Rac2, and RhoA treated with anti-ROR1 mAb were found in KYSE410 ESCC cells (Figure 4A). To elucidate which specific Rho GTPases were involved in both ROR1 and ROR2 signaling pathways, we performed Rho GTPase activation assays after ROR2 knockdown. The activation of RhoA, not Rac1 or Rac2, was downregulated in ROR2

**Table I** Associations between Wnt5a expression and clinicopathological features of esophageal squamous cell carcinoma

Characteristics	Cases	Wnt5a IRS <sup>a</sup>		$\chi^2$	P-value
		Low	High		
Age (years)					
≤64	11	11	0	7.875	0.005
>64	11	5	6		
Tumor					
T1–T2	15	11	4	0.008	0.927
T3	7	5	2		
Lymph node metastasis					
N0	17	15	2	4.886	0.027
N1–N2	5	2	3		
Distant metastasis					
M0	22	16	6	N/A	N/A
M1	0	0	0		
Pathologic grade					
I–II	14	11	3	0.633	0.426
III	8	5	3		

**Notes:** <sup>a</sup>Low expression: IRS≤4, high expression: IRS>4. Bold values indicate significance of P-values (P<0.05).

**Abbreviation:** IRS, immunoreactivity score.

knockdown cells, which could be rescued by the WT ROR2 overexpression (Figure 4B and C). Moreover, the combination of ROR1 blockage and ROR2 knockdown showed a similar effect on the decrease of RhoA activity to the single treatment of ROR1 blockage or ROR2 knockdown (Figure 4C).

CCG-1423 (a RhoA-specific inhibitor) was used to block the RhoA activity and to study its role in cell invasion. Treatment with 1 μmol/L CCG-1423 inhibited the cell invasion of ESCC cells by approximately 59.3% and 66.3% reduction for KYSE410 and KYSE520 cells, respectively (Figure 4D). Thus, these results suggested that RhoA functions as a downstream target of ROR1/ROR2 and is participated in cell invasion of ESCC cells.

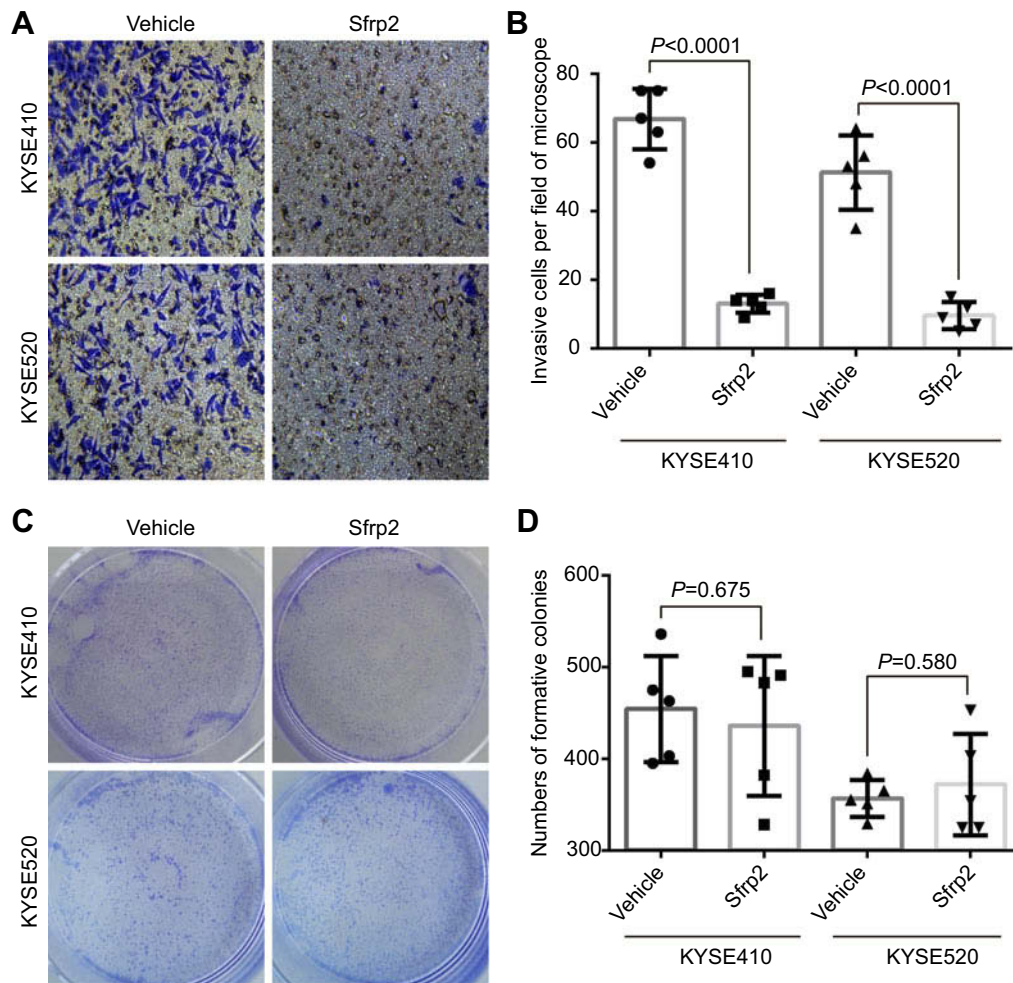
### ROR1/ROR2-dependent DAAM1 activity is involved in cell invasion

To study whether RhoA acted as the downstream of DAAM1, we silenced DAAM1 using shRNA and measured the RhoA activity. A ~0.5-fold decrease was shown in DAAM1-knockdown ESCC cells (Figure 5A). The blockage of RhoA activity by CCG-1423 treatment was unable to change DAAM1 activity (Figure 5B), suggesting the role of DAAM1 functioning as the upstream of RhoA.

Next, we examined whether DAAM1 activity was dependent on ROR1/ROR2 receptors in ESCC cells. A significant decrease in active DAAM1 levels was shown after ROR1 blockage or ROR2 knockdown (Figure 5C). The invasion of DAAM1 knockdown cells was largely inhibited, which could be rescued by the overexpression of WT DAAM1 (Figure 5D). Taken together, these experiments demonstrated that ROR1/ROR2-dependent DAAM1 activity acts as the upstream of RhoA and is involved in cell invasion of ESCC cells.

### ROR1 and ROR2 regulate the reassembly of microfilaments

We next examined whether ROR1 and ROR2 signaling regulate the reassembly of actin filaments in ESCC cells. Fluorescent phalloidin staining was performed to investigate the distribution of filamentous actin (F-actin) in KYSE410 cells. We found that the formation of microfilaments was disrupted by anti-ROR1 mAb treatment, and the length of microfilaments was largely decreased (Figure 6A–C). We also found that the knockdown of ROR2 disrupted the formation of microfilaments and decreased the length of microfilaments (Figure 6A–C). Thus, the findings indicated that ROR1 and ROR2 signaling regulates the reassembly of microfilaments in ESCC cells.



**Figure 2** Wnt5a stimulates the invasion of ESCC cells. **(A and B)** KYSE410 and KYSE520 ESCC cells were treated with 1  $\mu$ g/mL secreted Frizzled-related protein 2 (sfrp2), an antagonist of Wnt5a, and/or vehicles and allowed to invade the matrigel for 6 hrs. The invasive cells on the lower sides of Boyden chamber membranes were counted under a microscope field. Objective lens, magnification,  $\times 20$ ; numerical aperture, 0.75. **(C and D)** Wnt5a did not alter the ability of the colony formation of KYSE410 and KYSE520 cells. The single-cell suspensions of KYSE410 and KYSE520 cells were seeded on 6-well plates and were treated with 1  $\mu$ g/mL sfrp2 and/or vehicles. They were allowed to grow in a normal cell-cultural condition for 15 days. Tumor cell colonies were stained with crystal violet and counted the number.

**Abbreviation:** ESCC, esophageal squamous cell carcinoma.

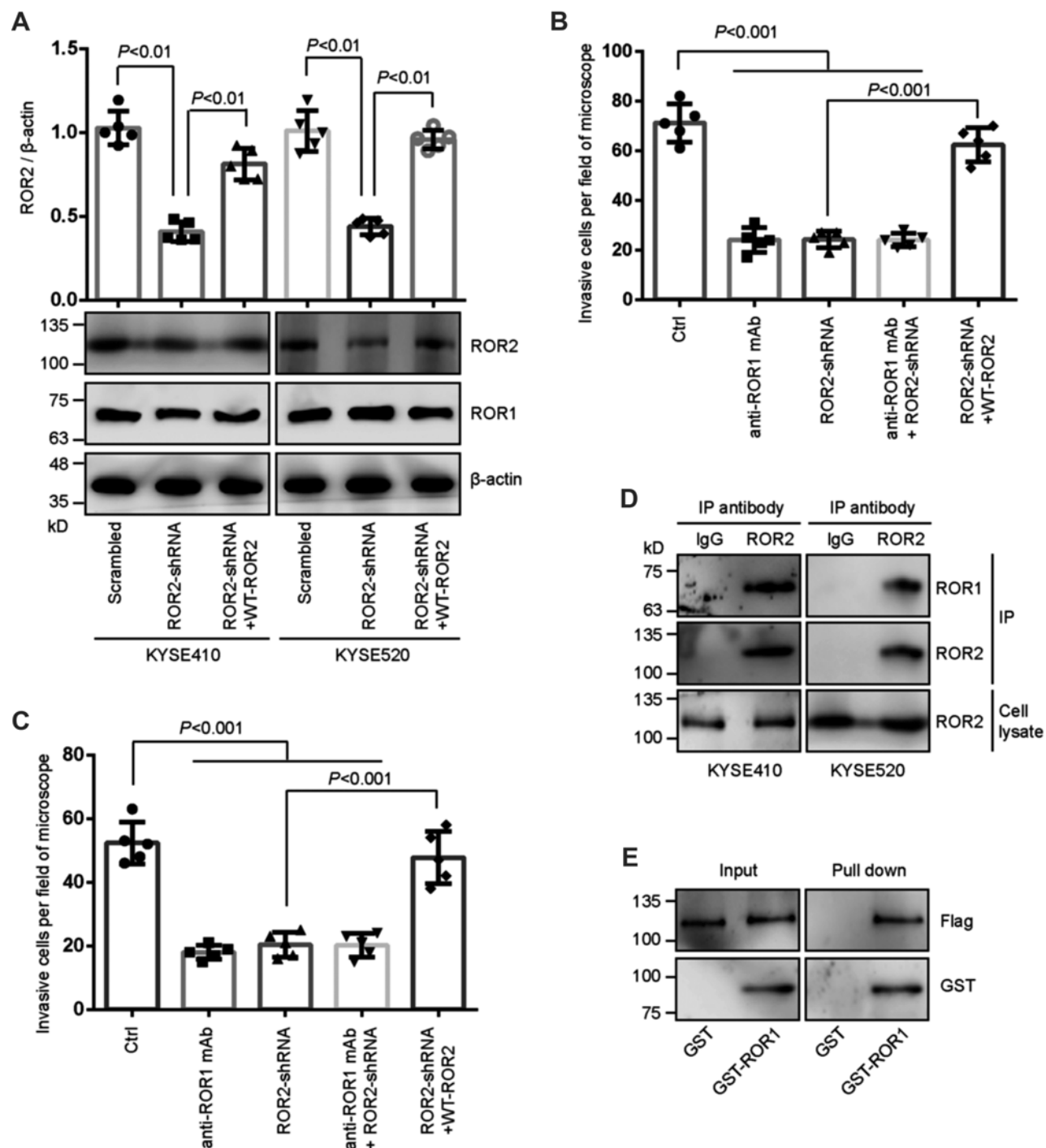
## Discussion

We previously found that Wnt5a stimulates the migration of breast cancer cells and promotes the invasion of glioblastoma cells via Rho signaling pathways.<sup>15,16,19</sup> Liu et al reported that Wnt5a enhances the ability of cell mobility via RhoA signaling pathways in gastric cancer.<sup>23</sup> In this study, we demonstrated that Wnt5a was highly expressed in invasive ESCC tissues, and the blockage of Wnt5a signaling by sfrp2 treatment largely inhibited the invasion of ESCC cells. These evidence support that Wnt5a acts as an oncogene in multiple solid tumours, including esophageal cancer, breast cancer, glioblastoma, and gastric cancer.

Evolutionarily conserved ROR1/2 are regarded as distinct receptors for Wnt5a and are involved in noncanonical

Wnt signaling in organogenesis and cancer metastasis.<sup>24</sup> Cirmtuzumab (UC-961) is a first-in-class humanized IgG1 monoclonal antibody, which is designed and developed to bind with high affinity to the extracellular domain of ROR1.<sup>25–27</sup> Cirmtuzumab is safe and effective in a Phase I trial for patients with progressive, relapsed, or refractory chronic lymphocytic leukemia (CLL).<sup>28</sup> Consistent with the studies for CLL, we found that anti-ROR1 mAb markedly inhibits the DAAM1/RhoA activity, microfilament formation, and the invasion of ESCC cells. This study identifies ROR1 as a novel pharmaceutical target to postpone the invasion of ESCC.

Wnt5a induces ROR1/2 heterooligomerization to strengthen leukemia chemotaxis and proliferation.<sup>24</sup>



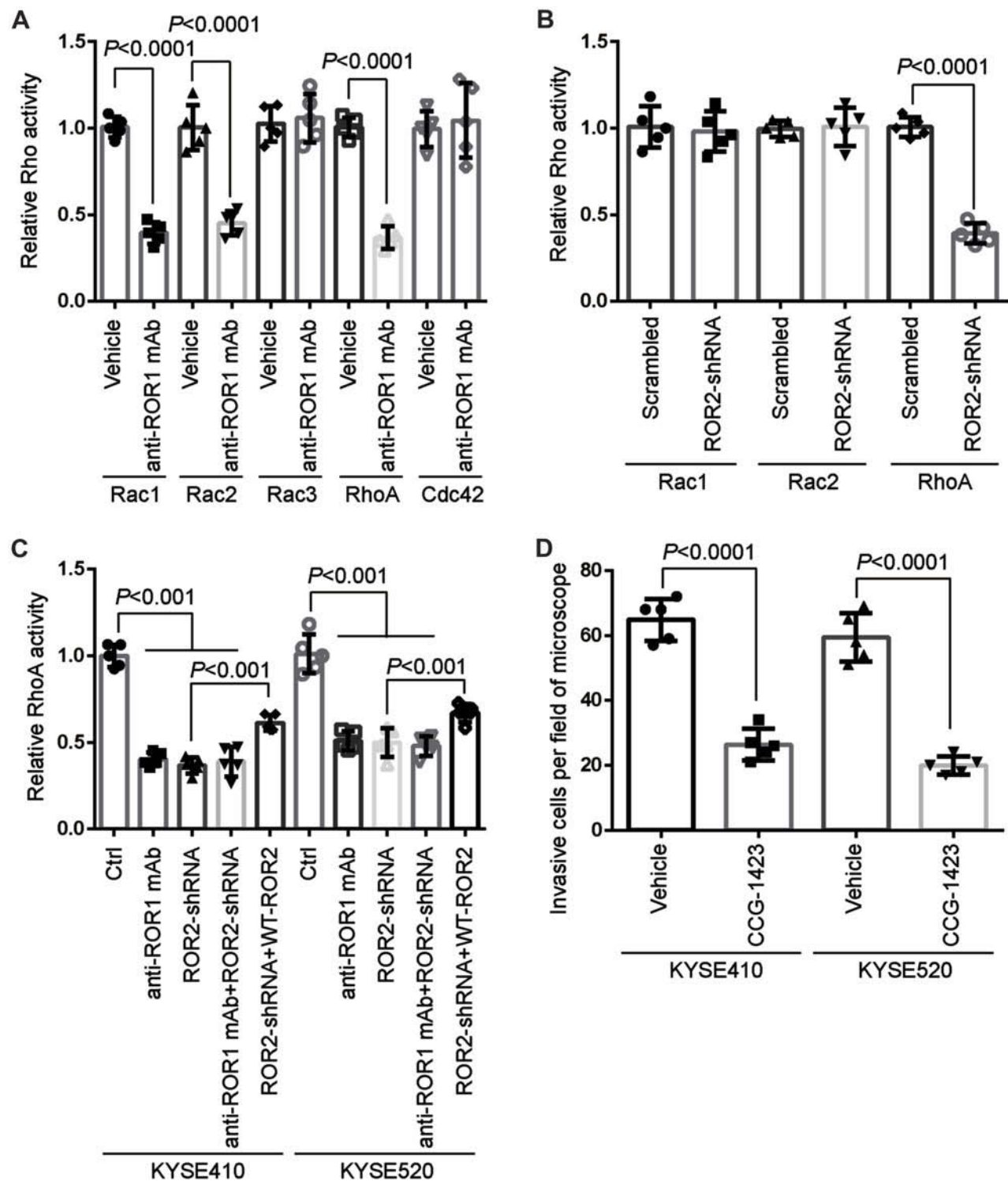
**Figure 3** ROR1 and ROR2 receptors are required for the invasion of ESCC cells. **(A)** KYSE410 and KYSE520 were transfected with scrambled shRNA, shRNA against ROR2 (ROR2-shRNA), or ROR2-shRNA plus shRNA-resistant WT ROR2, and then subjected to the Western blotting assays.  $\beta$ -actin as the loading control. **(B and C)** Cell invasion rate was largely abolished by anti-ROR1 mAb, ROR2-shRNA, or ROR2-shRNA plus shRNA-resistant WT ROR2. KYSE410 **(B)** and KYSE520 **(C)** cells were incubated with anti-ROR1 mAb, transfected with ROR2-shRNA or ROR2-shRNA plus shRNA-resistant WT ROR2, and then subjected to cell invasion assays. **(D)** The lysate of KYSE410 cells was subjected to immunoprecipitation (IP) with antibody to ROR2, followed by immunoblotting with antibody to ROR1 or ROR2. **(E)** ROR1 was directly binding to ROR2. Purified GST or GST-ROR1 was incubated with purified Flag-tagged full-length ROR2. The amounts of Flag-ROR2 co-purified with GST or GST-ROR1 (pull down) were analyzed by immunoblotting for anti-Flag or anti-GST.

**Abbreviations:** ESCC, esophageal squamous cell carcinoma; WT, wild type; ROR, receptor tyrosine kinase-like orphan receptor.

ROR2 expressed on the plasma membrane is essential for ligand binding and Wnt signal transduction.<sup>29,30</sup> In this study, an in vitro binding assay and immunoprecipitation

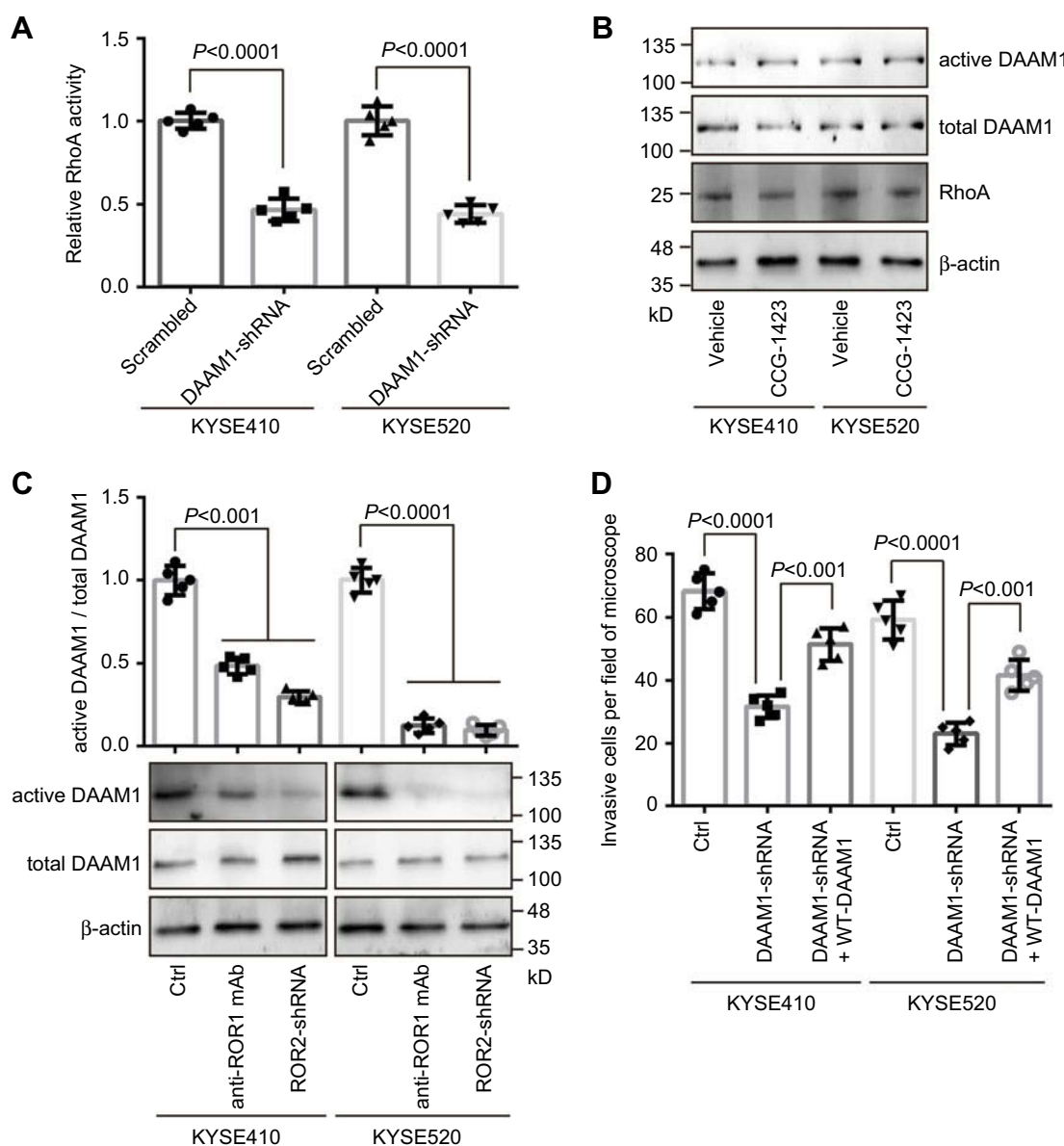
assay demonstrated that purified bacterially expressed recombinant ROR1 is directly binding to purified ROR2. Knockdown of ROR2 markedly inhibits the DAAM1/





**Figure 4** RhoA activation is regulated by ROR1/ROR2 and required for ESCC cell invasion. **(A)** KYSE410 cells were treated with anti-ROR1 mAb or vehicle and then measured the activation of Rac1, Rac2, Rac3, RhoA, and Cdc42 by G-LISA assays. **(B)** KYSE410 cells were transfected with ROR2-shRNA or scrambled shRNA and then measured the activation of Rac1, Rac2, and RhoA by G-LISA assays. **(C)** ROR2-knockdown KYSE410 and KYSE520 cells by shRNA transfection were treated with anti-ROR1 mAb, or rescued by transfected with shRNA-resistant WT-ROR2. The lysates of these cells were assayed for the evaluation of RhoA activation by G-LISA assays. **(D)** KYSE410 and KYSE520 cells were seeded on the Boyden chambers and treated with CCG-1423 (RhoA-specific inhibitor, 1  $\mu\text{mol/L}$ ) or vehicle for 1 hr, and then subjected to cell invasion assays.

**Abbreviations:** ESCC, esophageal squamous cell carcinoma; ROR, receptor tyrosine kinase-like orphan receptor.



**Figure 5** DAAM1 acts as the upstream of RhoA and mediates cell invasion. **(A)** KYSE410 and KYSE520 cells were transfected with DAAM1-shRNA or scrambled shRNA and then examined the RhoA activity by G-LISA assays. **(B)** DAAM1 activation was not altered by RhoA-specific inhibitor CCG-1423 treatment. KYSE410 and KYSE520 cells were treated with 1  $\mu$ mol/L CCG-1423 or vehicle for 1 hr. Cellular lysates were assayed for the active DAAM1 by a pull-down assay using a GST-RhoA as a bait. **(C)** DAAM1 acted as the downstream target of ROR1/ROR2. KYSE410 and KYSE520 cells were treated with anti-ROR1 mAb or transfected with ROR2-shRNA. Cellular lysates were assayed for the active DAAM1 by a pull-down assay using a GST-RhoA as a bait. **(D)** KYSE410 and KYSE520 cells were transfected with DAAM1-shRNA and rescued by shRNA-resistant wildtype (WT) DAAM1 overexpression, and then subjected to invasion assays.

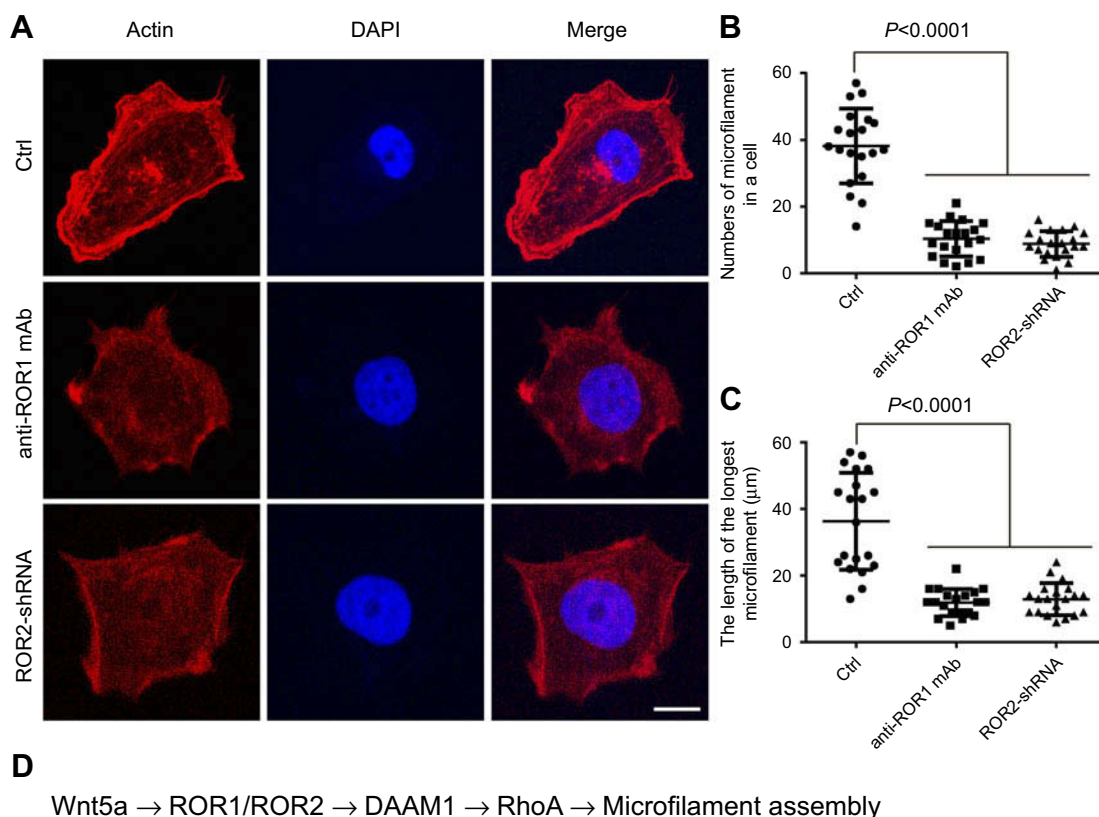
**Abbreviations:** ROR, receptor tyrosine kinase-like orphan receptor; WT, wild type.

RhoA activity, microfilament formation, and the invasion of ESCC cells. Moreover, anti-ROR1 mAb and ROR2 shRNA alone and combination of anti-ROR1 mAb with ROR2 shRNA play an equal effect on the invasion of ESCC cells, indicating ROR1/2 heterooligomers mediate Wnt5a-induced cell invasion.

DAAM1 is a formin protein, which links Wnt5a signal with Rho small GTPases and controls the rearrangement of microfilament.<sup>13,31</sup> Our results demonstrate that DAAM1

functions as the downstream target of Wnt5a and ROR1/2 signaling in ESCC cells. shRNA-mediating silence of DAAM1 inhibits RhoA activity and the invasion of ESCC cells. The decrease of RhoA activity and cell invasion in DAAM1-knockdown cells indicates that DAAM1 has the ability to interact with ROR1/2 in response to Wnt5a to activate RhoA and facilitate cell invasion in ESCC (Figure 6D).

CCG-1423, a small chemical component, is a Rho inhibitor involved in potentially blocking RhoA signaling through



**Figure 6** ROR1 and ROR2 participate in the rearrangement of microfilaments. **(A)** Blocking ROR1/ROR2 signaling by ROR2-shRNA transfection or anti-ROR1 mAb treatment disrupted the formation of microfilaments. KYSE410 cells treated with anti-ROR1 mAb or transfected with ROR2-shRNA were grown on coverslips. Subsequently, cells were fixed and stained with phalloidin and DAPI to reveal filamentous actin (F-actin) organization and nucleus location. Bar = 10  $\mu\text{m}$ . Objective lens, magnification,  $\times 40$ ; numerical aperture, 0.95. **(B and C)** The number of microfilaments in a cell and the length of the longest microfilament were determined in KYSE410 cells. Data was shown in the above scatter diagram.  $n=20$ . **(D)** The scheme of signaling pathway involving in the invasion of ESCC cells.

**Abbreviations:** ESCC, esophageal squamous cell carcinoma; ROR, receptor tyrosine kinase-like orphan receptor.

short-term treatment with CCG-1423 ( $<1$  mmol/L).<sup>32</sup> High dose of CCG-1423 inhibits Rho-dependent invasion of PC-3 prostate cancer cells.<sup>32</sup> Preincubation with CCG-1423 (1  $\mu\text{mol/L}$ ) completely inhibits the collagen-induced haptotaxis of breast MDA-MB-231 and MDA-MB-453 cancer cells.<sup>18</sup> CCG-1423 (0.022  $\mu\text{mol/L}$ ) blocks Wnt5a-induced glioblastoma U251 and T98MG cell invasion.<sup>16</sup> Here, we found that treatment with CCG-1423 (1  $\mu\text{mol/L}$ ) inhibited the cell invasion of ESCC KYSE410 and KYSE520 cells. Based on these literature, CCG-1423, a pharmacological inhibitor to prevent cancer cellular events via blocking RhoA signaling, has an extremely broad range of doses.

## Conclusion

This study clarified the connection between Wnt5a signaling and ESCC invasion. We demonstrated that Wnt5a mediates the invasion of ESCC cells, at least in part via ROR1/2 receptors and DAAM1/RhoA signaling pathway. Therefore,

Wnt5a/ROR1/2/DAAM1/RhoA signaling pathway may be the potential therapeutic targets for human esophageal cancer.

## Abbreviation list

ESCC, esophageal squamous cell carcinoma; ROR1, receptor tyrosine kinase-like orphan receptor 1; ROR2, receptor tyrosine kinase-like orphan receptor 2; DAAM1, disheveled-associated activator of morphogenesis 1; Sfrp2, secreted frizzled-related protein 2; mAb, monoclonal antibody.

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## Disclosure

The authors report no conflicts of interest in this work.

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