Prevalence and predictors of occult hepatitis C virus infection among Egyptian patients who achieved sustained virologic response to sofosbuvir/daclatasvir therapy: a multi-center study

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Background: Occult hepatitis C virus (HCV) infection (OCI) is characterized by the detection of HCV-RNA in non-serum reservoirs, such as peripheral blood mononuclear cells (PBMCs) and/or hepatocytes with undetectable HCV-RNA or antibodies in the serum. In this study, we tried to evaluate the prevalence and possible predictors of OCI in patients who achieved sustained virologic response (SVR) post sofosbuvir/daclatasvir (SOF/DCV) therapy.

Patients and methods: A cross-sectional multicenter study was designed to enroll 1,280 HCV-infected patients who received SOF (400 mg) plus DCV (60 mg) once daily ± ribavirin regimen for 12 weeks and achieved SVR 12 weeks post treatment. They were randomly recruited from three dedicated Egyptian centers for management of HCV. Real-time PCR was performed to detect HCV-RNA in serum and PBMCs and to evaluate the different risk factors pertaining to the existence of OCI.

Results: HCV-RNA was detected in PBMCs of 50 (3.9%) of them. All OCI cases exhibited significant fibrosis score and raised pre-treatment alanine aminotransferase (ALT) levels. Logistic regression analysis comparing OCI with non-OCI revealed that high pre-treatment viral load, raised ALT, advanced fibrosis score, prolonged prothrombin time, low albumin, Child B score, antiviral experienced patients, and raised bilirubin are the most significant predictor for the possibility of OCI presence with Odds Ratio as 7.03, 5.13, 4.4, 2.68, 2.52, 1.9, 1.5, and 1.2, respectively.

Conclusion: In spite of its remote possibility, OCI post SOF/DCV therapy may be present in some cases, and this may entail a re-auditing for the definition of SVR by dual testing in both serum and PBMCs.

Keywords: Occult, hepatitis C, Direct acting, Sofosbuvir

Introduction

The landscape of hepatitis C virus (HCV) management has evolved substantially since the emergence of interferon (INF)-based regimens and the subsequent all-oral treatment protocols with several waves of guidelines to tailor the best regimen.1-3 The optimal goal of all these guidelines is to achieve a sustained virologic response (SVR) which is defined as an undetectable HCV-RNA, using a sensitive assay (typically with a lower limit of 25 IU/mL), 12 weeks after completion of therapy (SVR12).1 The use of direct acting antiviral drugs (DAAs), especially the second generation with sofosbuvir (SOF)-based regimens, markedly improves the SVR rates.4 In Egypt, the real-life experience of...
using SOF plus daclatasvir (DCV), as adopted by the National Committee for the Control of Viral Hepatitis, proved to be a successful regimen with SVR rates approaching 95%.5,6

The concept of SVR was appraised with the emergence of a relatively new entity of detecting replicating viral genomic particles at extra-hepatic compartments, such as in peripheral blood mononuclear cells (PBMCs), hepatic tissue, or even in colonic mucosa.1,7–10 Therefore, the term occult C infection (OCI) was raised and represents the greatest challenge against the argument of SVR.

Occult C was considered when HCV-RNA was detectable in PBMCs, and/or liver tissue after repeated serum HCV-RNA–non-reactivity by sensitive assays (ranging between 52 and 1,000 vge or 10 and 615 IU/mL).2 Literature defined two different forms of OCI; the cryptogenic form in which patients have no anti-HCV antibodies, and the secondary form in which patients have anti-HCV antibodies and had cleared HCV infection either spontaneously or after therapy.11,12 The assessment of OCI among patients who received antiviral regimens was investigated in many clinical scenarios; such as in patient with spontaneously cleared virus,13 in hemodialysis regimens was investigated in many clinical scenarios; such as in patient with spontaneously cleared virus,13 in hemodialysis patients,14 in liver transplant settings,15 and also in patients with treatment-induced SVR, but data in this context were conflicting, with some studies finding genomic and antigenic HCV-RNA in PBMCs,1 while others did not.2 The presence of post-treatment viral particles in patients with SVR may carry a potential risk for disease relapse, disease progression, and/or a hidden re-infection foci that may affect the disease epidemiologic pattern and distribution.

Therefore, we herein, aimed to determine the prevalence of OCI in PBMCs among patients who achieved SVR12 after DAAAs and to determine the different predictors of OCI existence.

Patients and methods

Patients’ recruitment and study design

Between January 2017 and December 2017, a cross-sectional study was designed to enroll randomly selected patients in whom SVR12 was achieved at three dedicated Viral Hepatitis Management Centers in three different governorates; Assiut, Sohag, and Qena at Upper Egypt. Randomization was done through computer-based random allocation software that randomly selected cases from the pool within a specified time using patients’ ID number. All patients received a combination of SOF; 400 mg plus DCV; 60 mg once daily ± ribavirin (RBV) regimen for 12 weeks and achieved SVR12. Patients were either treatment naive or experienced to previous Interferon/RBV regimen. All patients had compensated liver cirrhosis as assessed by Fibroscan (EchoSens, Paris, France) prior to treatment.

All the following data were assessed at the pre-treatment phase in all patients: body mass index calculation, serum aspartate aminotransferase (IU/L), serum alanine aminotransferase (ALT, IU/L), serum albumin, serum bilirubin, serum alpha-fetoprotein, prothrombin time and concentration, international normalized ratio, blood urea, and serum creatinine. Also, liver stiffness score measurements were done by Fibroscan® (EchoSens) in kilopascals (kPa) with the following scaling: scores less than 7.4 kPa were equal to F0-F2, 9.5–12.4 kPa were equal to F3, and 14.5 kPa or greater were equal to F4 on META VIR pathologic scoring system.10,17

RNA isolation from plasma and PBMCs

Automatic extraction of viral RNA from thawed plasma was done according to the manufacturer’s protocol using the QIAamp1 Viral RNA Mini-Kit (cat#1048147, QIAGEN1, Qiagen, Hilden, Germany). Separation of PBMCs from blood was done using Ficoll Hypaque (Lonza, Verviers, Belgium) according to the manufacturer’s instructions then lysed by lysis buffer provided in the kits, then automated extraction of the total RNA from lysed PBMCs was performed according to the manufacturer’s protocol using QIAamp1 RNA Blood Mini-Kits (cat#52304, QIAGEN1, Qiagen).

Detection of HCV-RNA in plasma and PBMCs by quantitative real-time PCR

Detection of HCV viral load in the plasma and PBMCs was performed with the Artus1HCV-RG RT-PCR Kit (cat no. 4512865, QIAGEN1, Qiagen) by standardized quantitative real-time PCR according to the manufacturer’s protocol, and amplification was done by the ABI 7500 Fast Real-Time PCR Thermal cycler (Applied Biosystems, Foster City, CA, USA). Negative and positive controls were obtained from healthy volunteers and HCV-infected patients, respectively.

Data analysis and ethical considerations

 Frequencies, percentages, and means were used, as appropriate, for descriptive analysis. Univariate and multivariate logistic regression analyses were performed to assess the significant predictors of occult HCV persistence between groups. All statistical analyses was conducted by SPSS software for Windows, release 11(SPPS Inc., Chicago, IL, USA). A P-value<0.05 was considered significant.

The study protocol conformed to the ethical guidelines of the 1975 Declaration of Helsinki and was approved by
Results

Demographic and laboratory data of enrolled patients

A total of 1,280 patients (96.1% were males [n=1,100], mean age was 52.6±7.9 years) were recruited. Of them, 940 patients (76.4%) were treatment naïve and the remaining 26.5% were treatment experienced to INF/RBV. Cirrhosis was observed in 60% of patients (n=760). The basic demographic and laboratory data of patients are shown in Table 1.

Prevalence of OCI and characteristics

Among the 1,280 SVR patients, HCV-RNA was detected in PBMCs in 3.9% (n=50). Comparing positive and negative cases with OCI prior to therapy, there was a significant difference with regard to age (P=0.01), pre-treatment experience (P=0.001), pre-treatment prothrombin activity (P=0.01), pre-treatment serum bilirubin level (P=0.0001) and pre-treatment albumin level (P=0.001). However, we could not find any statistically significant difference with regard to gender, Fibroscan score pre-treatment viral load, and Child–Pugh score.

Post therapy (at SVR12), there was a statistical difference between two groups with regard to ALT levels (P=0.01), prothrombin activity (P=0.001) while, serum bilirubin and albumin levels did not show any statistical difference between two groups. The detailed comparison between positive and negative OCI cases is shown in Table 2.

Predictors of OCI among studied population

Logistic regression analysis for prediction of occult C revealed the following risk factors (OR): raised pre-treatment viral load (OR =7.03), raised ALT (OR =5.13), Fibroscan grade 4 (OR =4.4), abnormal prothrombin time 4–6 seconds (OR =2.68), low albumin (OR =2.52), Child B score (OR =1.9), previous treatment experience (OR =1.5), and raised bilirubin (OR =1.2) as shown in Table 3.

Discussion

The term OCI is now a challenging entity in the field of post hepatitis management and follow-up.11,18–21 It is described when HCV-RNA is detectable in liver and/or PBMCs in patients with undetectable serum HCV-RNA.22 On the level of molecular pathology, it is well known that exposure of the intracellular HCV-RNA strands to the stress of antiviral therapy is associated with disappearance of the antisense strand that leads to interruption of the virus life cycle.18–21 Concordant clearance of intracellular HCV-RNA strands and hence, the whole virus particles from serum are a promising sign of cure. On the contrary, the persistence of the intra-PBMCs strands and its reappearance or the presence of non-responding antisense strand to antiviral therapy is a predictor of non-responder to therapy or relapse.18–20 Therefore, OCI may be considered as a hidden source for HCV
reappearance or a remote source of re-infection in patients in whom SVR was assigned and now it is a debatable point of discussion, especially in localities with a high endemicity of HCV infection. From this respect, we tried to investigate the frequency of OCI in form of HCV persistence in the PBMCs in a relatively large well-defined cohort.

In the current study, OCI was detected in about 4% of patients, in whom, SVR12 was assigned. It was noted that all of them were experienced to prior IFN treatment failure. Conflicting results were found in literature. Abd Alla et al reported a high prevalence of OCI among treatment experienced and naive cases.23 Also, in the study of Khadr et al, secondary OCI was found in about 25% of cases treated with SOF/DCV.24 But, this study was criticized by the small sample size (40 patients). In the study of Radkowski et al they tested OCI after INF-based therapy and found positive OCI in PBMC in most of their patients and even on long-term follow-up post SVR.25 Also, Cavalheiro et al, emphasized the same concept in post-INF therapy.26 With the era of DAAs, re-evaluation of the existence of OCI post treatment was also detected in the study of Yousif et al, who tested OCI among different regimens of all-oral treatment and found a relatively higher prevalence of OCI (about 12%).27 On the other hand, others could not find any detectable HCV genomic RNA in their plasma or PBMCs; such as Maylin et al, study in which they tested 156 successfully treated immune-competent patients with INF-based therapy.28 Likewise, Bernardin et al tested HCV viremia in PBMCs of anti-HCV-positive/HCV-RNA-negative blood

Table 2 Univariate analysis of factors associated with the presence of OCI

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Negative OCI (n=1,230)</th>
<th>Positive OCI (n=50)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, mean ± SD</td>
<td>52.89±8.020</td>
<td>46.00±0</td>
<td>0.012a</td>
</tr>
<tr>
<td>Sex, n (%) male female</td>
<td>1,050 (85.4%)</td>
<td>50 (100%)</td>
<td>0.463a</td>
</tr>
<tr>
<td>Pre-treatment status, n (%)</td>
<td>Naive</td>
<td>Experienced (INF therapy)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>940 (76.4%)</td>
<td>290 (23.6%)</td>
<td>0.001a</td>
</tr>
<tr>
<td>Fibroscan score, n (%)</td>
<td>F1</td>
<td>F2</td>
<td>F3</td>
</tr>
<tr>
<td></td>
<td>90 (7.3%)</td>
<td>430 (35.0%)</td>
<td>260 (21.1%)</td>
</tr>
<tr>
<td></td>
<td>0 (0%)</td>
<td>0 (0%)</td>
<td>34 (68%)</td>
</tr>
<tr>
<td>Child–Pugh score, n (%)</td>
<td>None cirrhotic</td>
<td>A</td>
<td>B</td>
</tr>
<tr>
<td></td>
<td>520 (42.3%)</td>
<td>350 (28.5%)</td>
<td>360 (29.3%)</td>
</tr>
<tr>
<td></td>
<td>0 (0%)</td>
<td>30 (60%)</td>
<td>20 (40%)</td>
</tr>
<tr>
<td>Pre-treatment ALT, n (%)</td>
<td>Normal</td>
<td>Raised</td>
<td></td>
</tr>
<tr>
<td></td>
<td>350 (28.5%)</td>
<td>880 (71.5%)</td>
<td>11 (22%)</td>
</tr>
<tr>
<td></td>
<td>30 (2.4%)</td>
<td>1,200 (97.6%)</td>
<td>2 (4%)</td>
</tr>
<tr>
<td>Post-treatment ALT, n (%)</td>
<td>Normal</td>
<td>Not normalized</td>
<td></td>
</tr>
<tr>
<td></td>
<td>1,200 (97.6%)</td>
<td></td>
<td>42 (84%)</td>
</tr>
<tr>
<td></td>
<td>30 (2.4%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pre-treatment prothrombin time (seconds), n (%)</td>
<td>Normal</td>
<td>&lt;4 seconds</td>
<td>&gt;4 seconds</td>
</tr>
<tr>
<td></td>
<td>570 (46.3%)</td>
<td>300 (24.4%)</td>
<td>360 (29.3%)</td>
</tr>
<tr>
<td></td>
<td>3 (6%)</td>
<td>33 (66%)</td>
<td>14 (28%)</td>
</tr>
<tr>
<td>Post-treatment prothrombin time (seconds), n (%)</td>
<td>Normal</td>
<td>&lt;4 seconds</td>
<td>&gt;4 seconds</td>
</tr>
<tr>
<td></td>
<td>630 (51.2%)</td>
<td>300 (24.4%)</td>
<td>300 (24.3%)</td>
</tr>
<tr>
<td></td>
<td>0 (0%)</td>
<td>36 (72%)</td>
<td>14 (28%)</td>
</tr>
<tr>
<td>Pre-treatment bilirubin level (mg/dL), mean ± SD</td>
<td>1.35±0.6575</td>
<td>0.720±0.0837</td>
<td>0.001a</td>
</tr>
<tr>
<td>Post-treatment bilirubin level (mg/dL), mean ± SD</td>
<td>0.9±0.04</td>
<td>1±0.07</td>
<td>0.341</td>
</tr>
<tr>
<td>Pre-treatment albumin level (g/dL), mean ± SD</td>
<td>3.637±0.5009</td>
<td>4.280±0.1789</td>
<td>0.001a</td>
</tr>
<tr>
<td>Post-treatment albumin level (g/dL), mean ± SD</td>
<td>3.9±0.4</td>
<td>4±0.5</td>
<td>0.132</td>
</tr>
<tr>
<td>Pre-treatment viral load (log10 RNA by IU/mL), mean ± SD</td>
<td>5.4±2</td>
<td>4.9±1</td>
<td>0.24</td>
</tr>
</tbody>
</table>

Notes: *t-Test. *Chi-squared test.

Abbreviations: ALT, alanine aminotransferase; INF, interferon; OCI, occult C infection.
donors who had spontaneously or therapeutically cleared their plasma viremia found that the clearance of HCV from PBMC was relatively complete in both.29

In the current study, OCI was tested in the immediate phase of post SVR achievement (SVR12), which made the possibility of re-infection or a new infection somewhat impossible. In their interesting study, Ibarra et al documented the impaired cellular uptake of RBV into PBMCs with time. This may provide an explanation for the question why PBMCs could become a reservoir of HCV, especially with INF/RBV therapy and then the subsequent relapse, and treatment failure.30

Studies tackled this issue summarized the possible mechanism of OCI by the replication of HCV quasispecies in the PBMCs and then become as reservoir for virus.31

With regard to the clinical relevance of patients with positive OCI, all positive OCI cases were cirrhotic with a significant fibrosis score (F3 or F4) and also abnormal liver function tests, especially raised ALT prior to treatment. These findings were also reported in other studies of Rahman et al and Sood et al, which found a definite percentage of OCI that occurs more often in patients with overt cirrhosis.32,33 Elmasry et al examined the occurrence of OCI in HCV-infected liver transplant recipients who achieved SVR with recent DAAs and found an altered aminotransferase levels in 11% of their patients.34 However, their results may be influenced by the effect of immunosuppressive regimens as they tested OCI in post-liver transplant settings.

Our study proposed a strong recommendation of testing OCI in those with SVR as a routine part of post-treatment workup. Dual testing for both conventional PCR in the serum and PBMCs may be an essential endpoint for the precision of SVR. The need for a tailored protocol for retreatment of such patients with OCI may need further intense research.

Study limitations
This study has some limitations. First, the lack of sequential and serial follow-up by testing HCV-RNA in PBMCs post SVR24 in some patients. This weakness is considered and ongoing new study, by the same authors, is now on plan to test OCI post SVR24 in different treatment protocols and different patients’ inclusions. Second, the lack of simultaneous testing OCI in liver tissue may be considered as a weak point. However, performing liver biopsy is difficult both technically and ethically to be applied in such study design. The third weakness is the lack of genotype testing. Genotype testing was not mandated in the Egyptian protocol for treating HCV and was not considered as a prerequisite for enrolling patients. Moreover, most of studies testing HCV genotype documented genotype 4 as the main genotype in Egypt.35

Conclusion
In spite of its remote possibility, OCI post SOF/DCV therapy may be present in some cases, and this may entail a re-auditing for the definition of SVR by dual testing in both serum and PBMCs.

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Disclosure

The authors report no conflicts of interest in this work.

References


