Clinical significance of miR-195 in hepatocellular carcinoma and its biological function in tumor progression

Xiaoyan Chen1
Angang Wang2

1Department of Laboratory Medicine, Women and Children’s Hospital of Linyi City, Shandong 276000, People’s Republic of China; 2Department of Laboratory Medicine, People’s Hospital of Yutai County, Shandong 272300, People’s Republic of China

Introduction

Hepatocellular carcinoma (HCC) is one of the most common primary liver malignancy, with high mortality worldwide. The development of HCC is thought to be multifactorial, and seem to be closely related to chronic hepatitis B virus (HBV), hepatitis C virus infections.1–4 Most HCC patients are usually asymptomatic at an early stage, leading to initial diagnosis at an advanced stage with shorter overall survival.5,6 Surgical resection is the main treatment for HCC at present. Although multimodal treatment of HCC has made significant progress, the prognosis of HCC patients remains unsatisfactory due to high recurrence and early distant metastasis.7,8 Thus, special and effective cancer-related biomarkers for early diagnosis of HCC from healthy individuals, as well as from hepatitis patients are urgently required.

miRNAs are small endogenous noncoding RNAs of ~22 nucleotides in length, which regulate gene expression by binding to 3’UTR of target mRNA to initiate cleavage or translational repression of target transcripts.9,10 Numerous studies have...
shown that miRNAs played important roles in biological processes, such as cell differentiation, proliferation, migration, invasion, apoptosis, and tumor metastasis.11,12 miRNAs can be oncogenes or tumor suppressor genes in a majority of cancer during tumorigenesis, which are considered as novel biomarkers for tumor diagnosis and therapy. miR-195 has been reported to be downregulated in the tissues of HCC patients.13,14 However, the expression pattern of miR-195 in HCC cell lines and serum of HCC patients is still elusive.

In this study, we detected the expression of miR-195 in the serum of HCC patients, hepatitis only patients, and healthy individuals, as well as in cell lines. Its diagnostic value for cancer patients was also assessed. In addition, we investigated the effects of miR-195 on biological behaviors of HCC cells.

Patients and methods

Patients and specimens

From June 2012 to July 2017, a total of 120 HCC patients, 64 hepatitis only patients, and 118 healthy individuals at the People’s Hospital of Yutai County were recruited into this study. Blood samples were collected during the HCC diagnosis or physical examination. A total of 10 mL of whole blood was collected from each participant and the serum was separated from whole blood by centrifugation, and then stored at −80°C until RNA extraction. All the patients had not received any treatment before sampling. The clinicopathological information from all the HCC patients was collected and is summarized in Table 1. The protocols of this study were approved by the Ethics Committee of the People’s Hospital of Yutai County and adhered to the tenets of the Declaration of Helsinki. All the participants signed the written informed consent.

Cell lines and transfection

Human normal liver epithelial cell (THLE3) and HCC cell lines (HepG2 and SK-Hep1) were purchased from the American Type Culture Collection (ATCC), Manassas, VA, USA. HCC cell lines (Huh7 and PLC) were purchased from China Center for Type Culture Collection (CCTCC, Hubei, China). All cells were cultured in DMEM (Thermo Fisher Scientific, Waltham, MA, USA) with 10% FBS at 37°C in a humid incubator with 5% CO2. HCC cells were seeded into 6-well plates at a density of 2×104 and incubated 24 hours, and then transfected with miR-195 mimic, inhibitor, or their negative control (NC) using Lipofectamine 2000 reagent (Thermo Fisher Scientific) following the instructions of the manufacturer. Untreated cells were used as blank control.

Table 1 The relationship between miR-195 expression and clinicopathological parameters of HCC patients

<table>
<thead>
<tr>
<th>Parameters</th>
<th>No. of cases (n=120)</th>
<th>miR-195 expression</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>98</td>
<td>Low (n=67)</td>
<td>45</td>
</tr>
<tr>
<td>Female</td>
<td>22</td>
<td>High (n=53)</td>
<td>8</td>
</tr>
<tr>
<td>Age (years)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>≥55</td>
<td>59</td>
<td></td>
<td>24</td>
</tr>
<tr>
<td>&lt;55</td>
<td>61</td>
<td></td>
<td>29</td>
</tr>
<tr>
<td>Tumor size (cm)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>≥5</td>
<td>72</td>
<td></td>
<td>29</td>
</tr>
<tr>
<td>&lt;5</td>
<td>48</td>
<td></td>
<td>24</td>
</tr>
<tr>
<td>Hepatitis history</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>64</td>
<td></td>
<td>28</td>
</tr>
<tr>
<td>No</td>
<td>56</td>
<td></td>
<td>25</td>
</tr>
<tr>
<td>AFP (ng/mL)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>≥20</td>
<td>58</td>
<td></td>
<td>22</td>
</tr>
<tr>
<td>&lt;20</td>
<td>62</td>
<td></td>
<td>31</td>
</tr>
<tr>
<td>Lymph node metastasis</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>51</td>
<td></td>
<td>17</td>
</tr>
<tr>
<td>Negative</td>
<td>69</td>
<td></td>
<td>36</td>
</tr>
<tr>
<td>Differentiation</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Poor</td>
<td>63</td>
<td></td>
<td>24</td>
</tr>
<tr>
<td>Moderate + well</td>
<td>57</td>
<td></td>
<td>29</td>
</tr>
<tr>
<td>TNM stage</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>I–II</td>
<td>48</td>
<td></td>
<td>27</td>
</tr>
<tr>
<td>III–IV</td>
<td>72</td>
<td></td>
<td>26</td>
</tr>
</tbody>
</table>

Abbreviations: AFP, α-fetoprotein; HCC, hepatocellular carcinoma.

RNA isolation and quantitative real-time (qRT)-PCR analysis

Total RNA was isolated from serum and cell lines by using Trizol reagent (Thermo Fisher Scientific) according to the manufacturer’s instruction. Reverse transcription was performed using miScript (SYBR™) Green PCR kit (Qiagen NV, Venlo, the Netherlands) according to the manufacturer’s protocol. Total RNA concentration was assessed by measuring absorbance at 260 and 280 nm (A 260/280 ratio) and checked by gel electrophoresis individually. In general, 800 ng of RNA from 1 mL of serum was used for further use. The relative expression levels of miR-195 were detected by qRT-PCR, which was performed with the SYBR™ Green PCR master mix (Thermo Fisher Scientific) on an ABI 7900 real-time PCR system (Thermo Fisher Scientific). The relative quantification of miR-195 expression was calculated with the 2−ΔΔCt methods normalized to miR-16.

Cell proliferation assay

The viability of HCC cells was measured with the colorimetric MTT assay. Cells were plated in 96-well plates (1×104 cells per well) and transfected with miR-195 mimic,
inhibitor, or their NC. After transfection, 48 hours later, the medium was added with 20 µL MTT solution (5 mg/mL; Sigma-Aldrich Co., St Louis, MO, USA) every 24 hours and incubated for 4 hours at 37°C. The supernatant was discarded, and 100 µL dimethyl sulfoxide (Sigma-Aldrich Co.) was added to dissolve formazan crystals. Cell proliferation was determined using the colorimetric analysis (wavelength, 490 nm) by a spectrophotometer (Multiskan MK3; Thermo Fisher Scientific) at 24, 48, 72, and 96 hours. Each treatment group was measured at least three times.

Cell migration and invasion assays

Cell migration and invasion assays were assessed with 24-well Transwell chamber (8-µm pore size, Multiskan MK3; Thermo Fisher Scientific). For invasion assay, the top chamber was coated with Matrigel (BD biosciences, Bedford, MA, USA) on the upper side. Hundred microliter cell suspension with 1x10⁶ cells were plated in the top chamber with serum-free DMEM at 37°C for 24 hours. The bottom chamber was filled with 600 µL DMEM containing 20% FBS. Then non-invading cells on the upper surface were removed using cotton-tipped swabs, while the cells that invaded the lower side of the membrane were fixed in 3.7% formaldehyde for 5 minutes and stained with 0.1% crystal violet for 15 minutes. The cells were counted using a microscope (Olympus Corporation, Tokyo, Japan). For migration assay, the same procedure was followed as with invasion assay except that no Matrigel was used and the permeating time was 12 hours.

Statistical analysis

All statistical analyses were performed using the SPSS 21.0 software (IBM Corporation, Armonk, NY, USA) and GraphPad Prism 5.0 software (GraphPad Software, Inc., La Jolla, CA, USA). Data are presented as the mean ± SD. The significance between groups was analyzed by Student’s t-test or one-way ANOVA. The relationship between the expression of miR-195 and clinicopathological features was analyzed by the chi-square (χ²) test. The receiver operating characteristic (ROC) curves were generated to evaluate the diagnostic potential of serum miR-195 by calculating the area under the ROC curve (AUC). Each experiment was repeated at least three times. A P-value <0.05 was considered to be of statistical significance.

Results

Expression of miR-195 in the serum of HCC patients and HCC cell lines

A previous study by Sohn et al have shown that the serum levels of miR-195 were lower in HCC patients than in patients with chronic HBV. To further investigate the roles of miR-195 in the development of HCC, we detected the expression levels of miR-195 in the serum samples of HCC patients (64 HCC patients with hepatitis and 56 HCC patients without hepatitis), hepatitis only patients, and healthy control, as well as in the cell lines, using qRT-PCR. As shown in Figure 1A, the relative serum expression of miR-195 was significantly downregulated in HCC patients compared to healthy control (2.430±0.83) (P<0.001). There is no significant difference between healthy control and hepatitis only patients (2.341±0.75) (P>0.05). Compared to healthy control, the relative expression levels of miR-195 were significantly lower in HCC patients without hepatitis (1.371±0.48) (Figure 1B, P<0.001). The result of Figure 1C showed that the relative expression of miR-195 was significantly lower in HCC patients with hepatitis (1.340±0.45) than in hepatitis only patients (P<0.001). These results suggest miR-195 may have diagnostic value in distinguishing HCC patients from healthy individuals, as well as from hepatitis patients. Meanwhile, compared to the normal liver epithelial cell THLE3, the expression levels of miR-195 were decreased in HCC cell lines (all P<0.001, Figure 1D).

The relationship between miR-195 expression and clinicopathological characters of HCC patients

To investigate whether miR-195 expression was involved in the development of HCC, we analyzed the relationship between its expression and patients’ clinicopathological features. We divided the HCC patients into a low-expression group (n=67) and high-expression group (n=53) according to the mean value of miR-195 expression level (1.355). As shown in Table 1, the expression of miR-195 was significantly associated with lymph node metastasis (P=0.040) and TNM stage (P=0.030). However, there was no significant relationship with other parameters, such as gender, age, tumor size, hepatitis history, α-fetoprotein (AFP), and differentiation (all P>0.05).

Diagnostic value of miR-195 for HCC

We plotted the ROC curve to assess the potential usefulness of serum miR-195 as a noninvasive biomarker for the early diagnosis of HCC from healthy individuals, as well as from hepatitis patients. As shown in Figure 2A, the AUC value was 0.876 (95% CI =0.817–0.907). Using the optimum cutoff value of 1.685 according to the Youden index, the sensitivity and specificity were 76.7% and 77.0%, respectively. The miR-195 was effective in differentiating HCC patients from healthy individuals with high specificity and


sensitivity (Figure 2A). The AUC value was 0.859 (95% CI = 0.805–0.913) in Figure 2B, which suggests miR-195 can also distinguish HCC patient without hepatitis from healthy individuals yielding the sensitivity of 71.4% and specificity of 74.7% at the optimum cutoff value of 1.665. Results in Figure 2C showed that miR-195 expression had a higher accuracy in differentiating HCC patients from hepatitis patients based on AUC of 0.868 (95% CI = 0.806–0.930).

**Figure 1** Expression of miR-195 was determined by qRT-PCR, and the miR-16 was used as an internal control.

**Notes:** (A) The serum expression of miR-195 was downregulated in HCC patients compared to healthy control (***P < 0.001). There is no significant difference between healthy control and hepatitis only patients (ns P > 0.05). (B) The serum expression of miR-195 was lower in HCC patients without hepatitis than healthy control (**P < 0.01). (C) The serum expression of miR-195 was lower in HCC patients with hepatitis than hepatitis patients (***P < 0.001). (D) The relative expression of miR-195 was decreased in HCC cell lines compared to the normal liver epithelial cell (**P < 0.01).

**Abbreviations:** HCC, hepatocellular carcinoma; qRT, quantitative real-time; ns, no statistical significance.

**Figure 2** Diagnostic value of miR-195 for HCC by establishing a ROC curve.

**Notes:** (A) ROC curve of the diagnostic value of miR-195 in differentiating HCC patients from healthy individuals. The AUC is 0.862, with a sensitivity of 76.7% and a specificity of 77.0% at the optimum cutoff value of 1.685. (B) ROC curve of the diagnostic value of miR-195 in distinguishing HCC patients without hepatitis from healthy individuals. The AUC is 0.859, with a sensitivity of 71.4% and a specificity of 74.7% at the optimum cutoff value of 1.450. (C) ROC curve of the diagnostic value of miR-195 in distinguishing HCC patients with hepatitis from hepatitis patients based on AUC of 0.868. The sensitivity and specificity were 89.1% and 71.9%, respectively, at the optimum cutoff value of 1.960.

**Abbreviations:** AUC, area under the ROC curve; HCC, hepatocellular carcinoma; ROC, receiver operating characteristic.
In addition, the sensitivity and specificity for serum miR-195 were 89.1% and 71.9%, respectively, at the cutoff point of 1.960. Taken together, miR-195 can differentiate HCC patients, healthy individuals, and hepatitis patients.

Effects of miR-195 on HCC cell proliferation, migration, and invasion

To investigate the functional role of miR-195 in HCC, the effects of miR-195 on HCC cell proliferation was performed in vitro. We transfected miR-195 mimic or miR-195 inhibitor, or their NC into HepG2 and Huh7 cells, which exhibit a low miR-195 expression. Then, we analyzed miR-195 expression levels after transfection by qRT-PCR and found that the expression of miR-195 was significantly increased in the cells transfected with miR-195 mimic but decreased in those transfected with miR-195 inhibitor, compared to the controls (all \( P < 0.01 \), Figure 3A). To examine the role of miR-195 in the proliferation of HCC cells, MTT assay was performed. The results showed that cell proliferation of HepG2 and Huh7 cells transfected with miR-195 mimic was significantly lower but in those transfected with miR-195 inhibitor was higher, compared to their controls (\( P < 0.05 \), Figure 3B). In addition, Transwell assays were employed to evaluate the impact of miR-195 on migration and invasion. The results showed that miR-195 mimic inhibited the migration and invasion, but miR-195 inhibitor promoted the migration and invasion, compared to their controls (all \( P < 0.05 \), Figure 4). Taken together, these results demonstrate that miR-195 inhibits HCC cell proliferation, migration, and invasion.

Discussion

The detection of HCC at an early stage have a relatively longer survival rate, which can significantly impact curative treatment. Currently, the diagnosis of HCC is mainly based on assessing serum AFP levels, computed tomography detection, and magnetic resonance imaging. However, a lower rate of early diagnosis and lack of effective biological indicators for monitoring tumor occurrence may be the

![Figure 3](https://example.com/figure3.png)

**Figure 3** Effects of miR-195 on cell proliferation in HepG2 and Huh7 cells.  
**Notes:** The cells were transfected with miR-195 mimic, inhibitor, or their NCs. (A) The expression of miR-195 was significantly increased in cells transfected with miR-195 mimic but decreased in cells transfected with miR-195 inhibitor. (B) The proliferation ability was significantly reduced by miR-195 mimic but was promoted by the miR-195 inhibitor (*\( P < 0.05 \), and ***\( P < 0.001 \)).  
**Abbreviations:** NC, negative control; OD, optical density.
Current major problem for poor prognosis of HCC. Thus, the noninvasive sensitive markers for early HCC detection are needed.

miRNAs dysregulation has been observed in cancer research, which serves as potential biomarkers of diagnosis and prognosis as well as therapeutic targets for cancers.\textsuperscript{19-23} Numerous studies have proven that miRNAs play a regulatory effect on the wide aspects of biological processes, such as cell proliferation, migration, and invasion.\textsuperscript{24,25} In HCC, some miRNAs have also been identified as dysregulated in tissues or serum samples.\textsuperscript{26-28} For instance, Suehiro et al analyzed serum exosomal miR-122 and miR-21 expression levels in HCC patients that underwent transarterial chemoembolization (TACE), demonstrating that the exosomal miR-122 level alterations may represent a predictive biomarker in HCC patients with liver cirrhosis treated with TACE.\textsuperscript{28} A study by Yoon et al found that miR-21 expression was significantly increased in HCC tissues compared to nontumor tissues, and pointed that elevated miR-21 expression might represent a biomarker for HCC prognosis.\textsuperscript{29} Overexpression of miR-26b was found by Li et al inhibited the proliferation, invasion, and migration of HCC cells by targeting EphA2.\textsuperscript{30} Taken together, these studies demonstrated that the cancer-related miRNAs and corresponding direct target genes are essential for HCC processes.

In this study, by using qRT-PCR, we detected the expression of miR-195 in the serum of HCC patients, hepatitis patients, and healthy control, as well as in HCC cell lines. The results showed that miR-195 expression was significantly decreased in the serum of HCC patients compared to healthy controls. The serum expression levels of miR-195 were lower in HCC patients with hepatitis than the hepatitis patients, and lower in HCC patients without hepatitis than healthy individuals. Similarly, the expression of miR-195 was downregulated in HCC cell lines compared to healthy control.
HCC cell lines compared to the normal liver epithelial cell (THLE3). The analysis results of the correlation between miR-195 expression and clinicopathological characteristics showed that the miR-195 expression was significantly associated with lymph node metastasis and TNM stage. However, no significant relationship was found with other parameters, such as gender, age, tumor size, hepatitis history, AFP, and differentiation. The results showed miR-195 expression has no relationship with hepatitis history. Taken together, the results indicated that miR-195 may act as a suppressor gene and involved in the development of HCC. Then by using ROC curve analysis, we further analyzed the potential diagnostic value of miR-195 in HCC. The higher AUC results and promising sensitivity and specificity showed miR-195 had a relatively high accuracy in differentiating HCC patients from healthy individuals, as well as in differentiating HCC patients from hepatitis patients, which indicated miR-195 may be a diagnostic biomarker for HCC.

In addition to HCC, the role of miR-195 was studied in other malignancies, such as gastric cancer, breast cancer, colon cancer, and cholangiocarcinoma (CCA).31–34 Li et al indicated that miR-195 expression was downregulated in colon cancer tissues and associated with the enhanced proliferation, migration, and invasion of colon cancer, which is a potential diagnostic marker and therapeutic target for improving the clinical management of colon cancer.35 The study in CCA by Chen et al revealed that miR-195 expression was significantly lower in CCA patients than in normal control, and it was reference prognosis marker of CCA.36 These previous studies showed that miR-195 plays a crucial role in the progression of cancers. Previous study by Wang et al verified that miR-195 is a vital regulator in the lung metastasis of HCC by using a lung metastasis HCC cell line BEL-7402.13 The results of Yan et al’s study showed the expression of miR-195 is downregulated in HCC tissues and cell lines, and miR-195 inhibits cell proliferation via targeting AEG-1 in HCC.14 These studies suggested that miR-195 is associated with progression of HCC. To verify the functional role of miR-195 in HCC, the effects of miR-195 on HCC cell proliferation, migration, and invasion were explored by regulating miR-195 expression using miR-195 mimic or inhibitor. The results indicated that, compared to controls, miR-195 mimic obviously inhibited the cell proliferation, migration, and invasion, conversely, miR-195 inhibitor promoted biological behaviors in both HepG2 and Huh7 cells, which confirmed the functional role of miR-195 in HCC. In a previous study, Li et al identified LncRNA UCA1 enhanced mitochondrial function and cell viability through the miR-195/ARL2 signaling pathway in bladder cancer, and provided UCA1 as a potential diagnosis and treatment target for bladder cancer.37 However, the potential molecular mechanism of miR-195 in the processes of HCC remain unconfirmed in our present study. Further studies are needed to confirm the role of miR-195 in HCC progression.

Conclusion
Our data in this study revealed that the downregulated expression of miR-195 may serve as a potential noninvasive diagnostic biomarker in differentiating HCC patients from healthy individuals, as well as from hepatitis patients. miR-195 is associated with the development of HCC and may be involved in the tumor progression of HCC, and has the potential to be used in the targeted therapy for HCC patients.

Disclosure
The authors report no conflicts of interest in this work.

References


