

# Recent development in the strategies projected for chikungunya vaccine in humans

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**Abstract:** The unprecedented epidemic spread of chikungunya worldwide illustrates the critical need for potent vaccines and therapeutic interventions. The morbidity and mortality associated with this arboviral infection has become a major public health problem in many countries across different continents. Increasing public-private partnerships have opened new avenues in research and development of vaccines. This review mainly focuses on the recent advances in patented approaches for chikungunya vaccine development and the forthcoming challenges.

**Keywords:** chikungunya, epidemics, CHIKV vaccine candidates, clinical trials, humans

## Introduction

Chikungunya is an emerging arthritogenic arboviral illness caused by chikungunya virus (CHIKV), a member of the genus Alphavirus in the *Togaviridae* family. CHIKV is an enveloped, positive-sense, single-stranded RNA with 11.8 kb long linear genome with two open reading frames encoding four nonstructural replicase proteins (nsP1, nsP2, nsP3, and nsP4) and five structural proteins (three glycosylated envelope glycoproteins E1, E2, and E3, nonglycosylated nucleocapsid and cleavage product 6 k). CHIKV was first isolated during an epidemic in Tanganyika in 1953. In fact, the name “chikungunya” pronounced as “CHI-kən-GUUN-yə” is derived from the Kimakonde language of the Makonde tribe, which means “the bent walker/stooped walking” describing the contorted posture of a patient suffering with severe joint pain and other arthritic symptoms caused by this disease.<sup>1,2</sup> The virus has evolved into three distinct geographical lineages with distinct genotypes – East/Central/South African (ECSA), Asian, and West African.<sup>3</sup> Explosive outbreaks during the last decade worldwide have prompted National Institute of Allergy and Infectious Diseases (NIAID) to add CHIKV in the category C priority pathogen biodefense list (<https://www.niaid.nih.gov/research/emerging-infectious-diseases-pathogens>).

The virus transmission takes place by the arthropod vectors, female *Aedes aegyptica* or *Aedes albopictus* mosquitoes; aggressive daytime feeders that also transmit dengue, zika and yellow fever.<sup>2</sup> The routine sylvatic or nonhuman cycle involves other primates like apes and lemurs but during epidemics, chikungunya transmission can be continued by interhuman spread.<sup>4</sup> *A. albopictus* – the Asian tiger mosquito – is a more cold-tolerant mosquito and adaptation of a single amino-acid genetic mutation in the E1 glycoprotein of CHIKV strains resulted in the epidemiologic shift of chikungunya from tropical to temperate regions of the world.<sup>5</sup>

## Search strategy

This review is based on all types of studies related to CHIKV vaccine development in humans and discussed the various vaccine approaches in detail. The articles

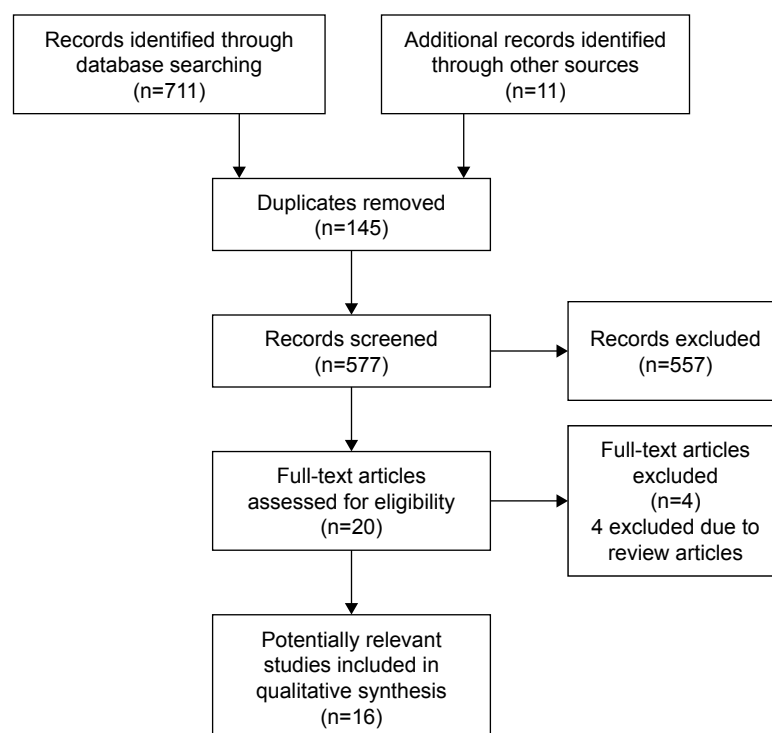
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were retrieved by PubMed, OVID, EMBASE, Google search, and clinical trials registries for relevant studies using search strategy (Search (((((((chikungunya) OR chikungunya) OR ((Chikungunya Fever) OR Chikungunya virus))) AND (((((((inactivated vaccine) OR live attenuated vaccine) OR virus like particle vaccine) OR recombinant vaccine) OR chikungunya virus vaccine)) OR (((chikungunya virus vaccine) OR vaccine) OR conjugate vaccine) OR virus like protein))) AND (((((((clinical trials) OR random) OR randomly) OR phase 1) OR phase 2) OR phase 3) OR randomised controlled trial) OR randomized))) NOT ((animals) NOT humans)). The search included publications that are recent till August, 2018. We have searched for studies to be included irrespective of any language or publication status. We have also manually searched the references and bibliographies of all the included studies and related review articles for any additional studies. Figure 1 shows the flowchart for screening and study selection process.

## Epidemiology

The dissemination of the CHIKV is geographically distributed in tropical and subtropical regions of East Africa, Southeast Asia, and Western Pacific. The persistent increase in unchecked vector population, immunologically naïve population, climatic changes, urbanization, and ease of global travel has elevated the risk for geographical expansion

of CHIKV to nonendemic regions. The first outbreak of chikungunya was documented in late 1950s to early 1960s in Bangkok and from 1963 to 1973 in India, affecting thousands of people. The next two major outbreaks were reported in coastal regions of Kenya in 2004 and consequently spread to numerous islands of Indian Ocean over a period of next 4 years. The largest outbreak occurred in islands of La-Reunion, in the western part of Indian Ocean in 2005 in which 266,000 cases and 254 deaths were reported by French public health authorities.<sup>6</sup> Several other countries including Mauritius, Seychelles, Madagascar, Gabon, Comoros and Mayotte, India, Sri Lanka, Indonesia, Thailand, Senegal, Philippines, Laos, Guinea, Taiwan, Vietnam, Myanmar, Singapore, Malaysia, Cambodia, and Pakistan also reported chikungunya cases. Massive chikungunya outbreaks were documented in India during 2006–2007, affecting >1.5 million people.<sup>7,8</sup> The disease then spread rapidly toward Pacific islands causing significant outbreaks in the Caribbean Islands in 2011.<sup>9</sup> Post the upsurge of Indian Ocean outbreak, the disease spread to a few places in Europe also. The CHIKV was later introduced in Americas in late 2013. The number of chikungunya cases spiraled during the devastating 2014–2015 epidemic in several North, Central, and South American countries resulting in millions of reported cases.<sup>10,11</sup> This was one of the most prominently reported outbreaks of CHIKV in Americas. The series of



**Figure 1** Prisma flowchart for the selection of studies.

chikungunya epidemics globally has shown dramatic increase in incidence rate of CHIKV over the last decade. The virus has been reported in more than 60 countries from various WHO regions with approximately one million cases and fatality rate of 0.1% per year globally.<sup>12–14</sup>

## Clinical presentation

The pathologic mechanism of this virus leads to acute infection of several weeks, which can persist for months with incubation period of 2–12 days.<sup>15</sup> CHIKV affects all age groups with dengue-like symptoms characterized by nausea, fever, chills/rigors, headache, polyarthralgia, myalgia, photophobia, and skin rash.<sup>8,16</sup> The hallmark characteristics of CHIKV infection are saddle-back fever, polyarthralgia (mainly in small joints – ankles, toes, wrists, and phalanges), and a petechial or maculopapular rash (mainly on face, limbs, and trunk).<sup>17,18</sup> Strikingly, recovery from an acute arthritic illness (high viral load  $\sim 10^8$ – $10^9$  viral copies/mL) takes few days to couple of weeks, though convalescence can be prolonged. In some chronic cases, the clinical manifestations may persist for several months or even years disquieting quality of life. The chronic symptoms mainly include recurrent low-grade body ache, fatigue, neuritis, and persistent swelling and incapacitating inflammation of the joints.<sup>19</sup> The major factors for CHIKV persistence may include genetic susceptibility, reinfection, exaggerated proinflammatory response leading to tissue damage, evasion of host defense machinery, activation of cryptic virus molecules, and autoimmune reactivity with viral and host antigens. Serious complications are uncommon. However, in rare cases CHIKV can be fatal, primarily in the elderly (>65 years), pregnant women, neonates exposed intrapartum (vertical transmission), young children (especially under 1 year), and persons with preexisting medical conditions like hypertension, diabetes, and heart disorders. Children atypically suffer from severe manifestations of CHIKV disease with neurologic complications like seizures, impaired consciousness, blindness due to retro-bulbar neuritis, and acute flaccid paralysis.<sup>20</sup>

## Immunologic basis of CHIKV vaccine development

CHIKV infection confers life-long immunity.<sup>21</sup> The activation of innate immune system, in particular, Type I interferons, is fundamental to clearance of acute CHIKV infections. Besides, numerous other innate immune cells like macrophages, natural killer cells, and dendritic cells also play pivotal role as robust responders to early CHIKV infections. However, recently, the protective and pathologic

attributes of the innate immunity have been implicated in the pathogenesis of CHIKV infections.<sup>22</sup> The components of adaptive immunity – both humoral and cellular immunity – also imply a crucial role in the protection and elimination of CHIKV-induced infections. However, the humoral immune response (B cell response) plays decisive role in protection against CHIKV infection compared to cellular immunity in human and mouse models.<sup>23</sup> There are neutralizing antibodies produced through humoral immune mechanism against glycoproteins on the outer surfaces of CHIKV envelope that mediate the clearing of viremia.<sup>24</sup> Generally, IgM is detectable between days 2 and 8 of illness and is normally present for only up to 3 months, while convalescent IgG is measurable from day 4 and persists for years. Recently, Chua et al described that anti-CHIKV IgM targets E1–E2 surface glycoproteins simultaneously rather than E1 or E2 individually, and thus helps in designing more sensitive and reliable IgM assay using an optimized native antigen.<sup>25</sup> Additionally, CD8<sup>+</sup> T cells in cell-mediated immunity play an important role in development of robust protective immunity in controlling acute infections (early phase), whereas CD4<sup>+</sup> T cells mediate immune response during chronic phase.<sup>26</sup> However, there is an urgent need for future research to understand the mechanism of immunopathogenesis of adaptive immunity during the sequel of chronic CHIKV infection in humans and vaccine development.

## Recent developments in CHIKV vaccines

The inadequate traditional mosquito control measures, dearth of herd immunity, rapid globalization, sporadic and episodic outbreaks worldwide, and high disease-associated morbidity emphasize the need for research and development of effective vaccines for this neglected tropical disease – “Chikungunya”. Vaccines are one of the most important medical contributions and cost-effective public health interventions of the 20th century to prevent communicable diseases. Unlike dengue, the development of an ideal CHIKV vaccine is pragmatic owing to the low antigenic variability of the virus. Despite vigorous efforts and support by researchers, vaccine producers, policy makers, and funding agencies across the globe, no vaccine has been licensed to combat CHIKV infections hitherto. Advances in the field of vaccinology in 21st century have led to the development of >20 CHIKV vaccine candidates using diverse vaccine strategies. These include live-attenuated vaccines, whole-virus inactivated vaccines, recombinant virus-vectored vaccines, inactivated virus-like particles (VLPs), DNA vaccines, mRNA-based vaccines, synthetic vaccines, and subunit formulations of CHIKV.<sup>27–29</sup> These CHIKV

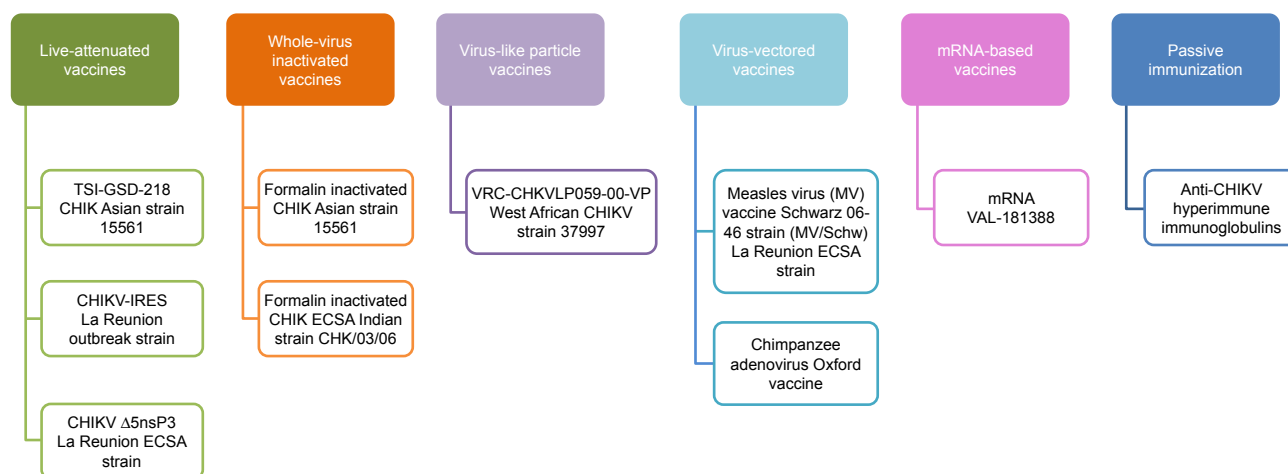
vaccine candidates exhibit wide array of advantages as well as disadvantages in manufacturing, dosage, safety, efficacy, scalable production, and cost effectiveness during preclinical and clinical phases of development. Recently, chikungunya has also been added in the revised list of WHO Research and Development Blueprint preparedness for priority diseases/pathogens most likely to cause epidemics in the annual review meeting held at Geneva, Switzerland, in January 2017.<sup>30</sup>

In this review, the major breakthroughs in the development of CHIKV vaccines that have entered human clinical trial testing have been discussed in detail. Figure 2 illustrates the various strategies projected for CHIKV vaccine development in humans.

## Live-attenuated CHIKV vaccines

The early attempts to develop CHIKV vaccine began in 1962 with the isolation of a CHIKV strain 15,561 from a viremic patient in Thailand. The wild-type strain was serially passaged by investigators in African green monkey kidney cells at the Walter Reed Army Institute of Research (WRAIR) and in human MRC-5 cells by the US Army Medical Research Institute of Infectious Diseases (USAMRIID) and attenuated strain CHIK 181/Clone 25 was selected as the “vaccine seed”. Chikungunya vaccine was manufactured at The Salk Institute-Government Services Division (TSI-GSD) in 1984 in collaboration with the US Department of Defense (DoD).<sup>31,32</sup> The attenuated TSI-GSD-218 revealed promising results in Phase I and Phase II trials conducted at USAMRIID and the University of Maryland Center for Vaccine Development, respectively.<sup>33,34</sup> No significant difference in local and systemic reactogenicity parameters between alphavirus-naïve

volunteers (n=15) and placebo groups was observed in Phase I study. In Phase II randomized, double-blind trial, 73 volunteers (age 18–40 years) were recruited, of which 59 volunteers were immunized with single subcutaneous dose of vaccine and 14 placebos received MRC-5 tissue culture fluid. Forty out of 58 serologically evaluable volunteers (69%) were seroconverted within 14 days, 57 (98%) were seroconverted ( $\geq 1:20$  by PRNT50) by 28 days, and 85% at 1 year. Five of 59 (8%) vaccinees developed transient mild arthralgia and none was reported by the placebo group (n=14). However, the number and severity of local and systemic reactions were similar in both the groups. Despite safe and highly immunogenic nature in clinical trials, TSI-GSD-218 development was terminated long before origination of deadly outbreaks in 2005 due to lack of funds, uncertainty about safety of bulk production, ambiguity about its misuse as a biologic warfare threat, and limited market for vaccine supply. However, the resurgence of chikungunya in islands of Caribbean and Indian Ocean raised the urgent need of technology transfers of this potential US military CHIKV vaccine. Five companies across the globe signed the material transfer agreement, of which only one pharmaceutical company has successfully developed the vaccine, which shows promising results in preclinical studies, and is ready for Phase I clinical trial.<sup>32</sup> Strikingly, a recent study by Gorchakov et al has raised the concern of safety issues on the use of attenuated 181/clone 25 CHIKV vaccine strain.<sup>35</sup> Attenuation of strain 181/clone 25 is attained by only two individual point mutations in structural E2 gene and reversions to wild-type sequences may result in phenotypic instability, accounting for mild reactogenic symptoms (transient arthralgia in 8% of



**Figure 2** Various strategies projected for chikungunya vaccine development in humans.

**Abbreviations:** CHIKV-IRES, chikungunya virus-internal ribosomal entry site; ECSA, East/Central/South African; TSI-GSD, The Salk Institute-Government Services Division.

alphavirus-naïve volunteers), thus emphasizing the need for improvement of 181/clone 25 CHIKV vaccine.

Another novel live-attenuated CHIK/internal ribosomal entry site (IRES) vaccine candidate derived from La Reunion outbreak strain has been developed based on engineering two IRES elements in the CHIKV genome in order to overcome reversion issues of previous vaccine. This vaccine has shown very promising safety and efficacy outcomes in multiple pre-clinical models.<sup>36–39</sup> Takeda has recently decided to project this potential CHIK/IRES vaccine candidate toward clinical evaluation.<sup>40</sup> Over the past year, Zydus Cadila, an Indian health care company, collaborated with the Japanese, Takeda Pharmaceutical Company Limited for conducting joint research to accelerate the development of CHIKV vaccine. The joint venture has raised the hope of rapid advancement of CHIKV/IRES (v1/v2) to human trials in the near future.

Another attenuated vaccine strategy developed to limit reversions is the deletion in structural and nonstructural genes of La Reunion (LR-CHIKV) strain of ECSA genotype. Three different CHIKV vaccine candidates were developed; Δ5nsP3 an attenuated virus with a large deletion of 180 nucleotides (60 amino acids) from the nsP3 replicase region; DREP-E DNA-launched replicon vaccine with deletion of the region encoding the capsid protein; MVA-CE recombinant vaccinia Ankara virus expressing entire structural (C-E3-E2-6K-E1) protein. A recent study showed that the monovalent, single dose of attenuated Δ5 virus vaccine was the safest, most efficacious, durable, and generated protective antibody and T cellular responses with long-term memory in preclinical models.<sup>41,42</sup> These promising results of attenuated Δ5nsP3 vaccine candidate named VLA1553 have recently marked its entry into a Phase I trial sponsored by a European biotech company – Valneva, Austria GmbH.<sup>43</sup>

## Whole-virus-inactivated CHIKV vaccines

The early work on the production of inactivated CHIKV vaccine began in 1967. The whole virus “seed culture” of CHIKV strain 15,561 grown in frozen African green monkey kidney tissue culture cells was inactivated using formalin, followed by purification.<sup>44</sup> Harrison et al administered formalin-inactivated vaccine in two doses given twice (either 0.5 mL or 1 mL) in 16 healthy volunteers (aged 21–55 years) divided equally into 2 groups (n=8).<sup>45</sup> The results showed 100% sero-conversion rate (plaque reduction neutralization test [PRNT], complement fixation and hemagglutination assays) in both the groups without eliciting any local and systemic reactions. Despite excellent immunogenic response, the development of formalin-killed CHIKV vaccine was halted due to high manufacturing costs involving high biocontainment facilities.

The major drawback was difficulty in comparison of results with other vaccine studies due to difference in dosage of vaccine inoculum used.

After more than three decades, another novel purified, noninfectious, inactivated CHIKV vaccine candidate has been developed using ECSA clinical virus isolates (CHK/03/06) obtained during 2006 outbreak in India and propagated in Vero and MRC-5 cells. The chemical inactivation of stabilized virus formulation was done using formalin or beta-propiolactone.<sup>8,46</sup> Bharat Biotech International Limited filed several patents (US 8865184, EP 2720715 B1, WO 2017123932 A1) related to inactivated CHIKV vaccine formulation. Subsequently, after successful production and preclinical animal testing, the approved vaccine has advanced for Phase I clinical trial (CTRI/2017/02/007755).<sup>47</sup>

## VLP CHIKV vaccines

In contrast to traditional attenuated and inactivated vaccine approaches, use of VLP has proven to be a new highly efficacious vaccine approach as these are nonreplicating, noninfectious, and self-assembled multiprotein viral constructs resembling native alphavirus. The novel strategy involves transfection of embryonic kidney (HEK-293) cells with optimized codons encoding structural protein genes from West African CHIKV strain 37997, resulting in the production of VLPs with selective expression of E1 and E2 glycoproteins in cell culture supernatant. In preclinical animal model, immunization with these VLPs induces high-titer neutralizing antibodies against enveloped glycoproteins that provide protection against viremia after high-dose challenge with wild-type virus.<sup>48,49</sup> The novel VLPs vaccine platform became the first recombinant vaccine prototype to enter the Phase I clinical trial (NCT01489358) in 2011. VRC 311 was a nonrandomized, open-label trial in 25 adult volunteers (aged 18–50 years) enrolled at the National Institutes of Health, USA. Participants receive three intramuscular injections of VLP vaccine (VRC-CHKVLP059-00-VP) without adjuvant in three dosage groups 10, 20, or 40 µg at weeks 0, 4, and 20.<sup>50</sup> Overall, vaccine was well tolerated, safe, and highly immunogenic with no serious adverse events (arthralgia). Remarkably, neutralizing antibodies were immunogenic after first and second vaccination in all participants with a significant boost observed 4 weeks after the third vaccination. However, the result of neutralization titers of this study cannot be extrapolated with previous clinical studies as these were measured by high-throughput flow cytometry-based assay as compared to the traditional PRNT assay. Based on the promising results, VLP vaccine has been advanced to



Phase-IIa randomized multicenter trial involving six cities in endemic regions of the Caribbean (NCT02562482).<sup>51</sup> The study is sponsored by the National Institutes of Health and designed to recruit 400 healthy volunteers (aged 18–60 years) to evaluate the safety and immunogenicity of 20 mcg dose in a two-injection regimen at days 0 and 28. The primary endpoint results are anticipated to get published soon. Over the past year, PaxVax, a “US based vaccine manufacturing company,” has been working to develop a commercial manufacturing process of this potential CHIKV vaccine in alliance with the National Institute of Health and the DoD, USA. PaxVax has in-licensed the chikungunya VLP vaccine platform (PXVX0317) that has entered Phase IIb randomized clinical trial in 415 healthy adults to mitigate the high cost and associated risks (NCT03483961).<sup>52</sup> Hence, the encouraging results from preclinical and Phase I trial represents VLPs as nonadjuvanted, safe, and highly immunogenic vaccine delivery platform capable of inducing humoral immune response. However, the demerit of this approach is that VLPs require multiple dosages to induce maximal immunogenicity, thereby increasing the cost of manufacturing and limiting the use in resource-constrained countries. Another drawback is their low-yield production. To overcome this issue, Metz and Pijlman in 2016 produced CHIKV-enveloped VLPs in insect cells using recombinant baculovirus expression platform, generating high yields on large scale with high immunogenicity and protection against CHIKV infection in experimental model systems.<sup>53</sup>

## Live-attenuated, recombinant virus-vectored CHIKV vaccines

Another modern vaccine approach uses attenuated measles virus (MV) Schwarz strain (MV/Schw), an RNA virus containing a large 3.7 kb insert of structural coding gene (C-E3-E2-6KE1) derived from La Reunion ECSA CHIKV strain. This recombinant MV-CHIKV vaccine generates VLPs under culture conditions. During preclinical testing, MV-CHIKV induces 100% seroprotection in mice with strong neutralizing and cellular immune response after second dose.<sup>54</sup> This novel vaccine is developed by Themis Bioscience GmbH based on company's proprietary Themaxyn® platform in collaboration with the Institut Pasteur. Based on the promising preclinical data (in mice and unpublished work in nonhuman primates), a Phase I, randomized clinical trial in 42 healthy volunteers (18–45 years) was done to assess the safety, tolerability, and immunogenicity of the recombinant vaccine (EudraCT no 2013-001084-23).<sup>55</sup> The vaccine was given in three dose groups – low, medium, and high in two immunization

schedules. In all dose groups, the candidate vaccine generated adequate neutralizing antibodies after first vaccination and 100% seroconversion rate following the booster vaccination. The mild-to-moderate adverse events were noticed in all vaccinees; however, no vaccination-related serious adverse events were observed. After the successful Phase I trial, MV-CHIKV is being tested in three Phase II trials conducted simultaneously at three different locations (Europe, USA, and Caribbean island). Recently, the development of MV-CHIKV vaccine is being funded by Innovate UK (a nondepartmental public body, sponsored by the UK Department for Business, Innovation & Skills) for rapid clinical evaluation at large scale. Themis Bioscience has started a Phase II trial in 2016 to determine the optimum dose of MV-CHIKV vaccine in 320 participants (18–55 years) (NCT02861586).<sup>56</sup> The study results are anticipated in 2018. Another Phase II trial has also been prompted by Themis Bioscience in collaboration with WRAIR, USA in a previously endemic area in Caribbean island (NCT03101111).<sup>57</sup> A third Phase II trial of MV-CHIKV vaccine has been actively initiated by NIAID (NCT03028441) in three different states of USA (Georgia, Iowa, and Texas).<sup>58</sup> Overall, MV-CHIKV vaccine will be tested in more than 600 healthy volunteers. Recently, another Phase II block-randomized, prospective trial has commenced to compare the different formulations and dosages of MV-CHIKV vaccine in 60 healthy volunteers (NCT).<sup>59</sup> These active ongoing trials will facilitate the novel MV-CHIKV vaccine candidate toward Phase III trials. The major advantage of this vaccine is its cost-effectiveness in manufacturing on large scale and requires comparably low effective doses in two-dose immunization schedule. However, interference of the preexisting immunity against MV infection or vaccination must be taken into account in upcoming trials.

A novel CHIKV vaccine using advanced recombinant adenoviral technology, known as chimpanzee adenovirus Oxford 1 and 2 (ChAdOx1 and ChAdOx2), expressing structural proteins of CHIKV has been designed by Jenner Institute. Chimpanzee adenoviruses are nonenveloped, genetically stable, complex replication-deficient vectors, thermostable for storage at 45°C for 6 months, or ambient temperatures for much longer, and relevant among all age groups. The single dose of this vaccine has elicited safe and robust immune response against the encoded antigens in preclinical models and successfully advanced to good manufacturing practice production.<sup>60</sup> The research group headed by Prof Arturo Reyes-Sandoval at Jenner Institute, University of Oxford financially supported by Innovate UK has recently projected ChAdOx 1 vaccine to enter nonrandomized Phase I

clinical trial to evaluate its safety and immunogenicity.<sup>61,62</sup> This vaccine platform has been recognized by the WHO R&D Blueprint process for acceleration of large-scale production of CHIKV vaccine in response to outbreak preparedness.

## mRNA-based CHIKV vaccines

Moderna Therapeutics – a US-based Biotech Company (originally named ModeRNA), develops drugs and vaccines based on mRNA technology. The innovative idea behind this promising drug approach is to deliver engineered mRNAs that direct cells to produce and express viral antigenic proteins, emulating a viral infection that could elicit the immune response to generate antibodies. This multidisciplinary mRNA platform overcomes an array of challenges – evasion of immune system against engineered mRNAs, ample synthesis of desired protein inside target cells, and so on. An array of potent vaccines has been developed against emerging infectious diseases like influenza, chikungunya, ebola, and zika using this approach. Single dose of CHIKV mRNA vaccine encoding structural polyprotein of CHIKV provoked 100% protection in mice and strong antibody responses in nonhuman primates. Based on these promising results, CHIKV vaccine mRNA-1388 has advanced for Phase I study in humans (NCT03325075).<sup>63</sup>

## Passive immunization

The passive immunization by transfer of purified polyclonal immunoglobulins from human sera has been widely demonstrated in many viral infections. Couderc et al used human immunoglobulins purified from convalescent phase plasma samples of CHIKV-infected patients in two susceptible mouse models.<sup>64</sup> These immunoglobulins show high neutralizing activity in vitro, prevent viral dissemination in tissues, and show prophylactic and therapeutic efficacy from lethal CHIKV infections. However, over the past years, a number of studies have also established that monoclonal antibodies also show protective efficacy in CHIKV-infected mice.<sup>65–68</sup> These studies suggested that CHIKV immunoglobulins may act as antiviral therapy for high-risk human population exposed to CHIKV.<sup>69</sup> Keeping these studies in view, anti-CHIKV hyperimmune intravenous immunoglobulins therapy undergoes an open, nonrandomized Phase I/II trial (CHIKVIG-01) in high-risk neonates born to CHIKV-viremic mothers (NCT02230163).<sup>70</sup>

## Vaccine trials in clinical pipeline

Work over the past 5 years has brought revolution in the CHIKV vaccine development; predominantly 2017/2018

has marked the crucial year of this decade in which eleven promising clinical trials have been instigated in different phases of clinical pipeline worldwide. Table 1 summarizes the various chikungunya vaccine strategies in human clinical trials.

## Ongoing Phase I clinical trials

1. A novel CHIKV live-attenuated vaccine VLA1553 ( $\Delta 5nsP3$ ) has been projected for Phase I, randomized, multicenter trial in 120 healthy volunteers of age 18–45 years to be enrolled across three locations in USA ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT03382964) NCT03382964). Single immunization of three dose levels – low (n=30), medium (n=30), and high (n=60) will be given intramuscularly to assess the safety and immunogenicity. This study has been sponsored by Valneva Austria GmbH.<sup>43</sup>
2. Another Phase I, randomized, open-label, placebo-controlled, multicentric clinical trial has started recruiting healthy adults (n=60) of age 18–50 years to evaluate the safety, tolerability, and immunogenicity of inactivated CHIKV vaccine BBV87 in prime-boost regimen. This trial has been registered by Bharat Biotech International Ltd, Hyderabad with Clinical Trial Registry India (CTRI/2017/02/007755) in early 2017.<sup>47</sup>
3. A Phase I/II, double blinded, placebo-controlled, dose comparison trial ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT03028441) NCT03028441) has started recruiting healthy adults aged 18–45 years (n=180) to evaluate the safety, immunogenicity, and schedule of live-attenuated, measles-vectored CHIKV vaccine (MV-CHIKV) developed by Themis Bioscience GmbH. The different dosage and immunization schedules will be evaluated. Themis' prophylactic vaccine candidate (MV-CHIKV) is being tested and sponsored by NIAID, USA.<sup>58</sup>
4. The Jenner Institute, University of Oxford has designed a novel chimpanzee adenoviral vector-based vaccine, ChAdOx1- and ChAdOx2-expressing chikungunya structural antigens (capsid and envelope glycoprotein). After the preclinical validation of the single-dose, adjuvant-free CHIKV vaccine, this novel candidate vaccine has now entered into a Phase I clinical trial funded by UK's Innovate agency.<sup>61</sup> An open-label, nonrandomized, dose-escalation trial has started recruiting healthy volunteers (age 18–50 years) to assess the safety and immunogenicity of ChAdOx1 CHIK vaccine (CHIK001) ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT03590392) NCT03590392).<sup>62</sup>
5. In late 2017, a Phase I, randomized, placebo-controlled, dose-escalation clinical trial ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT03590392)

**Table I** Chikungunya vaccine strategies in human clinical trial testing

Type of vaccine	Manufacturer	Phase of clinical trial	Trial registration no	Recruitment status	Reference
Live-attenuated vaccine					
TSI-GSD-218	USAMRIID/Salk Institute for Biological Studies	Phase I	NA	Completed with results	33
		Phase II	NA	Completed with results	34
Δ5nsP3 VLA1553	Valneva Austria GmbH	Phase I	<a href="#">ClinicalTrials.gov</a> NCT03382964	Recruiting	43
Inactivated vaccine					
Formalin-inactivated vaccine	United States USAMRIID and WRAIR, USA	Phase I	NA	Completed with results	45
Inactivated chikungunya vaccine BBV87	Bharat Biotech International, Ltd, India	Phase I	Clinical Trial Registry India (CTRI/2017/02/007755)	Active, recruiting	47
VLP vaccine					
VRC311: VRC-CHKVLP059-00-VP	NIAID	Phase I	<a href="#">ClinicalTrials.gov</a> NCT01489358	Completed with results	50
VRC704: VRC-CHKVLP059-00-VP	NIAID	Phase IIa	<a href="#">ClinicalTrials.gov</a> NCT02562482	Active, not recruiting	51
PXVX0317 CHIKV-VLP	Pax Vax	Phase IIb	<a href="#">ClinicalTrials.gov</a> NCT03483961	Active, not recruiting	52
Live, measles-virus-vectored vaccine					
MV-CHIKV	Themis Bioscience GmbH/ Institut Pasteur in Paris, in collaboration with University of Vienna, Austria	Phase I	EudraCT 2013-001084-23	Completed with results	55
MV-CHIK-202	Themis Bioscience GmbH, in collaboration with University of Vienna, Austria	Phase II	Edura CT 2015-004037-26	Completed	56
			<a href="#">ClinicalTrials.gov</a> NCT02861586	Results awaited	
MV-CHIK-204	Themis Bioscience GmbH in collaboration with WRAIR, USA	Phase II	<a href="#">ClinicalTrials.gov</a> NCT03101111	Active, not recruiting	57
MV-CHIKV	Themis Bioscience GmbH in collaboration with NIAID	Phase I/II	<a href="#">ClinicalTrials.gov</a> NCT03028441	Active, not recruiting	58
MV-CHIKV	Themis Bioscience in collaboration with Celerion Belfast	Phase II	<a href="#">ClinicalTrials.gov</a> NCT03635086	Active, not recruiting	59
Chimpanzee adenovirus-vectored vaccine	University of Oxford UK Research and Innovation	Phase I	UK Research and Innovation 972212	Active	62
mRNA-based vaccine					
mRNA VAL-181388	Moderna Therapeutics, USA and Defense Advanced Research Projects Agency	Phase I	<a href="#">ClinicalTrials.gov</a> NCT03325075	Recruiting	63
Passive immunization					
Anti-CHIKV hyperimmune immunoglobulins	Centre Hospitalier Universitaire de, Pointe-à-Pitre, Caribbean	Phase I/II	<a href="#">ClinicalTrials.gov</a> NCT02230163	Unknown	70

**Abbreviations:** MV, measles virus; NA, not applicable; NIAID, National Institute of Allergy and Infectious Diseases; USAMRIID, United States Army Medical Research Institute of Infectious Diseases; TSI-GSD, The Salk Institute-Government Services Division; VLP, virus-like particle; WRAIR, Walter Reed Army Institute of Research.

NCT03325075) to evaluate the safety and immunogenicity of CHIKV vaccine candidate dubbed mRNA VAL-181388 in healthy adults (n=60) in a nonendemic chikungunya region has been registered. The study

involves three dosages in two immunization schedules. This study is sponsored by Moderna therapeutics supported by Defense Advanced Research Projects Agency, an agency of the US DoD.<sup>63</sup>



6. An open, nonrandomized Phase I/II trial ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT02230163) NCT02230163) has been initiated in 2014, with an aim to evaluate the safety and clinical evaluation of anti-CHIKV hyperimmune intravenous immunoglobulins (CHIKIVIG-01) therapy to high-risk neonates born to CHIKV-viremic mothers. Unfortunately, very few neonates have been recruited till date and study has not been verified since last 2 years.<sup>70</sup>
7. Another commercial organization, Indian Immunologicals Ltd, Hyderabad has received chikungunya vaccine seed materials (TSI-GSD-218 or 181/clone 25) from the USAMRIID. After the completion of initial preclinical toxicity studies and reinitiating the inactivated CHIKV vaccine development, the Phase I clinical trial is in progress.<sup>71</sup>
4. Another Phase II, comparator-controlled, randomized, double-blinded clinical trial ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT03101111) NCT03101111) has started recruiting (n=100) healthy volunteers aged 21–50 years in a previously epidemic area (Puerto Rico) to evaluate the safety of live-attenuated MV-vectored CHIKV vaccine (MV-CHIK204). This trial is also sponsored by Themis Bioscience GmbH in collaboration with WRAIR.<sup>57</sup>
5. Themis Bioscience has initiated an additional Phase II randomized trial to investigate the safety, tolerability, and long-term durability of licensed MV-CHIKV in different formulations (lyophilized, liquid frozen, and liquid SPS) and dose regimens.<sup>59</sup>

## Ongoing Phase II clinical trials

1. After the promising results of Phase I trial of VLP vaccine that consists of the E1, E2, and capsid proteins published by Chang et al,<sup>50</sup> a Phase II, multicenter, randomized, placebo-controlled, double-blind trial of CHIKV VLP vaccine has been registered by NIAID ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT02562482) NCT02562482) in 400 healthy adults (age 18–60 years) in CHIKV endemic areas. The primary objectives are to evaluate safety and tolerability of a two-injection vaccine regimen of VRC-CHKVLP059-00-VP at 20 mcg compared to placebo (PBS) in healthy adults.<sup>51</sup>
2. Pax Vax has recently announced a Phase II parallel-group, randomized, double-blind trial ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT03483961) NCT03483961) to evaluate the safety and immunogenicity of VLP vaccine, PXVX0317 CHIKV-VLP, to different doses/formulations and schedules in 415 healthy adults (age 18–45 years) at three locations in USA. This trial has received the designation of priority access “Fast Track Program” from Food and Drug Administration of USA.<sup>52</sup>
3. Data from the Phase I trial of MV-based CHIKV vaccine trial are greatly anticipated. Recently, a Phase II, double-blinded, block-randomized, and placebo-controlled trial has been registered ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT02861586) NCT02861586) to evaluate the optimal dose of measles-virus vaccine (MV-CHIK-202) by assessing immunogenicity, safety, and tolerability in 320 healthy volunteers (age 18–55 years). Two dose levels (low and high) at three immunization schedules will be assessed. This trial is being sponsored by Themis Bioscience GmbH in collaboration with University of Vienna, Austria. Very recently, the ongoing trial has completed the recruitment of the participants

and 263 healthy adults have been enrolled. The results are expected to be published soon.<sup>56</sup>

## Challenges in CHIKV vaccine development

The attempts to develop CHIKV vaccine began in the early 1950s. The development of a safe and effective vaccine has been plagued by several challenges and quandaries. Unlike global dengue vaccine development for which more than 60 clinical trials have been registered so far, only few CHIKV vaccine development initiatives have been completed, with limited vaccine candidates in preclinical and clinical pipeline using different technologic approaches. The challenges in developing a CHIKV vaccine can be categorized into three broad groups: clinical/therapeutic, social/demographic, and economic.

The major issue in the CHIKV vaccine field is the selection of biologic assays used to evaluate vaccine immunogenicity. The standard protection assay, that is, PRNT, is usually based on the measurement of neutralizing antibodies at baseline and post-vaccination. The interlab and interassay cutoff variation (two to threefold) in seroconversion rates and antibody titers exhibit significant shortcoming in vaccine trials. The low and waning immunogenicity of candidate vaccines in vaccinees and healthy controls is also a matter of great concern. The candidate vaccines in pipeline mainly focus on adult population, so their efficacy in pediatric population remains uncertain. The appropriate dosing requirement of varied CHIKV vaccine formulations, that is, multiple doses or single dose is better to maintain immunity against CHIKV, is also vacillating. Also, the substantial gap in our understanding of innate and adaptive immune responses that confer protection or pathologic immune response following a CHIKV infection poses another major challenge in vaccine development. The codistribution and coinfection of chikungunya and dengue virus outbreaks is another bottleneck.

Despite the substantial increase in global travel to many CHIKV-endemic areas, the unpredictable epidemiology of chikungunya (sporadic outbreaks interspersed by long periods of quiescence) makes it less attractive for travel and tourism industry unlike diseases like yellow fever. The prevalence of chikungunya among the developing and underdeveloped countries with low literacy levels, low per capita income, low living standards, and poor sanitation reduces the probability of marketing and commercialization of CHIKV vaccine in developed nations. Finally, a successful CHIKV vaccine candidate to enter the Phase III clinical trials and licensure is challenging due to selection of a suitable location with adequate incidence of chikungunya in target population to demonstrate clinical efficacy. The huge capital investment, uncertain profits, and redundant regulatory pathways further reduce the interest of pharmaceutical companies and investors in CHIKV vaccine development. However, to overcome the financial risks and regulatory challenges, there is an urgent need for the global partnerships including national and international public, private, and government funding agencies, industry, researchers, clinicians, public health officials, and other global health stakeholders with an aim to develop new vaccines.

## Summary

This review gives an insight about several CHIKV vaccines in clinical pipeline and puts into perspective the findings, merits, and limitations of all clinical trials currently available. Although no vaccine is licensed at present, an effective vaccine for chikungunya will not only deter the rising trend of this infection but also contribute in limiting the disease-associated morbidity and health care costs involved in its management. In general, the ideal CHIKV vaccine to be licensed and ready for commercial use should be well tolerated, safe, adjuvant-free, and cost-effective, devoid of major vaccine-related adverse events such as arthralgia, have robust efficacy, and provide long-lasting immunity after a single dose in the general population. The efficacy of these vaccines in an endemic setting with limited resources would be greatly valuable as a significant burden of chikungunya lies in such regions.

This study will be helpful in suggesting which type of vaccine is clinically relevant and could be adopted for future clinical trials for estimation of safety, immunogenicity, tolerability, and efficacy. The rapid ongoing progress has raised hope for large-scale Phase III efficacy trials in endemic settings to begin soon. Although there is still a long way to go before a prophylactic vaccine for chikungunya is available, the vigorous restorative advancement during the last year

makes the likelihood of having an effective vaccine in the near future high.

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