Gastric adenocarcinoma and proximal polyposis of the stomach: diagnosis and clinical perspectives

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Abstract: Gastric adenocarcinoma and proximal polyposis of the stomach (GAPPS) is a recently described, rare gastric polyposis syndrome. It is characterized by extensive involvement of the fundus and body of the stomach with fundic gland polyps sparing the antrum and lesser curvature, an autosomal dominant inheritance pattern with incomplete penetrance, and a significant predisposition for the development of gastric adenocarcinoma. Due to the recent discovery of APC promotor IB mutations (c.-191T>C, c.-192A>G, and c.-195A>C), which reduce binding of the transcription factor Yin Yang 1 (YY1) and transcriptional activity of the promotor, as its underlying genetic perturbation, GAPPS has been added to the growing molecular class of APC-associated disorders. Recent reports on family members afflicted by gastric polyposis due to GAPPS have described the development of metastatic cancer or the presence of invasive gastric adenocarcinoma in total gastrectomy specimens after variable periods of endoscopic surveillance emphasizing the need for an improved understanding of the to-date poorly characterized natural history of the syndrome. There are, however, currently no guidelines on screening, timing of prophylactic gastrectomy, or endoscopic surveillance for GAPPS available. In this review, we summarize the clinical, pathological, and genetic aspects of GAPPS as well as management approaches to this rare cancer predisposition syndrome, highlighting the need for early recognition, a multidisciplinary approach, and the creation of prospective family registries and consensus guidelines in the near future.

Keywords: gastrointestinal polyposis, fundic gland polyp, APC promotor IB variant, APC-associated disorder, gastric adenocarcinoma predisposition

Diagnosis

In 2012, Worthley et al reported in three families the clinicopathological features of a novel gastric polyposis syndrome termed gastric adenocarcinoma and proximal polyposis of the stomach (GAPPS).1 The authors described in a large Australian family and two smaller families from USA and Canada that multiple family members afflicted by fundic gland polyposis (>100 polyps carpeting the fundus and body of the stomach sparing the antrum and lesser curvature in the index proband) multiple cases of intestinal-type adenocarcinoma of the stomach arising in regions of fundal gland polypsis with high-grade dysplasia and adenomatous polyps. Within the same issue, researchers from Japan described two additional families with a similar phenotype.2 Genetic testing for known gastrointestinal (GI) polyposis and gastric cancer predisposition syndromes was negative. Together with an autosomal dominant inheritance pattern, the authors proposed a series of endoscopic and pathological criteria for the diagnosis of GAPPS, which are listed in Table 1.1
The initial endoscopic appearance of massive gastric polyposis of family members afflicted by GAPPS can resemble polyposis as a part of other gastric or GI polyposis syndromes. Figure 1 shows the pathognomonic lack of involvement of the antrum by fundic gland polyps (FGPs) on upper endoscopy in GAPPS, which can facilitate differential diagnosis to other GI polyposis syndromes. Generous biopsy sampling for an accurate histopathological diagnosis of the encountered lesions is paramount. In addition, in view of the overlapping phenotype with other GI polyposis syndromes, the criteria proposed by Worthley et al require the exclusion of other heritable gastric polyposis syndromes. These should include MUTYH-associated polyposis (due to germline variants in the MUTYH gene; GI phenotype of multiple adenomatous colon polyps with greatly increased lifetime risk of colorectal cancer, followed by duodenal polyps and elevated duodenal cancer risk), juvenile polyposis syndrome (mutations in the BMPR1A or SMAD4 genes; multiple initially benign hamartomatous juvenile polyps across the GI tract with colorectal cancer the most common type of cancer), Peutz–Jeghers syndrome (loss of tumor suppressor STK11 [LKB1]), hamartomatous polyps in the GI tract, elevated cancer risk in many other organs), and Cowden syndrome (loss of tumor suppressor PTEN; hamartomatous polyps, colorectal cancer most frequent GI cancer; breast, thyroid, and endometrial cancers and extraintestinal manifestations). Table 2 lists cancer manifestations and initial phenotypes of common hereditary GI polyposis syndromes in comparison to the clinicopathological phenotype of GAPPS. To rule out germline variants associated with other gastric polyposis syndromes, commercially available multi-cancer gene panels for genetic testing are available, which include above polyposis-disposition genes and in addition can capture variants associated with known hereditary gastric cancer predisposition syndromes, which include CDH1 (hereditary diffuse gastric cancer [HDGC]), CTNNA1 (HDGC), TP53 (Li–Fraumeni syndrome), MLH1, PMS2, MSH2, and MSH6 (Lynch syndrome). Panel testing should follow current guidelines established by the American College of Medical Genetics and American Society of Clinical Oncology including ethical and policy issues in the genetic testing and screening of children.1–7

While awaiting results of genetic testing, the initial clinicopathological information from the proband can be used to aid differential diagnosis of other gastric polyposis syndromes: for example, hamartomatous polyps in Peutz–Jeghers, juvenile polyposis, or Cowden syndrome are distinct from FGPs with dysplasia due to their disorganized growth of tissue and initially benign character. However, securing a diagnosis of the underlying GI polyposis syndrome based

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### Table 1 Diagnostic criteria for GAPPS

<table>
<thead>
<tr>
<th>Diagnostic Criteria</th>
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<tr>
<td>1. Gastric polyps restricted to the body and fundus with no evidence of colorectal or duodenal polyposis</td>
</tr>
<tr>
<td>2. &gt;100 polyps carpeting the proximal stomach in the index case, or &gt;30 polyps in a first-degree relative of another case</td>
</tr>
<tr>
<td>3. Predominantly FGP histology, some having regions of dysplasia (or a family member with either dysplastic FGPs or gastric adenocarcinoma)</td>
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<tr>
<td>4. An autosomal dominant pattern of inheritance</td>
</tr>
<tr>
<td>5. Exclusion of other heritable gastric polyposis syndromes* and the use of PPIs*</td>
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*Abbreviations: FGP, fundic gland polyps; GAPPS, gastric adenocarcinoma and proximal polyposis of the stomach; GJPS, generalized juvenile polyposis syndrome; MAP, MUTYH-associated polyposis; PJS, Peutz–Jeghers syndrome; PPIs, proton pump inhibitors.*
Table 2: Hereditary GI polyposis syndromes

<table>
<thead>
<tr>
<th>Syndrome</th>
<th>Gene(s)</th>
<th>Cumulative cancer risk</th>
<th>Pattern of inheritance</th>
<th>Predominant initial phenotype</th>
<th>References</th>
</tr>
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<tbody>
<tr>
<td>MUTYH-associated polyposis</td>
<td>MUTYH/MYH</td>
<td>Colon cancer – 43%–100% Small bowel (duodenum) – 4% Stomach – 1%</td>
<td>Autosomal recessive</td>
<td>&gt;10 colorectal polyps (adenomatous and serrated)</td>
<td>58–61</td>
</tr>
<tr>
<td>Juvenile polyposis syndrome</td>
<td>SMAD4, BMPR1A</td>
<td>Colon cancer – 38%–68% Gastric – up to 21% Pancreas – elevated, small Small bowel – elevated, small</td>
<td>Autosomal dominant</td>
<td>Elevated risk for cancer as a result of hamartomatous polyps in the GI tract including colon, rectum, and stomach</td>
<td>9,60,62,65</td>
</tr>
<tr>
<td>Peutz-Jeghers syndrome</td>
<td>STK11</td>
<td>Breast – 45%–50% Colon – 39% Gastric – 29% Pancreas – 11%–36% Ovarian – 18%–21% Lung – 15%–17% Small bowel – 13% Cervical – 10% Endometrial – 9% Testicular – elevated, small</td>
<td>Autosomal dominant</td>
<td>Hamartomatous gastrointestinal polyps with distinctive Peutz–Jeghers histology in small bowel, stomach, colon, and nasal passages. Pigmented spots around the mouth, eyes, nostrils, and anus and on the fingers during childhood</td>
<td>9,13,60,64</td>
</tr>
<tr>
<td>PHTS; Cowden syndrome</td>
<td>PTEN</td>
<td>Breast – 25%–85% Thyroid – 3%–38% Kidney (renal cell) – 15%–34% Endometrium – 5%–28% Colon 9%–16% Melanoma 6%</td>
<td>Autosomal dominant</td>
<td>Multiple hamartomatous polyps across the colon as well as other areas within the gastrointestinal tract. Other types of polyps include ganglioneuromatous, adenomatous, and lymphoid polyps. Non-malignant features include macrocephaly, hamartomatous brain tumors, and distinctive skin findings such as trichilemmomas, acral keratoses, and papillomatous papules. Developmental delay and/or autism spectrum disorders may be present</td>
<td>65–69</td>
</tr>
<tr>
<td>FAP</td>
<td>APC</td>
<td>Colon – 100% Small bowel (duodenum) – 3%–10% Thyroid – 2% Liver (hepatoblastoma) – 1%–2% Brain/central nervous system – 1%–2% Pancreas – 1.7% Stomach – &lt;1%</td>
<td>Autosomal dominant</td>
<td>Large numbers of adenomatous polyps in the GI system, particularly in the colon, rectum, stomach, and small bowel. Gastric polyps are common. Desmoid tumors are the most serious cause of morbidity and mortality among affected individuals after colorectal cancer</td>
<td>60,70–72</td>
</tr>
<tr>
<td>AFAP</td>
<td>APC</td>
<td>Colon –69% Small bowel (duodenum) – 4%–12% Thyroid – 1%–2%</td>
<td>Autosomal dominant</td>
<td>Fewer than a hundred polyps, polyps more frequently located in right side of the colon</td>
<td>60,73</td>
</tr>
<tr>
<td>GAPPS</td>
<td>APC/promotor IB variants</td>
<td>Intestinal-type of gastric cancer – significantly elevated, exact risk unknown</td>
<td>Autosomal dominant</td>
<td>Fundic gland polyps with dysplasia in the absence of colonic or duodenal polyps</td>
<td>1,46</td>
</tr>
</tbody>
</table>

Note: Data from Snygal et al.57

Abbreviations: AFAP, attenuated FAP; FAP, familial adenomatous polyposis; GAPPS, gastric adenocarcinoma and proximal polyposis of the stomach; GI, gastrointestinal; PHTS, PTEN hamartoma tumor syndrome.

on morphological characteristics of gastric polypectomy specimens alone is unlikely; for example, histopathological differences of gastric hamartomatous polyps among GI polyposis syndromes are difficult to diagnose, have a low diagnostic accuracy (41% for juvenile polyposis and 54% for Peutz–Jeghers syndrome), and in general, do not allow to distinguish the underlying genetic syndrome without any genetic or clinical corroboration.57 Distinctive features reported for
Rudloff

Peutz–Jeghers polyps are a cytoarchitectural pattern of pits and glands grouped or packeted together with intervening septations of smooth muscle strands not connected to the muscularis mucosae and an unremarkable lamina propria mucosae, whereas juvenile polyps more frequently have an overall disorganized pit and gland architecture with varying cystic glands and form spherical, club-shaped, or irregular villiform structures due to overgrowth of an edematous lamina propria with a mononuclear inflammatory cell infiltrate.8,9 Of note, hybrid polyps with varying degrees of hamartomatous and FGP features have been described in FGPs arising in familial adenomatous polyposis (FAP) and attenuated FAP (AFAP) patients similarly underscoring the need of clinical and genetic corroboration to secure a diagnosis.10,11 Another entity with histopathological similarities to hamartomatous polyps are hyperplastic polyps where pits are connected to deeper portions in a linear trajectory with overall reduced architectural disorganization and varying degrees of stromal edema and inflammatory cell infiltrate.8,12 In contrast to GAPPS, MUTYH-associated polyposis is an autosomal recessive condition and is frequently associated with colonic polyposis, an exclusion criteria of GAPPS, with rare FGP involvement of the stomach.9,13 Other conditions with possible endoscopic resemblance of GAPPS are Ménétrier’s disease or hypoproteinemic hypertrophic gastropathy forming giant folds in the stomach lining, which are due to an overgrowth of mucous cells in the stomach wall. Cronkhite–Canada syndrome is another GI polyposis syndrome, which is characterized by protein-losing gastroenterocolopathy, which typically affects adults older than 50 years.9,14 Two-thirds of cases are reported from Japan with a male:female ratio of 2:1.14 The histological hallmark of Cronkhite–Canada syndrome polyps resembles juvenile polyposis syndrome or hyperplastic polyps histologically characterized by expanded edematous lamina propria containing a predominantly mononuclear inflammatory cell infiltrate and tortuous, dilated to cystic glands/foveolae or crypts.9,15 Rather than occurring due to a hereditary predisposition, its etiopathogenesis is suspected to be of autoimmune origin.16 Thus, in contrast to hereditary GI polyposis syndromes, the intestinal mucosa not involved by hamartomatous polyps is histologically not normal but shows lamina propria edema, inflammatory cell infiltration, and gland/crypt dilation.9,15 Clinical symptoms are driven by the GI symptoms of protein losing enteropathy or complications from hamartomatous polyps and include unique skin changes in the form of alopecia, hyperpigmentation, onychodystrophy (loss of fingernails), or the loss of taste (ageusia). Patients afflicted by Cronkhite–Canada syndrome carry a poor prognosis predominantly due to the difficult to control enteropathy. Linitis plastica, the submucosal invasion of the diffuse form of gastric cancer, can predominantly involve the upper parts of the stomach and resemble “polypoid” enlargement of the stomach mucosa. Figure 2 shows total gastrectomies specimens of patients with

Figure 2  Gastric polyposis of the fundus and body of the stomach with pathognomonic sparing of the antrum and lesser curvature in a GAPPS proband with confirmed c.-191T>C APC gene promoter IB variant.

Notes: (A) Carpet of polyps involving upper four-fifth of the stomach including cardia, fundus, and body; arrow indicates gastroesophageal junction. (B) Uninvolved antrum, forceps lifts pylorus. (C) Gastric polyps at cardia not extending into esophagus (arrow indicates esophageal mucosa). (D) Linitis plastica due to HDGC causing enlarged folds and “polypoid” appearance without involvement of the distal stomach. (E) Irregular enlarged folds with intact mucosa due to HDGC. (F) Sagittal section shows tumor submucosal tumor infiltration (arrow).

Abbreviations: GAPPS, gastric adenocarcinoma and proximal polyposis of the stomach; HDGC, hereditary diffuse gastric cancer.
florid gastric polyposis due to GAPPS and with limitis plastic due to HDGC involving the upper four-fifth of the stomach. The endoscopic appearance will depend on the extent of the polyposis, and in case of confluent, “carpet-like” involvement with limited disruption of the mucosa, the endoscopist may need to rely on the unique localization of the disease sparing the antrum and parts of the lower curvature and the histopathological diagnosis of FGPs on obtained biopsies to make the connection to GAPPS. Most reports on families afflicted by GAPPS report the presence of small fundal and body polyps (3–12 mm in size).

An important other benign cause for gastric polyposis on endoscopy is the chronic use of proton pump inhibitors (PPIs) inducing oxyntic cell hyperplasia, glandular dilatations, and FGPs. In this case, endoscopy is recommended to be repeated after discontinuation and an appropriate off-treatment interval. There is currently insufficient evidence to consider PPI therapy, a precipitating factor for the development of GAPPS. Serum gastrin levels, which are usually elevated in chronic PPI usage, were found to be normal in affected family members including obligate carriers (4/9) tested positive (P=0.007) suggesting a protective role of H. pylori in the development of GAPPS. These findings mirror observations in FAP where patients with FGPs had lower rates of H. pylori compared to patients without FGPs (13% vs 67%). In contrast, H. pylori infection and degree of atrophic gastritis were positively correlated with the incidence of gastric adenomas in FAP patients.

In contrast, there are several clinical observations, which imply gastric acid homeostasis and milieu in the natural history of FGPs in family members with a genetic predisposition to GI cancers: while chronic PPI usage has been observed as a risk factor for the development of FGPs, inversely Helicobacter pylori (H. pylori) infections have been reported to be negatively associated with the occurrence of FGPs both in healthy individuals and FAP patients. In family members afflicted by FAP, regression of FGPs has been described upon acquisition of H. pylori infection. A possible protective role of H. pylori infection in the APC-related disorder GAPPS has also been suggested in the initially described large Australian GAPPS family when comparing H. pylori status of affected and unaffected family members (P=0.007). Thus, in light of opposing associations of PPI usage and H. pylori infections on FGP incidence and in the absence of more natural history information derived from families afflicted by GAPPS, an approach of caution including early discontinuance of PPIs if clinically feasible and heightened surveillance of family members at risk for GAPPS with a history of chronic PPI usage appears to be the safest course of action. Discontinuation of PPI therapy and regression of FGPs may aid cases of diagnostic uncertainty where a florid polyposis picture with the unique sparing of the gastric antrum and lesser curvature with dysplasia on final pathology of generous biopsy sampling of FGPs is not present. FGPs due to PPI usage are significantly fewer, rarely harbor dysplasia, and do not spare the antrum. With the recent identification of APC promotor IB variants and ability to conduct timely germline genetic testing after genetic counseling stoppage and re-scoping of PPIs will rarely be required to secure the diagnosis and is not a must when other criteria have been met.

The possible association of H. pylori infection rates and the GAPPS phenotype including fundic gland polyposis was evaluated by Worthley et al in greater detail; in 27 family members with known H. pylori status, all evaluated family members with the GAPPS phenotype were negative for H. pylori (0/18) whereas four of the nine unaffected family members including obligate carriers (4/9) tested positive (P=0.007) suggesting a protective role of H. pylori in the development of GAPPS. These findings mirror observations in FAP where patients with FGPs had lower rates of H. pylori compared to patients without FGPs (13% vs 67%).

In contrast, H. pylori infection and degree of atrophic gastritis were positively correlated with the incidence of gastric adenomas in FAP patients. It is currently not known whether H. pylori infection rates are associated with gastric adenoma or adenocarcinoma in GAPPS or whether reduced H. pylori infection rates in patients with the GAPPS phenotype are a result of the disturbed intraluminal milieu due to fundic gland polyposis. Without a causative implication of H. pylori in GAPPS progression, clinical management of GAPPS patients is currently not affected by H. pylori status. In addition to the exclusion of other heritable gastric polyposis syndromes listed in Table 2, Worthley et al require as a part of their diagnostic criteria for GAPPS the endoscopic exclusion of colonic and duodenal polyposis as a part of FAP and AFAP. Fundal gland polyps are frequently present in FAP and AFAP, which are GI polyposis syndromes involving the colon and rectum but frequently harbor FGPs.

Histopathology
The polyposis and cancer predisposition syndrome with the greatest phenotypic overlap with GAPPS and most frequently cited as a diagnostic challenge are FAPs. Fundal gland polyps are one of the sentinel features of the GAPPS phenotype; FGPs most commonly occur as sporadic FGPs where they occur in small numbers, are usually small (<5 mm), are associated with chronic PPI therapy, and frequently harbor in the sporadic setting somatic CTNNB1 (β-catenin) mutations (Table 3). Sporadic FGPs rarely harbor dysplasia and are not connected to an increased risk of cancer. They are more frequently found in probands in their forties and fifties and more commonly in women.
Gastric FGPs are also a frequent phenotypic manifestation of the APC-associated disorders FAP and AFAP. While widely varying rates of FGPs and fundal gland polyposis in individuals with FAP and AFAP have been reported, low-grade dysplasia is a common feature (up to 96%) of FAP-associated FGPs. FAP-associated FGPs preferentially harbor APC mutations and have an, albeit low, potential of malignant transformation toward adenocarcinoma of the stomach. While gastric carcinoma arising in FGPs of FAP patients has been described, the lifetime risk of gastric cancer in FAP patients from Western countries has been reported to be low (0.6%) but has been suggested to be rising. Higher rates of gastric cancer have been described in FAP patients from countries with higher gastric cancer risks including Japan and Korea. Table 3 summarizes common clinicopathological characterizations and clinical implications of FGPs detected within sporadic vs within polyposis-associated contexts. Figure 3 shows GAPPS-associated FGPs with low-grade and focal high-grade dysplasia.

A more systematic review on the histopathology of GAPPS has recently been released. While limited to the review of multiple biopsies and gastrectomy specimens from the initially described, large Australian family, the detailed pathological and immunohistochemical review made important findings: hyperproliferative aberrant pits (HPAPs) were found to be the most frequent and most early histopathological abnormality in a wider histopathological spectrum of GAPPS. HPAPs describe the disorganized hyper-proliferation of oxyntic glands of the stomach mucosa around gastric pits, which give rise to polyoid lesions. The finding of concomitant neoplastic elements in the form of FGPs with multifocal “flat” dysplasia, gastric adenomatous polyps, which were associated with gastric adenocarcinoma together with an immunohistochemical profile, in particular the gastric lineage marker MUC5AC shared by both precancerous lesions and adenocarcinoma, suggests a dysplasia–adenoma–adenocarcinoma sequence in the development of the malignant phenotype of GAPPS. Whether the absence or presence of some these precancerous lesions including their histopathological characteristics (HPAPs, FGPs with or without dysplasia, presence of adenomatous lesions)

Table 3 Disease associations determine biological behavior of fundic gland polyps

<table>
<thead>
<tr>
<th>Fundic gland polyps</th>
<th>Sporadic</th>
<th>FAP/AFAP</th>
<th>GAPPS</th>
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<tbody>
<tr>
<td><strong>Clinicopathological characteristics</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Number</td>
<td>Few found in (0.8–1.9%) esophagogastroduodenoscopies</td>
<td>Many hundreds of polyps in 12.5–84% of FAP patients</td>
<td>&gt;100 or more polyps in index patients</td>
</tr>
<tr>
<td>Age of onset</td>
<td>Middle age</td>
<td>Younger age, 20–30 years of age</td>
<td>Reported in teens</td>
</tr>
<tr>
<td>Size</td>
<td>Small (&lt;5 mm)</td>
<td>1–5 mm, sessile, domed shape with color of normal mucosa</td>
<td>Variable</td>
</tr>
<tr>
<td>Location</td>
<td>Body, fundus</td>
<td>Any location, fundic gland polyps may involve antrum</td>
<td>Fundus and body sparing antrum and lesser curvature</td>
</tr>
<tr>
<td>Dysplasia</td>
<td>Infrequent, low-grade dysplasia</td>
<td>Low grade in 25–50%</td>
<td>Focal high grade</td>
</tr>
<tr>
<td>Somatic mutations</td>
<td>&gt;90% CTNNB1 (β-catenin)</td>
<td>Biallelic APC inactivation</td>
<td>APC, TP53, GNAS, FBXW7</td>
</tr>
<tr>
<td>Gastric cancer risk</td>
<td>None</td>
<td>Low</td>
<td>High</td>
</tr>
</tbody>
</table>

Notes: FAP=plastennstated FAP due to APC germline variants. GAPPS due to APC promoter IB germline variants. More common in middle age women. More equal gender distribution. Variants detected in polyp tissue in addition to the APC germline variants in FAP/AFAP and APC promoter IB variants in GAPPS. Gastric cancer risk reported to be higher in afflicted patients from areas with high gastric cancer rates such as Japan and Korea. 

Abbreviations: AFAP, attenuated FAP; FAP, familial adenomatous polyposis; GAPPS, gastric adenocarcinoma and proximal polyposis of the stomach.

Figure 3 Histopathology of fundic gland polyps in GAPPS.

Notes: (A) Fundic gland polyp with flattened cuboidal epithelium with focal area of low-grade dysplasia (black arrows) and high-grade dysplasia (white arrows) from total gastrectomy specimens of a GAPPS proband with confirmed c.-191T>C APC gene promoter iB variant. (B) Fundic gland polyp with low-grade and (C) high-grade dysplasia in gastrectomy specimens (inset 20×).
can be used to triage patients to undergo total gastrectomy vs endoscopic surveillance has to await the collection of larger case series. The concerns of adequate sampling via endoscopic biopsies in the face of hundreds of heterogeneous polypoid lesions, which may include FGPs with different degrees of dysplasia, adenomas, or mixed histology polyps, pose significant challenges.

Gastric hyperplastic polyps (GHPs) account for the most frequent type of polyp in the stomach, and based on the above unique features of FGPs due to GAPPS, an expeditious distinction to GHPs based on clinicopathological criteria should be possible. GHPs more frequently occur as solitary lesions (65%–75%). Common sites include the antrum, and in more recent reports, the body of the stomach; however, GHPs can occur anywhere. GHPs are intimately linked to gastric inflammation and gastric mucosal injury with chronic gastritis (with or without H. pylori) being the most common (up to 85%). Other conditions associated with GHPs include autoimmune gastritis, bile reflux, and post-gastrectomy status. GHPs are understood as a hyper-regenerative, hyper-proliferative repair response to an underlying mucosal injury. GHPs carry a risk of intraepithelial dysplasia and cancer, which is most strongly related to polyp size and age. Rates of dysplasia in removed GHPs range from 0.4% to 10%, and gastric adenocarcinoma is found less frequent (<1%). GHPs greater than 5 mm should be removed, and oncologic post-polypectomy surveillance is guided by findings on histopathological examination of the polyp and the stage of the surrounding chronic gastritis stage. GHPs result from excessive proliferation of foveolar cells and are histopathologically characterized by surface pits connected to deeper portions of glands in a linear trajectory, which can take on a serrated or star-like appearance on cross-sections. In addition, GHPs not infrequently have infiltration of the stroma by a variable mononucleolar cell infiltrate. The antral involvement, presentation as solitary or multifocal lesions, and the unique histopathological characteristics of foveolar cell hyperplasia with cystic dilatations, smooth muscle strands extending from muscularis mucosa towards surface, and a variable acute and chronic inflammatory stromal infiltrate are key differences to FGPs occurring in GAPPS.

Genetics
The underlying genetic aberration of GAPPS was recently discovered by Investigators from the Queensland Institute of Medical Research, Adelaide, Australia. This advancement already had significant implications both for an improved understanding of the molecular perturbations present in GAPPS and for clinical management with regard to the screening of family members at risk for GAPPS. Initial whole exome and whole genome analyses of affected family members in the first two families described in the original study in 2012 failed to produce a variant, which co-segregated with afflicted family members by GAPPS. A loss of heterozygosity analysis in FGPs on a 46 Mb region of chromosome 5 previously identified by linkage analysis narrowed the region to 12.7 Mb, which was consequently subject to Sanger sequencing. Sequencing identified variants c.-195A>C and c.-125delA in the IB promoter of the APC gene, which completely co-segregated with the multiple affected family members in the large Australian family. APC promoter IB variant c.-191T>C co-segregated in affected family members of families 2, 4, 5, and 6, and promoter variant c.-192A>C co-segregated in family 3 with the GAPPS phenotype. These hotspot APC promoter IB variants were consequently identified in other GAPPS families reported on more recently. Molecularly, the identified point mutations in the APC IB promoter significantly reduce the binding of the transcription factor Yin Yang 1 (YY1) in gastric and colon cancer cells providing functional validation of the identified germline mutation. Importantly, methylation studies on both uninvolved stomach mucosa and FGPs in GAPPS specimens showed methylation of the promoter IA isoform of the APC gene, which the authors discussed as a possible cause for the selective gastric phenotype of the syndrome compared to the other APC-associated disorders FAP and AFAP. It is estimated that promoter IB-driven transcription of the APC gene is ∼15-fold higher than transcripts originating from the IA promoter, which is nearly universally methylated in gastric cancer and noninvolved gastric mucosa. Thus, reduced promoter IB activity due to the loss of the enhancer function of the transcriptional enhancer YY1 leads to a selective tumor suppressive phenotype in the stomach, which does not occur in the colon where intact promoter IA-driven APC gene transcription is preserved and able to compensate for the lost promoter IB function explaining why APC promoter IB point mutations are not associated with a colonic phenotype typically found in FAP or AFAP. Structural variants differently affecting APC promoter IA and IB function hence may form the basis for important unique phenotypic traits in APC-associated disorders adding to the many described genotype–phenotype associations described for FAP. Figure 4 illustrates the position of promoter IA and IB on the APC gene locus, which are ∼29 kb apart, in relation to several large genomic deletions described in FAP families. FAP families with underlying noncoding deletions in the APC...
gene promoter regions more commonly affect the IB isoform and have been reported to have a greater variability of upper GI manifestations, possibly including a higher prevalence of FGPs, compared to the FAP families with the common exomic APC mutations. It is thought that interference with predominantly promotor IB regulated enhancer elements and thus IB activity in APC-mutation carriers with large genomic deletions manifests with different upper GI phenotypes including a greater prevalence of FGPs. These promotor IB structural variants, either in the form of select point mutations in the YY1 binding motif in GAPPS or large deletions affecting regulatory elements of promotor IB, have a unique gastric phenotype due to methylation of the IA promotor in the upper GI tract but preserved promotor IA function in the colon. APC gene perturbations due to variants in the codon regions or rare structural variants affecting the IA promotor in contrast present with the classic FAP and AFAP phenotypes with the classic colonic polyposis and extracolonic manifestations. That APC promotor IB variants are part of a greater spectrum of APC-associated disorders and are related to FAP and AFAP with its classical colonic polyposis phenotype is supported by the presence of a mild colonic phenotype in the form of a greater frequency of colonic polyps or a prior report of a point mutation in the APC IB promotor in a FAP family with colonic phenotype. It is tempting to speculate that the colonic phenotype in this family was due to promotor IA function not completely compensating for the loss of the impaired IB promotor.

There are several phenotypic variabilities in GAPPS, most notably age of onset, penetrance, or degree of dysplasia to name the clinically most relevant ones. The recent genetic and molecular pathological findings may now provide some explanation for the observed heterogeneity in the clinical phenotype and manifestations of GAPPS. For example, the synchronous promotor IB variants c.-125delA and c.-195A>C found in the original large Australian family had a greater impact on APC IB promotor activity in gastric and colonic cancer cells than the c.-191T>C variant. In addition, a number of second hits have been identified in GAPPS specimens, which might differently augment the impact of APC gene germline haploinsufficiency. Reported second hit events in GAPPS in the form of somatic mutations in FGPs or gastric adenocarcinoma of individuals affected by GAPPS include truncating APC mutations, or mutations in TP53, GNAS, or FBXW7. It is tempting to speculate that the presence and type of these second hit events might be responsible for different rates of malignant transformation possibly manifesting in the accelerated development of high-grade FGP dysplasia, the development of adenomatous changes, and the clinical expression of the syndrome in general.

Clinical course
There currently is a large void in the understanding of the natural history of GAPPS, and at least two disconcerting reports have emerged, which in general, do not support prolonged endoscopic surveillance in family members with
gastric polyposis due to GAPPS. Most recently, Repak et al described as a part of their European family a 54-year-old index proband with GAPPS who was diagnosed after a 19-month of endoscopic surveillance with liver metastasis and who died shortly thereafter. Of note, there was no change in the endoscopic appearance or the histopathology of the proband’s gastric polyposis. Similarly, one of the daughters of the proband after being surveilled for 5 years showed an increase in the size of FGPs with low-grade dysplasia and focal high-grade dysplasia on repeat endoscopy. While awaiting total gastrectomy, the patient was diagnosed with widespread metastatic disease and succumbed thereafter. Both her sisters when undergoing gastrectomy after a variable period of surveillance were found to harbor invasive stage I gastric adenocarcinoma showing again FGPs with low-grade and focal high-grade dysplasia on their last endoscopy. Worthley et al described a 33- and 48-year-old relative of the initial proband in their large Australian GAPPS family who died from metastatic intestinal-type adenocarcinoma of the stomach after being diagnosed with GAPPS previously and undergone a period of prior endoscopic observation. In the case of the 33-year-old index patient, both adenomatous polyps and polyps with mixed FGP and adenomatous histopathology were found at total gastrectomy 2 years after the initial diagnosis. While histopathological findings as known to-date indicate a dysplasia–adenoma–carcinoma sequence in the syndrome, these reports suggest that reliance on unchanged endoscopic appearance and histopathology of sampled polyps carries the risk to miss occult sites of malignant transformation and focal progression. The inability to adequately sample the upper four-fifth of the stomach covered by hundreds of polyps by endoscopy, which are frequently large, and harbor a heterogeneous pattern of dysplasia and adenomatous changes should trigger an expedited referral to a clinical geneticist and surgical oncologist. Total gastrectomy should be considered in all GAPPS patients with fundal gland polyposis and the presence of dysplasia on gastric biopsy or polypectomy specimens, who are able to undergo major surgery. Heterogeneity in the impact of the individual primary promoter variants (c.-125 delA c.-195A>C vs c.-191T>C) differently affecting the recruitment of YY1, differences in the frequency and type of second hit events, and any underlying perturbations of the APC promoter IA are likely drivers of the malignant phenotype including rate of progression toward invasion and metastasis. As their impact on malignant transformation and cancer progression is currently unknown, the interindividual heterogeneity within GAPPS will require an individual approach to family members at risk and affected by the phenotype.

The other area of uncertainty is the incomplete understanding of other organs at risk for cancer. Table 4 lists previously described GAPPS families with the age of youngest affected family member by gastric polyposis and gastric adenocarcinoma. With regard to extra gastric manifestations of GAPPS, colonic involvement in GAPPS has undergone additional investigation. McDuffie et al reported an increased incidence of colonic polyps in afflicted family members, and there is agreement on the need of colonscopic screening with regular follow-up colonoscopic examinations guided by the initial findings in these patients. It is thought that the mild colonic phenotype generally with small, <20 in number hyperplastic polyps or small adenomas in GAPPS family members is due to an incomplete protection of APC promoter IA activity. In contrast, it is interesting to note that colon cancer has been described in almost half of the GAPPS families reported to-date where family history information was available (including families with reported but not confirmed colon cancer incidences). While consistent with the phenotype of other APC-associated disorders, further follow-up of current GAPPS families and the identification of new families have to be awaited prior to including adenocarcinoma of the colon into the cancer phenotype of GAPPS. Pathological examination of colonic polyps from patients with GAPPS did not reveal any unique features, and there has been no detailed pathological review of colon cancers arising within the context of GAPPS.

Conclusion

GAPPS is a novel, autosomal dominant gastric polyposis syndrome with a significant predisposition for adenocarcinoma of the stomach, metastasis, and death. With the recent identification of APC gene promoter IB variants as its underlying genetic aberration and a phenotypic overlap with FAP and AFAP caused by the loss of the tumor suppressor APC, GAPPS has been included as the most recent addition into APC-associated disorders. Diagnosis is made by the presence of FGPs sparing the antrum and lesser curvature of the stomach, the absence of colonic and duodenal polyposis, an autosomal dominant inheritance pattern, and the exclusion of other gastric polyposis syndromes and chronic use of PPIs. Family members at risk should be screened by germline mutation testing for the presence of APC promoter IB variants to initiate timely endoscopic evaluation and surveillance. GAPPS has a to-date poorly defined natural history and the
### Table 4 Clinicopathological characteristics of previously reported families with GAPPS

<table>
<thead>
<tr>
<th>Family</th>
<th>Reference</th>
<th>APC promotor IB mutation</th>
<th>Age of youngest reported affected family member by gastric polyposis (years)</th>
<th>Gastric polyposis phenotype</th>
<th>Age of youngest reported affected family member by gastric adenocarcinoma (years)</th>
<th>Gastric adenocarcinoma</th>
<th>Colon cancer</th>
<th>Other cancers</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Worthley et al, Li et al⁶⁶</td>
<td>c.-125delA; c.-195A&gt;C</td>
<td>10</td>
<td>GAPPS</td>
<td>33</td>
<td>Yes</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>II</td>
<td>Worthley et al, Li et al⁶⁶, McDuffie et al⁵⁵</td>
<td>c.-191T&gt;C</td>
<td>21</td>
<td>GAPPS</td>
<td>50</td>
<td>Yes</td>
<td>Yes (unconfirmed)</td>
<td>Nonmelanoma skin cancer; thyroid</td>
</tr>
<tr>
<td>III</td>
<td>Worthley et al, Li et al⁶⁶</td>
<td>c.-192A&gt;G</td>
<td>34</td>
<td>GAPPS</td>
<td>34</td>
<td>Yes</td>
<td>No</td>
<td>Leukemia; brain; prostate; lung</td>
</tr>
<tr>
<td>IV</td>
<td>Yanaru-Fujisawa et al¹</td>
<td>Not reported</td>
<td>Daughter, son, nephew of 56-year-old proband</td>
<td>GAPPS</td>
<td>56</td>
<td>Yes</td>
<td>Unknown</td>
<td>Unknown</td>
</tr>
<tr>
<td>V</td>
<td>Yanaru-Fujisawa et al¹</td>
<td>Not reported</td>
<td>Daughter, niece of 42-year-old proband</td>
<td>GAPPS</td>
<td>Niece of 42-year-old proband</td>
<td>Yes</td>
<td>Unknown</td>
<td>Unknown</td>
</tr>
<tr>
<td>VI</td>
<td>Li et al⁴⁶</td>
<td>c.-191T&gt;C</td>
<td>Not reported</td>
<td>GAPPS</td>
<td>Not reported</td>
<td>No</td>
<td>Yes</td>
<td></td>
</tr>
<tr>
<td>VII</td>
<td>Li et al⁴⁶</td>
<td>c.-191T&gt;C</td>
<td>Not reported</td>
<td>GAPPS</td>
<td>Not reported</td>
<td>Yes</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>VIII</td>
<td>Li et al⁴⁶</td>
<td>c.-191T&gt;C</td>
<td>Not reported</td>
<td>GAPPS</td>
<td>Not reported</td>
<td>Yes</td>
<td>Yes</td>
<td>Prostate; three more unconfirmed cases of colon cancer</td>
</tr>
<tr>
<td>IX</td>
<td>McDuffie et al⁵⁵</td>
<td>Not detected</td>
<td>45</td>
<td>GAPPS</td>
<td>55</td>
<td>Yes</td>
<td>Yes</td>
<td>GE junction</td>
</tr>
<tr>
<td>X</td>
<td>Repak et al⁴⁷</td>
<td>c.-191T&gt;C</td>
<td>19</td>
<td>GAPPS</td>
<td>23</td>
<td>Yes</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>XI</td>
<td>Beer et al⁴⁸</td>
<td>c.-191T&gt;C</td>
<td>28</td>
<td>GAPPS</td>
<td>Not reported</td>
<td>Yes</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>XII</td>
<td>Mitsui et al⁹⁹</td>
<td>c.-191T&gt;C</td>
<td>Nice of 48-year-old proband</td>
<td>GAPPS</td>
<td>48</td>
<td>Yes</td>
<td>Unknown</td>
<td></td>
</tr>
</tbody>
</table>

**Note:** If family reported more than once, most recent and most comprehensive report was used.

**Abbreviations:** GAPPS, gastric adenocarcinoma and proximal polyposis of the stomach; GE, gastroesophageal.
rate of malignant transformation, and endoscopic surveillance in family members with gastric polyposis with high-grade dysplasia on biopsy and polyectomy specimens can miss invasive cancer. Colonoscopic surveillance is prudent while risk estimates of colon and other cancer phenotypes possibly associated with GAPPS are awaiting larger sample numbers. The creation of family registries and the formation of updated guidelines by expert consensus panels similar to the International Gastric Cancer Linkage Consortium for the screening and management of HDGC, together with enhanced education, are likely to reduce mortality in the future.

Disclosure
The author reports no conflicts of interest in this work.

References


Gastric adenocarcinoma and proximal polyposis of the stomach