Fecal carriage and phylodiversity of community-acquired \textit{bla}_\textup{TEM} Enteric bacilli in Southwest Nigeria

Paul Akinniyi Akinduti¹, Grace Iyabo Olasehinde¹, Oluwaseun Ejilude², Olugbenga Samson Taiwo¹, Yemisi Dorcas Obafemi³
¹Microbiology Unit, Department of Biological Sciences, Covenant University, Otta, Nigeria; ²Microbiology Unit, Sacred Heart Hospital, Lantoro, Abeokuta, Nigeria

Purpose: Increasing rates of clonal spread of fecal \textit{bla}_\textup{TEM} bacilli remains a huge concern to the community health with resultant high morbidity. The fecal carriage and clonal diversity of \textit{bla}_\textup{TEM} within the communities in Southwest Nigeria were surveyed.

Materials and methods: Enteric bacilli obtained from fresh fecal samples randomly collected from community residents were biotyped and profiled for antibiotic susceptibility. Resistant strains were typed for beta-lactamase, extended-spectrum beta-lactamases (ESBL), AmpC and carbapenemase production while the R-plasmid carriage was detected and mating activities were examined. The presence of \textit{bla}_\textup{TEM} gene was assayed by PCR and its phylodiversity determined with 16sRNA genomic profiling.

Results: \textit{Escherichia coli} have the highest (28.6%) occurrence rate and \textit{Pseudomonas aeruginosa} (20.5%) showing significant resistance to beta-lactamase inhibitors (ampicillin, cefuroxime and cefotaxime), and high-level multidrug resistance of more than 15.2% rate to ampicillin, cefuroxime, cefazidime, tetracycline and imipenem. \textit{E. coli} and \textit{Klebsiella oxytoca}, are the highest beta-lactamase, ESBL and AmpC producers encoded with high molecular weight R-plasmid (>11.0 kbp) and significant rate of conjugation and transformational activities. Only 2/14, 1/13 and 1/6 ESBL-type of \textit{E. coli}, \textit{K. oxytoca} and \textit{Enterobacter cloaca}, expressed \textit{bla}_\textup{TEM} gene, clustering into five different phylodiverse groups with close genomic relatedness with other bacilli.

Conclusion: This is an indication of clonal dissemination of ESBL \textit{bla}_\textup{TEM} encoded enteric bacilli having high phylodiverse characteristics through fecal carriage in the Nigerian community which requires public health education, food and environmental hygiene for its prevention.

Keywords: fecal, antibiotics, ESBL, R-plasmid, \textit{bla}_\textup{TEM}

Introduction

Increasing rates of clonal spread of beta-lactamase enteric bacilli remains a huge concern to the community health with resultant high morbidity. Fecal colonization rates with extended-spectrum beta-lactamases (ESBL)-producing isolates are becoming prevalent in most communities with poor detection making implementing epidemiological measures to curtail and control the spread remain unachievable. Nevertheless, the rate of fecal carriage among the community residents remains a public health problem which has not been addressed in a few national studies.

Fecal colonization with ESBL enteric bacilli is a risk factor for acquisition of severe gastrointestinal infection and a potential source of transmission among households. With the uncontrolled use of antibiotics in most communities in Nigeria, acquisition of resistant strains considerably accelerates the selection pressure for diversification.
and dissemination of mutant ESBL. Currently, the high prevalence of all bla genes in different communities is aided by continuous horizontal transfer of bla-encoded plasmids among clonally unrelated strains and also through local or international transfer which played a significant role in the persistence and spread of blaTEM which was not commonly found. A number of TEM derivatives have been found to reduce affinity for beta-lactamase inhibitors showing negligible hydrolytic activity against the extended-spectrum cephalosporins. However, interesting mutants of TEM beta-lactamases are now being recovered and could maintain the ability to hydrolyze third-generation cephalosporins which also demonstrate inhibitor resistance called complex mutants of TEM. Plasmid-mediated horizontal transfer of blaTEM genes has been demonstrated between poultry and humans with high conjugative mobility and transformational characteristics.

Furthermore, the increasing rates of ESBL-expression among Gram-negative bacilli characterized with multiple clonal strains have risen substantially in several communities causing focal outbreaks resulting in high mortality rate mostly in under-developed countries having sporadic dissemination. Several infection-control measures focused on reducing this spread and deadly impact of its clonal transmission yielded little or no impact in many developing countries as a result of persistent spread of high level multi-antibiotic resistant strain. Recent mutational changes resulting in phylo-diverse beta-lactamase strains are an indication of antibiotic overuse, prolonged intestinal carriage and competitive interactions of various enteric strains within a host. The available therapeutic options for the treatment of ESBL-associated enteric infections are limited due to drug resistance conferred by the blaTEM along with frequently observed co-resistance to various antibiotic classes, such as cephemycins, fluoroquinolones and aminoglycosides. A shift in the distribution of different ESBL types that show resistance to beta-lactam antibiotics among Gram-negative bacilli, usually confer resistance to carbapenems and are also known to emerge as metallo-beta-lactamases (MBL).

The recent trend in the prevalence of enteric pathogens producing blaTEM is associated with high rates of morbidity and health care costs. TEM gene mutations, which commonly spread with R-plasmids in enteric bacilli, sporadically reduce antibiotic efficacy. Therefore, this study was carried out to survey the fecal dissemination of blaTEM encoded enteric bacilli with diverse phylogenetic characteristic within the studied community.

Materials and methods
Sampling
Fresh fecal samples were randomly collected from 406 community residents who were attending the out-patient clinics. The fecal samples were transported and stored at 4°C in cold chain before analysis. For this study, approval was obtained from the Ethics committee of the Federal Medical Center, Abeokuta which also serves as referral center in Southwest, Nigeria (Ethical assign number: FMCA/470/HREC/02/2016) and also written informed consent from the individuals recruited for the study as they visited the out-patient clinics.

Culture and biotyping
Enteric bacilli were isolated from fecal samples cultured on MacConkey Agar (Oxoid CM 516, UK) and Salmonella-Shigella agar (Oxoid CM 543, UK), after incubation at 37°C for 18–24 hours. The enteric bacilli were selected according to their colony and cellular morphology and were biochemically characterized with API 24E test kit and interpreted using the manual for laboratory investigation of acute enteric infections.

Antimicrobial susceptibility testing
Susceptibility of the enteric bacteria isolates to various commonly used antibiotics was determined using disc diffusion assay with the following panel: ampicillin (10 µg), amoxicillin/clavulanic acid (20 µg/10 µg), cefotaxime (30 µg), ceftazidime (30 µg), imipenem (10 µg), cefuroxime (30 µg), gentamicin (10 µg), ciprofloxacin (5 µg), tetracycline (30 µg) and azithromycin (5 µg). Briefly, 0.5 McFarlan turbid broth of pure isolate was spread on Mueller Hinton agar and each antibiotic disc was placed at a distance of about 20 mm from each other on the inoculated agar and incubated at 37°C for 18–24 hours. The inhibition zones were measured and interpreted.

Minimum inhibitory concentration (MIC)
Resistant enteric bacilli were selected and their respective antibiotics inhibitory concentration was determined using the standard micro-dilution broth assay and the respective MIC of antibiotics to each isolate was determined for identification of multidrug resistant (MDR) enteric bacilli.

Phenotypic assay for beta-lactamase and ESBL production
All MDR strains were tested for the production of beta-lactamase using a modified method of starch-iodide paper
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acidametric method.21 The overnight broth culture of beta-lactamase producing isolates was assayed for ESBL types using the double disc method using ceftazidime (30 µg), cefotaxime (30 µg) discs, and amoxicillin-clavulanate (20/10 µg).22

Inhibitor based test for AmpC determination and Hodge test

All MDR isolates were tested for AmpC beta-lactamase inhibition on cefoxitin (30 µg) disc alone and cefoxitin (30 µg) containing boronic acid (400 µg).23 Production of carbapenemase by each strain was determined with imipenem disc (10 µg) according to the modified Hodge test (MHT) method.24

R-Plasmid profiling and mating analysis

Carriage of extra-chromosomal resistant plasmid (R-Plasmid) was investigated and profiled with alkaline lysis method25 and photo-documented with ultraviolet light to determine the DNA band sizes. Conjugative activities of the ESBL types was confirmed by mating Escherichia coli recipient strains, ATCC 25567 (penicillin-susceptible and lactose fermentation-positive strain) containing 2M calcium chloride (Oxoid). Transconjugants growing in the selection plates were subjected to double disc diffusion test method26 prescribed by Apfalter et al., (2007) to confirm the presence of ESBL resistance determinants present in the recipient isolates.

PCR assays for TEM beta-lactamase genes

Chromosomal DNA from ESBL strains were extracted by boiling colonies of bacteria emulsified in RNAse-free water at 55°C for 10 minutes, centrifuged at 12,000 rpm for 5 minutes and the supernatant was carefully removed into another clean tube as DNA template for amplification. The extracted chromosomal DNA was adjusted using Thermoscientific Nanodrop Spectrophotometer of absorbance of 260 nm filter before adding to PCR mix. Each PCR assay was carried out in a volume of 2.5 µL 10× PCR buffer, 1.5 µL 25 mM MgCl₂, 0.5 µL of 10 mM dNTPs (dATP, dGTP, dCTP, dTTP), 0.2 µL Forward primer of 2500 pmole (5-AGATCCGATCAGACAGAATA-3), 0.2 µL of Reverse primer of 2500 pmole (5-TCTAGGCTAGTACTCTGTAT-3), 0.2 µL Taq polymerase enzymes, 17.9 µL distilled water and 2.0 µL of each extracted chromosomal DNA to make final reaction volume of 25.0 µL and amplified accordingly (Table 1).

Genotypic characterization of TEM beta-lactamase strains

Extraction of chromosomal DNA of each ESBL strain was carried out using QIAamp DNA Mini Kit (Qiagen, USA) and estimated on Thermoscientific Nanodrop Spectrophotometer at an absorbance of 260 nm filter. Amplification for the 16sRNA assay was carried out in a volume of 25 µL reaction containing 2.5 µL of 10× Bioline Taq reaction buffer, 2 µL of 2.5 mM dNTPs (dATP, dGTP, dCTP, dTTP), 1 µL of 5 µM Forward primer 16sRNA-U1 (CCACGACGCCGG AATACG), 1 µL of 5 µM Reverse primer, 16sRNA-U2 (ATC GGTACC TTT TACGACTTC),28 10 ng/µL DNA template of 3 µL, 0.4 µL DNA Taq polymerase (5 unit/µL) and nuclease-free water to make up to the volume of 25 µL. The amplification reaction was carried out in 32 repeated temperature cycles in a thermal cycler block (ATC 401; Nyx Technik) at a temperature of 94°C for 5 minutes, denaturation at 94°C for 30 seconds, annealing at 56°C for 30 seconds and extension at 72°C for 45 seconds.

Amplicon sequencing and BLAST

Amplicon products were purified with QIAquick PCR purification kit (Qiagen, USA) and 6 µL diluted DNA amplicon was added to 4 µL of 16sRNA primer at a concentration of 0.8 pmol/µL and the reaction mixtures were added to Taq sequencing preparations (containing tag-oligonucleotide primer gene 16sRNA U1/lower primer 16sRNA U2). Amplification was carried out in a thermal cycler block (ATC 401; Nyx Technik) at 94°C for 5 min, 35 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 30 s, followed by a final extension at 72°C for 10 min.

Table 1 PCR conditions used for the detection of TEM beta-lactamase genes

<table>
<thead>
<tr>
<th>Target gene</th>
<th>PCR conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>bl TEM</td>
<td>Denaturation for 5 minutes at 94°C; 35 cycles of 94°C for 60 seconds, 55°C for 60 seconds, 72°C for 60 seconds; and a final extension of 72°C for 10 minutes.</td>
</tr>
</tbody>
</table>
Taq polymerase) according to the manufacturer (Applied Biosystem Company, USA) instructions and sequenced using ABI Prism Big Dye Terminator version 3.0 cycle and the products were subsequently analyzed on an ABI PRISM 3700 DNA. The nucleotide sequences were translated for molecular characterization on GeneBank (www.ncbi.org) and more than 98% similarity was considered.

**Phylogenetic analysis**

Nucleotides obtained from 16sRNA sequences from ESBL type enteric bacilli were analyzed for diversity with highly similar sequences from different geographical locations having highest percentage coverage query and identity from Gen Bank database with their repository accession numbers. Nearest phylodiversity was determined by manually aligning the ESBL type sequences with homologous sequences configured into multiple sequence alignment in MEGA 6 software version 69 (http://www.megasoftware.net) to construct dendrogram by Maximum Likelihood method as independent, unordered and equally weighted according to Fitch parsimony with 1,000 heuristic bootstrap replicates and substitution model as “p” distance.

**Data analysis**

The significance of the antibiotic profile of the isolates was analyzed with SPSS software version 20.0 using Chi-square to evaluate the differences between susceptibility, intermediate and resistance level of fecal bacilli and the significant differences by ANOVA for R-plasmid carriage, conjugants and transformants at $P<0.05$.

**Results**

**Antibiogram of isolated bacilli**

Of all the enteric isolates obtained from the community subjects, *E. coli* showed highest occurrence rate (28.6%), followed by *Pseudomonas aeruginosa* (20.5%) while *Salmonella* spp. and *Shigella* spp. were the least found isolates (Figure 1). A significant rate of 52.2%, 53% and 56% enteric bacilli showed resistant to beta-lactamase antibiotics (ampicillin, cefuroxime and cefotaxime), and low resistance to ciprofloxacin (18.8%) and gentamycin (18.2%), respectively ($P<0.001$) (Table 2). Majority of the enteric bacilli were observed to be MDR to ampicillin, cefuroxime, amoxicillin-clavulanic acid and azithromycin whereas more than 15.2% of resistant bacilli reveal high MDR profile to more than two classes of antibiotics such as ampicillin, cefuroxime, cefazidime, tetracycline and imipenem (Table 3).

**Beta-lactamase production, ESBL-types and mating activities of resistant enteric bacilli**

Comparative distribution of beta-lactamase producers were highest among *E.coli* and *Klebsiella oxytoca*, and the similar high rate was observed in ESBL assay while AmpC and carbapenamase production were low and no *Salmonella* spp. produce AmpC beta-lactamase and carbapenamase (Figure 2). In Table 4, the highest rate of 52.6% harbored high molecular weight R-plasmid (>11.0 kbp) and 2.6% express low molecular weight R-plasmid (<5 kbp) while 21.1% possessed no R-plasmid. Mating activity revealed the significant rate of 29% and 34.4% of the isolates having high plasmid weight to conjugant and transform recipient isolates effectively ($P<0.05$).

**Genotyping and clonal diversity of *bla* TEM isolates**

Genomic amplification of *bla* TEM genes recovered from ESBL type (Figure 3) showed 2/14, 1/13 and 1/6 of *E. coli*, *K. oxytoca* and *Enterobacter cloaca* expressing *bla* TEM (Figure 4), respectively. Clonal diversity of the 16sRNA gene sequence of ESBL-types carrying *bla* TEM gene recovered from community residents in Southwest Nigeria revealed cluster into five diverse phylogroups (G) with other bacilli deposited in GenBank. *E. coli* F1 from Nigerian community clustered with *Enterobacter* spp. from China, India and Portugal into phylogroup 4 (G4). High-level phylodiversity was shown by *E. coli* F2 clustering to G3 with *Klebsiella michiganensis* found in Port-Harcourt, Nigeria and *Enterobacter hormaechei* (Mexico) while similar clonal diversity was expressed...
Table 2 Antimicrobial susceptibility pattern of enteric bacilli (N=479)

<table>
<thead>
<tr>
<th>Antibiotics</th>
<th>Susceptible n (%)</th>
<th>Intermediate n (%)</th>
<th>Resistant n (%)</th>
<th>χ²</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ampicillin (10 µg)</td>
<td>103 (21.5)</td>
<td>126 (26.3)</td>
<td>250 (52.2)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Augmentin (20/10 µg)</td>
<td>198 (41.3)</td>
<td>61 (12.7)</td>
<td>220 (45.9)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cefuroxime (30 µg)</td>
<td>135 (28.2)</td>
<td>90 (18.8)</td>
<td>254 (53.0)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Azithromycin (50 µg)</td>
<td>202 (42.2)</td>
<td>101 (21.1)</td>
<td>176 (36.8)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ciprofloxacin (5 µg)</td>
<td>261 (54.5)</td>
<td>128 (26.7)</td>
<td>90 (18.8)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Imipenem (10 µg)</td>
<td>329 (68.7)</td>
<td>87 (18.2)</td>
<td>63 (13.2)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cefotaxime (30 µg)</td>
<td>118 (24.6)</td>
<td>93 (19.4)</td>
<td>268 (56.0)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tetracycline (30 µg)</td>
<td>253 (52.8)</td>
<td>164 (34.2)</td>
<td>62 (13.0)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ceftazidime (30 µg)</td>
<td>254 (53.0)</td>
<td>110 (23.0)</td>
<td>115 (24.0)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gentamycin (10 µg)</td>
<td>216 (45.1)</td>
<td>176 (36.8)</td>
<td>87 (18.2)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Note: Significant P<0.05.

Table 3 Antibiotic profile of resistant enteric isolates obtained (N=198)

<table>
<thead>
<tr>
<th>nAB</th>
<th>Antibiotic resisted</th>
<th>Rate n (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>AMP, AMC, CFX, AZT, CFX, IMP, CTX, TET, CFZ, GN</td>
<td>7 (3.5)</td>
</tr>
<tr>
<td>9</td>
<td>AMP, AMC, CFX, AZT, CFX, IMP, CTX, TET, CFZ</td>
<td>7 (3.5)</td>
</tr>
<tr>
<td>8</td>
<td>AMP, AMC, CFX, AZT, CFX, IMP, CTX</td>
<td>14 (7.1)</td>
</tr>
<tr>
<td>8</td>
<td>AMP, AMC, CFX, AZT, IMP, CTX, CFZ</td>
<td>12 (6.1)</td>
</tr>
<tr>
<td>6</td>
<td>AMP, AMC, CFX, AZT, IMP</td>
<td>10 (5.1)</td>
</tr>
<tr>
<td>5</td>
<td>AMP, AMC, CFX, AZT, CFZ</td>
<td>17 (8.6)</td>
</tr>
<tr>
<td>4</td>
<td>AMP, AMC, CFX, AZT</td>
<td>11 (5.6)</td>
</tr>
<tr>
<td>4</td>
<td>AMP, AMC, CFX, CTX, IMP, CFZ, TET, CFZ</td>
<td>30 (15.2)</td>
</tr>
<tr>
<td>3</td>
<td>AMP, AMC, CFZ</td>
<td>34 (17.2)</td>
</tr>
<tr>
<td>3</td>
<td>AMP, CFX, CTX</td>
<td>56 (28.3)</td>
</tr>
</tbody>
</table>

Abbreviations: %, percentage resisted; AMC, amoxicillin/clavulanate; AMP, Ampicillin; AZT, Azithromycin; CFX, Cefuroxime; CFZ, Ceftazidime; CFX, Ciprofloxacin; CTX, Cefotaxime; GN, Gentamicin; IMP, Imipenem; nAB, number of antibiotics resisted with MIC >32 µg/mL; TET, Tetracycline.

Figure 2 Comparative distribution of beta-lactamase, ESBL, AmpC and carbapenemase types among the multi-antibiotics resistant isolates.

Abbreviation: ESBL, extended-spectrum beta-lactamases.

Table 4 Investigation of R-Plasmid carriage and mating rate of multi-antibiotic resistant bacteria isolates (N=38)

<table>
<thead>
<tr>
<th>Number of plasmid band</th>
<th>R-plasmid carriage n (%)</th>
<th>Conjugants n (%)</th>
<th>Transformants n (%)</th>
<th>F-test</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>None</td>
<td>8 (21.1)</td>
<td>2 (5.3)</td>
<td>1 (2.6)</td>
<td>1.25</td>
<td>0.860</td>
</tr>
<tr>
<td>Low MW (&lt;5 kbp)</td>
<td>1 (2.6)</td>
<td>6 (15.8)</td>
<td>7 (18.4)</td>
<td>3.27</td>
<td>0.530</td>
</tr>
<tr>
<td>Normal MW (6-10 kbp)</td>
<td>9 (23.7)</td>
<td>4 (10.5)</td>
<td>13 (34.2)</td>
<td>2.86</td>
<td>0.020</td>
</tr>
<tr>
<td>High MW (&gt;11 kbp)</td>
<td>20 (52.6)</td>
<td>11 (29.0)</td>
<td></td>
<td>9.25</td>
<td>0.032</td>
</tr>
</tbody>
</table>

Note: P<0.05 is significantly different.

Abbreviations: kbp, kilo base pair; MW, molecular wt.

Discussion

Investigation of disseminating ESBL types is now a recognized issue in emerging global health threat due to unfavorable treatment outcome of common enteric infectious diseases particularly in several communities. High occurrence rates of MDR enteric bacilli (E. coli, Pseudomonas aeruginosa, Salmonella spp. and Shigella spp.) to beta-lactamase antibiotics (ampicillin, cefuroxime and cefotaxime) suggest an increasing prevalence of resistant strains which is gradually becoming a nightmare and a fatal challenge for the community health. Unfortunately, reports regarding community bacterial resistance to antimicrobial agents are limited in Southwest Nigeria. Therefore, the increasing trends and a recorded upsurge in resistance and prevalence of community-acquired ESBL-types is not yet clear, but associations with foodstuffs, consumption of animals fed with antibiotics and frequent contact with patients during their illness could be major factors to consider.

Comparatively, high distribution of E. coli and K. oxytoca ESBL and AmpC types is a reflection of emerging community-acquired ESBL strains which could be elicited by K. oxytoca clustered together with seven other bacilli in G2 but closely related to Klebsiella spp. and phylogroup G5 clustered E. cloaca from community subjects with Kosakonia cowani from China (Figure 5).
by self-medication, uncontrolled use of antibiotics and unhygienic eating habits in numerous poorly located restaurants and street food vendors in many communities.\textsuperscript{31} This high level production of AmpC beta-lactamases and carbapenemase by community \textit{E. coli}, \textit{K. oxytoca} and \textit{Citrobacter freundii} poses a grave danger to the residents.\textsuperscript{32,33} Furthermore, increased use of broad-spectrum antibiotics during prolonged infections usually aids resistance with severe complications and financial loss.

Fecal carriage of high molecular weight R-plasmid (>11.0 kbp) among ESBL types correlated with plasmids encoding resistance to other antibiotics such as aminoglycosides, sulfoxamides, tetracyclines and other antibiotics.\textsuperscript{34,35} The unforeseen trend in plasmid-mediated resistance via heavy plasmid coding for ESBL carriage and intestinal colonization of MDR strains to other class of antibiotics may mask the spread of these clones among the residents. The plasmid-mediated ESBL mainly confer resistance on oxyimino-\(\beta\)-lactams, such as cefotaxime, ceftazidime, ceftriaxone and aztreonam, causing selective pressure for antibiotic resistant transfer to other isolates. Conjugative property of the ESBL bacilli further confirms the role of food hawked in the community as a potential reservoir of antibiotic resistance bacteria that contained a pool of mobile genetic elements (plasmid), which are readily disseminated to other human pathogens.\textsuperscript{36} Having high degree of transformation activity, identified ESBL types could propagate spread of resistant genes at high frequency that could be promoted by constant release of naked DNA from dying bacilli which can be taken up by other isolates, integrating into the cell genome and enhancing undetected spread within the community.\textsuperscript{37}

Clonal transmission of antibiotic resistance may be not only due to dissemination of different \(\beta\)-lactamases\textsuperscript{38} but also by spread of clonal diverse variants with high potential plasmid-mediation and mating activity. The study shows only 2 of 14, 1 of 13 and 1 of 6 ESBL-type of \textit{E. coli}, \textit{K. oxytoca} and \textit{E. cloaca} expressed \(\text{bla}_{\text{TEM}}\) gene, respectively. Clonal diversity of the 16sRNA gene sequence of ESBL-types carrying \(\text{bla}_{\text{TEM}}\) gene recovered from community residents in Southwest Nigeria revealed close relatedness with other enteric pathogens from other countries. TEM-type ESBL (\(\text{bla}_{\text{TEM}}\) Gene) known to hydrolyze ampicillin at a greater rate but with less activity against extended-spectrum cephalosporins, is observed to be responsible for resistance in commonly used third generation cephalosporins in this locality.\textsuperscript{39}

Expression of \(\text{bla}_{\text{TEM}}\) gene observed in community ESBL isolates reveals resistance to ceftazidime with very high MIC values. Therefore, molecular epidemiological studies need to be carried out to prevent dissemination of this strain in order to curtail the spread of enteric infections and prevention of major outbreaks of \(\text{bla}_{\text{TEM}}\) ESBL strains in the Nigerian community. High-level phylodiversity observed among the Southwest Nigeria isolates reveals transfer of 16sRNA genotyped bacilli harboring \(\text{bla}_{\text{TEM}}\) gene which might have been transmitted from clones of different origins.\textsuperscript{3}

Frequent household contact with pets,\textsuperscript{40} animal food\textsuperscript{41} and in-door reared birds,\textsuperscript{42} as well as horizontal transfer of gene within the commensal microflora\textsuperscript{43} could necessitate clonal diversity of the \(\text{bla}_{\text{TEM}}\) ESBL types and also serve as vehicles for undetected clonal spread of antimicrobial-resistant bacilli.\textsuperscript{44} Consumption of poultry products such as birds treated with antibiotic or fed with antibiotic-supplemented feed, imported from neighboring countries into Southwest Nigeria could be implicated as source of \(\text{bla}_{\text{TEM}}\) clones circulating within the community, thereby increasing intestinal colonization and resultant carriage level of \(\text{bla}_{\text{TEM}}\) bacilli among the community residents. This poses a grave danger to the populace and the community residents, making routine surveillance of antimicrobial susceptibility difficult and formulation of public health guidelines unachievable.

\textbf{Conclusion}

Identification of \(\text{bla}_{\text{TEM}}\) carriage among enteric bacilli circulating in the Nigerian community, having high phylodiverse
characteristic needs concerted effort to prevent its dissemination and practicable public health education on food and environmental hygiene.

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Disclosure
The authors report no conflicts of interest in this work.
References


Fecal carriage of phylodiverse blaTEM bacilli


