Influence of MSI and 18q LOH markers on capecitabine adjuvant monotherapy in colon cancer patients

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Purpose: The aim of this study was to evaluate whether pretreatment analysis of selected molecular markers can be used for the prediction of disease-free survival (DFS)/overall survival (OS) of capecitabine adjuvant monotherapy in colon cancer patients.

Patients and methods: A total of 126 patients enrolled in a capecitabine Phase IV clinical trial were analyzed for microsatellite instability (MSI), 18q loss of heterozygosity (LOH), thymidylate synthase (TYMS) 5′ variable number of tandem repeat (VNTR), and methylene tetrahydrofolate reductase (MTHFR) C677T variants. The significance in predicting 5-year DFS/OS was assessed by Kaplan–Meier and Cox regression analyses.

Results: The MSI-high (MSI-H) genotype was significantly associated with DFS (HR 0.205, 95% CI 0.05–0.88, P=0.033) and OS (HR 0.208, 95% CI 0.05–0.89, P=0.035) compared to the microsatellite stable genotype. In models stratified according to clinicopathologic characteristics, the MSI-H genotype remained a positive predictive factor for DFS/OS only in patients with stage III (P=0.023) and patients with tumors localized proximally to the splenic flexure (P=0.004). Distal colon cancers with 18q LOH have a greater survival rate when treated with capecitabine than patients with stable tumors (81.3% vs 50.0%, HR for relapse 0.348, 95% CI 0.13–0.97, P=0.043). TYMS 5′ VNTR and MTHFR C677T variants were not associated with DFS or OS.

Conclusion: MSI and 18q LOH markers have the potential to be utilized in the selection of colon cancer patients eligible for capecitabine adjuvant monotherapy.

Keywords: gastrointestinal cancer, microsatellite instability, 18q allelic imbalance, prognostic marker

Introduction

Colorectal cancer (CRC) is the third most common cancer in men (746,000 cases, 10.0% of the total) and the second in women (614,000 cases, 9.2% of the total) worldwide.¹ Over the past decade, CRC mortality rates have declined in many longstanding economically developed countries as a consequence of improved screening programs and the availability of newer and more effective agents for adjuvant and palliative chemotherapies. On the other hand, this is not the case in low-resource countries of Central and Eastern Europe where CRC mortality rates continue to increase.¹,²

The current standard primary treatment of CRC is complete surgical resection of the tumor followed by systemic chemotherapy in patients with a high risk of recurrence. 5-Fluorouracil (5-FU) in combination with leucovorin (LV), with or without oxaliplatin (FOLFOX), is the most widely used treatment for an adjuvant setting of CRC.¹ All of these agents are administered intravenously (by bolus or infusion), thereby causing
significant inconvenience to patients. Xeloda® (capecitabine; Hoffmann-La Roche Ltd., Basel, Switzerland), an oral fluoroopyrimidine, is a prodrug of 5-FU. Phase III clinical trials demonstrated that capecitabine is at least as effective as bolus 5-FU/LV in terms of time to disease progression and overall survival (OS), but achieves significantly higher response rates, has improved tolerability with significantly lower incidence of stomatitis, nausea, and alopecia, and has the advantage of oral administration.4,5 Furthermore, some pharmacoeconomic analyses have highlighted that capecitabine is more cost-effective when compared with 5-FU/LV.5,7 Most cost savings are attributable to reduced administration costs and lower incidence of adverse effects, that is, hospitalization for treatment-related adverse events. The only clinical adverse event seen more commonly with capecitabine is the hand-foot syndrome, which, however, is never life threatening.4

Interindividual differences in toxicity and response to chemotherapy are currently observed in practically all available CRC treatment regimens. Thus, to improve the outcome of the treatment, efforts must be directed toward the identification of patients who are likely to respond to a specific therapy, those who will experience severe toxicities, and those who will benefit from chemotherapy in the adjuvant setting. Stage II CRC patients present with no lymph node or distant metastases and therefore have a better overall prognosis. Surgical resection is highly effective for a localized disease, but nevertheless, a significant portion of these patients develop recurrence. Currently, these high-risk patients are identified by tumors that not only penetrate the bowel wall but also show evidence of adhesion to or invasion of the surrounding structures, free perforation, or obstruction.8 Clearly, there is a need for novel predictive factors, which will be used together with nodal involvement in the recognition of stage II patients who are likely to experience relapse and therefore, in need of adjuvant treatment. Most of the studies evaluating molecular markers for the prediction of efficacy of the adjuvant therapy in CRC evaluate patients receiving either 5-FU/LV monotherapy or a combination of 5-FU/LV with irinotecan or oxaliplatin. However, there are very scarce data on capecitabine-based protocols. Identifying the subset of patients who will most probably respond to capecitabine monotherapy is imperative since oral administration is preferred by patients, is more cost-effective, and has less severe adverse events and inconveniences attributable to combinational therapies.

The main aim of this study was to evaluate whether selected molecular parameters can be used as predictive markers for disease-free survival (DFS) and OS in colon cancer patients treated with capecitabine adjuvant monotherapy. Molecular markers that were hypothesized as predictive for the efficacy of the therapy were microsatellite instability (MSI), allelic imbalance at the 18q chromosomal arm (demarcated as a loss of heterozygosity [LOH] – 18q LOH), and thymidylate synthase (TYMS) 5′ variable number of tandem repeat (VNTR) and methylene tetrahydrofolate reductase (MTHFR) C677T variants.

Materials and methods
Study design
The study was designed as a local observational open-label trial (EU PAS register number ML20122). A total of 126 patients with histologically proven stage III or high-risk stage II colon cancer were recruited between January 1, 2006 and December 31, 2009, at the University Clinic for Oncology and Radiotherapy, Skopje, Macedonia. Patients with rectal cancer, previous malignancies, previous chemotherapy or immunotherapy, moderate or severe renal impairment, clinically significant cardiovascular disease, central nervous system disorders, chronic inflammatory bowel disease, bowel obstruction or active peptic ulcer, and patients with known dihydropyrimidine dehydrogenase polymorphisms were not included in the study. The study was approved by the ethical review board of the Faculty of Medicine, Skopje (03-525; February 16, 2006) and written informed consent and a questionnaire that contained the most relevant data on family history were obtained from each patient. All recruited patients underwent surgical resection of the tumors and were treated with capecitabine adjuvant monotherapy according to the following protocol: capecitabine 1,250 mg/m², twice daily 12 hours apart (total daily dose 2,500 mg/m²) for 14 days. The treatment was repeated every 3 weeks for a total of eight cycles (24 weeks). A total of 119 (94.44%) patients completed the treatment protocol. The treatment was stopped in seven patients due to severe adverse events (diarrhea, n=1) or patient choice or frailty (n=6). After completion of the therapy, follow-up visits were scheduled every 3 months in the first year, every 6 months in the next 2 years, and once a year in the following 2 years until a total of 60 months since the completion of chemotherapy. According to the Schedule of Assessments, each follow-up visit included a blood hematology test (complete blood count), blood chemistry tests (serum creatinine, urea and electrolytes, total bilirubin, aspartate aminotransferase, alanine aminotransferase, alkaline phosphatase, total protein, and albumin), tumor marker tests (CEA and CA 19-9), and abdominal ultrasound. The final evaluation of each patient was done at the end of the trial, 60 months since the completion of treatment, or earlier in case of premature withdrawal.
**MSI and 18q LOH analysis**

Paired peripheral blood (EDTA) and formalin-fixed, paraffin-embedded (FFPE) tumor tissue samples were obtained from each patient. DNA from both samples was extracted using the QIAamp DNA Blood Mini Kit and QIAamp DNA FFPE Tissue Kit (Qiagen NV, Venlo, the Netherlands) according to the manufacturer’s protocols. MSI and 18q LOH were analyzed with multiplex fluorescent PCR followed by capillary electrophoresis on 310 or 3500 Automated Genetic Analyzers (Thermo Fisher Scientific, Waltham, MA, USA). The MSI status was determined using 10 short tandem repeat markers: BAT25, BAT26, D2S123, D5S107, D5S346, D17S250, D18S46, D18S58, D18S61, and D18S535 derived from the panel of microsatellite loci defined by the National Cancer Institute. The tumor samples were classified as MSI-high (MSI-H) if instability was present at >30% of the loci screened, MSI-low (MSI-L) if at least one but <30% of the loci showed instability, or microsatellite stable (MSS) if all loci were stable. For statistical analysis, MSI-L and MSS patients were combined in one group since tumors with low-frequency MSI are not biologically distinct from those exhibiting microsatellite stability. Four markers were used for the 18q LOH analysis: D18S46, D18S58, D18S61, and D18S535. 18q LOH was scored only in MSS tumors and was defined as the peak ratio of tumor to normal >1.35 or <0.65 for each marker. Markers with monomeric results or MSI were determined to be noninformative. Positivity for 18q LOH was defined if at least one of the four markers showed the presence of LOH. Negativity for 18q LOH was defined as the presence of at least two informative markers with the absence of LOH.

**MTHFR C677T genotyping**

MTHFR C677T polymorphism (rs1801133) was genotyped by allele discrimination PCR on a Stratagene Mx3005P (Agilent Technologies, Santa Clara, CA, USA) real-time PCR system using TaqMan® SNP genotyping assay (reference C____1202883_20; Thermo Fisher Scientific). The genotypes were determined in a reaction mix containing 20 ng DNA in a total volume of 25 µL according to the manufacturer’s recommended protocol. Positive and negative controls were included on each plate and reproducibility was checked by recommended protocol. Positive and negative controls were determined in a reaction mix containing 20 ng DNA (Agilent Technologies, Santa Clara, CA, USA) real-time PCR by allele discrimination PCR on a Stratagene Mx3005P. The genotypes for the 18q LOH analysis were: 3RC/3RC, 3RC/3RG, 3RG/3RG, and 3RG/3GC. The germline wild-type genotype was labeled 3RG/3RG. The genotype 3RC/3RG was defined as the peak ratio of tumor to normal >1.35 or <0.65 for each marker. Markers with monomeric results or MSI were determined to be noninformative. Positivity for 18q LOH was defined if at least one of the four markers showed the presence of LOH. Negativity for 18q LOH was defined as the presence of at least two informative markers with the absence of LOH.

**TYMS 5’VNTR analysis**

TYMS 5’VNTR variants were analyzed using a custom designed assay. The 5’-TSER of TYMS gene was amplified using the following primers: 5’GTG-GCTCCCTGTTTCCCC3’ (forward primer) and 6-FAM-5’GCTCCAGCCCGACACAGGCA3’ (6-FAM labeled reverse primer). A total of 100 ng of DNA was used in 25 µL final volume including 1x Hot Start Buffer B2, 1x solution S, 2 mM Mg2+, 0.2 mM of each dNTP, 1 µM of both primers, and 1 U of HOT FIREPol® DNA Polymerase (Solis BioDyne, Tartu, Estonia). PCR was performed on Veriti (Thermo Fisher Scientific) using a touchdown program: initial denaturation at 95°C for 12 minutes; 10 cycles of 1 minute at 95°C, 1.5 minutes at 75°C with 1°C reduction per cycle; 25 cycles of 1 minute at 95°C and 1.5 minutes at 65°C, and final elongation at 72°C for 10 minutes. Five microliters of the PCR product was digested with the restriction enzyme HaeIII according to the manufacturer’s instructions (New England Biolabs, Ipswich, MA, USA). The digest is used to discriminate the 3RC allele from the 3RG allele since G>C SNP in the 5’-TSER 3RC allele affects the HaeIII site, PCR and the digested product were combined in a 1:5 ratio and detected by capillary electrophoresis on a 3500 Automated Genetic Analyzer (Thermo Fisher Scientific). The results were reported as high expression alleles (2R/3G, 3C/3G, and 3G/3G genotypes) and low expression alleles (2R/2R, 2R/3C, and 3C/3C genotypes).

**Statistical analysis**

Descriptive data were tabulated as mean ± standard deviation and range for continuous variables and number (and percentage) for categorical variables. Differences in demographic and clinical characteristics of patients were tested using the unpaired Student’s t-test (continuous variables) or Fisher’s exact test (categorical variables). The primary end point of the study was DFS, defined as the time from the end of the adjuvant therapy to the first recurrence. OS, which was defined as the time from the end of the adjuvant therapy to colon cancer-specific death, was the secondary end point. In detail, relapse-free patients in this study were: 1) those who were alive and without recurrences during follow-up (5 years after therapy, N=45); 2) those who died of causes not related to colon cancer and without recurrences during follow-up (the end of follow-up is the date of death, N=45); 3) those who were alive and without recurrences, but because of personal reasons decided not to adhere to regular follow-up visits (the end of follow-up is the last follow-up control, N=24, DFS avg =45 months). Only patients who completed the treatment protocol and had follow-up data for at least 18 months were evaluated for DFS and OS (n=110). Survival curves were generated according to the Kaplan–Meier method and univariate survival distributions were compared.
with the use of the log-rank test. HR and 95% CI for univariate and multivariate models were computed with the use of Cox proportional hazards regression model. All statistical analyses were conducted using the IBM SPSS software v22.0 (IBM Corporation, Armonk, NY, USA).

Results

Disease relapse was observed in 38 patients (34.55%). The mean DFS for the relapse group was 17.92 months (95% CI 13.47–22.63) and for the relapse-free group 60.14 months (95% CI 54.39–65.44). None of the clinicopathologic variables (gender, age, tumor localization, and stage) was predictive for DFS/OS in the study group (log-rank test for DFS P-values: 0.65, 0.64, 0.31, and 0.42, respectively; data not shown).

The general characteristics of the study participants and the association of the tested markers with the clinicopathologic variables are presented in Table 1. MSI was observed in 17 patients (17.71%), whereas 18q LOH was present in 44 (55.70%) MSS patients. MSI-H was associated with localization of the tumors to a site proximal to the splenic flexure (P=0.012) (Table 1). In all other aspects, there was no statistical significance among patients exhibiting MSI-H and MSS, as well as between patients with 18q LOH and 18q intact tumors (Table 1).

Significant interaction was observed between the MSI status and benefit of treatment. The rate of 5-year DFS was 50.0% in 18q LOH patients with tumors exhibiting MSI-H (88.2%) was significantly higher than DFS in patients with MSS tumors (60.8%; P=0.032) (Table 2; Figure 1). In the multivariate analysis adjusted for age, gender, stage, and tumor localization, the MSI-H genotype was significantly associated with DFS (HR 0.205, 95% CI 0.05–0.88, P=0.033) and OS (HR 0.208, 95% CI 0.05–0.89, P=0.035) compared to the MSS genotype (Table 2). In models stratified according to clinicopathologic characteristics, the MSI-H genotype remained a positive predictive factor for DFS/OS only in patients with stage III tumors (P=0.023) and patients with tumors localized proximally to the splenic flexure (P=0.004, Table 2; Figure 2). None of the MSI-H patients with stage III or proximal tumors developed relapse.

In MSS patients, there was no significant difference in DFS (HR 0.618, 95% CI 0.30–1.29, P=0.199) or OS (HR 0.628, 95% CI 0.31–1.31, P=0.213) according to the 18q LOH status (Table 3; Figure 3). We further examined the effect of 18q LOH in strata according to tumor stage, localization, gender, and age. Patient mortality did not significantly differ according to any clinicopathologic characteristic except for tumor localization. We found that patients with 18q LOH distal colon cancer have a significantly higher survival rate than patients with chromosomal stable tumors (81.3% vs 50.0%, P=0.027, Figure 4). Multivariate analyses on the effect of 18q LOH in patients treated with capcitabine adjuvant monotherapy showed better survival only in patients with distal localization of their tumors (HR for relapse 0.348, 95% CI 0.13–0.97, P=0.043), but not in patients with proximal localization.

Table 1 Demographics and clinical characteristics of the study participants and the association of MSI and 18q LOH status with clinicopathologic variables

<table>
<thead>
<tr>
<th>All patients (N=110)</th>
<th>MSI status*</th>
<th>18q LOH status*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Patients with MSI-H tumors (N=17)</td>
<td>Patients with MSS tumors (N=93)</td>
</tr>
<tr>
<td></td>
<td>Patients with 18q LOH tumors (N=44)</td>
<td>Patients with 18q intact tumors (N=34)</td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>60 (54.6)</td>
<td>8 (15.4)</td>
</tr>
<tr>
<td>Female</td>
<td>50 (45.4)</td>
<td>9 (20.5)</td>
</tr>
<tr>
<td>Age (years)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean ± SD</td>
<td>59.5 ±9.2</td>
<td>59.3 ±9.3</td>
</tr>
<tr>
<td>Range</td>
<td>36–81</td>
<td>40–81</td>
</tr>
<tr>
<td>Tumor localization</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Proximal</td>
<td>42 (38.2)</td>
<td>11 (29.7)</td>
</tr>
<tr>
<td>Distal</td>
<td>68 (61.8)</td>
<td>6 (10.2)</td>
</tr>
<tr>
<td>Tumor stage</td>
<td></td>
<td></td>
</tr>
<tr>
<td>II</td>
<td>62 (56.4)</td>
<td>10 (19.2)</td>
</tr>
<tr>
<td>III</td>
<td>48 (43.6)</td>
<td>7 (15.9)</td>
</tr>
</tbody>
</table>

Notes: Data are presented as numbers with percentages in parentheses unless otherwise indicated. *Missing data are due to the low quality of FFPE tumor specimens (N=14). †Data are shown only for patients with MSS tumors and informative 18q LOH (N=78). Bold value indicates significance.

Abbreviations: FFPE, formalin-fixed, paraffin-embedded; LOH, loss of heterozygosity; MSI, microsatellite instability; MSS, microsatellite stable; MSI-H, MSI-high.
Table 2 Summary of statistically significant effects of MSI status on DFS and OS

<table>
<thead>
<tr>
<th>Multiple regression (adjusted for covariates)</th>
<th>Univariate analyses for survival</th>
<th>5-year survival (%)</th>
<th>Mean survival ± SD (months)</th>
<th>Hazard ratio (95% CI)</th>
<th>P-valuea</th>
<th>P-valueb</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-year survival</td>
<td>No covariates</td>
<td>86.2</td>
<td>97.81 ± 7.81</td>
<td>0.973</td>
<td>0.29</td>
<td>0.035</td>
</tr>
<tr>
<td>Univariate analyses for survival</td>
<td>adjusted for covariates</td>
<td></td>
<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Mean survival ± SD (months)</td>
<td></td>
<td>86.2 ± 7.81</td>
<td>97.81 ± 7.81</td>
<td>0.973</td>
<td>0.29</td>
<td>0.035</td>
</tr>
<tr>
<td>Hazard ratio (95% CI)</td>
<td></td>
<td>0.810 (0.50–1.31)</td>
<td>1.000</td>
<td>0.810</td>
<td>0.50</td>
<td>0.50</td>
</tr>
</tbody>
</table>

Notes: Log-rank test. aCox proportional hazards model. This test is not applicable since all cases in MSI-H stage III are censored. Bold values indicate significance.

Abbreviations: DFS, disease-free survival; MSI, microsatellite instability; MSI-H, MSI-high; MSS, microsatellite stability; NA, not applicable; OS, overall survival.

Discussion

Colon cancer arises through at least two distinct pathways of genetic instability: one involving a chromosomal instability presenting as LOH, and the other involving MSI. Patients with MSI-H tumors have distinct clinical and pathological features irrespective of the tumor origin and more favorable prognosis in terms of survival if treated with surgery alone.11–15 Regarding the relative benefits of chemotherapy among MSI-H and MSS patients, a current consensus has been reached regarding the lack of benefit of 5-FU-based therapies in patients with the MSI-H genotype.16 Several studies have shown that only patients with MSS tumors have significant survival benefit with the 5-FU-based treatment13,17–19 and that this treatment might even be harmful to patients harboring MSI-H tumors since 5-FU masks the overall good prognostic impact of the MSI-H genotype.20,21 However, the predictive utility of MSI still remains controversial given that some studies have failed to demonstrate the negative predictive impact of MSI,22–25 and some have suggested that MSI-H is strongly prognostic for DFS and OS and that patients with MSI-H tumors maintain their survival advantage in comparison with MSS patients in the presence of 5-FU.26–32 Correspondingly, our data showed that the MSI-H genotype is a positive predictive factor for DFS and OS in patients treated with capecitabine adjuvant monotherapy. The only two cases of MSI-H-positive patients with recurrence within the 5-year follow-up have similar clinicopathologic characteristics: age at diagnosis >60 years, adenocarcinoma with distal localization (sigmoid colon) and local invasion in the pericolic tissues (stage T4). In contrast, the relapse-free patients with T4 tumor stage have very different characteristics: age at diagnosis <60 years, BRAF-mutated mucinous adenocarcinoma located proximal to the splenic flexure. This suggests that the present tumor invasion in the pericolectum cannot be a sole reason for the decreased benefit of the capecitabine monotherapy, but rather the recurrence is most likely influenced by different
molecular changes that arise due to different mechanisms of MSI occurrence. This is clinically important since it gives an opportunity for further stratification of patients with the MSI-H genotype. Patients with proximal MSI-H tumors that arise from serrated adenomas have a better outcome regardless of the stage of the disease and are candidates for capecitabine monotherapy. Thus, these patients would be spared of the unnecessary burden of combination chemotherapy, that is, treatment-related toxic effects and reduced quality of life. In contrast, the positive predictive effect of the MSI-H genotype is not apparent in patients with distally located tumors treated with capecitabine monotherapy and they should be considered for the standard FOLFOX protocol. However, although this is a prospective study and all patients received the same clinical management through the treatment courses, our findings need to be interpreted with caution since the small number of patients made the subgroup analysis difficult. For example, the most probable reason for stage III MSI-H patients showing better survival than high-risk stage II MSI-H patients is the enrichment of patients with proximal tumors who have better prognosis within the stage III group due to the small sample size. Furthermore, since we did not have a control group comprising patients who did not receive adjuvant chemotherapy, it remains unclear if this association is due to the treatment alone or to the combination with the preferable prognostic impact of the MSI-H genotype. Nonetheless, it is worth noting, based on our results, that the MSI-H genotype could not be associated with a reduced survival rate in patients treated with capecitabine, as reported by some research groups,20,21 and therefore, these patients should not be excluded from 5-FU-based adjuvant therapy.

Allelic loss at the 18q chromosomal arm occurs with a great frequency in the carcinogenic process of colon cancer. This region contains several important tumor suppressor genes, such as DCC, SMAD4 (DPC4), SMAD2, and CABLES1.33–35 Given its potential use as a prognostic marker, a number of studies have evaluated the association between 18q LOH and CRC outcome. However, the results are difficult to interpret since reports are almost evenly divided between those finding that 18q LOH predicts poor survival34–40 and those showing no association.41–46 One possible explanation for the inconsistency of the association is that many of the initial studies did not consider the potential confounding effect of MSI-H, which is typically associated with improved patient survival. Because chromosomal instability and MSI are almost mutually exclusive,47 the association between 18q LOH and poor prognosis in these studies might be simply due to the enrichment of MSI-H tumors within the group of 18q stable patients. In contrast to the extensive exploration of the influence of the 18q LOH markers on the CRC patients’ prognosis, there are still limited data regarding the effect of these markers on survival in patients receiving 5-FU-based adjuvant chemotherapy. Results from these studies generally demonstrate that the retention of the

![Figure 1](https://www.dovepress.com/)

**Figure 1** Kaplan–Meier estimates for disease-free survival (A) and overall survival (B) by MSI status in patients treated with capecitabine adjuvant monotherapy. **Abbreviations:** MSI, microsatellite instability; MSI-H, MSI-high; MSI-L, MSI-low; MSS, microsatellite stable.
Markers for capecitabine monotherapy in colon cancer patients

Figure 2 Kaplan–Meier curves of disease-free survival rates according to MSI status in CRC patients receiving capecitabine adjuvant monotherapy stratified according to the stage (A, B) and localization (C, D) of the tumors.

Abbreviations: MSI, microsatellite instability; MSI-H, MSI-high; MSI-L, MSI-low; MSS, microsatellite stable; CRC, colorectal cancer.

18q alleles in MSS cancer points to a favorable outcome after adjuvant chemotherapy in colon cancer patients.\textsuperscript{35,48} The data from our study could not confirm these findings, since we did not find a statistically significant difference in survival between patients with 18q LOH and stable tumors when treated with capecitabine adjuvant monotherapy both with univariate and with multivariate analyses. However, in models stratified according to clinicopathologic features, surprisingly, we observed a positive predictive impact of 18q LOH on survival in patients with a distal localization of the tumors regardless of the tumor stage. We cannot give a definite explanation for these contradictory findings. However, all the studies (including ours) used different markers and criteria for assessing 18q LOH, which could confound the results and cause discrepant findings. Thus, it would be beneficial to develop a consensus panel of markers to assess...
<table>
<thead>
<tr>
<th>LOH Status</th>
<th>DFS(^a)</th>
<th></th>
<th></th>
<th>OS(^a)</th>
<th></th>
<th></th>
</tr>
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<tbody>
<tr>
<td></td>
<td>Univariate analyses for survival</td>
<td>Multiple regression (adjusted for covariates)</td>
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<td>Multiple regression (adjusted for covariates)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>5-year survival (%)</td>
<td>Mean survival ± SD (months)</td>
<td>(P)-value</td>
<td>Hazard ratio (95% CI)</td>
<td>5-year survival (%)</td>
<td>Mean survival ± SD (months)</td>
</tr>
<tr>
<td>18q LOH, all patients</td>
<td>70.5</td>
<td>73.5 ± 6.16</td>
<td>0.092</td>
<td>0.618 (0.30–1.29)</td>
<td>70.5</td>
<td>76.92 ± 5.40</td>
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<tr>
<td>18q stable, all patients</td>
<td>50.0</td>
<td>59.39 ± 7.78</td>
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<td></td>
<td>50.0</td>
<td>63.78 ± 6.69</td>
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<td>18q LOH, high-risk stage II</td>
<td>76.0</td>
<td>78.2 ± 7.75</td>
<td>0.444</td>
<td>0.708 (0.20–4.48)</td>
<td>76.0</td>
<td>80.97 ± 6.76</td>
</tr>
<tr>
<td>18q stable, high-risk stage II</td>
<td>62.5</td>
<td>69.0 ± 7.82</td>
<td></td>
<td></td>
<td>62.5</td>
<td>64.21 ± 6.46</td>
</tr>
<tr>
<td>18q LOH, stage III</td>
<td>63.2</td>
<td>60.91 ± 8.47</td>
<td>0.153</td>
<td>0.546 (0.20–1.46)</td>
<td>63.2</td>
<td>65.03 ± 7.21</td>
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<tr>
<td>18q stable, stage III</td>
<td>38.9</td>
<td>48.69 ± 10.65</td>
<td></td>
<td></td>
<td>38.9</td>
<td>56.61 ± 10.88</td>
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<tr>
<td>18q LOH, proximal localization</td>
<td>41.7</td>
<td>48.83 ± 12.57</td>
<td>0.719</td>
<td>1.140 (0.39–3.34)</td>
<td>41.7</td>
<td>56.00 ± 10.88</td>
</tr>
<tr>
<td>18q stable, proximal localization</td>
<td>50.0</td>
<td>52.50 ± 10.81</td>
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<td>50.0</td>
<td>56.79 ± 9.26</td>
</tr>
<tr>
<td>18q LOH, distal localization</td>
<td>81.3</td>
<td>81.53 ± 6.06</td>
<td>0.027</td>
<td>0.348 (0.13–0.97)</td>
<td>81.3</td>
<td>83.70 ± 5.26</td>
</tr>
<tr>
<td>18q stable, distal localization</td>
<td>50.0</td>
<td>60.16 ± 10.04</td>
<td></td>
<td></td>
<td>50.0</td>
<td>64.61 ± 8.52</td>
</tr>
</tbody>
</table>

Notes: \(^a\) Patients with MSI-H and noninformative 18q LOH tumors were excluded from the statistical analysis. \(^b\) Log-rank test. \(^c\) Cox proportional hazards model. Bold value indicates significance.

Abbreviations: DFS, disease-free survival; LOH, loss of heterozygosity; OS, overall survival; MSI-H, microsatellite instability-high.
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Author contributions
All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; gave final approval of the

Figure 3 Kaplan–Meier estimates for disease-free survival (A) and overall survival (B) by the 18q LOH status in patients treated with capecitabine adjuvant monotherapy. Abbreviation: LOH, loss of heterozygosity.

Figure 4 Kaplan–Meier curves of disease-free survival according to the 18q LOH status of the tumor among patients with microsatellite stable colon cancer with localization proximal (A) and distal (B) to the splenic flexure. Abbreviation: LOH, loss of heterozygosity.
version to be published; and agree to be accountable for all aspects of the work.

**Disclosure**

The authors report no conflicts of interest in this work.

**References**


