Novel vancomycin–peptide conjugate as potent antibacterial agent against vancomycin-resistant Staphylococcus aureus

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Background: Increase in vancomycin (Van)-resistant bacterial strains including vancomycin-resistant Staphylococcus aureus (VRSA) and lack of new effective antibiotics have become a formidable health problem.

Materials and methods: We designed a new conjugate composed of Van and a peptide Hecate (Hec; Van/Hec), and its potential antimicrobial activity was evaluated.

Results: Results from disk diffusion test, time-kill assay, determination of minimum inhibitory concentration (MIC), microscopy, and comet assay showed strong antimicrobial effects of Van/Hec against wild-type, methicillin-resistant Staphylococcus aureus (MRSA) and VRSA. Microscopy revealed that the exposure to Van/Hec results in disruption of bacterial cell integrity in all tested strains, which was not observed in case of Van or Hec alone.

Conclusion: Overall, we showed that the preparation of conjugates from antibiotics and biologically active peptides could help us to overcome the limitation of the use of antibiotic in the treatment of infections caused by multidrug-resistant bacteria.

Keywords: vancomycin, antibacterial, Staphylococcus aureus, antibiotic resistance, peptide
enzymes that determine the structure of peptidoglycan precursors but not by the affinity for a target enzyme because it does not interact with the biosynthetic enzymes present in cell wall but forms complexes with peptidoglycan precursors.10

Cell-penetrating peptides (CPPs), which facilitate cellular uptake or intake of different molecular equipment, have attracted a great deal of interest.12 Their ability is to form pores into the bacterial cell walls.13 The possibility of conjugation of the peptides with various nanomaterials14 or chemical activators15 offers a wide range of new opportunities for their use in medical applications. Therefore, we used antibacterial peptide Hecate (Hec; Gram+ and Gram− pathogens) for the conjugation with glycopeptide antibiotic Van, which is mostly used as a drug against Gram-positive-resistant bacterial strains.16 Hec (derivative of melittin, a component of honey bee (Apis mellifera) venom) was created from 23 amino acids (FALALHec (derivative of melittin, a component of honey bee (Apis mellifera) venom) was created from 23 amino acids (FALAL-Hec (derivative of melittin, a component of honey bee (Apis mellifera) venom) was created from 23 amino acids (FALAL-

Synthesis of Hec

Hec was synthesized using standard Fmoc solid phase synthesis on Liberty Blue peptide synthesizer (CEM, Matthews, NC, USA). The sequence and monoisotopic molecular weight of Hec were as follows: FALALKALKKKALKKALKKALKK at 64 Da, respectively. The purity of the peptide was evaluated using high-performance liquid chromatography with ultraviolet detection (HPLC–UV; ESA Laboratories Inc., Chelmsford, MA, USA). The final purity was >92%. The molecular weight of the peptide was verified by the high-performance liquid chromatography coupled with electrospray ionization quadrupole time-of-flight (HPLC–ESI–Q-TOF; Bruker Daltonik GmbH, Bremen, Germany) technique.

Conjugation of Van and Hec through 1,1′-carbonyldiimidazole (CDI)

The Van solution (0.5 mL, 2.70 mM) was mixed with Hec (0.5 mL, 1.58 mM) and a zero-length cross-linker (CDI – 6.2 μM in 20 μL dimethyl sulfoxide [DMSO]). The mixture was shaken overnight at 25 °C on Biosan Orbital Shaker (Biosan, Riga, Latvia).

Matrix-assisted laser desorption/ionization (MALDI)–time of flight mass spectrometry (TOF MS) analyses of Hec, Van, and Van/Hec

The samples were diluted in 0.1% trifluoroacetic acid (TFA), and 1 μL of each of the samples was spotted in duplicate onto the MALDI target (MTP 384 target polished steel plate; Bruker Daltonics, Bremen, Germany) and air-dried at 25 °C. Then, each spot was overlaid with 1 μL of 2,5-dihydroxybenzoic acid (DHB) or sinapic acid (SA) matrix solution (20 mg/mL) in 50% acetonitrile and 0.1% TFA and air-dried completely prior to the MALDI–TOF MS measurement on ultrafleXtreme (Bruker Daltonics, Bremen, Germany). Reflector positive mode was used for Van and Hec, and in case of Van/Hec analysis, linear positive mode was used. Each spectrum was taken in the m/z range 500–8000 Da; one sample mass spectrum was averaged from 1000 sub-spectra per spot.

Fourier transform infrared spectroscopy (FTIR)

All FTIR spectra were collected using a Nicolet iS10 FTIR spectrometer with a diamond attenuated total reflectance (ATR) attachment (Thermo Fisher Scientific, Waltham, MA, USA). The samples were supplied dropwise (5 μL) on the diamond crystal of the ATR cell first, and then, the film was measured after spontaneous evaporation of the solvent. The infrared (IR) spectra were recorded from 4000 to 650 cm⁻¹ at a resolution of 4 cm⁻¹. Each spectrum was acquired by adding together 64 interferograms. The spectra were acquired at 22°C.

Cultivation of bacterial strains

Bacterial strains (S. aureus NCTC 8511, methicillin-resistant Staphylococcus aureus [MRSA] ST239:SCCmec IIIA, VRSA, Van-resistant Enterococcus faecalis ATCC 51299, Escherichia coli NCTC 13216) were obtained from the Czech...
Collection of Microorganisms, Faculty of Science, Masaryk University, Brno, Czech Republic. Four human pathogenic MRSA were obtained from England with the cooperation of the University of Cambridge. The composition of cultivation medium was as follows: nutrient broth 13 g/L and 1000 mL sterilized MilliQ water with 18 MΩ. pH of the cultivation medium was adjusted to pH 7.4. Prior to experiments, the bacterial cultures were diluted by the cultivation medium to 

\[ \text{OD}_{600 \text{ nm}} = 0.5 \text{ McFarland}. \]

**Determination of inhibition zones using disk diffusion method**

The antimicrobial effect was analyzed using the measurement of inhibition zones by the agar microdilution method. The agar surface of Petri dish was covered with a mixture of 100 µL of overnight bacterial cultures and 3 mL of Luria–Bertani (LB) broth to get the final density of 0.5 McFarland standard. The excess volume of the mixture of the Petri dishes was aspirated. The disks (6 mm) were soaked with the appropriate concentrations of the solutions of antibiotics (lincomycin, amoxicillin, tetracycline, penicillin, erythromycin, and Van – antibiotics were tested at concentrations referred to as limits for the emergence of resistance) and antibacterial agents (Van, peptide Hec, and Van/Hec conjugate) by addition of 10 µL of the solutions. The disks were placed onto the Mueller–Hinton (MH) agar plates. The MH agar plates were protected from possible external contamination and placed in a thermostat (Tuttnauer 2450EL; Tuttnauer Ltd., Beit Shemesh, Israel) at 37 °C for 24 h. The results were evaluated the next day. The inhibition zones were measured by caliper and photographed.

**Time-kill analysis**

The Eppendorf tubes containing MH broth and antimicrobial agents with the concentration of 5 µM (Van, peptide Hec and Van/Hec conjugate) were inoculated by bacterial cultures (to a density of 0.5 McFarland), adjusted to \(1 \times 10^9\). Then, the samples were incubated for different time periods (0, 6, 12 and 24 h) in a shaker at 37 °C. Separate samples were prepared for each of the time points to be studied. The Eppendorf tubes with samples were removed from the shaker after a particular time and then serially diluted (10-fold dilutions) in sterile water to measure viable counts. In all, 100 µL of the each of the diluted samples was plated separately on blood agar and then incubated at 37 °C for 24 h. The number of colonies on blood agar plate was counted. The ultimate colony-forming units (CFUs) were the number of colonies multiplied by the dilution ratio. The experiment was repeated five times.

**Cell culture and testing of the cytotoxicity**

The PNT1A human cell line established by immortalization of normal adult prostatic epithelial cells, which were bought from Sigma-Aldrich Co., was cultured in Roswell Park Memorial Institute (RPMI)-1640 medium with 10% fetal bovine serum, supplemented with penicillin (100 U/mL) and streptomycin (0.1 mg/mL). The treatment was initiated after the cells reached ~60%–80% confluence. The cell viability was estimated using the MTT assay following the conditions used in our previous study.

**Estimation of hemocompatibility**

Fresh human blood was collected from the volunteer with signed informed consent by a venepuncture from an antecubital vein. This study was approved by the ethics committee of the University Hospital Brno (Brno, Czech Republic). Plasma and serum were removed and washed with 150 mM NaCl from the sample by centrifuging at 2000× g for 5 min three times. Red blood cells (RBCs) were diluted with PBS (pH 7.4), and subsequently, Van, Hec, and Van/Hec were added in different concentrations (20, 10, 5, and 2.5 µM) separately into the RBC solution. PBS and 0.1% Triton X-100 were used as a negative control and a positive control, respectively. The samples were mixed and incubated for 1 h at 37 °C. After that, the samples were centrifuged at 3000× g for 10 min, and the absorbance was measured at \(\lambda = 540\) nm. A percentage of hemolysis was calculated according to the following equation: 

\[ \%\text{hemolysis} = \left(\frac{(A_t - A_s)(A_{100\%} - A_s)}{A_{100\%}}\right) \times 100, \]

where \(A_t\) is the absorbance of the supernatant from the samples incubated with the Van,
Hec, or Van/Hec; \( A_{\text{r}} \) is the absorbance of the supernatant from negative control (PBS, pH 7.4); and \( A_{\text{100%}} \) is the absorbance of the supernatant of positive control (0.1% Triton X-100), which causes complete lysis of RBCs.

**Microscopy of antibacterial agents against bacteria in ambient light and live/dead cell assay**

The microscopic assay for the evaluation of live/dead bacterial cells was performed using an inverted Olympus IX 71S8F-3 fluorescence microscope (Olympus Corporation, Tokyo, Japan) equipped with Olympus UIS2 series objective LUCPlanFLN 40× (N.A. 0.6, WD 2.7–4 mm, F.N. 22) and a mercury arc lamp X-cite 12 (120 W; Lumen Dynamics, Mississauga, ON, Canada). Two fluorescent dyes were used for the live/dead cell assay: propidium iodide (PI) for staining of cells with damaged membranes and SYTO9 (Thermo Fisher Scientific) for permeating both intact and damaged membranes of the cells.20

**Single-cell gel electrophoresis (comet assay)**

The human cells were plated at a density of 10⁴ cells/well in six-well dishes (The bacterial cells were diluted at a density 0.5 McFarland.) and treated with Van, peptide Hec, and Van/Hec for 2 h. The applied concentration of Van, peptide Hec, and Van/Hec was 5 µM. As a control, 250 µM H₂O₂ was used. After harvesting (in the case of the bacterial cells, the lysozyme was added for destroying of bacterial cell wall in the concentration of 20 µg/ml; the samples were shaken together for 1 h, and after lysis reaction, the steps of the methods were followed as for the human cells). ~10 µL of the cell suspension was mixed with 75 µL of 0.8% low-melting point agarose (CLP, San Diego, CA, USA) and layered on one end of a frosted plain glass slide. Then, it was covered with a layer of the low-melting agarose (100 µL). After solidification of the gel, the slides were immersed in a lysing solution (2.5 M NaCl, 100 mM Na₂EDTA, 10 mM Tris, pH 10 containing 1% Triton X-100 and 10% DMSO) for 2 h incubation at 4 °C. A cold alkaline electrophoresis buffer was poured into the chamber and incubated for 30 min at 4 °C. The electrophoresis was carried at 4 °C for 30 min at 1.25 V/cm and 300 mA. The slides were neutralized (0.4 M Tris, pH 7.5) and then stained with ethidium bromide (EtBr; 2 µg/mL). The cells were analyzed using a fluorescence microscope (Olympus IX 71S8F-3; Olympus Corporation) and classified according to the shape of the fluorescence of the comet tail (0 [no visible tail] to 4 [significant DNA in the tail]).

**Statistical analysis**

Software Statistica (data analysis software system), version 10.0 (StatSoft, Tulsa, OK, USA) was used for data processing. The general regression model was used to analyze the differences between the measured values. To reveal differences between the cell lines, Tukey’s post hoc test within homogenous groups was used. Unless noted otherwise, \( p < 0.05 \) was considered as significant.

**Molecular characterization of VRSA**

**Preparation of samples for isolation of bacterial DNA**

The harvested bacterial pellets were resuspended in 400 µL of lysis buffer (6 M guanidine hydrochloride, 0.1 M sodium acetate). Then, the lysates were incubated for 30 min on a roller incubator at 25 °C. Subsequently, DNA was isolated from the lysates using the MagNA Pure Compact Nucleic Acid Isolation Kit following the manufacturer’s instructions on MagNA Pure Compact Instrument (Hoffman-La Roche Ltd., Basel, Switzerland).

**Detection of vanA gene**

Polymerase chain reaction (PCR) with the following set of primers designed in Primer3 online tool was used for detection of \( \text{vanA} \) gene: \( \text{vanA} \) fw 5′-CCATACAAGGTCATGGTTGAATTG-3′ and \( \text{vanA} \) rw 5′-GGGCATCATACGGGATACATC-3′. PCR amplification was performed with Mastercycler® nexus (Eppendorf, Hamburg, Germany) in a volume of 20 µL. PCR reaction mix includes the following components: 0.1 µg bacterial DNA; 1.5 mM MgCl₂; 200 µM each of dATP, dTTP, dGTP, and dCTP; 0.5 µM of each primer, and 1 unit Taq polymerase (NEB, Ipswich, MA, USA). The PCR program was carried out as follows: initial denaturation at 94°C for 2 min, 40 cycles of denaturation at 94 °C for 20 s, annealing at 55 °C for 20 s, and extension at 68 °C for 45 s, with the final extension at 68 °C for 5 min. After PCR, 650 bp amplicons were detected by gel electrophoresis and EtBr staining.

**Results and discussion**

The main objectives were to synthesize Van/Hec conjugate and test the antimicrobial effects against \( S. \) aureus and its resistant counterparts – MRSA, VRSA, and vancomycin-resistant \( Enterococcus \) faecalis (VRE). Individual parts of designated conjugate showed different antimicrobial activities depending on the susceptibility of bacterial strains. Van...
as an antibiotic has a good antimicrobial activity against *S. aureus* and MRSA but not against VRSA. The high resistance of VRSA toward Van limits its application. On the other hand, it has been reported that Hec has antimicrobial activity. Despite this, as most of CPPs, Hec has drawbacks resulting from pronounced unwanted toxicity in eukaryotic cells. We produced Van/Hec (Figure 1A) using Van and Hec as building blocks, which were conjugated through zero-length cross-linker. The Van/Hec was produced to increase the antimicrobial activity of Van through a CPP-based penetration of VRSA’s cell wall and to decrease the toxicity of the naked and linear CPP molecule. The conjugation site is most likely the carboxyl group of Van, which is similar to the product of Yarlagadda et al. The Van/Hec was characterized using MALDI–TOF MS (Figure 1B) and FTIR (Figures 1C and S1). The IR spectra of Van/Hec conjugate exhibit the presence of vibrations associated with both of the constituents. The identification of the newly formed (presumed) amide bond between Van and Hec is extremely hard because of the considerable amount of very similar functional groups (amide groups, hydroxyl groups). Despite this, the spectra are consistent with the expected product – Van/Hec, showing the vibrations attributable to either Van or Hec.

**Van resistance results in a pronounced cross-resistance to other antibiotics**

In order to obtain information about the possible Van-induced cross-resistance, we decided to test the susceptibility of established strains to selected conventional antibiotics (Figure S2A). The analysis of inhibition zones of applied antibiotics (lincomycin, amoxicillin, tetracycline, penicillin, erythromycin, and Van) revealed their pronounced antimicrobial activity only in *S. aureus* with inhibition zones between 16 and 34 mm (Figure S2Aa). In MRSA, the antibacterial effect of the antibiotics significantly decreased and only tetracycline (32 mm), Van, and amoxicillin (16 mm) retained their antimicrobial properties (Figure S2Ab). Finally, the testing of antibiotics in VRSA revealed the total absence of inhibition zones, but we can see a partial inhibition zone after the application of tetracycline on VRSA (Figure S2Ac). The values of inhibition zones are summarized in Table 1. Overall, it has to be noted that the VRSA owns superior properties, which predispose it as an exceptional model for studying novel antimicrobial agents. MICs for the selected antibiotics were studied by several other researchers, which are in good agreement with our results.

At first, we investigated the susceptibility of VRSA to Van, peptide Hec, and Van/Hec and then compared the data with the susceptibility of MRSA and *S. aureus*. As we expected, the inhibition zones showed that Van had the strongest antimicrobial activity in *S. aureus* (16 mm) and MRSA (16 mm; Figure S2Ba and b), while for VRSA, the application of Van showed no observable antimicrobial activity (Figure S2Bc). Small inhibition zones (8 mm) were found in the case of peptide Hec against *S. aureus* and MRSA strains (Figure S2Ba and b) but not in the case of VRSA (Figure S2Bc). However, a strong effect of Van/Hec was found for all the tested bacteria. The diameters of inhibition zones were 25, 24 and 12 mm for *S. aureus*, MRSA, and VRSA, respectively (Figure S2C). To validate the results, we determined the MICs of antimicrobial agents by broth microdilution method (Table 2). The MICs of Van for *S. aureus*, MRSA, and VRSA were 0.6, 5, and > 80 µM, respectively. All used bacterial strains in the study had a peptide Hec MIC higher than 80 µM, and MICs of Van/Hec for *S. aureus*, MRSA, and VRSA were 0.5, 3.5, and 5 µM respectively (Table 2). We also used VRE for testing the susceptibility against these agents. Only the Van/Hec conjugate had antimicrobial activity against VRE (Figure S3). The obtained results confirmed the successful preparation of Van/Hec as antimicrobial agents and led us to conclude that Van resistance can be suppressed by Van/Hec.

**Influence of Van/Hec on growth properties of bacteria and its undesired toxicity in eukaryotic cells (hemolysis assay)**

The effect of Van/Hec was investigated by time-kill assay against *S. aureus*, MRSA, and VRSA (Figure 2A and B). Van/Hec significantly inhibited bacterial growth of all bacterial strains when compared with the growth control. *S. aureus*, MRSA, and VRSA showed similar colony count reductions after 24 h of Van/Hec treatment (control 8 and after treatment 2 log10 in CFU/mL). The reduction in bacterial growth was > 3 log10 CFU/mL for all bacterial strains after treatment with Van/Hec, indicative of a bactericidal effect. Van inhibited the growth of *S. aureus* and MRSA, but not in the case of VRSA. We can also conclude that Hec did not have any beneficial antibacterial effect, or it is necessary to use a higher concentration (MIC > 80 µM). However, after the application of Van/Hec, the bacterial growth started to decrease even in the case of VRSA (Figure 2A and B). This confirms our presumptions that Van/Hec should have better antibacterial activity in comparison with its individual parts or other antibiotics. Finally, the antibacterial effect of Van, peptide Hec, and Van/Hec was confirmed by time-kill analysis against four pathogenic MRSA isolates, and
the effect of the agents was similar to that of MRSA from the Czech collection of microorganisms (Figure S4). The effect of the agents has also been established on VRE. The percentage of viable bacterial cells after treatment with Van/Hec confirmed its antimicrobial activity (Figure S3). These results show a significant impact of Van/Hec on weakening the bacterial resistance. The cross-resistance, which is caused by the reduction of membrane permeability to antibiotics

Table 1 Inhibition zones (mm) showing significant cross-resistance of VRSA strain

<table>
<thead>
<tr>
<th>Strain/ATB</th>
<th>Lincomycin</th>
<th>Amoxicillin</th>
<th>Tetracycline</th>
<th>Penicillin</th>
<th>Erythromycin</th>
<th>Van</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. aureus</td>
<td>22</td>
<td>18</td>
<td>34</td>
<td>18</td>
<td>20</td>
<td>16</td>
</tr>
<tr>
<td>MRSA</td>
<td>0</td>
<td>16</td>
<td>32</td>
<td>8</td>
<td>0</td>
<td>16</td>
</tr>
<tr>
<td>VRSA</td>
<td>0</td>
<td>0</td>
<td>29</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

Note: In case of tetracycline, VRSA showed a partial inhibition zone.

Abbreviations: VRSA, vancomycin-resistant Staphylococcus aureus; Van, vancomycin; S. aureus, Staphylococcus aureus; MRSA, methicillin-resistant Staphylococcus aureus.

Table 2 Comparison of antimicrobial agents using MICs determined by broth microdilution method.

<table>
<thead>
<tr>
<th>Bacterial strains/agents</th>
<th>Van/Hec (µM)</th>
<th>Hec (µM)</th>
<th>Van (µM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. aureus</td>
<td>0.5</td>
<td>&gt; 80</td>
<td>0.6</td>
</tr>
<tr>
<td>MRSA</td>
<td>3.5</td>
<td>&gt; 80</td>
<td>5</td>
</tr>
<tr>
<td>VRSA</td>
<td>5</td>
<td>&gt; 80</td>
<td>&gt; 80</td>
</tr>
</tbody>
</table>

Abbreviations: MIC, minimum inhibitory concentration; Van, vancomycin; Hec, Hecate; S. aureus, Staphylococcus aureus; MRSA, methicillin-resistant Staphylococcus aureus.
and drug inactivation or rapid efflux of the antibiotic, can be avoided using Van/Hec, which provides easier penetration and delivery of the antibiotics when conjugated to CPP. An experiment was also carried out to check the activity of the conjugate against Gram-negative bacterium E. coli (NCTC 13216). The viability of E. coli cells was studied after 24 h of the treatment with the conjugate using Multiskan EX (Thermo Fisher Scientific). The experiment was repeated five times. The result (Figure S5) indicated that 10 μM of the conjugate was able to kill almost 98% of the cells.

Furthermore, cytotoxicity of Van, Hec, and Van/Hec was evaluated using nonmalignant epithelial cell PNT1A (Figure 2C). The cell viability after the application of Van was not found for all the applied concentrations. In contrast, Hec caused significant toxic effects within the higher applied concentrations. However, similar results close to Van were shown after the applications of Van/Hec to eukaryotic cells; the toxicity was estimated to be ~10% for higher concentrations. Similarly, ambient microscopy revealed a significant increase in apoptotic features of Hec-treated cells compared to other treated cells (Figure 2E).

The hemolytic effects of Van were negligible and similar to negative control – PBS (Figure 2D). On the contrary, Hec alone induced a higher level of hemolysis (~15% after treatment with 20 μM). Such hemolytic effects are well known and described for Hec and are one of the major complications of its application in medicine. Despite this, Sanches et al described that the coupling of gallic acid at N-terminus of the peptide decreases its activity. This could be the possible explanation, not only for decreasing cytotoxicity in epithelial...
cells but also for decreased hemotoxicity. Our results showed that the conjugation pronouncedly enhances the biocompatibility of Van/Hec and its antimicrobial effects.

**Microscopic estimation of live/dead cells and damage to DNA (bacterial and human cells)**

The antibacterial effect of Van, peptide Hec, and Van/Hec after applications for *S. aureus*, MRSA, and VRSA were confirmed under ambient light illumination by an optical microscope (Figure 3). Decreasing numbers of all bacterial strains after treatment with agents were shown in comparison with those of the control groups. Van/Hec was found to rupture the cell wall of *S. aureus*, MRSA and VRSA.

The viability of the tested bacterial strains after the addition of the antimicrobial agents was further investigated using live/dead assay by fluorescence microscopy. To quantify live and dead bacterial cells, two fluorescent dyes were used. SYTO9 indicates live bacteria in green fluorescent color because it can permeate cell membrane, while PI showed dead bacteria in red fluorescent color with destroyed membrane. The bacterial strains without the application of Van, Hec, and Van/Hec were used as the control group. The staining of live cells (control group) can be observed by SYTO9 as green color. After the treatment of Van, Hec, and Van/Hec with bacterial strains, a decrease in cell viability can be seen as a red color (PI). It was confirmed that the combination of Van and peptide Hec after application on the bacteria (Figure S6) contributes to the inhibition of the growth of the bacterial cells and even for resistant strains of bacteria to Van (Figure 4A).

DNA damage of VRSA after treatment with antibacterial agents (positive control – H$_2$O$_2$ 150 µM, Van, peptide Hec, and Van/Hec) was shown by the single-cell gel electrophoresis assay (comet assay). Comet tails were formed 2 h after treatment of bacteria with agents indicating bacterial DNA damage. Normal bacterial DNA without treatment has been depicted as a negative control, which remained intact after the single cell gel electrophoresis (SCGE) assay. Damage of bacterial DNA was found in the case of each antibacterial agent for *S. aureus*, MRSA, and VRSA (Figures 4B and S7). Comet grade and index of damage for all bacterial strains were the largest (grade 4) and most visible after treatment with Van/Hec (Figures 4C and S7C and D).

Comet assay has been also done for testing genotoxicity in the human cells – PNT1A (Figure S8). The human cells were treated for 2 h with antibacterial agents and positive control (H$_2$O$_2$). Insignificant genotoxicity was found after treatment with the antibacterial agents in comparison with positive control (H$_2$O$_2$). Our results demonstrate that our designed Van/Hec is highly biocompatible with eukaryotic organisms. Therefore, the Van/Hec could be considered as a promising antimicrobial agent toward VRSA.

**Figure 3** Bacterial cells.

*Note:* (A) *S. aureus*, (B) MRSA, and (C) VRSA after applications of 5 µM antibacterial agents: (a) Van, (b) peptide Hec, and (c) Van/Hec conjugate using optical microscope in ambient light. Scale bar is 2 µm; magnification is 600×.

*Abbreviations:* *S. aureus*, *Staphylococcus aureus*; MRSA, methicillin-resistant *Staphylococcus aureus*; VRSA, vancomycin-resistant *Staphylococcus aureus*; Van, vancomycin; Hec, Hecate.
Confirmation of presence of vanA gene in VRSA

In our study, we focused on confirmation of the presence of vanA gene coding the ligase responsible for the synthesis of D-Ala-D-Lac dipeptide that seems to be essential for the development of high-level Van resistance in VRSA strains. We verified the presence of vanA gene in the VRSA strain (Figure S9). This result is in compliance with the data from MIC that showed high-level Van resistance of tested strain (MIC > 80 µM) typical for the presence of vanA gene cluster.

Conclusion

In the present experiment, we developed a new potential antimicrobial agent by conjugating a common antibiotic (Van) and a CPP peptide (Hec) linked through a zero-length linker (CDI) to overcome the limitation of the use of antibiotics in the treatment of the resistant and non-resistant S. aureus strains, including MRSA and VRSA. Results from disk diffusion test, time-kill assay, determination of MIC, microscopy, and comet assay showed strong antimicrobial effects of Van/Hec conjugate against wild-type, methicillin- and Van-resistant staphylococcal strains (MRSA and VRSA). Microscopic results revealed that the exposure to Van/Hec conjugate caused disruption of bacterial cell integrity in all the tested strains, which was not found in case of Van and Hec alone. The conjugate has been found to be nontoxic, which was confirmed by different assays on nonmalignant epithelial cells and human RBCs. Overall, Van/Hec conjugate appears to be a promising antibacterial agent, which can help us to overcome the limitation of the use of antibiotic for the treatment of the infections caused by resistant Staphylococcal strains including MRSA and VRSA.

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Disclosure

The authors report no conflicts of interest in this work.

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