

Five-year China Hospital Invasive Fungal Surveillance Net (CHIF-NET) study of invasive fungal infections caused by noncandidal yeasts: species distribution and azole susceptibility

Meng Xiao, 1,2 Sharon C-A Chen,3 Fanrong Kong,3 Xin Fan,^{2,4} ling-Wei Cheng,^{1,2} Xin Hou, 1,2 Meng-Lan Zhou, 1,2 He Wang, 1,2 Ying-Chun Xu1,2

On behalf of the China Hospital Invasive Fungal Surveillance Net (CHIF-NET) Study Group

Department of Clinical Laboratory, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences, Beijing, China; ²Beijing Key Laboratory for Mechanisms Research and Precision Diagnosis of Invasive Fungal Diseases, Beijing, China; ³Centre for Infectious Diseases and Microbiology Laboratory Services, Institute of Clinical Pathology and Medical Research, New South Wales Health Pathology, Westmead Hospital, University of Sydney, Sydney, NSW, Australia; ⁴Department of Infectious Diseases and Clinical Microbiology, Beijing Chaoyang Hospital, Beijing, China

Correspondence: Ying-Chun Xu Department of Clinical Laboratory, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences, No.1 Shuaifuyuan Street, Dongcheng District, Beijing 100730, China Tel +86 10 6915 9766 Fax +86 10 6915 9742 Email xycpumch@139.com

Sharon C-A Chen Centre for Infectious Diseases and Microbiology Laboratory Services, Institute of Clinical Pathology and Medical Research, New South Wales Health Pathology, Westmead Hospital, University of Sydney, Sydney, NSW 2145, Australia Tel +61 2 9845 6255 Fax +61 2 9893 8659 Email Sharon.Chen@health.nsw.gov.au

Purpose: In this study, we report results from a 5-year surveillance for noncandidal yeast species causing invasive infections from 65 hospitals in China.

Materials and methods: Species identification was carried out by matrix-assisted laser desorption/ionization-time of flight mass spectrometry (MALDI-TOF MS) supplemented by rDNA sequencing, and fluconazole and voriconazole susceptibilities of yeasts were determined by Clinical and Laboratory Standards Institute (CLSI) disk diffusion methods.

Results: Overall, 884 noncandidal isolates belonging to 38 species were collected. Cryptococcus neoformans was the most common (75.6%), which also comprised 96.5% of the isolates from cerebrospinal fluid (CSF) and 62.6% from blood, followed by Trichosporon asahii (6.9%) and Rhodotorula mucilaginosa (5.1%). Fluconazole susceptibility and resistant rates were 74.1% and 9.7% for C. neoformans and 81.0% and 5.2% for T. asahii. Voriconazole exhibited good activity in comparison to these two species (99.5% and 98.3% of the isolates, were susceptible). However, 100% of the R. mucilaginosa isolates were resistant to both azoles. Other noncandidal yeast species showed reduced susceptibility to fluconazole (53.3%) but most were susceptible to voriconazole (94.3%). Over the 5 years, a decrease in the proportion of fluconazole-susceptible isolates was observed for C. neoformans (90%-67%, P<0.001) and other noncandidal yeast species (91%-66%, P<0.001). Moreover, the prevalence of azole-resistant R. mucilaginosa increased from 1% to 7% (P<0.001).

Conclusion: The shift in azole susceptibilities in mainland China calls for continued surveillance for noncandidal yeasts.

Keywords: invasive fungal infections, noncandidal yeasts, epidemiology, azole susceptibility, China

Introduction

Invasive yeast infections are a major threat to patients, particularly the immunocompromised and critically ill, with high morbidity and mortality. 1-5 Although Candida species remain the major cause of such infections, noncandidal yeast species are increasingly encountered as pathogens. 1,5-8 However, knowledge of the clinical characteristics and epidemiology of these pathogens remains relatively limited. 1,6,9 Moreover, data on antifungal susceptibility profiles of noncandidal yeasts are relatively few. Even where antifungal susceptibility was performed, there are no clinical breakpoints (CBPs) established using standard broth microdilution methods to guide interpretation. 10,111 These limitations result in uncertainty in clinical management and best practice use of antifungal drugs.^{1,6}

China, as one of the most fast developing countries, also suffers from the challenges of relative lack of epidemiology and drug resistance data for invasive yeast infections.^{7,12} To close this knowledge gap and to assist clinical management, the China Hospital Invasive Fungal Surveillance Net (CHIF-NET) study was initiated in 2009, focusing on both invasive candidiasis (IC) and noncandidal infections.⁷ Up to the fifth surveillance year (2014), 65 hospitals from 27 of the 34 provinces in China participated, enabling over 8,000 yeast isolates being collected.

In this study, we summarize the overall comparative species distribution of noncandidal yeast isolates and their antifungal susceptibility to fluconazole and voriconazole as determined by the Clinical and Laboratory Standards Institute (CLSI) disk diffusion methodology. 10,13

Materials and methods Study design and isolates

The CHIF-NET study is a prospective, laboratory-based, multicenter study of invasive yeast infections.⁷ This study comprised data from August 1, 2009, to July 31, 2014, and study inclusion criteria have been described previously.⁷ Each surveillance year, all non-repetitive yeast isolates from eligible patients with invasive infections were forwarded to the central laboratory, the Department of Clinical Laboratory, Peking Union Medical College Hospital, for species confirmative identification and antifungal susceptibility testing. The study was approved by the Human Research Ethics Committee of Peking Union Medical College Hospital (S-263). The quality control strains for identification and antifungal susceptibility testing were *Candida parapsilosis* ATCC 22019 and *Candida krusei* ATCC 6258.

Species identification

All yeast isolates were identified to the species level in the central laboratory by sequencing of the fungal rDNA internal transcribed spacer (ITS) regions in year 1⁷ or by an algorithm of matrix-assisted laser desorption/ionization-time of flight mass spectrometry (MALDI-TOF MS, Vitek MS system; bioMérieux, Marcy-l'Étoile, France) supplemented by ITS sequencing.¹⁴

Antifungal susceptibility testing

Susceptibility to fluconazole and voriconazole was determined using the CLSI disk diffusion method,^{6,10} and the results were interpreted as per the CLSI M44-S3 document (for fluconazole, susceptible, ≥19 mm; susceptible dose-dependent [SDD], 15–18 mm; resistant, ≤14 mm;

for voriconazole, susceptible, \geq 17 mm; SDD, 14–16 mm; resistant, \leq 13 mm).¹³

Statistical analyses

All comparisons were performed using SPSS software version 12.0 (SPSS Inc., Chicago, IL, USA). Comparisons of continuous variables were performed by using the Mann–Whitney test, and comparisons of categorical variables were performed by using a chi-squared test or Fisher's exact test, as appropriate. A *P*-value of 0.05 was significant.

Results

Isolates and patients

Fifty-five of 65 participating hospitals submitted a total of 884 non-repetitive noncandidal yeast isolates from separate patients (the remaining 10 hospitals identified no episodes of noncandidal yeast infection cases during the study period) (Figure 1). Of the isolates, 300 (35.5%) were cultured from female patients and 544 (64.5%) from male patients. Patient age ranged from 0 to 91 years (median 58, IQR 30–44).

A total of 38 noncandidal yeast species were identified (Table 1). By genera, *Cryptococcus* species was most common (77.5% or 654/844 isolates), followed by *Trichosporon* species (74/844, 8.8%), *Rhodotorula* species (44/844, 5.2%), and other uncommon genera (<4%). At the species level, *Cryptococcus neoformans* was predominant, accounting for over 75.6% of the isolates (638/844), but *Cryptococcus gattii* was rare (7/844, 0.8%). *Trichosporon asahii* was the second commonest species (58/844, 6.9%), followed by *Rhodotorula mucilaginosa* (43/844, 5.1%), *Kodamaea ohmeri* (26/844, 3.1%), and *Saccharomyces cerevisiae* (16/844, 1.9%) (Table 1).

Although *Cryptococcus* spp. was predominant in all seven geographic regions, its frequency varied from 56.3% in northeast China to 89.4% in southwest China (Table 2). *Trichosporon* spp. was more prevalent in northwest and south China regions (frequency 21.7% and 20.9%, respectively), *Rhodotorula* spp. was more commonly seen in north and northeast regions (12.5% and 11.3%, respectively), whereas other noncandidal yeast species had a frequency of 16.3% and 15%, respectively, in the south and north China regions (Table 2).

Species distribution by hospital service

Overall, only 7.1% (60/844) and 6.8% (57/844) of the noncandidal yeast isolates were collected from emergency departments and outpatient clinics, respectively, and the majority of isolates (727/844, 86.1%) were cultured from patients in inpatient wards.

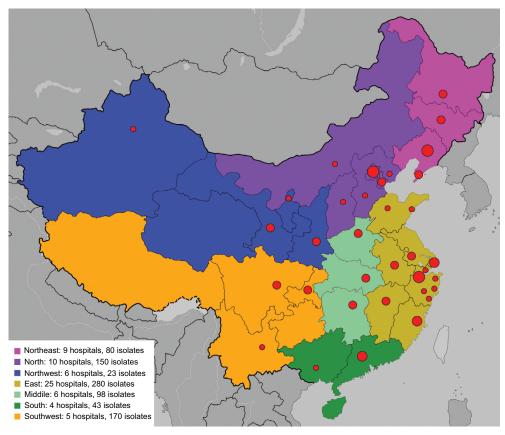


Figure I Noncandidal invasive yeast isolates collected in the five-year China Hospital Invasive Fungal Surveillance Net study.

Note: Seven geographic regions in China were labeled by different colors in the map, and locations of participant hospitals were labeled by red dots.

Of these 727 isolates, 487 (67.0%) were from inpatients in medical wards, 95 (13.1%) from surgical wards, 92 (12.7%) from patients in intensive care units (ICUs), and 53 (7.3%) from other ward types. Of note, the variation in specimen distribution among different inpatient departments largely stemmed from the proportions of the most common organism, *C. neoformans*, 67.2% (429/638) of which were isolated from medical wards. In comparison, isolate rates of other species from medical, surgical wards, and ICUs exhibited less variation (28%–32.9%).

Species distribution by specimen types

In this study, over 50% of the isolates (433/844) were cultured from cerebrospinal fluid (CSF), followed by blood (31.4%), ascitic fluid (4.1%), pus (3.7%), tissue (3%), venous catheter (2.5%), and pleural fluid (1.9%) (Table 3). The specimen distribution in noncandidal yeast infections was notably different to that in IC, among which blood samples predominated (3,858/8,829 isolates, 43.7% during the same period of time in CHIF-NET), and CSF samples only accounted for <2% (162/8,829) of the collection (Xiao M et al, unpublished data).

There was a high frequency of *Cryptococcus* spp. in CSF samples (428/433 isolates, 98.8%) (Table 3), whereas other species were rarely recovered from CSF (four isolates of *T. asahii* and one isolate of *Sporidiobolus* spp.) (Table 3). *Cryptococcus* spp. were also the most common pathogens identified in blood, pus, tissue, and pleural fluid samples (Table 3). However, non-CSF clinical samples comprised a broader range of noncandidal pathogens, with a total of 19 species identified from blood and 16 and 12 species from ascitic fluid and pus samples, respectively.

Antifungal susceptibilities

Overall, 576/844 (68.2%) isolates were susceptible to fluconazole, and 15.6% of the isolates (132/844) were fluconazole-resistant (Table 1). In comparison, voriconazole exhibited superior activity, with 93.7% (791/844) of the isolates being susceptible to the agent and resistance only occurred in 5.7% (48/844) of the cases (Table 1).

The azole susceptibilities varied between different species and for both the azoles tested. For the most common species, *C. neoformans* and *T. asahii*, 74.1% and 81.0% of the isolates were susceptible to fluconazole, respectively, while

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Table I Species distribution of noncandidal yeast isolates causing invasive infections and azole susceptibility of each species

| Species | Total | % | Antifungal susceptibility (%) | | | | | | |
|------------------------------|-------|------|-------------------------------|-------------|------|------|--------------|------|--|
| | | | Flucona | Fluconazole | | | Voriconazole | | |
| | | | S | SDD | R | s | SDD | R | |
| Cryptococcus spp. | 654 | 77.5 | 73.7 | 16.4 | 9.9 | 99.4 | 0.5 | 0.2 | |
| Cryptococcus neoformans | 638 | 76.4 | 74.1 | 16.1 | 9.7 | 99.5 | 0.5 | | |
| Cryptococcus gattii | 7 | 0.8 | 57.1 | 28.6 | 14.3 | 100 | | | |
| Cryptococcus laurentii | 4 | 0.5 | 50.0 | 25.0 | 25.0 | 100 | | | |
| Cryptococcus curvatus | 3 | 0.4 | 66.7 | 33.3 | | 100 | | | |
| Cryptococcus arboriformis | 1 | 0.1 | 100 | | | 100 | | | |
| Cryptococcus humicola | 1 | 0.1 | | | 100 | | | 100 | |
| Trichosporon spp. | 74 | 8.8 | 77.0 | 16.2 | 6.8 | 97.3 | | 2.7 | |
| Trichosporon asahii | 58 | 6.9 | 81.0 | 13.8 | 5.2 | 98.3 | | 1.7 | |
| Trichosporon mucoides | 3 | 0.4 | 66.7 | 33.3 | | 100 | | | |
| Trichosporon japonicum | 3 | 0.4 | 33.3 | 33.3 | 33.3 | 66.7 | | 33.3 | |
| Trichosporon asteroides | 3 | 0.4 | 100 | | | 100 | | | |
| Trichosporon inkin | 3 | 0.4 | 33.3 | 33.3 | 33.3 | 100 | | | |
| Trichosporon dermatis | 1 | 0.1 | 100 | | | 100 | | | |
| Trichophyton interdigitale | I | 0.1 | | 100 | | 100 | | | |
| Trichosporon jirovecii | I | 0.1 | 100 | | | 100 | | | |
| Trichosporon montevideense | 1 | 0.1 | 100 | | | 100 | | | |
| Rhodotorula spp. | 44 | 5.2 | | | 100 | | | 100 | |
| Rhodotorula mucilaginosa | 43 | 5.1 | | | 100 | | | 100 | |
| Rhodotorula diobovatum | 1 | 0.1 | | | 100 | | | 100 | |
| Other yeast spp. | 72 | 8.5 | 51.4 | 23.6 | 25.0 | 95.8 | 2.8 | 1.4 | |
| Kodamaea ohmeri | 26 | 3.1 | 38.5 | 42.3 | 19.2 | 100 | | | |
| Saccharomyces cerevisiae | 16 | 1.9 | 87.5 | | 12.5 | 93.8 | 6.3 | | |
| Dipodascus capitatus | 7 | 0.8 | 71.4 | 28.6 | | 100 | | | |
| Pichia caribbica | 5 | 0.6 | 20.0 | 60.0 | 20.0 | 80.0 | 20.0 | | |
| Arthrographis kalrae | 2 | 0.2 | | | 100 | 100 | | | |
| Aureobasidium pullulans | 1 | 0.1 | | | 100 | 100 | | | |
| Cyberlindnera rhodanensis | I | 0.1 | | | 100 | 100 | | | |
| Debaryomyces nepalensis | 1 | 0.1 | | 100 | | 100 | | | |
| Trichomonascus ciferrii | 1 | 0.1 | | | 100 | 100 | | | |
| Hanseniaspora opuntiae | I | 0.1 | 100 | | | 100 | | | |
| Kazachstania telluris | ı | 0.1 | | | 100 | 100 | | | |
| Pichia fabianii | 1 | 0.1 | 100 | | | 100 | | | |
| Pichia jadinii | I | 0.1 | 100 | | | 100 | | | |
| Pichia kluyveri | 1 | 0.1 | | | 100 | 100 | | | |
| Pichia manshurica | I | 0.1 | | | 100 | 100 | | | |
| Pichia sydowiorum | 1 | 0.1 | 100 | | | 100 | | | |
| Pseudozyma antarctica | I | 0.1 | 100 | | | 100 | | | |
| Pseudozyma spp. | 1 | 0.1 | 100 | | | 100 | | | |
| Quambalaria cyanescens | ı | 0.1 | 100 | | | 100 | | | |
| Rhodosporidiobolus fluvialis | ı | 0.1 | | | 100 | | | 100 | |
| Sporidiobolus spp. | ı | 0.1 | | | 100 | 100 | | | |
| Total | 844 | 100 | 68.2 | 16.1 | 15.6 | 93.7 | 0.6 | 5.7 | |

 $\textbf{Note:} \ \ \textbf{Bold} \ \ \textbf{data} \ \ \textbf{represented} \ \ \textbf{as summarized} \ \ \textbf{data}.$

 $\textbf{Abbreviations:} \ R, \ resistant; \ S, \ susceptible; \ SDD, \ susceptible \ dose-dependent.$

both species had susceptibility rates of >98% to voriconazole (Table 1). However, all isolates of *R. mucilaginosa* were cross-resistant to fluconazole and voriconazole (Table 1). Only around half of the other uncommon noncandidal yeast isolates (37/72, 51.4%) were susceptible to fluconazole, and one-fourth (18/72, 25.0%) were fluconazole resistant. However, these species were susceptible to voriconazole (69/72, 95.8%).

Five-year trends

Over 5 years, the frequency of isolation of *Cryptococcus* spp., *Trichosporon* spp., and other noncandidal yeast spp. varied between 73.6%–82.1%, 5.8%–11.3%, and 6.6%–10.5%, respectively, with no significant trend. However, *Rhodotorula* spp. increased significantly from 1.3% in year 1 to 7.0% in year 5 (P<0.001).

Table 2 Geographic distribution of noncandidal yeast genera in mainland China

| Geographic region | Number of isolates (%) | | | | | | |
|-------------------|------------------------|-------------------|------------------|------------------|--|--|--|
| | Cryptococcus spp. | Trichosporon spp. | Rhodotorula spp. | Other yeast spp. | | | |
| East | 230 (82.1) | 15 (5.4) | 13 (4.6) | 22 (0.1) | | | |
| Middle | 80 (81.6) | 8 (8.2) | 1 (1.0) | 9 (0.1) | | | |
| North | 106 (70.7) | 19 (12.7) | 17 (11.3) | 8 (0.1) | | | |
| Northeast | 45 (56.3) | 13 (16.3) | 10 (12.5) | 12 (0.2) | | | |
| Northwest | 16 (69.6) | 5 (21.7) | 0 (0) | 2 (0.1) | | | |
| South | 25 (58.1) | 9 (20.9) | 2 (4.7) | 7 (0.2) | | | |
| Southwest | 152 (89.4) | 5 (2.9) | I (0.6) | 12 (0.1) | | | |

Table 3 Species distribution by specimen types

| Specimen type | Number of isolates (%) | | | | | | |
|------------------------------|------------------------|-------------------|-------------------|------------------|------------------|--|--|
| | Total | Cryptococcus spp. | Trichosporon spp. | Rhodotorula spp. | Other yeast spp. | | |
| Cerebrospinal fluid | 433 (51.3) | 428 (98.8) | 4 (0.9) | | I (0.2) | | |
| Blood | 265 (31.4) | 169 (63.8) | 28 (10.6) | 34 (12.8) | 34 (12.8) | | |
| Ascitic fluid | 35 (4.1) | 9 (25.7) | 13 (37.1) | 2 (5.7) | 11 (31.4) | | |
| Pus | 31 (3.7) | 11 (35.5) | 10 (32.3) | 2 (6.5) | 8 (25.8) | | |
| Tissue | 25 (3.0) | 21 (84.0) | | I (4.0) | 3 (12.0) | | |
| Venous catheter | 21 (2.5) | 3 (14.3) | 11 (52.4) | 2 (9.5) | 5 (23.8) | | |
| Pleural fluid | 16 (1.9) | 8 (50.0) | 3 (18.8) | | 5 (31.3) | | |
| Bronchoalveolar lavage fluid | 5 (0.6) | 2 (40.0) | 2 (40.0) | | I (20.0) | | |
| Hydrarthrosis | 5 (0.6) | | 3 (60.0) | | 2 (40.0) | | |
| Peritoneal dialysate | 4 (0.5) | | | 2 (50.0) | 2 (50.0) | | |
| Bone marrow | 3 (0.4) | 2 (66.7) | | I (33.3) | | | |
| Bile | I (0.I) | I (100) | | | | | |

Cryptococcus spp. exhibited a significantly decreased fluconazole susceptibility from 90.5% in year 1 to 66.0% in year 5 (P<0.001) (Figure 2). In addition, there were no fluconazole-resistant *Trichosporon* species strains in years 1–3, but 21.4% of the strains were resistant in year 4, and 6.3% in year 5 (P<0.001) (Figure 2). Other noncandidal species also exhibited decreased susceptibility from 90.5% in year 1 to 66.0% in year 5 (P<0.001) (Figure 2).

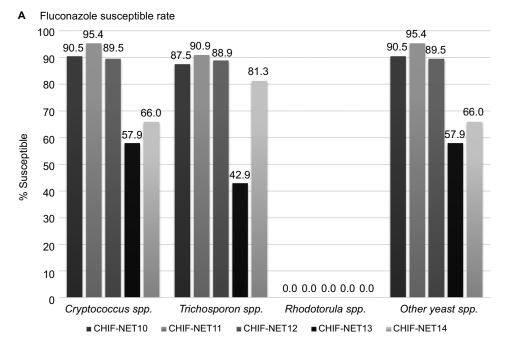
Discussion

The growing population of immunosuppressed patients and increase in medical interventions have resulted in the rise of invasive fungal infections and emergence of novel opportunistic pathogen species. 1,4,5,15 Although epidemiology and antifungal susceptibility data on IC, which account for a large proportion of invasive fungal infections, are well established, knowledge of infections caused by noncandidal yeasts remain limited in China.

Of note, the significance of antifungal resistance is well acknowledged in non-Candida albicans species and increasing trend in species distribution from C. albicans to non-C. albicans species. ^{1,16,17} However, similar issues in noncandidal yeast species have been relatively understudied, despite the fact that noncandidal yeast species may be less susceptible to

antifungal drugs. ^{1,8,18,19} One difficulty in assigning susceptibility is the absence of CBPs for noncandidal yeast species based on broth microdilution methods, ^{10,11} with epidemiologic cutoff values (ECVs) only developed for *C. neoformans*. ²⁰ In the CHIF-NET study, CLSI disk diffusion methods were employed, as interpretative criteria have been well studied and verified in the ARTEMIS global surveillance program and provide a less expensive and more flexible antifungal susceptibility testing alternative. ^{6,10,17} Disk diffusion assays have exhibited good correlation with broth microdilution methods. ^{21,22}

Of the 844 isolates collected, *C. neoformans* was the most common organism (>75%), predominating in both CNS (>98%) and bloodstream infections (~63%). In comparison, non-*C. neoformans* species, including *C. gattii*, were sporadically discovered (<1%). Although globally *Cryptococcus* spp. cause infections mainly in HIV/AIDS patients,⁵ a large proportion of cryptococcal infections occur in non-HIV infected patients in China²³ and are predominantly caused by *C. neoformans* ST5/VNI/ genotype.²⁴ Although as shown in our previous reports, *Cryptococcus* spp. remained highly susceptible to amphotericin B and 5-flucytosine (>98% of the isolates had wild-type phenotype to these two agents),²⁴ as azoles are still the mainstay of treatment for



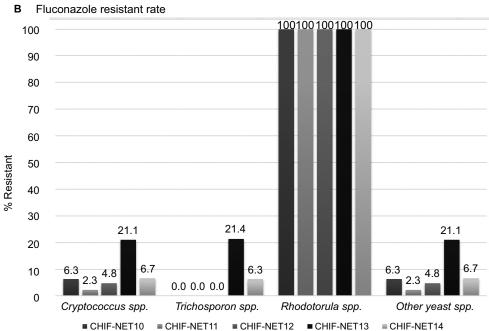


Figure 2 Trends of fluconazole susceptibility over 5 years. Notes: (A) Trends of fluconazole susceptible rate. (B) Trends of fluconazole resistant rate. Abbreviation: CHIF-NET, China Hospital Invasive Fungal Surveillance Net.

cryptococcosis, 25,26 the decreasing trend of susceptibility to fluconazole observed in this study is clinically relevant.

Trichosporon spp. was the third most common noncandidal yeast genus reported in the ARTEMIS global study,6 and in this study, the second most common. The genus can been found in the environment and is associated with summer-type hypersensitivity pneumonitis mostly reported in Japan.^{1,27} Invasive fungal infections caused by Trichosporon spp., particularly fungemia, most commonly affect patients with

hematological diseases. 1,19 Although the *Trichosporon* spp. was all formerly classified as Trichosporon beigelii, molecular assays had reclassified the genus, and T. asahii remained most common human pathogenic species. 19,27

Rhodotorula species are also emerging opportunistic pathogens, with a higher prevalence in the Asia-pacific regions (17%) than in other regions (5%-14%).6 As found in this study, fungemia is typically the predominant clinical manifestation for *Rhodotorula* infection, ²⁸ although the species can also cause central nervous system infection.²⁹ The major risk factors for *Rhodotorula* infection include patient immunosuppression and the presence of a central venous catheter.^{28,29} In addition, the genus is notable because of its intrinsic resistance to both echinocandins, fluconazole, and often to other azoles.^{1,8,29} Although susceptibility to voriconazole may be variable,¹ in this study, all *Rhodotorula* isolates were cross-resistant to voriconazole. The frequency of *Rhodotorula* spp., over 95% of which were *R. mucilaginosa*, significantly increased over 5 years.

Other noncandidal yeast species, although accounting for less than 4% of the collection, also exhibited decreased fluconazole susceptibility (susceptible rate of around 50% overall), but remaining susceptible to voriconazole. There are no robust guidelines to inform antifungal therapy for these infections. Better diagnostics coupled with surveillance data such as that from the CHIF-NET study could benefit selection of initial antifungal therapy.

As a limitation of this study, CBPs used for CLSI disk diffusion testing were not species-specific adjusted for non-candidal yeast species, as previously noted.⁶ In addition, only two azole agents were studied, as the CLSI disk diffusion methodology was only established for fluconazole and voriconazole when the CHIF-NET study was initiated.^{6,17} Although many noncandidal yeast species, eg, *Cryptococcus*, *Trichosporon*, and *Rhodotorula* species, are echinocandin-resistant, data on susceptibility profiles to a broad range of antifungal agents remain a guide to antifungal therapy testing for such susceptibility, including the newer azoles, echinocandins, amphotericin, and flucytosine, will be undertaken in the next stage of the CHIF-NET study.

Conclusion

Our surveillance provided accurate epidemiology and robust antifungal susceptibility data on noncandidal yeast causing invasive infections in China, which was useful for guiding the selection of adequate antifungal therapy. In addition, the notable trends of decreased fluconazole susceptibility in noncandidal yeast species warranted further continued surveillance and essential stewardship interventions.

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14) Qing Yang, The First Affiliated Hospital of Zhejiang University School of Medicine, Hangzhou, Zhejiang Province; 15) Hai-Feng Shao, General Hospital of Nanjing Military Area Command, Nanjing, Jiangsu Province; 16) Wen-En Liu, Hong-Ling Li, Xiangya Hospital Central South University, Changsha, Hunan Province; 17) Huo-Xiang Lv, Qu-Hao Wei, Zhejiang Province People's Hospital, Hangzhou, Zhejiang Province; 18) Yong Wang, Yan Jin, Shandong Provincial Hospital, Qingdao, Shandong Province; 19) Li-Wen Liu, The People's Hospital of Liaoning Province, Shenyang, Liaoning Province; 20) Dan-Hong Su, The First Affiliated Hospital of Guangzhou Medical Collage, Guangzhou, Guangdong Province; 21) Yu-Xing Ni, Shanghai Ruijin Hospital, Shanghai; 22) Gui-Ling Zou, Xue-Fei Du, The Fourth Affiliated Hospital of Harbin Medical University, Harbin, Heilongjiang Province; (23) Xin-Lan Hu, Ning Li, Fujian Provincial Hospital, Fuzhou, Fujian Province; 24) Ling Ma, Shuai-Xian Du, Union Hospital Tongji Medical College of Huazhong University of Science and Technology, Wuhan, Hubei Province; 25) Xiu-Lan Song, The First Hospital of Jiaxing, Jiaxing, Zhejiang Province; 26) Hua Yu, Xiang-Ning Huang, Sichuan Academy of Medical Sciences and Sichuan Provincial People's Hospital, Chengdu, Sichuan Province; 27) Tie-Li Zhou, Qing Wu, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou, Zhejiang Province; 28) Wei-Jia, Gang Li, The General Hospital Affiliated to Ningxia Medical University, Yinchuan, Ningxia Province; 29) Qiang-Qiang Zhang, Huashan Hospital, Fudan University, Shanghai; 30) Zhi-Jie Zhang, The Second Hospital Affiliated to China Medical University, Shenyang, Liaoning Province; 31) Zhi-Yong Zhang, Southwest Hospital Affiliated to the Third Military Medical University, Chongqing; 32) Rong Zhang, Hong-Wei Zhou, The Second Affiliated Hospital of Zhejiang University School of Medicine, Hangzhou, Zhejiang Province; 33) Xiu-Li Xu, Xiao Chen, Xijing Hospital, Fourth Military Medical University, Xi'an, Shaanxi Province; 34) Li-Ping Zhang, Li Yan, The First Affiliated Hospital of Chongqing Medical Hospital, Chongqing; 35) Xue-Song Xu, Wei Li, China-Japan Union Hospital of Jilin University, Changchun, Jilin Province; 36) Tie-Ying Hou, Li-Yan Zhang, Guangdong Provincial People's Hospital, Guangzhou, Guangdong Province; 37) Lin-Qiang Deng, Hui Chen, Jiangxi Province People's Hospital, Nanchang, Jiangxi Province; 38) Ke-Cheng Li, Fei Xia, Third Affiliated Hospital of Wenzhou Medical College, Wenzhou, Zhejiang Province; 39) Wei Song, Yong-Xin Shi, The Affiliated Qingdao Municipal Hospital of Qingdao University Medical College, Qingdao, Shandong Province; 40) Yuan-Hong Xu, Ji-Lu Shen, The First Affiliated Hospital of AnHui University of Science And Technology, Hefei, Anhui Province; 41) Xiao-Min Xu, Ningbo Second Hospital, Ningbo, Zhejiang Province; 42) Guo-Xiong Li, Hui Ding, Central Hospital of Lishui City, Lishui, Zhejiang Province; 43) Rong Tang, Xing Ding, Shanghai first People's Hospital, Shanghai; 44) Jian-Hong Zhao, Dong-Yan Shi, The Second Hospital of Hebei Medical University, Shijiazhuang, Hebei Province; 45) Jing Wang, Xiao-Guang Xiao, First Affiliated Hospital of Dalian Medical University, Dalian, Liaoning Province; 46) Ling Meng, Second Affiliated Hospital of Lanzhou University, Lanzhou, Gansu Province; 47) Xiao-Ming Wang, Xu-Feng Ji, The First Hospital of Jilin University, Changchun, Jilin Province; 48) Su-Fei Yu, Chun-Yan Xu, Zhejiang Taizhou Hospital, Taizhou, Zhejiang Province; 49) Qiong Zhang, Ping Ji, The First Hospital of Xinjiang Medical University, Urumchi, Xinjiang Province; 50) Long-Hua Hu, Bai-Ling Zhang, The Second Affiliated Hospital of Nanchang University, Nanchang, Jiangxi Province; 51) Bin Yang, Yu-Lan Lin, First Affiliated Hospital of Fujian Medical University, Fuzhou, Fujian Province; 52) Jin-E Lei, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an, Shaanxi Province; 53) Hai-Bin Wang, Jing Zhu, First Affiliated Hospital of PLA General Hospital, Beijing; 54) Hong-Jie Liang, The First Affiliated Hospital of Guangxi Medical University, Nanning, Guangxi Province; 55) Xiao-Ling Ma, Huai-Wei Lu, Anhui Provincial Hospital, Hefei, Anhui Province; 56) Wen-Cheng Xue, General Hospital of Shenyang Military Area, Shenyang, Liaoning Province; 57) Bin Shan, Yan Du, The First Affiliated Hospital, Kunming Medical University, Kunming, Yunnan Province; 58) Xiang-Yang Chen, People's Hospital of Zhengzhou, Zhengzhou, Henan Province; 59) Run-Mei Zhang, Jian-Bang Kang, The Second Hospital of Shanxi Medical University, Taiyuan, Shanxi Province; 60) Jian-Lei Zhang, Tianjin First Center Hospital, Tianjin; 61) Wei Cao, The Second Xiangya Hospital of Central South University, Changsha, Hunan Province; 62) Jun-Lin Zhang, Quang Fu, The Affiliated Hospital of Inner Mongolia Medical University, Hohhot, Inner Mongolia Province; 63) Yan-Ping Fan, Dalian Municipal Central Hospital, Dalian, Liaoning Province; 64) Lian-Hua Wei, Feng-Mei Zou, Gansu Province People's Hospital, Lanzhou, Gansu Province; and 65) Yan-Yan Guo, Tangshan Gongren Hospital, Tangshan, Hebei Province. 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Author contributions

All authors contributed toward data analysis, drafting, and revising the study and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

References

- 1. Miceli MH, Díaz JA, Lee SA. Emerging opportunistic yeast infections. *Lancet Infect Dis*. 2011;11(2):142–151.
- Schwartz S, Kontoyiannis DP, Harrison T, Ruhnke M. Advances in the diagnosis and treatment of fungal infections of the CNS. *Lancet Neurol*. 2018;17(4):362–372.
- Pande A, Non LR, Romee R, Santos CAQ. Pseudozyma and other non-Candida opportunistic yeast bloodstream infections in a large stem cell transplant center. Transplant Infectious Disease. 2017;19(2): e12664.
- Kullberg BJ, Arendrup MC. Invasive Candidiasis. N Engl J Med. 2015;373(15):1445–1456.
- Limper AH, Adenis A, Le T, Harrison TS. Fungal infections in HIV/ AIDS. Lancet Infect Dis. 2017;17(11):e334–e343.
- Pfaller MA, Diekema DJ, Gibbs DL, et al. Results from the ARTEMIS DISK Global Antifungal Surveillance Study, 1997 to 2007: 10.5-year analysis of susceptibilities of noncandidal yeast species to fluconazole and voriconazole determined by CLSI standardized disk diffusion testing. J Clin Microbiol. 2009;47(1):117–123.
- Wang H, Xiao M, Chen SC, et al. *In vitro* susceptibilities of yeast species to fluconazole and voriconazole as determined by the 2010 National China Hospital Invasive Fungal Surveillance Net (CHIF-NET) study. *J Clin Microbiol*. 2012;50(12):3952–3959.
- Spiliopoulou A, Anastassiou ED, Christofidou M. *Rhodotorula* fungemia of an intensive care unit patient and review of published cases. *Mycopathologia*. 2012;174(4):301–309.
- Pfaller MA, Diekema DJ. Rare and emerging opportunistic fungal pathogens: concern for resistance beyond *Candida albicans* and *Aspergillus fumigatus*. J Clin Microbiol. 2004;42(10):4419–4431.

- CLSI. M60. Performance Standards for Antifungal Susceptibility Testing of Yeasts, First Edition. Wayne, PA: Clinical and Laboratory Standards Institute (CLSI); 2018:28.
- EUCAST. Antifungal Agents, Breakpoint Tables for Interpretation of MICs, Version 8.1. 2017. Available from: http://www.eucast.org/clinical_breakpoints/. Accessed September 13, 2018.
- 12. Guo F, Yang Y, Kang Y, et al. Invasive candidiasis in intensive care units in China: a multicentre prospective observational study. *J Antimicrob Chemother*. 2013;68(7):1660–1668.
- CLSI. M44-S3. Zone Diameter Interpretive Standards, Corresponding Minimal Inhibitory Concentration (MIC) Interpretive Breakpoints, and Quality Control Limits for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Third Informational Supplement. Wayne, PA: Clinical and Laboratory Standards Institute (CLSI); 2011.
- 14. Zhang L, Xiao M, Wang H, et al. Yeast identification algorithm based on use of the Vitek MS system selectively supplemented with ribosomal DNA sequencing: proposal of a reference assay for invasive fungal surveillance programs in China. J Clin Microbiol. 2014;52(2):572–577.
- Borman AM, Linton CJ, Miles SJ, Johnson EM. Molecular identification of pathogenic fungi. *J Antimicrob Chemother*. 2008;61(Suppl 1):i7–i12.
- Perlin DS, Rautemaa-Richardson R, Alastruey-Izquierdo A. The global problem of antifungal resistance: prevalence, mechanisms, and management. *Lancet Infect Dis*. 2017;17(12):e383–e392.
- Pfaller MA, Diekema DJ, Gibbs DL, et al. Results from the ARTEMIS DISK Global Antifungal Surveillance Study, 1997 to 2007: a 10.5-year analysis of susceptibilities of *Candida* Species to fluconazole and voriconazole as determined by CLSI standardized disk diffusion. *J Clin Microbiol*. 2010;48(4):1366–1377.
- Denning DW. Echinocandin antifungal drugs. *Lancet*. 2003;362(9390):1142-1151.
- de Almeida Júnior JN, Hennequin C. Invasive *Trichosporon* Infection: a Systematic Review on a Re-emerging Fungal Pathogen. *Front Microbiol*. 2016;7:1629.
- Pfaller MA, Castanheira M, Diekema DJ, Messer SA, Jones RN. Wildtype MIC distributions and epidemiologic cutoff values for fluconazole, posaconazole, and voriconazole when testing *Cryptococcus neoformans* as determined by the CLSI broth microdilution method. *Diagn Microbiol Infect Dis*. 2011;71(3):252–259.

- 21. Espinel-Ingroff A, Canton E, Gibbs D, Wang A. Correlation of Neo-Sensitabs tablet diffusion assay results on three different agar media with CLSI broth microdilution M27-A2 and disk diffusion M44-A results for testing susceptibilities of Candida spp. and *Cryptococcus neoformans* to amphotericin B, caspofungin, fluconazole, itraconazole, and voriconazole. *J Clin Microbiol*. 2007;45(3):858–864.
- Xiao M, Fan X, Chen SC, et al. Antifungal susceptibilities of *Candida glabrata* species complex, *Candida krusei*, *Candida parapsilosis* species complex and *Candida tropicalis* causing invasive candidiasis in China:

 year national surveillance. *J Antimicrob Chemother*. 2015;70(3): 802–810.
- Feng X, Yao Z, Ren D, Liao W, Wu J. Genotype and mating type analysis
 of *Cryptococcus neoformans* and *Cryptococcus gattii* isolates from
 China that mainly originated from non-HIV-infected patients. *FEMS Yeast Res*. 2008;8(6):930–938.
- 24. Fan X, Xiao M, Chen S, et al. Predominance of *Cryptococcus neoformans* var. *grubii* multilocus sequence type 5 and emergence of isolates with non-wild-type minimum inhibitory concentrations to fluconazole: a multi-centre study in China. *Clin Microbiol Infect*. 2016;22(10):887. e1–887.e9.
- Chen SC, Sorrell TC, Chang CC, Paige EK, Bryant PA, Slavin MA. Consensus guidelines for the treatment of yeast infections in the haematology, oncology and intensive care setting, 2014. *Intern Med J.* 2014;44(12b):1315–1332.
- Arendrup MC, Boekhout T, Akova M, Meis JF, Cornely OA, Lortholary O; European Society of Clinical Microbiology and Infectious Diseases Fungal Infection Study Group, European Confederation of Medical Mycology. ESCMID and ECMM joint clinical guidelines for the diagnosis and management of rare invasive yeast infections. Clin Microbiol Infect. 2014;20(Suppl 3):76–98.
- Sugita T, Ikeda R, Nishikawa A. Analysis of *Trichosporon* isolates obtained from the houses of patients with summer-type hypersensitivity pneumonitis. *J Clin Microbiol*. 2004;42(12):5467–5471.
- Tuon FF, Costa SF. Rhodotorula infection. A systematic review of 128 cases from literature. Rev Iberoam Micol. 2008;25(3):135–140.
- Tsiodras S, Papageorgiou S, Meletiadis J, et al. *Rhodotorula mucilagi-nosa* associated meningitis: A subacute entity with high mortality. Case report and review. *Med Mycol Case Rep.* 2014;6:46–50.

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