Chronic postcataract endophthalmitis caused by *Penicillium* species in an immunocompetent patient

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Objective: *Penicillium* species are abundant, generally saprophytic, and widely distributed in the environment. On rare occasions, they cause endophthalmitis in immunocompromised patients or in immunocompetent patients with uncontrolled diabetic mellitus or immunosuppressive agents. We report a case of endophthalmitis caused by *Penicillium* species in an immunocompetent patient without any disorders.

Case presentation: A 67-year-old woman presented with ocular pain, hyperemia, and blurred vision in her right eye. She was in an immunocompetent condition and had received cataract surgery without complications 3 months ago. Severe cell infiltration and a clump of 3 mm diameter were found in the anterior chamber of the right eye, and there was no abnormality in the posterior segment. Serum β-d-glucan was not elevated; bacterial culture, histological analysis, and PCR using aqueous humor did not detect any pathogen (28S rDNA $2.4 \times 10^2$ copies/mL). However, because the clinical findings suggested fungal endophthalmitis, antifungal medication was initiated. The inflammation was gradually resolved with contraction of the clump; however, it recurred 47 days after the initiation by discontinuation of antifungal medication. Examination of culture identified *Penicillium* species in samples from both the anterior chamber and the vitreous.

Conclusion: Chronic postoperative endophthalmitis by *Penicillium* species can occur in an immunocompetent patient without any systemic diseases.

Keywords: endophthalmitis, immunocompetent, *Penicillium*, postcataract

Introduction

*Penicillium* species are abundant, generally saprophytic, and widely distributed in the environment.¹,² They rarely cause disease in humans, although they are increasingly recognized as a potential pathogen in immunocompromised hosts.³ In the eye, *Penicillium* species can cause infectious keratitis,⁴ but rarely is endophthalmitis developed, even in patients who are immunocompromised or using an immunosuppressive agent.⁵ Exogenous fungal endophthalmitis accounts for ~10% of cases of postoperative endophthalmitis and mainly occurs in immunosuppressed patients.⁶ *Candida*, *Aspergillus*, and *Fusarium* species are the most common culture-positive isolates from such patients.⁷ We report a case of chronic postoperative endophthalmitis after cataract surgery caused by a filamentous fungus, *Penicillium* species, in an immunocompetent patient without any systemic diseases.

Case report

A 67-year-old woman was referred to our department with ocular pain, hyperemia, and blurred vision in the right eye. Cataract surgery had been performed for both eyes 3 months ago. Severe cell infiltration and a clump of 3 mm diameter were found in the anterior chamber of the right eye, and there was no abnormality in the posterior segment. Serum β-d-glucan was not elevated; bacterial culture, histological analysis, and PCR using aqueous humor did not detect any pathogen (28S rDNA $2.4 \times 10^2$ copies/mL). However, because the clinical findings suggested fungal endophthalmitis, antifungal medication was initiated. The inflammation was gradually resolved with contraction of the clump; however, it recurred 47 days after the initiation by discontinuation of antifungal medication. Examination of culture identified *Penicillium* species in samples from both the anterior chamber and the vitreous.
in a clinic at 3 months ago. She had no history of systemic disease and was in an immunocompetent condition. At first presentation, best-corrected visual acuity (BCVA, measured with a standard Japanese visual acuity chart and converted to Snellen visual acuity) and intraocular pressure (IOP) in the right eye were 20/20 and 17 mmHg, but slit-lamp biomicroscopy revealed a white clump of 3 mm diameter, anterior inflammatory cells, and an iris posterior synechia in the anterior segment (Figure 1A and B). There was no inflammation in the posterior segment (Figure 1C). Serum examination showed a slight increase in white blood cells (8,600/mm³) but not in β-D-glucan (9 pg/dL) or inflammatory proteins. Bacterial culture of the anterior humor and histological analysis of the white clump did not detect the cause of the endophthalmitis. As well, increase of 18S/28S rDNA (18S rDNA, not detected; 28S rDNA 2.4×10² copies/mL) was not presented by broad-range PCR using the aqueous humor. Because the white clumps in the anterior chamber strongly suggested fungal endophthalmitis, treatment with systemic and topical antifungal agents (systemic fosfluconazole [504 mg/d, 4 days] and topical pimaricin ophthalmic ointment [4 times/day]) was initiated in addition to antibacterial treatment (systemic cefepime [1,000 mg/d, 4 days] and topical moxifloxacin hydrochloride [6 times/day], vancomycin eyedrops [6 times/day], and ceftazidime eyedrops [6 times/day]). These medications were tapered according to the remission of the clump and inflammation in the anterior segment. At 14 days, the endophthalmitis was resolved (Figure 1D) and the subjective symptoms had disappeared.

On day 47 after the initiation of treatment, she visited our hospital for headache, recurrence of ocular pain, hyperemia, and blurred vision in the right eye. BCVA was decreased to hand motion, and IOP was elevated to 40 mmHg in the right eye. Slit-lamp biomicroscopy showed reappearance of a white clump in the anterior segment, anterior inflammatory cells, large iris nodules, fibrin formation, and a posterior synechia (Figure 2A). The posterior segment was not visible due to inflammation in the anterior segment. Serum examination did not present an increase in β-D-glucan. Vitrectomy was emergently performed, and the aqueous humor and vitreous humor samples were obtained. Examination of microbial culture using samples from both the anterior chamber and the vitreous identified *Penicillium* species. We diagnosed fungal endophthalmitis caused by *Penicillium* species, and treatment with systemic micafungin (100 mg/d, 9 days), oral voriconazole (200 mg/d, 9 days) and flucytosine (6,000 mg/d, 9 days), and voriconazole eyedrop (4 times/day) was initiated. The ocular inflammation by fungal endophthalmitis was resolved (Figure 2B and C). The subjective symptoms disappeared (Figure 2D). The examination of the anterior segment revealed no white mass (Figure 2E). The posterior segment showed no inflammation (Figure 2F).

Figure 1 Photographs of the right eye on days 1 and 17.
Notes: Slit-lamp photographs (A, B, D) and a fundus photograph (C). On the first visit, a white clumped mass (white arrow head) (A), iris synechia (yellow arrow head), and inflammatory cells were seen in the anterior chamber (B), and no inflammation was seen in the posterior segment (C). After treatment, the anterior chamber showed no inflammation or white mass (D).
endophthalmitis was gradually resolved with contraction of the clump after 14 days (Figure 2B), although the iris nodules remained. BCVA was improved to 20/16, and IOP was decreased to 16 mmHg. No recurrence was found 16 months after the second treatment.

Informed consent
Written informed consent has been provided by the patient for the publication of her case details and images.

Discussion
Postoperative fungal endophthalmitis shows a variable presentation, ranging from localized in the anterior chamber to more diffuse intraocular inflammation. The present case had a prolonged latency period, as in previous reports of fungal endophthalmitis, and followed these trends with diffuse anterior segment inflammation. The presence of iris nodules, as shown in this case, has been also documented in other cases of inflammation with fungal etiologies, including Candida, Fusarium, and Coccioidoides species. Therefore, endophthalmitis by Penicillium species was suspected due to the clinical findings along with the latency period.

Since Penicillium species are sometimes encountered in the clinical laboratory as culture contaminants, diagnosis is established by histological demonstration of fungus-invaded tissue or PCR. In this case, biopsy of the iris nodules and broad-range PCR for 18S/28S rDNA were performed, as in previous reports. However, these methods could not detect Penicillium infection. We diagnosed Penicillium species infection from the results of microbial cultures using both the anterior humor and the vitreous samples and the efficacy of antifungal agents. We set the positive threshold value in PCR for 18S/28S rDNA because a trace amount of contamination can be detected.

Fungal contamination sometimes detected by elevation of 28S rDNA by PCR using the intraocular fluids. Although 2.4x10^2 copies/mL of 28S rDNA was detected by PCR using the aqueous humor in this case, we determined the result was negative based on these reports. However, it should be finally considered as positive.

Treatment of fungal endophthalmitis is difficult; no clear protocols exist because of the lack of well-controlled double-blind studies and the low incidence of the diseases. In the present case, systemic fosfluconazole and topical pimaricin ophthalmic ointment did not lead to complete remission, whereas systemic flucytosine indicated a splendid effect. For Penicillium infection in HIV patients, antifungal medication including flucytosine is selected, and flucytosine is highly active against Penicillium species. It has been also reported that flucytosine penetrates into vitreous with higher concentration than other antifungal agents, although the eye is a protected compartment and penetration of systemically administered antifungal agents are highly variable, particularly in the posterior segment.

Conclusion
This is a rare case of postoperative fungal endophthalmitis caused by Penicillium species in an immunocompetent patient without any systemic diseases. In these cases, if examination of microbial culture or PCR for 18S/28S rDNA using intraocular fluids are negative, antifungal treatment based on the clinical findings should be performed, which leads to the diagnosis.

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Disclosure
The authors report no conflicts of interest in this work.

References