A new SERS strategy for quantitative analysis of trace microalbuminuria based on immunorecognition and graphene oxide nanoribbon catalysis

Guqing Wen1,2,*
Qi Jing1,2,*
Aihui Liang1,2
Zhiliang Jiang1,2

1Key Laboratory of Ecology of Rare and Endangered Species and Environmental Protection (Guangxi Normal University), Ministry of Education, Guangxi Key Laboratory of Environmental Pollution Control Theory and Technology, Guilin, China; 2Department of Environment Science, College of Environment and Resources, Guangxi Normal University, Guilin, China

*These authors contributed equally to this work

Background: Microalbuminuria (mAlb) detection is essential for the diagnosis and prognosis of nephrotic patients and hypoproteinemia. In this article, we develop a new surface-enhanced Raman scattering (SERS) quantitative analysis method to detect mAlb in urine.

Methods: Combined the mAlb immunoreaction with gold nanoreaction of graphene oxide nanoribbons (GONR)-HAuCl4-H2O2, and used Victoria blue B (VBB) as molecular probe with a SERS peak at 1,615 cm⁻¹, a new SERS strategy for quantitative analysis of trace mAlb in urine was established.

Results: The linear range of SERS quantitative analysis method is from 0.065 to 2.62 ng/mL, with a detection limit of 0.02 ng/mL. The SERS method was applied to analysis of mAlb in urine with good accuracy and reliability, the relative standard deviation is 0.49%–2.28% and the recovery is 96.9%–109.8%.

Conclusion: This study demonstrated that the new SERS quantitative analysis method is of high sensitivity, good selectivity and simplicity. It has been applied to analysis of mAlbs in urine, with satisfactory results.

Keywords: graphene oxide nanoribbon, nanocatalysis, microalbumin immunoreaction, gold nanoreaction, SERS

Introduction

Nanoparticles in solutions not only have novel surface nanoplasmon effect but also exhibit high catalytic activity as natural enzymes and mimic enzymes with stabilization and economic characteristics, and it has been used in different fields such as materials science, physics, chemistry, biology, and environmental science.¹–⁴ In analytical chemistry, catalysis such as molecular reaction and nanoparticle reaction can be used to amplify signals to enhance sensitivity and have been utilized in absorption, fluorescence, resonance Rayleigh scattering (RRS), and surface-enhanced Raman scattering (SERS).⁵–⁷ A label-free DNAzyme-cleaving fluorescence method was developed for the determination of trace Pb²⁺ based on the catalysis of AuPd nanoalloy on the reduction of rhodamine 6G.⁵ Qu et al reported a colorimetric platform for visual detection of 0.1–10 ng/mL cancer biomarker based on intrinsic peroxidase activity of graphene oxide (GO).⁶ He et al prepared Au@Pt nanorods, which showed multiple enzyme properties and were used for spectrophotometric determination of 4.5×10⁻⁵–1×10⁻³ mol/L glucose (Glu).⁷ The nanocatalytic particle reaction is very interesting due to the novel surface plasmon resonance of Au and Ag nanoparticle that can be utilized to develop surface plasmon resonance absorption, RRS, and SERS methods with good features. A facile and sensitive peptide-modulating GONR catalytic nanoplasmon analytical
platform was reported for human chorionic gonadotropin.\textsuperscript{8} Recently, non-metal nanoparticles are interesting to analysts. Carbon nanotube (CNT) is a one-dimensional nanomaterial with a complete molecular structure at the nanoscale and is a good precursor to prepare water-soluble and stable GONR. CNTs have been used in the field of chemical sensing for its excellent physical and chemical properties. Ye et al\textsuperscript{9} produced MnO\textsubscript{2}/CNT composites with KMnO\textsubscript{4} oxidizing multi-walled carbon nanotubes (MWCNTs) to have strong electrocatalytic oxidation properties to detect H\textsubscript{2}O\textsubscript{2} as low as 0.1 \textmu M. Qu et al\textsuperscript{10} investigated the catalytic activity of the peroxidase mimetics of single-walled carbon nanotubes (SWCNTs) and achieved the Cu\textsuperscript{2+} visual detection. Cui et al\textsuperscript{11} prepared helical CNTs by hydrothermal-hydrogen reduction and investigated its catalytic activity as peroxidase mimetics. An electrochemical biosensor for H\textsubscript{2}O\textsubscript{2} was developed with a linear range (LR) of 0.5–115 \textmu mol/L. Zhang et al\textsuperscript{12} formed composite nanomaterials by combining positively charged gold nanoparticles (AuNPs) with SWCNTs and found that the material had a strong peroxide mimetic enzyme activity, thereby established a labeled DNA hybridization colorimetric detection method. However, CNTs are not water soluble that limits its application, and GONRs overcome this problem and provide the conditions for its analytical application without organic solvents. Zhang\textsuperscript{13} developed a bioelectrochemical sensor based on GONR modified for rapid detection of L/D-amino acids (AA) with an LR of 0.25–1.25 mmol/L and a detection limit (DL) of 100 and 60 \textmu mol/L, respectively. Dong et al\textsuperscript{14} used GONR to build biosensors to detect 5–100 \textmu mol/L adenosine triphosphate. Zhu et al\textsuperscript{15} developed a novel MWCNTs@GONR core-shell heterostructure and a sensitive electrochemical sensor for the detection of 8–500 nmol/L polycyclic aromatic amines. So far, no SERS was used to track GONR-catalytic AuNP reaction that can be regulated by mAlb immunoreaction for assay of trace mAlb.

SERS is a kind of selective and sensitive molecular spectral technique, which has attracted much attention in analysis, biology, and medical treatment.\textsuperscript{16} Generally, SERS signal depends upon a number of factors. However, the substrate-adsorbed molecular probe can greatly amplify the signals and it is linear to the SERS signal at some certain conditions,\textsuperscript{17,18} and an SERS method can be developed for the detection of the molecular probe. Javier and Ronel\textsuperscript{17} studied a paper-based portable SERS method for the detection of uric acid, with an LR of 0–3.5 mmol/L and a DL of 0.11 mmol/L. Wang et al\textsuperscript{18} used the catalytic activity of GO to catalyze the H\textsubscript{2}O\textsubscript{2}–HAuCl\textsubscript{4} system, and then ligands regulated the use of SERS to detect human chorionic gonadotropin, and Hg\textsuperscript{2+} in concentrations ranging from 0.25 to 10 ng/mL, 0.25–10 nmol/L, was tested on samples which showed good recovery. Frost et al\textsuperscript{19} developed a SERS sensor based on citrate-functionalized AuNP for 50–1,000 ng/mL Pb\textsuperscript{2+}. Gao et al\textsuperscript{20} immobilized the peptide nucleic acid with the target DNA on a slide, and 1.0×10\textsuperscript{10}–1.0×10\textsuperscript{6} mol/L DNA can be detected by SERS. Another type of SERS method was reported according to the change in the concentration of nanosol substrate in the analytical system. For example, a facile aptamer-regulating gold nanoplasmonic SERS detection strategy was proposed for trace lead ions based on the nanocatalytic particle reaction and nanoplasmon. Immunoassay is a kind of analytical method based on the specific reaction of antigen and antibody (Ab). It has the characteristics of high sensitivity and specificity and has been used in the fields of disease diagnosis, food safety, and environmental protection.\textsuperscript{21} In recent years, highly sensitive SERS technology combining with specific immunoreaction has been favored to analysts.\textsuperscript{22–26} Ma et al\textsuperscript{26} reported an SERS quantitative analysis method for trace human chorionic gonadotropin using a label-free Victoria blue B (VBB; C\textsubscript{33}H\textsubscript{30}CIN\textsubscript{5}) as probe in the aggregated immunonanogold sol substrate. She et al\textsuperscript{27} combined Hg\textsuperscript{2+} with the double-labeled Raman active 4-mercaptobenzoic acid and gold nanoparticles monoclonal antibodies on immunochromatographic test strips to obtain SERS immunoprobe to detect as low as 0.45 pg/mL of Hg\textsuperscript{2+}. To our best knowledge, there are no SERS quantitative analysis methods for trace mAlb without preparation of immunonanoprobe and label-free SERS molecular probes, based on the coupling of GONR-catalytic nanoreaction with immunoanalysis.

Albumin is a normal protein in the blood, but there is only a small amount of albumin in the urine under physiological conditions (<20 mg/L), because it is usually reabsorbed by glomerular filtration and renal proximal tubule.\textsuperscript{28} Urinary albumin that is normally in the range of 20–200 mg/L is called as microalbuminuria (mAlb).\textsuperscript{29} So it should be noted when the mAlb concentration is sustained excess, because it shows that nephrotic patients have a large number of albumin leakage and may be hypoproteinemia. And the development of kidney disease is possibly irreversible. Therefore, proteinuria is an important clinical symptom of nephropathy, and the control of mAlb concentration has a great clinical reference value to determine the degree of disease and prognosis. The rise of mAlb concentration is a reflection of early glomerular lesions sensitive indicators, especially in chronic renal injury such as diabetic nephropathy, hypertension, and systemic lupus erythematosus, and its detection is clinically significant.\textsuperscript{30} At present, mAlb detection methods are mainly electrochemical immunosensor,\textsuperscript{31}...
radioimmunoassay, ELISA, turbidity analysis, high-performance liquid chromatography, and so on. In this paper, a new gold nanoplasmon molecular spectral platform was established for rapid and selective quantitative detection of mAlb based on the immunoreaction-regulation of the GONR catalytic AuNP reaction.

**Experimental Instruments and reagents**

A DXR smart Raman spectrometer (Thermo Fisher Scientific, Waltham, MA, USA) with laser wavelength of 633 nm, power of 2.5 mW, slit of 50 µm, and acquisition time of 5 seconds; Hitachi F-7000 fluorescence spectrophotometer (Hitachi Ltd, Tokyo, Japan); TU-1901 dual-beam UV-visible spectrophotometer (Beijing Puxi General Equipment Limited Company, Beijing, China); and constant temperature water bath were used.

A 97 mmol/L HAuCl₄ solution, 10 µmol/L VBB, 10 mol/L H₂O₂ solution, 3.4 mmol/L trisodium citrate, 10 mmol/L AgNO₃, 0.1 mol/L Glu, 10 mg/mL mAlb, and 5 mg/mL mAlb Ab were prepared. 98% H₂SO₄, KMnO₄, GO (Nanjing Xianfeng Nanomaterial Technology Co. Ltd, Nanjing, China) and MWCNTs (No XFM12; Nanjing Xianfeng Nanomaterial Technology Co. Ltd) were used. The used reagents were of analytical pure grade, and the experimental water was the secondary distilled water.

**Preparation of GONR**

GONR was prepared by chemical melting MWCNTs. 50 mg of MWCNT powder was added into a 50 mL round bottom flask which containing 10 mL of concentrated H₂SO₄ and the reaction took place for 1 hour. Then, a certain amount (100, 200, 250, 300, 400, 500, 750, or 1,000 mg) of KMnO₄ was added and mixed well before the solution being heated for 2 hours in a 60°C water bath. The product was added into 200 mL ice water containing 5 mL of 30% H₂O₂ before being dispersed for 10 minutes by ultrasonic and centrifuged at 7,000 rpm for 10 minutes. The supernatant was obtained which contained 230 µg/mL GONR. Thus, GONR of different oxidation degrees (GONR1, GONR2, GONR3, GONR4, GONR5, GONR6, GONR7, and GONR8) were prepared by changing the amount of KMnO₄. The supernatant was neutralized to pH 7 with 50 mmol/L NaOH and diluted to the desired concentration before use.

**Procedure**

A suitable concentration of mAlb, 50 µL of 60 ng/mL mAlb and 60 µL of 47.6 ng/mL GONR were added into a 5 mL test tube successively. After mixing well for 10 minutes, 100 µL of 2.9 mmol/L HAuCl₄ and 30 µL of 0.1 mol/L H₂O₂ solution were added and diluted to the volume of 1.5 mL with water. The mixture was heated in a 60°C water bath for 10 minutes, then cooled with ice water to terminate the reaction, and 50 µL 10 µmol/L VBB was added. The SERS intensity (I₁,615 cm⁻¹) and the blank without mAlb (I₀,615 cm⁻¹) was measured to calculate the value of ΔI₁,615 cm⁻¹ = I₁,615 cm⁻¹ − (I₀,615 cm⁻¹) of DL was three times of SD which obtained from parallel determination of blank samples (n=10).

**Results and discussion**

**Principles of analysis**

The catalytic reaction of H₂O₂–HAuCl₄ increased with the increase of GONR concentration. The more the nanoparticle catalyst is added, the higher SERS intensity is obtained. In the system with Ab, the protein macromolecules bound to GONR and inhibited its catalytic activity. When the mAlb antigen was added, it specifically bound to the Ab, and the nanoenzyme GONR was released. With the increase of the amount of mAlb, the SERS peak increased due to the restoration of GONR catalysis. The mAlb concentration and SERS peak were of a linear relationship, which could be established to detect mAlb by SERS technique (Figure 1).

**SERS spectra**

Under the normal pressure and temperature, the reaction of H₂O₂ and HAuCl₄ was slow. GONR had strong catalytic effect on the reaction to form AuNPs that exhibited five strong SERS peaks in the presence of VBB molecular probes. The characteristic peaks of the above system include 1,164 (C—O stretching), 1,199 (C—H bending), 1,362 (ring stretching), 1,393 (C—N stretching), and 1,615 (C=O and N—H stretching). After adding Ab, it could inhibit the catalytic activity of GONR by binding to the surface of GONR catalyst, and the SERS signal linearly decreased (Figure S1). After the addition of mAlb, the stable immunocomplexes formed to release free GONR, and the SERS signal enhanced due to the catalysis recovering. With the increase of the amount of mAlb, the catalysis increased to cause the SERS intensity to gradually increase for the GONR6 (Figure 2), GONR3 (Figure S2A), and GO (Figure S2B) analytical systems. Results showed that the GO peak was the strongest, and the GONR6 system for detection of mAlb was the most sensitive. The blank spectra of pure GONRs, antibodies, and micro-albumin of the test concentrations were recorded respectively, and the results showed that all the blank spectra were very weak and that meant that the SERS peaks mainly attributed to the product of the nanocatalytic reaction. The SERS peak at 1,615 cm⁻¹
Figure 1 Principles of SERS immunoanalysis of mAlb coupled with GONR catalysis.
Abbreviations: SERS, surface-enhanced Raman scattering; GONR, graphene oxide nanoribbon; VBB, Victoria blue B; Ab, antibody; mAlb, microalbumin; MWcNT, multi-walled carbon nanotube.

Figure 2 SERS spectra of GONR6-Ab-mAlb-HAuCl4-H2O2.
Notes: a: 0.21 mmol/L H2AuCl4 + 0.006% H2O2 + 2.34 ng/mL GONR6 + 0.1 mmol/L HCl + 3.3×10⁻⁷ mol/L VBB + 3.3 ng/mL Ab; b: a + 0.26 ng/mL mAlb; c: a + 0.665 ng/mL mAlb; d: a + 1.33 ng/mL mAlb; e: a + 2 ng/mL mAlb; f: a + 2.62 ng/mL mAlb.
Abbreviations: SERS, surface-enhanced Raman scattering; GONR, graphene oxide nanoribbon; Ab, antibody; mAlb, microalbumin; VBB, Victoria blue B.
A new SERS strategy for quantitative analysis

was chosen for mAlb assay because it was the most sensitive and the intensity $\Delta I_{1,615 \text{ cm}^{-1}} = I_{1,615 \text{ cm}^{-1}} - (I_{1,615 \text{ cm}^{-1}})_0$ was linear to the mAlb concentration.

**RRS and UV-Vis spectra**

According to the procedure, the RRS spectra were obtained by synchronous scanning with the fluorescence spectrophotometer under the conditions of voltage $=450$ V, excited slit = emission slit = 5 nm, emission filter = 1% T attenuator, and $\lambda_{ex} - \lambda_{em} = \Delta \lambda = 0$. The results showed that GONR had a strong resonance scattering peak at about 310 nm. With the increase of mAlb concentration, more GONR was released and more AuNPs were produced which lead to the linear increase in the RRS peak (Figure S2). In the GONR systems, GONR6 was the most sensitive, and a simple and sensitive RRS method could also be developed to detect the concentrations of mAlb with an LR of 0.08–3.18 ng/mL (Figure 3A). Compared to the SERS method, the RRS method was less sensitive, but the procedure of RRS method is simpler than the SERS method without VBB. The TU-1901 dual-beam

![Figure 3 RRS and absorption spectra of GONR6-Ab-mAlb-HAuCl₄-H₂O₂](image)

**Notes:** (A) ± 0.21 mmol/L HAuCl₄ + 0.006% H₂O₂ + 2.34 ng/mL GONR6 + 3.3×10⁻⁷ mol/L VBB + 3.3 ng/mL Ab; b: ± 0.53 ng/mL mAlb; c: ± 1.06 ng/mL mAlb; d: ± 2.12 ng/mL mAlb; e: ± 2.65 ng/mL mAlb. (B) ± 0.21 mmol/L HAuCl₄ + 0.006% H₂O₂ + 2.34 ng/mL GONR6 + 3.3×10⁻⁷ mol/L VBB + 3.3 ng/mL Ab; a: ± 0.53 ng/mL mAlb; c: ± 1.06 ng/mL mAlb; d: ± 2.12 ng/mL mAlb; e: ± 2.65 ng/mL mAlb.

**Abbreviations:** RRS, Rayleigh scattering; GONR, graphene oxide nanoribbon; Ab, antibody; mAlb, microalbumin; VBB, Victoria blue B.
UV-Vis spectrophotometer was used to measure the absorbance of the system, and the results (Figure 3B, Figure S4) showed that GONR6 system had an obvious absorption peak at about 580 nm. With the increase of mAlb concentration, the Abs peak gradually increased.

**Catalysis and Inhibition**

Under the experimental conditions, MWCNT has no catalysis because it is insoluble in water. GONR and GO catalyzed \( \text{H}_2\text{O}_2\)–HAuCl\(_4\) reaction to generate nanoparticles (Figure 4). As the oxidation–reduction pair of \( \text{Au}^{3+}/\text{Au} \) has a low potential (1.401 V),\(^{36} \) it is difficult to form AuNPs from the \( \text{Au}^{3+} \) ions in one step. The reaction is easier to proceed with the addition of GONR as a catalyst. Since GONR has rich surface electrons, the electrons are transferred from the \( \text{Au}^{3+} \) ions and converted into \( \text{Au}^{+} \) ions. The oxidation–reduction pair of \( \text{Au}^{+}/\text{Au} \) has higher potential (1.692 V) and AuNPs are easier to be obtained. The catalytic activities of eight kinds of different GONR were compared mainly by three parameters (LR, linear equation, and coefficient), and the inhibition of Ab was studied (Table 1). The results showed that the linear equation of GONR6 system had

### Table 1 Comparison of catalysis and Ab inhibition by SERS method

<table>
<thead>
<tr>
<th>Catalysis system</th>
<th>Linear range</th>
<th>Linear equation</th>
<th>Coefficient</th>
</tr>
</thead>
<tbody>
<tr>
<td>GONR1</td>
<td>0.91–3.06 ng/mL GONR1</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 120.01 \text{c} + 4.5 )</td>
<td>0.9321</td>
</tr>
<tr>
<td>GONR2</td>
<td>0.91–3.06 ng/mL GONR2</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 198.4 \text{c} - 32.1 )</td>
<td>0.9643</td>
</tr>
<tr>
<td>GONR3</td>
<td>0.91–3.06 ng/mL GONR3</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 346.0 \text{c} - 139.2 )</td>
<td>0.9537</td>
</tr>
<tr>
<td>GONR4</td>
<td>0.91–3.06 ng/mL GONR4</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 412.7 \text{c} - 90.9 )</td>
<td>0.9293</td>
</tr>
<tr>
<td>GONR5</td>
<td>0.91–3.06 ng/mL GONR5</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 437.6 \text{c} - 137.3 )</td>
<td>0.9551</td>
</tr>
<tr>
<td>GONR6</td>
<td>0.91–3.06 ng/mL GONR6</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 782.5 \text{c} - 271.5 )</td>
<td>0.9458</td>
</tr>
<tr>
<td>GONR7</td>
<td>0.91–3.06 ng/mL GONR7</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 428.9 \text{c} - 71.7 )</td>
<td>0.9689</td>
</tr>
<tr>
<td>GONR8</td>
<td>0.91–3.06 ng/mL GONR8</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 275.3 \text{c} - 52.0 )</td>
<td>0.9644</td>
</tr>
<tr>
<td>Ab-GONR1</td>
<td>0.26–5.3 ng/mL Ab</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 60.8 \text{c} + 61.8 )</td>
<td>0.904</td>
</tr>
<tr>
<td>Ab-GONR2</td>
<td>0.26–5.3 ng/mL Ab</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 94.5 \text{c} + 85.3 )</td>
<td>0.9356</td>
</tr>
<tr>
<td>Ab-GONR3</td>
<td>0.26–5.3 ng/mL Ab</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 161.0 \text{c} + 60.7 )</td>
<td>0.9562</td>
</tr>
<tr>
<td>Ab-GONR4</td>
<td>0.26–5.3 ng/mL Ab</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 174.7 \text{c} + 33.2 )</td>
<td>0.9764</td>
</tr>
<tr>
<td>Ab-GONR5</td>
<td>0.26–5.3 ng/mL Ab</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 197.5 \text{c} + 153.0 )</td>
<td>0.929</td>
</tr>
<tr>
<td>Ab-GONR6</td>
<td>0.26–5.3 ng/mL Ab</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 363.5 \text{c} - 511.8 )</td>
<td>0.9776</td>
</tr>
<tr>
<td>Ab-GONR7</td>
<td>0.26–5.3 ng/mL Ab</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 206.8 \text{c} + 160.5 )</td>
<td>0.9339</td>
</tr>
<tr>
<td>Ab-GONR8</td>
<td>0.26–5.3 ng/mL Ab</td>
<td>( \Delta I_{1,615 \text{ cm}^{-1}} = 130.2 \text{c} + 113.2 )</td>
<td>0.9204</td>
</tr>
</tbody>
</table>

**Abbreviations:** Ab, antibody; GONR, graphene oxide nanoribbon; SERS, surface-enhanced Raman scattering.
the highest slope. This shows that GONR6 had the strongest catalysis because MWCNT had the most suitable oxidation degree and the carboxyl numbers were the most suitable. When the Ab was added into the system, the Ab would attach to the GONR surface, blocking the contact between the catalyst and \( \text{H}_2\text{O}_2/\text{HAuCl}_4 \) to inhibit its catalytic activity. With the increase of Ab, the system catalytic effect reduced and the SERS intensity weakened. The system reduction \( \Delta I_{1,615 \text{ cm}^{-1}} \) has a linear relationship with Ab concentration, and the inhibition of Ab-GONR6 system was the strongest because the slope is biggest.

**Laser scattering**

The size distribution of nanoparticles of mAlb-Ab-GONR6-\( \text{H}_2\text{O}_2/\text{HAuCl}_4 \) system was detected (Figure 5). The results showed that the average particle size of the non-mAlb system was 240±12 nm (Figure 5A). With the increase of mAlb concentration, the average particle sizes of the nanoparticles were 190.1±9.5 and 170.5±8.5 nm, respectively (Figure 5B and C), and the particle size was generally uniform.

**Transmission electron microscopy (TEM)**

In addition to laser scattering, TEM was also used to record the shape and size of the particles. According to the procedure, TEM of the different systems were recorded (Figure 6). The catalytic activity of the blank system without mAlb was weak, so the reaction of \( \text{H}_2\text{O}_2/\text{HAuCl}_4 \) was slow and there were rare big particles in the system (Figure 6A). When mAlb was added, it reacted specifically with the Ab to form free GONR6 and more nanoparticles were produced due to the restoration of GONR6 catalysis (Figure 6B). For the lower oxidation degree of GONR2, the catalytic effect was weaker and the AuNPs produced by the reaction were less. GONR2 with low oxidation degree contained a small amount of nanobelts. GONR6 solution with higher oxidation degree showed good catalytic activity, and the reaction resulted in more nanoparticles with comparatively more uniform particle size.

**Optimization of analytical conditions**

The analytical conditions of \( \text{HAuCl}_4/\text{H}_2\text{O}_2/\text{GONR6}-\text{Ab-mAlb} \) system were optimized (Figure S5). The results showed that \( \Delta I_{1,615 \text{ cm}^{-1}} \) reached the maximum at 1.84 ng/mL of GONR6. When the concentration of Ab was 3.3 ng/mL, \( \Delta I_{1,615 \text{ cm}^{-1}} \) reached the maximum. When the concentration of \( \text{H}_2\text{O}_2 \) was 2 mmol/L, \( \Delta I_{1,615 \text{ cm}^{-1}} \) was maximum. When the \( \text{HAuCl}_4 \) concentration was 0.152 mmol/L, \( \Delta I_{1,615 \text{ cm}^{-1}} \) reached the maximum, and so 0.152 mmol/L HAuCl4 was selected. The HCl concentration was optimized too, and 0.11 mmol/L HCl was selected. Under the chosen conditions at 60°C water bath, the reaction time of 10 minutes was good. The
Table 2 Analysis feature of the SERS system

<table>
<thead>
<tr>
<th>GONR system</th>
<th>LR (ng/mL)</th>
<th>Linear equation</th>
<th>Coefficient</th>
<th>DL (ng/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GONR1</td>
<td>0.33–6.6</td>
<td>$\Delta I_{1615 \text{cm}^{-1}} = 107.6C + 159.0$</td>
<td>0.9016</td>
<td>0.1</td>
</tr>
<tr>
<td>GONR2</td>
<td>0.33–3.2</td>
<td>$\Delta I_{1615 \text{cm}^{-1}} = 108.2C + 40.9$</td>
<td>0.9586</td>
<td>0.1</td>
</tr>
<tr>
<td>GONR3</td>
<td>0.33–3.2</td>
<td>$\Delta I_{1615 \text{cm}^{-1}} = 380.6C + 70.4$</td>
<td>0.9805</td>
<td>0.1</td>
</tr>
<tr>
<td>GONR4</td>
<td>0.33–3.2</td>
<td>$\Delta I_{1615 \text{cm}^{-1}} = 276.6C + 80.0$</td>
<td>0.9699</td>
<td>0.1</td>
</tr>
<tr>
<td>GONR5</td>
<td>0.24–3.2</td>
<td>$\Delta I_{1615 \text{cm}^{-1}} = 398.6C - 55.7$</td>
<td>0.9821</td>
<td>0.1</td>
</tr>
<tr>
<td>GONR6</td>
<td>0.065–2.6</td>
<td>$\Delta I_{1615 \text{cm}^{-1}} = 914.2C + 20.0$</td>
<td>0.9874</td>
<td>0.02</td>
</tr>
<tr>
<td>GONR7</td>
<td>0.13–2.6</td>
<td>$\Delta I_{1615 \text{cm}^{-1}} = 558.4C + 109.0$</td>
<td>0.9317</td>
<td>0.04</td>
</tr>
<tr>
<td>GONR8</td>
<td>0.13–2.6</td>
<td>$\Delta I_{1615 \text{cm}^{-1}} = 311.1C + 71.1$</td>
<td>0.978</td>
<td>0.04</td>
</tr>
</tbody>
</table>

Abbreviations: LR, linear range; DL, detection limit; GONR, graphene oxide nanoribbon.

Working curve

According to the procedure, the working curves of different SERS systems were drawn. For the GONR6 system, which was the most sensitive, the concentration of mAlb in the LR of 0.065–2.6 ng/mL has a good linear relationship with the $\Delta I_{1615 \text{cm}^{-1}}$, with the DL of 0.02 ng/mL. By comparing three analytical techniques of SERS/RRS/Abs (Tables 2, S1, and S2), the SERS method of GONR6 system had the highest slope (914.19), which shows that it has the highest sensitivity because of being strongest catalytic amplification, so it was selected to measure mAlb. Compared with the reported mAlb analysis method (Table S3),31–35,37–39 this method is highly sensitive and easy to operate, and it is one of the best methods for detecting mAlb.

Interference effects

The effects of coexisting substances in HAuCl$_4$-H$_2$O$_2$-GONR6-Ab-mAlb system on the determination of 10 ng/mL mAlb were investigated. The results showed that 100 times of K$^+$, Zn$^{2+}$, Ca$^{2+}$, S$_2$O$_3^{2-}$, SO$_4^{2-}$, NO$_3^-$, CO$_3^{2-}$, tyrosine, lysine, and phenylalanine; 50 times of Cu$^{2+}$, Mg$^{2+}$, tryptophan, and glutamic acid; and 10 times of Fe$^{3+}$ and cysteine did not interfere with the determination in the relative error range of no more than ±10.0%, and the method has good selectivity.

Sample analysis

Five fresh urine samples including two from diabetic patients were obtained from the Yanshan District People’s Hospital of Guilin City, and the samples were detected according to the experimental procedures. Then, different standard mAlb solution was added and detected. The results (Table S4) showed that the relative SD was between 0.49% and 2.28%, and the recoveries were between 96.9% and 109.8%.

Conclusion

The as-prepared GONR has strong catalytic effect on HAuCl$_4$-H$_2$O$_2$ nanoreaction to form AuNPs with nanoplasmon effect such as SERS, RRS, and Abs, and Ab could adsorbed on the surface of GONR, which blocks the redox electron transfer of HAuCl$_4$ and H$_2$O$_2$ to inhibit its catalytic action. Experiments show that mAlb enhances the signals of Ab-GONR-HAuCl$_4$-H$_2$O$_2$ nanoanalytical system. Based on this principle, an immunoregulation gold nanoplasmon method for the rapid detection of mAlb has been established, with simplicity, high selectivity, and sensitivity.

Acknowledgment

This work was supported by the National Natural Science Foundation of China (No 21767004, 21667006, 21465006, 21477025), the Key Laboratory of Ecology of Rare and Endangered Species and Environmental Protection (ERESEP2017Z09), and the University Key Laboratory of Karst Ecology and Environmental Change of Guangxi Province (YRHJ16Z009).

Disclosure

The authors report no conflicts of interest in this work.

References

