IncRNA LOC100132354 promotes angiogenesis through VEGFA/VEGFR2 signaling pathway in lung adenocarcinoma

Yumin Wang1,*, Fan Zhang1,*, Junjun Wang1, Lijuan Hu1, Feng Jiang1, Jian Chen1, Jie Chen2, Liangxing Wang3

1Department of Laboratory Medicine, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou China; 2Intensive Center Unit, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou China; 3Department of Respiratory Medicine, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China

*These authors contributed equally to this work

Purpose: This study aimed to study the biological function and the molecular mechanisms associated with the promotion of angiogenesis by IncRNA LOC100132354 in lung adenocarcinoma (LAD).

Patients and methods: The mRNA expression levels of 100 pairs of LAD and normal tissue samples of LOC100132354, vascular endothelial growth factor A (VEGFA), VEGF receptor-2 (VEGFR2), basic fibroblast growth factor (bFGF), and thrombospondin-1 (TSP-1) were analyzed by qPCR. LOC100132354 was knockdown and overexpressed in SPCA-1 and A549 cell lines to analyze the protein and mRNA expression levels of VEGFA, VEGFR2, bFGF, TSP-1, and changes in protein expression levels of Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mek1/2, and P-Erk1/2. Tumor microvessel density (MVD) was analyzed in experimental nude mice.

Results: The qPCR results showed that the mRNA expression levels of LOC100132354, VEGFA, VEGFR2, and bFGF mRNA in LAD tissues were significantly increased, while TSP-1 mRNA was significantly decreased compared with the adjacent tissues. Survival analysis showed that VEGFA, VEGFR2, and bFGF were poor predictors, while TSP-1 was a good predictor in LAD. Knockdown or overexpression of LOC100132354 affected the expression levels of bFGF, VEGFA/VEGFR2 signaling pathway, and downstream target molecules, such as Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mek1/2, and P-Erk1/2, while decreased TSP-1. After knockdown or overexpression of VEGFA expression, no significant changes in the expression level of LOC100132354 were found. Tumorigenesis of nude mice confirmed that LOC100132354 can significantly increase the tumor MVD.

Conclusion: These findings suggest VEGFA was a downstream target gene of LOC100132354, promoting angiogenesis through VEGFA/VEGFR2 signaling pathway and downstream target molecules in LAD. So, LOC100132354 is considered as an antiangiogenic target in LAD.

Keywords: lung cancer, IncRNA, angiogenesis, signaling pathway

Introduction

Lung cancer ranks first in terms of both morbidity and mortality of all cancers.1 Of all lung cancers, non-small-cell lung cancer (NSCLC) accounts for approximately 85%. According to the histopathological classification, NSCLC is divided into lung adenocarcinoma (LAD) and squamous cell carcinoma. Despite several advancements in chemotherapy, radiation, and surgery, lung cancer still remains aggressive and resulted in mortality.2 The average 5-year survival of lung cancer is below 15%.3–6 Recently, an increasing proportion of LAD has drawn a great attention on the socioeconomic environments.7 However, the mechanisms of LAD have not yet been uncovered.
angiogenesis.27 Bonspondin-1 (TSP-1) is a strong negative regulator of tumor growth and is determined by the tumor microenvironment. Multiple angiogenic growth factors that can stimulate endothelial cell proliferation, and angiogenesis in tumor tissues is directly related to VEGF. VEGF consists of five family members, namely VEGFA, VEGFB, VEGFC, VEGFD, and VEGFE. VEGF binds to its receptors to induce endothelial cell proliferation, and angiogenesis in tumor tissues is directly related to VEGF. VEGF consists of five family members, namely VEGFA, VEGFB, VEGFC, VEGFD, and VEGFE. VEGF binds to its receptors to induce the biological effects. The VEGF receptors (VEGFRs) mainly include Flt-1, KDR/Flk-1 (VEGFR2), Flt-4, and so on.20,21 VEGF, VEGFR, and their downstream signaling transduction pathways are used as specific tumor angiogenesis markers. Also, VEGFA/VEGFR2 signaling transduction plays a leading role in the formation of vascular endothelial cells. VEGF/VEGFR2 is a key factor for promoting angiogenesis in lung cancer,22 and it can promote tumor growth.23 Therefore, inhibition of VEGFA/VEGFR2 signaling pathway can inhibit tumor cell growth by blocking angiogenesis.24,25 Basic fibroblast growth factor (bFGF) is also one of the angiogenic growth factors that can stimulate endothelial cells to synthesize collagenase and promote invasion of the surrounding stroma during angiogenesis. Moreover, VEGF can induce endothelial cell production and release bFGF, further promoting the role of angiogenesis.26 While thrombospondin-1 (TSP-1) is a strong negative regulator of tumor angiogenesis.27 VEGF/VEGFR2 can activate Ras by multiple pathways28 and in turn activate Raf/Mek/Erk to exert the biological functions. Studies have shown that specific inhibitors of Mek1/2 can prevent the activation of Erk, thereby inhibiting the formation of blood vessels in vitro. Therefore, we speculate that LOC100132354 may regulate VEGF/VEGFR2 signaling pathway to promote angiogenesis in LAD, but the specific regulatory mechanisms need to be elucidated.

Patients and methods

Patient samples

We collected 100 LAD tissue samples and corresponding normal tissue (NT) samples in the First Affiliated Hospital of Wenzhou Medical University, from January 2013 to August 2015. The TNM stage was determined according to the American Joint Committee on Cancer (AJCC) and the Union for International Cancer Control (UICC) in 2002. The diagnosis of LAD was confirmed using histopathology results. The clinical and characteristic data of these LAD patients are presented in Table 1. We followed-up the survival of these cases by telephone or other ways, and the range of follow-up time was 3–23 months. This study was approved by the Institutional Ethics Review Board of the First Affiliated Hospital of Wenzhou Medical University, and all patients provided written informed consent.

Quantitative PCR

Total RNA was obtained from fresh LAD tissues and cell lines with TRIzol Reagent kit (Invitrogen, Waltham, MA, USA) and was reverse-transcribed into cDNA with a reverse-transcribed kit (TaKaRa, Shanghai, China) according to the manufacturer’s instructions. LOC100132354, VEGFA, VEGFR2, bFGF, TSP-1, and β-actin mRNA expressions were detected by qPCR in ABI 7300 analyzer. The primers of six genes are shown in Table 2. Two micrograms of total RNA was reverse-transcribed into cDNA. qPCR was performed in a total reaction volume of 20 µL, which consists of 2 µL cDNA, 1 µL PCR forward primer (10 mM), 10 µL SYBR Premix (2×), 1 µL PCR reverse primer (10 mM), and 6 µL distilled water. The qPCR program was as follows: 10 minutes denaturation at 95°C, 40 cycles for 5 seconds at 95°C, 30 seconds at 60°C, and 5 minutes extension at 72°C. All the results were normalized to β-actin, and all the experiments were performed in triplicate. The median in each triplicate for calculating the relative lncRNA expressing concentrations (ΔCt= Ct median Ct median β-actin), and 2−ΔΔCt formula for the relative expression.29
Two human LAD cell lines (SPCA-1, A549) were purchased from the Cell Bank of the Chinese Academy of Sciences and were cultured with complete medium (90% RPMI1640 supplemented with 10% FBS) at 37°C in 5% CO2. The complete medium was replaced every 2–3 days.

**Cell culture**

Two human LAD cell lines (SPCA-1, A549) were purchased from the Cell Bank of the Chinese Academy of Sciences and were cultured with complete medium (90% RPMI1640 supplemented with 10% FBS) at 37°C in 5% CO2. The complete medium was replaced every 2–3 days.

**Lentivirus-mediated siRNA and overexpression vector construction**

We constructed siRNA GV248 vector and overexpression GV303 vector for LOC100132354 (GeneChem, Shanghai, China). SiRNA sequences of LOC100132354 were as follows: siRNA1: CCACCTGGGATAGGACAAA, siRNA2: GAAGGAGAAATTGACAGGTT, siRNA3: CTGG AGTAGAGAAGA GAAT. SiRNA sequences of VEGFA were as follows: siRNA1: AAGCCGTCCT GTGTCGCGCTG, siRNA2: AACGATGAAGCCCTGGAGTGC, siRNA3: AAG TTAAAAGTGCCTGAACTG. Transfection was performed by adding $2 \times 10^8$ cells into 6-well plate. After 24 hours, the medium was changed, and the cells were incubated with the transfection complex according to the manufacturer’s standard operating procedure. The multiplicity of infection values

### Table 1 The clinical features of 100 patients with LAD

<table>
<thead>
<tr>
<th>Cases (n)</th>
<th>VEGFA*</th>
<th>P-value</th>
<th>VEGFR2*</th>
<th>P-value</th>
<th>bFGF*</th>
<th>P-value</th>
<th>TSP-1*</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>40</td>
<td>8.12±0.57</td>
<td>0.876</td>
<td>6.23±0.32</td>
<td>0.765</td>
<td>4.01±0.22</td>
<td>0.543</td>
<td>0.62±0.02</td>
</tr>
<tr>
<td>Female</td>
<td>60</td>
<td>7.99±0.65</td>
<td></td>
<td>6.16±0.27</td>
<td></td>
<td>3.98±0.20</td>
<td></td>
<td>0.61±0.01</td>
</tr>
<tr>
<td>Clinical stage</td>
<td>0.001</td>
<td>0.008</td>
<td>0.019</td>
<td>0.021</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>la</td>
<td>22</td>
<td>4.01±0.12</td>
<td></td>
<td>2.34±0.07</td>
<td></td>
<td>1.23±0.13</td>
<td></td>
<td>0.91±0.12</td>
</tr>
<tr>
<td>lb</td>
<td>31</td>
<td>4.19±0.14</td>
<td></td>
<td>2.46±0.12</td>
<td></td>
<td>1.65±0.12</td>
<td></td>
<td>0.86±0.11</td>
</tr>
<tr>
<td>Ila</td>
<td>15</td>
<td>5.23±0.13</td>
<td></td>
<td>3.25±0.13</td>
<td></td>
<td>2.02±0.14</td>
<td></td>
<td>0.71±0.09</td>
</tr>
<tr>
<td>IIb</td>
<td>16</td>
<td>5.89±0.21</td>
<td></td>
<td>4.42±0.18</td>
<td></td>
<td>2.32±0.14</td>
<td></td>
<td>0.35±0.05</td>
</tr>
<tr>
<td>IIa</td>
<td>15</td>
<td>9.86±0.71</td>
<td></td>
<td>7.78±0.32</td>
<td></td>
<td>5.24±0.25</td>
<td></td>
<td>0.15±0.01</td>
</tr>
<tr>
<td>Differentiation degree</td>
<td>0.008</td>
<td>0.002</td>
<td>0.021</td>
<td>0.032</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Low</td>
<td>13</td>
<td>9.34±0.70</td>
<td></td>
<td>7.24±0.42</td>
<td></td>
<td>7.23±0.35</td>
<td></td>
<td>0.90±0.11</td>
</tr>
<tr>
<td>Low-moderate</td>
<td>14</td>
<td>7.23±0.35</td>
<td></td>
<td>5.21±0.34</td>
<td></td>
<td>5.12±0.21</td>
<td></td>
<td>0.71±0.10</td>
</tr>
<tr>
<td>Moderate</td>
<td>36</td>
<td>4.45±0.32</td>
<td></td>
<td>4.24±0.21</td>
<td></td>
<td>3.45±0.16</td>
<td></td>
<td>0.50±0.08</td>
</tr>
<tr>
<td>High-moderate</td>
<td>19</td>
<td>3.83±0.25</td>
<td></td>
<td>2.34±0.18</td>
<td></td>
<td>1.45±0.12</td>
<td></td>
<td>0.31±0.05</td>
</tr>
<tr>
<td>High</td>
<td>18</td>
<td>2.56±0.18</td>
<td></td>
<td>2.01±0.09</td>
<td></td>
<td>1.34±0.09</td>
<td></td>
<td>0.17±0.03</td>
</tr>
<tr>
<td>LN metastasis</td>
<td>0.000</td>
<td>0.000</td>
<td>0.002</td>
<td>0.012</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>28</td>
<td>11.21±1.21</td>
<td></td>
<td>9.34±0.54</td>
<td></td>
<td>8.43±0.46</td>
<td></td>
<td>0.11±0.03</td>
</tr>
<tr>
<td>No</td>
<td>72</td>
<td>3.21±0.23</td>
<td></td>
<td>2.24±0.12</td>
<td></td>
<td>1.87±0.16</td>
<td></td>
<td>0.91±0.15</td>
</tr>
<tr>
<td>Smoking</td>
<td></td>
<td>0.854</td>
<td>0.734</td>
<td>0.521</td>
<td>0.773</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>37</td>
<td>8.23±0.56</td>
<td></td>
<td>6.34±0.33</td>
<td></td>
<td>4.11±0.23</td>
<td></td>
<td>0.65±0.02</td>
</tr>
<tr>
<td>No</td>
<td>63</td>
<td>8.16±0.53</td>
<td></td>
<td>6.21±0.29</td>
<td></td>
<td>4.02±0.21</td>
<td></td>
<td>0.60±0.01</td>
</tr>
</tbody>
</table>

**Notes:** *rneRNA relative expression level.

**Abbreviations:** bFGF, basic fibroblast growth factor; LAD, lung adenocarcinoma; LN, lymph node; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor A; VEGFR2, vascular endothelial growth factor receptor-2.

### Table 2 The primers of five genes

<table>
<thead>
<tr>
<th>Gene ID</th>
<th>Sense primer</th>
<th>Antisense primer</th>
<th>Product size</th>
</tr>
</thead>
<tbody>
<tr>
<td>LOC100132354</td>
<td>TTTgggCTgagCacagagaag</td>
<td>gTaggagTgTgTgTgTgg</td>
<td>198 bp</td>
</tr>
<tr>
<td>VEGFA</td>
<td>TCCACGACGCAACGCACATAG</td>
<td>GAGGGTCGCGCCGATTAGAC</td>
<td>174 bp</td>
</tr>
<tr>
<td>VEGFR2</td>
<td>CGTCAACAAAGTTGGGAGGA</td>
<td>CACATGACCCCAACAGCAG</td>
<td>153 bp</td>
</tr>
<tr>
<td>bFGF</td>
<td>CCTACCAACAATGCTGGGAGA</td>
<td>CACGTGACCCACACGAACG</td>
<td>187 bp</td>
</tr>
<tr>
<td>TSP-1</td>
<td>GCTTTGACATGTTGGACT</td>
<td>ATGCCGCTCAACTGACACC</td>
<td>193 bp</td>
</tr>
<tr>
<td>β-actin</td>
<td>CCTGGCACAACCAGCACATT</td>
<td>GCTGTACACATCCTGCTGGAA</td>
<td>157 bp</td>
</tr>
</tbody>
</table>

**Abbreviations:** bFGF, basic fibroblast growth factor; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor A; VEGFR2, vascular endothelial growth factor receptor-2.
lentivirus-mediated vectors. The study involved SPCA-1 LOC100132354, VEGFA siRNAs cell lines (siRNAs group, including siRNA1, siRNA2, and siRNA3). SPCA-1 was transfected with lentivirus-negative control LVCON077 vector (siRNA NC group), SPCA-1 (siRNA control group), and the A549 LOC100132354. VEGFA overexpressing cell line (OE group) and A549 were transfected with lentivirus negative control LVCON145 vector (OE NC group) and A549 (OE control group). We have previously constructed the best LOC100132354 interference sequence for SPCA-1 cell line and LOC100132354 overexpression level in A549 cells.14

Western blotting
The cell lysate was placed on ice for 30 minutes. The supernatant was electrophoresed on SDS-PAGE and then transferred onto a nitrocellulose membrane. Following the molecular cloning procedure, the primary antibody was blocked overnight and horseradish peroxidase secondary antibody was allowed to bind for 1 hour.31 The protein levels of VEGFA, VEGFR2, bFGF, TSP-1, Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mek1/2, and P-Erk1/2 in these groups were then analyzed.

Nude mice tumor formation and microvessel density (MVD) detection
The cells include SPCA-1 LOC100132354 siRNA2 cell line (siRNA group), siRNA NC group, OE group, and OE NC group. These cells at a density of $2 \times 10^6$ cells were inoculated into the axillary region of BALB/c nude mice subcutaneously. MVD was measured according to the method reported by Weidner et al and Maeda et al, and the sections were analyzed to determine the highest blood vessel density in the tumor, necrosis area with lesser blood vessels, and no blood vessels at the junction with NTs under low magnification microscope (100×). Any brown-yellow stained endothelial cells or cell plexuses that distinguish from the surrounding tumor tissues and connective tissue components were considered as blood vessels and confirmed after finding the highest density of blood vessels under the high magnification microscope (200×). If the structures are not connected, its branching structure also serves as a blood vessel.32,33 The number of microvessels of the five fields of vision was recorded for each sample, and the average number of groups was taken as the MVD value for the group. The investigation confirms with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication No. 85–23, revised 1996). The study protocol was approved by the Institutional Animal Experimental Ethics Review Board of Wenzhou Medical University.

Statistical methods
Differences in variables between the groups were assessed using Kruskal–Wallis test for non-normal distribution data or one-way ANOVA test for normal distribution data. Comparison between the two groups was evaluated by Student’s $t$-test or Mann–Whitney $U$ test or least significant difference test. Survival curve was analyzed with GraphPad Prism software 6.0, and chi-squared test analyzed the differences. $P<0.05$ was considered to be statistically significant.

Results
The expression level and survival analysis of LOC100132354, VEGFA, VEGFR2, bFGF, and TSP-1 in LAD
The results of qPCR showed that the relative mRNA expression level of LOC100132354, VEGFA, VEGFR2, and bFGF was obviously higher than the NT tissues (Mann–Whitney $U=103.00, 87.00, 79.00, 96.00$ and $P=0.0040, 0.0018, 0.0019, 0.0021$), while TSP-1 mRNA was obviously lower than the NT tissues (Mann–Whitney $U=165.00, P=0.0000$) in LAD (Figure 1). This suggested significantly enhanced angiogenic capacity in the LAD tissues. One-way ANOVA test or $t$-test showed that the relative mRNA expression levels of VEGFA, VEGFR2, bFGF, and TSP-1 ($P=0.001, 0.008, 0.019, 0.021, 0.008, 0.021, 0.032, 0.000, 0.002, 0.012$) were obviously different in clinical stage, differentiation degree, and lymph node metastasis, while VEGFA, VEGFR2, bFGF, and TSP-1 ($P=0.876, 0.765, 0.543, 0.762, 0.854, 0.734, 0.521, 0.773$) showed no differences in sex and smoking (Table 1).

The survival analysis showed that VEGFA, VEGFR2, and bFGF ($\chi^2=16.26, 10.78, 16.22$ and $P<0.0001, P=0.0010, P<0.0001$) were considered as poor predictors, while TSP-1 ($\chi^2=17.50, P<0.0001$) was a good predictor in LAD (Figure 2). Studies have confirmed that VEGFA and VEGFR2 played a pivotal role in angiogenesis and bFGF was considered as an angiogenic factor, while TSP-1 was a negative regulator of vascular growth. So, our results confirmed that the growth of blood vessels was significantly increased in the LAD tissues.

LOC100132354 significantly enhanced angiogenic capacity in the LAD cell lines through VEGFA/VEGFR2 signaling pathway and downstream Ras/Raf/Mek/Erk pathway
We further confirmed that the protein expression levels of VEGF, VEGFR2, bFGF, Ras, P-A-Raf, P-B-Raf, P-C-Raf,
Promotion of angiogenesis by lncRna LOC100132354 in LAD

P-Mek1/2, and P-Erk1/2 \( t=7.258, 8.850, 8.994, 4.920, 9.032, 7.334, 9.141, 7.143; \ P=0.018, 0.013, 0.012, 0.039, 0.012, 0.024, 0.012, 0.019 \) were significantly higher in the OE group than in the OE NC group, while TSP-1 protein expression levels were lower \( t=−7.471, \ P=0.017 \). The protein expression levels of VEGF, VEGFR2, bFGF, Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mek1/2, and P-Erk1/2 \( t=8.701, 7.574, 6.491, 5.421, 5.578, 5.443, 10.140, 8.424, 12.689; \ P=0.013, 0.021, 0.023, 0.032, 0.031, 0.032, 0.010, 0.014, 0.006 \) in the siRNA group were lower than those of the siRNA NC group, but the TSP-1 protein expression levels were higher than those of the siRNA NC group \( t=−24.005, \ P=0.002 \) (Figure 3).
Compared with the OE NC group, the expression levels of VEGFA, VEGFR2, and bFGF mRNA (t=15.796, 16.041, 15.578; P=0.003, 0.002, 0.003) in the OE group were increased, while TSP-1 mRNA level was decreased (t=6.244; P=0.025). Compared with the siRNA NC group, the expression levels of VEGFA, VEGFR2, and bFGF mRNA (t=5.399, 5.903, 5.556; P=0.032, 0.023, 0.036) were decreased in the siRNA group, while TSP-1 mRNA level was increased (t=14.477; P=0.005) (Figure 4A).

These results confirmed that after inhibition or overexpression of LOC100132354, the key molecular proteins and mRNA levels in the VEGFA/VEGFR2 signaling pathway of the cell line were significantly changed.

**VEGFA is a downstream target gene of LOC100132354**

LOC100132354 RNA expression level showed no obvious changes after VEGFA knockdown (t=0.538; P=0.644) or...
Figure 3: The proteins of VEGFA, VEGFR2, bFGF, TSP-1, Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mek1/2, and P-Erk1/2 from different cell lines by Western blotting. 
Notes: (A) The Western blotting figure of these proteins. (B) The results of VEGFA, VEGFR2, bFGF, TSP-1 in the four groups. (C) The results of Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mek1/2, and P-Erk1/2 in the four groups. *P<0.05, **P<0.01.
Abbreviations: bFGF, basic fibroblast growth factor; OE, overexpression; NC, negative control; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor A; VEGFR2, vascular endothelial growth factor receptor-2.
overexpression \( (t=0.144; P=0.898) \) (Figure 4B, C), suggesting that VEGFA was a downstream target gene of LOC100132354.

**LOC100132354 significantly enhanced the MVD value of the nude mice**

Our previous work showed that LOC100132354 significantly enhanced the tumorigenic ability in the nude mice experiment.\(^1^4\) Tumorigenesis of nude mice confirmed that the tumor MVD was decreased significantly \( (t=10.765; P=0.012) \) after inhibition of LOC100132354 and increased significantly \( (t=15.879; P=0.009) \) after overexpression of LOC100132354 (Figure 5 and Table 3).

**Discussion**

Tumor cells secrete VEGFA and bind to the VEGFR2 on the surface of endothelial cells to exert their biological effects.\(^1^7,1^8,3^4\) Studies have shown that the expression levels of VEGFA and VEGFR2 were significantly elevated in NSCLC tissues,\(^3^5,3^6\) indicating that angiogenesis was significantly enhanced in NSCLC tumors. Steady expression of VEGFA can lead to increase in tumor volume in lung cancer in mice,\(^3^6\) while inhibition of VEGFA signaling pathways resulted in the inhibition of tumor growth and tumor angiogenesis.\(^3^7,3^8\)

In this study, the mRNA expression levels of LOC100132354, VEGFA, VEGFR2, and bFGF were significantly higher than the corresponding NT tissues, while TSP-1 mRNA are significantly lower than the corresponding NT tissues in LAD. The survival analysis showed that VEGFA, VEGFR2, and bFGF were poor predictors, while TSP-1 was a good predictor in LAD. These results revealed that the angiogenic capacity was enhanced significantly and could predict the prognosis of LAD.
and after overexpression of LOC100132354, the MVD was increased significantly ($t=15.879; P<0.009$). 

Table 3: Comparison of MVD in nude mouse tumors ($\pm$SD) (n=25)

<table>
<thead>
<tr>
<th>Group</th>
<th>MVD</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>siRNA NC</td>
<td>14.2±1.32</td>
<td>0.012</td>
</tr>
<tr>
<td>siRNA</td>
<td>6.0±0.92</td>
<td></td>
</tr>
<tr>
<td>OE NC</td>
<td>10.0±1.01</td>
<td></td>
</tr>
<tr>
<td>OE</td>
<td>18.1±1.21</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviations: MVD, microvessel density; OE, overexpression; NC, negative control.

In our previous studies, we found that LOC100132354 increased the ability of colony formation, cell migration, adhesion, proliferation, invasion, S phase, and G2/M phase cells, while decreased G0/G1 phase cells and apoptosis. 

In this study, we further confirmed that the expression levels of VEGFA, VEGFR2, bFGF, Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-MekI/2, and P-Erk1/2 were significantly higher from the OE group than those of the OE NC group, while TSP-1 expression levels were increased. So, the knockdown and overexpression experiments consistently confirmed that LOC100132354 regulated the angiogenic capacity by VEGFA/VEGFR2 pathways in LAD.

LOC100132354 RNA expression levels showed no changes after VEGFA knockdown or overexpression, otherwise LOC100132354 could significantly enhance the MVD value of the nude mice. This suggested that VEGFA was a downstream target gene of LOC100132354.

Conclusion

Our study ascertained that LOC100132354 increased the angiogenic capacity and the mechanism of regulation of angiogenic capacity in LAD. Our results confirmed that lncRNA LOC100132354 can act as an antiangiogenic target in LAD.

Acknowledgments

This study was financially supported by the National Natural Science Foundation of China (81672088), Zhejiang Provincial Natural Science Foundation (LQ16H160020), Zhejiang Provincial Health Planning Commission (2018KY514), and the Wenzhou Municipal Science and Technology Bureau of China (Y20170208, Y20170718).

Disclosure

The authors report no conflicts of interest in this work. There are not any of the family relationship between such authors.

References