Main trends of immune effects triggered by nanomedicines in preclinical studies

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Abstract: The application of nanotechnology to emerging medicinal products is a crucial parameter for the implementation of personalized medicine. For example, sophisticated drug delivery systems can target the diseased tissue by recognizing patient-specific biomarkers while carrying pharmacologically active molecules. However, such nanomedicines can be recognized by the immune system as foreign triggering unexpected biological reactions. The anticipation of the immunogenic potential of emerging nanotechnology-based products in the preclinical phase is challenging due to high interspecies variations between the immune systems of laboratory animals and humans. A close monitoring of the scientific literature is required to better understand the relationship between various immune reactions and the diversity of nanomedicines currently in the development pipeline. We have reviewed the most frequent immune reactions induced by the nanomaterials in vivo and have identified the main effects triggered by lipid-based, polymer-based and inorganic nanoparticles, as the main categories of nanomaterials used in medicine. According to our results, almost 50% of the investigated nanomaterials induced effects related to the activation of the immune system. Among them, complement activation-related hypersensitivity reactions and activation of adaptive immune response were the most frequent effects reported for the lipid-based nanoparticles. However, many of these effects are not or are only partially covered by the current regulatory framework applicable for nanomedicines. In addition, we extracted the most relevant nanospecific properties responsible for the observed biological effects. Our analysis led to identification of the most prevalent measurement endpoints relevant for the assessment of the immunotoxic potential of the nanotechnology-based products and will support the smooth and safe translation of the new formulations to clinical applications.

Keywords: immune reactions, nanomaterials, preclinical testing, in vivo, personalized medicine

Introduction

The application of nanotechnology in health care holds promise for the development of innovative medical products addressing, in particular, unmet medical needs. The possibility to design nanomedicines that are able to transport biologically active molecules and target specifically the diseased tissue makes them a promising tool for the implementation of personalized medicines. However, innovative materials as well as the use of foreign proteins used for the design of nanomedicines can be potentially immunogenic. In particular, when intravenously (IV) administered, nanomedicines interact immediately with the blood and immune system. The human immune system comprises innate and adaptive immune mechanisms involving a network of interdependent signaling pathways. The innate immune system generates immediate unspecific defense reactions to encounter microbial invasions, whereas the adaptive immune...
system engenders delayed but more specific immune response involving the activation of lymphocytes and production of specific antibodies.

Nanoparticles (NPs) were shown to interact with both innate and adaptive immune mechanisms. However, the mechanisms and biological consequences of this interaction are not well understood yet. Once in the blood system, NPs interact with plasma proteins which bind to NP surface, forming so-called protein corona (PC). This process can in a significant way impact the interaction of NPs with the components of the immune system, for example, binding of the immunoglobulins to NP surface facilitates their recognition and uptake by the phagocytic cells in the process called opsonization.

In the area of drug delivery, the combined effect of both nanocarrier and encapsulated drug has to be considered. In several cases, the NP shell protecting the active pharmaceutical ingredient may reduce the toxic effect of the drug on the immune system; in other cases, it can act in synergy with it, for example, stimulating the immune response against the tumor. However, NP-induced effect can also lead to unwanted immune reactions impacting the safety of nanoformulations. In addition, the immune system is very species specific, and interindividual diversity of the human immune system makes preclinical safety assessments of nanomedicines challenging. Particularly, knowledge of which physicochemical properties of NPs can influence a biological effect is needed for the design of safe pharmaceutical products based on nanotechnology.

We have reviewed the most frequent immune reactions induced by the nanomaterials in vivo and identified the main effects triggered by lipid-based, polymer-based and inorganic NPs. In addition, we have extracted the most relevant nano-specific properties responsible for the observed biological effect. Our analysis led to identification of the most prevalent endpoints relevant for the assessment of the immunotoxic potential of the nanotechnology-based products. A detailed knowledge on the interaction of the NPs with the immune system will support the translation of emerging nanomedicines to clinical applications.

Methodological approach

Search management

The present review has to be seen as narrative and not as a systematic review. As the first step, we performed the literature search by using Scopus, Web of Science and PubMed search engines, the most commonly known search engines in the biomedical field. All studies investigating the effects in vivo of all nanomaterials potentially relevant for the nanomedicine field were considered.

The initial search was carried out using the prefix “nano*” AND “immunotoxicity” OR “immune response” OR “hypersensitivity” AND “in vivo” keywords and excluding “vaccine” keyword. The additional search was done using “nano*” AND “complement activation” AND “in vivo” keywords. Other additional searches were done using “liposome” OR “graphene” AND “immunotoxicity” AND “in vivo” keywords.

In the second step, the results were manually curated. Double hits and entries that were out of scope were removed. Reviews, book chapters and conference proceedings were also not considered. Reported effects resulting from the nanoformulations containing active pharmaceutical ingredients were only taken into account when the study indicated the impact of the nanocarrier on the observed biological effect. The results stating absence of significant immune response of the investigated formulations were classified separately. The search was performed from December 2016 till March 2017, and the last update was performed at the end of June 2017.

Quality of research studies

An important aspect to evaluate the quality of the studies reporting an immune effect was the description of the physicochemical characterization of the investigated nanomaterials. Ninety-two percent of the publications provided the physicochemical characterization of the investigated nanomaterial or referred to the previously published physicochemical characterization. Moreover, 59% of them performed additionally the in vitro studies to complete the information obtained from in vivo experiments.

The ability of NPs to interfere with the immune assays, as well as their contamination by the endotoxin should be evaluated before the experiments, since it can highly influence the assay results as well as the reliability of data in independent runs due to batch-to-batch variances. However, reports on the reliability of the measurements as well as information on interference with the test reagents were rarely provided. Therefore, this parameter can be considered as the confounding factor of the performed analysis.

Overview of the results

In our search, we considered all categories of nanomaterials potentially used for medical applications. According to the classification of nanomedicines proposed by Wicki et al, we grouped the various NPs into lipid-based NPs, polymer-based NPs and inorganic NPs. Among the 229 original
research articles investigating the effect on the immune system, 138 (60%) were related to inorganic NPs, 47 (20%) to lipid-based NPs and 46 (20%) to polymer-based NPs (Figure 1A). The majority of these reports (~60%) demonstrated an interaction with the immune system either in terms of the stimulation (47%), suppression (10%) or dysregulation (2.5%) covering the ambiguous effects on the immune system (Figure 1B). In 40% of the studies (91 articles), no significant effect on the immune system was found. Inorganic NPs were responsible for 68% of all described adverse effects on the immune system (not shown).

We also extracted the most frequent immune effects related to each category of medical NPs (Figure 2). The majority (61%) of reports related to lipid-based NPs demonstrated an effect on the immune system, including mainly activation of the complement system, complement activation-related pseudoallergy (CARPA) and activation of the adaptive immune response (in particular, poly(ethylene glycol) [PEG] ylated liposomes). Around 65% of tested polymer-based NPs did not show any significant immunotoxicity. The most frequent reaction induced by polymer-based NPs was antigenicity accompanied by the production of specific antibodies and leading to accelerated blood clearance (ABC) of the nanoformulation. Seventy percent of the investigated inorganic NPs induced an adverse effect on the immune system, demonstrating in particular an increased risk of inflammation associated with either innate or adaptive immune response.

**Figure 1** Number of studies in vivo related to the main categories of investigated nanomaterials (A), and general effects on the immune system including activation, suppression or dysregulation or absence of the immune reaction (B).

**Abbreviation:** NP, nanoparticle.

**Figure 2** Most frequently reported in vivo immune effects related to each of the NP category employed in the nanomedicine, such as lipid-based, polymer-based and inorganic NPs.

**Notes:** “CARPA” effect is a subcategory of “complement activation” effect, “inflammation” effect is a subcategory of “activation of the innate immune cells” effect and “accelerated blood clearance” effect is a subcategory of the “adaptive immune response” effect. The subcategories are highlighted for a more detailed view on the specific immune effects induced by different NP categories.

**Abbreviations:** CARPA, complement activation-related pseudoallergy; NP, nanoparticle.

**Activation of the immune system**

Activation of the immune system was the most frequently observed immune reaction after the administration of nanomaterials in animal studies, independent of the category of the NPs. Stimulation of the immune response can be beneficial to the organism since it supports the recognition and elimination of foreign materials and the defense against bacterial and viral infection. For example, gold nanorods were shown to inhibit respiratory syncytial virus and were able to stimulate antiviral response in mice. Currently, a number of studies exploit the immunostimulating properties of NPs used as anticancer therapies and as vaccine vehicles. In this review, though, we focused on unintended immune reactions triggered by NPs as they are not always detected on time. Uncontrolled and excessive immunostimulation can lead to autoimmune disorders, or alternatively can induce inflammation and damage the tissues even a long time after the exposure. Among the 108 publications reporting on the
On the other hand, depending on the surface functionalization, gold nanorods were shown to induce either M1 or M2 macrophage polarization and promote or not inflammatory injury to the liver. These findings indicate that by tuning the surface properties, it is possible to reverse the adverse effect of NPs in order to achieve the expected biological profile.

Complement activation

The system of complement is an important part of the innate immunity, designated as first-line defense against pathogenic infections. It can be activated via antibodies binding to the antigen (classical pathway), mannose binding lectin pathway or alternative pathway initiated by direct binding of the pathogen to the complement protein. Even if each pathway is triggered differently, they converge at the step where the third complement protein (C3) is cleaved into anaphylatoxin C3a and opsonic component C3b. The cascade of different reactions generates three major effects: 1) proinflammatory process with the release of anaphylatoxins, 2) opsonization of pathogen, which is subsequently eliminated by the phagocytic cells and 3) membrane attack complex leading directly to lysis of the targeted pathogen cell. All three pathways classical, lectin and alternative pathway, can be activated in contact with NPs, leading to the inflammatory process accumulation in the liver and spleen macrophages or hypersensitivity reactions (Figure 4). Furthermore, particle recognition by the reactive intravascular macrophages can additionally enhance the release of anaphylatoxin and the proinflammatory reactions as recently confirmed by Wibroe et al. The resulting CARPA is a frequent side effect of IV administered liposomal and micelle drugs already on the market or in the development stage (Figure 3).

Occasionally, it can lead to life-threatening
conditions including pulmonary edema, cardiovascular stress, hypoxia and other hypersensitivity symptoms.

Lipid-based NPs with the functionalized surface were the main category of NPs reported to induce CARPA effect in vivo. However, an appropriate animal model must be used to observe hypersensitivity reactions. More frequently, the in vitro and in vivo methods for the blood markers of complement activation were employed to assess the effect of investigated NPs. Though it has to be noted that in most cases, a strong complement activation leads to hypersensitivity reactions, interindividual variations in human complement activation responses exist. Apart from the lipid-based formulations, polymeric and inorganic NPs also, such as iron oxide NPs and gadolinium conjugates, were reported to activate the complement system. Nevertheless, the exact molecular mechanism initiating this interaction still remains to be elucidated.

**Activation of the adaptive immune response**

The capacity to increase the adaptive immune response via activation of T-cells and B-cells and the subsequent production of specific antibodies are usually associated with biotechnology-derived products. Therapeutic proteins can be recognized as foreign and are presented to T-cells, leading to activation of the cascade of cell-mediated and humoral responses. Upon activation and depending on the type of encountered pathogen, T-cells proliferate and differentiate into T helper 1 cells (Th1), T helper 2 cells (Th2), Th17 or regulatory T lymphocytes. Evaluation of Th1 to Th2 balance following interaction with an investigated nanomaterial can provide insights on the proinflammatory potential and the subsequent immune pathway. Th1 cells play a major role in

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**Figure 4** Schematic illustration of the impact of physicochemical properties of NPs on the interaction with the complement system and the induction of CARPA.

**Abbreviations:** CARPA, complement activation-related pseudoallergy; NP, nanoparticle.
the cellular immunity and are associated with acute inflammation; Th1 cytokines can induce macrophage polarization toward M1 phenotype. Such Th1-polarized inflammatory response was induced in mice by intratracheal administration of iron oxide NPs, leading to pulmonary inflammation.\(^\text{47,48}\) Th2 cells promote humoral response, but are also related to allergy disorders, for example, silica NPs were reported to induce allergen-specific Th2-type allergic immune responses in a mouse model.\(^\text{49}\) In addition, several medical NPs including polymeric micelles and dextran-coated iron oxide NPs were able to stimulate the adaptive immune response inducing production of corresponding immunoglobulins.\(^\text{50–52}\) A particularly interesting observation is related to PEG coating frequently used in nanomedicine to provide better stabilization and a lower recognition by the immune system in order to increase the drug circulation time.\(^\text{53–55}\) Interestingly, PEGylated liposomes, iron oxide NPs and gold NPs were reported to lose their long circulating properties when they were administered repeatedly to the same animal. This unexpected phenomenon called “accelerated blood clearance” (ABC) occurred due to the production of specific anti-PEG IgM antibodies labeling the NPs, followed by the increased uptake of NPs by the macrophages and their accumulation in the liver and spleen (Figure 5).\(^\text{56–60}\) Several attempts were made to find an alternative NP coating which could keep stealth properties of PEG without inducing the ABC phenomenon and compromising the efficacy of drug.\(^\text{61,62}\)

**Immunosuppression**

Suppressive effect on innate and acquired immune responses can lead to less-effective responses of the organism to pathogen infection. Such effect was observed for 10% of investigated nanomaterials. A decreased production of the specific antibodies in response to antigen, reduced activity of natural killer cells and slower proliferation of lymphocytes have been demonstrated in vitro and in vivo for inorganic NPs, mainly fullerene derivatives\(^\text{63–66}\) and several metallic NPs.\(^\text{67–71}\) The diminution of the host immunity against *Listeria monocytogenes* infection was noted in mice administered with liposome-encapsulated hemoglobin, but this issue has been overcome by the steric stabilization of the formulation and adequate changes in lipid composition.\(^\text{72,73}\)

An important aspect in the assessment of the drug delivery systems is the influence of the encapsulated drug. Depending on the liposome composition and other physicochemical properties of the formulation, the myelosuppressive effect of the encapsulated doxorubicin was shown to be attenuated\(^\text{74}\) or, on the contrary, amplified, when cumulated with the immunotoxic effect of the carrier.\(^\text{75}\) Therefore, a simultaneous safety testing of the NP formulation with and without the encapsulated drug could provide more accurate information.

**Influence of the physicochemical parameters**

A big part of the ongoing research effort concentrates on determining the key physicochemical properties of NPs that can induce or modulate a biological effect. This information has a fundamental importance for drug developers since it allows designing of efficacious and safe therapeutics displaying a reduced risk of immune response, unless such response is needed (immunostimulatory agents, vaccine vehicles etc.). Correlation of the physicochemical characteristics with the biological effect is also the first step for determining the so-called critical quality attributes, which are formally designated parameters of manufactured pharmaceutical products enabling evaluation of their quality.\(^\text{76}\) Among the selected scientific articles, 64% tried to identify such key physicochemical properties of NPs responsible for the observed immune reaction. Surface-related properties such as coating, surface functionalization and charge were among the most critical parameters as reported by the researchers (Figure 6). Indeed, coating of NPs with the hydrophilic polymer not only ensures better steric stabilization, but also has a direct impact on the opsonization process and subsequent interaction with the immune cells. Increased PEG density on the surface of lipid NPs reduced their hemolytic and immunostimulating activity.\(^\text{77}\) But, on the other hand, the second administration of PEGylated formulations to the same animal resulted in the production of anti-PEG antibodies and the ABC (Figure 5).

In addition, the size, the chemical structure and hydrophobicity were also recognized as important properties in many
Immune effects triggered by nanomedicines

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studies. Smaller NPs were, in general, found to have higher immunotoxic potential than the bigger ones, whereas hydrophobicity influenced activation of the complement system and cytokine secretion by the splenocytes.

Finally, it has to be noted that in the biological environment, physical and chemical properties of NPs, such as size, shape or surface properties, can change following NP agglomeration or dissolution. Likewise, the NP PC composition also can undergo dynamic changes influencing subsequent interaction with the immune components. In addition, some studies seem to indicate that the conformational changes of the proteins involved in the formation of the PC are the key events initiating immunogenic reactions.

Testing of the nanomedicine immunotoxicity

Given the increasing number of nanotechnology-based products in development stage, and the risk of interspecies variations limiting utility of animal studies, cost- and time-effective, humanized in vitro screening methods are needed to detect possible immune-related toxicities of nanomedicines. The evaluation of the state of the macrophages involved in the first-line interaction with NPs and mediating specific and nonspecific immune responses has been suggested as valuable testing approach for predicting the immunotoxic potential of NPs. Macrophage viability, generation of reactive oxygen and nitrogen species, as well as uptake of NPs were among the most tested endpoints studied in vitro (Figure 7B).

Another function of phagocytic cells, that is, the release of cytokines/chemokines, leading to inflammation and activation of the immune system, was intensively studied in vivo and in vitro (Figure 7A and B). The amount of secreted cytokines analyzed in relevant in vitro models can reflect the in vivo situation and particularly predict severe immune reactions such as cytokine storm. However, certain types of NPs can adsorb measured cytokines on their surface, reducing their content in the supernatant and generating false-negative results of the assays. Evaluation of the cytokine expression at the protein level or at the gene level (Figure 7B), usually less subject to interferences, can provide a valuable option. The gene expression of the range of cytokines was used by Moyano et al. to establish a link between NP hydrophobicity and activation of the immune system.

Measurement of the complement activation is particularly important in case of IV administered nanomedicines due to its implications to several other immune effects such as enhanced uptake by the macrophages, hypersensitivity reactions and inflammatory process (Figure 4). Several methods for testing of the complement activation in vivo and in vitro in human blood are available or in the phase of development. The major issue related to in vivo models is their high interspecies variation and the difficulty to find an appropriate animal model sufficiently predictive for the...
hypersensitivity reactions in humans. Due to their availability and cost-effectiveness, in vitro methods for assessing complement activation are more and more used to design and predict biocompatibility and the circulation time of developed nanoformulations for IV use.

The binding of the specific plasma proteins to NP surface (opsonization) is the first reaction of the organism to IV administered NPs and contributes to their recognition and uptake by the immune cells. Detection of the immunogenic potential of NPs at this early step would be optimal to avoid subsequent immunotoxic reactions such as complement activation, activation of macrophages and lymphocytes, proinflammatory reactions etc. Some attempts have been already made to correlate the PC “fingerprint” of gold NPs with the subsequent internalization by the cells. However, much more information needs to be gathered before we are able to properly link the composition of NP PC with the specific reactions of the immune system.

Finally, independent of the investigated endpoint, the reliability and suitability of the method should be evaluated. Due to their specific properties, NPs often interfere with the immune assays. They can adsorb cytokines on their surface, reducing their amounts in the supernatant, or interfere with

Figure 7 Most frequently studied endpoints for the in vivo (A) and in vitro/ex vivo (B) assessment of the immunotoxic potential of nanomaterials.

Abbreviations: NK, natural killer; NO, nitric oxide.
the absorbance and fluorescence measurements. Another issue is the contamination by the endotoxin, which activates the immune response leading to false-positive result. A thorough testing of the eventual sample contamination, appropriate controls and the use of several complementary assays should improve the quality of results and help with their interpretation.

Considerations for regulatory evaluations
Currently, nanotechnology-based medicinal products aiming to enter into clinical application have to follow the regulatory path of medicinal products. Relevant guidelines for their evaluation on quality, efficacy and safety have been published by the International Conference on Harmonization of Technical Requirements for Registration of Pharmaceuticals for Human Use (ICH). The potential of these innovative products to interfere with the immune system is assessed with the information based on the preclinical in vivo standard toxicity studies. Immunogenicity assessments are required for biotechnology-derived pharmaceuticals. For other nanotechnology-based products not involving biotechnology, changes in relevant parameters including hematology, globulin level in serum, immune organ weight and histology and incidences of infections or tumors can trigger additional immunotoxicity studies. However, even though this strategy is functioning for the majority of xenobiotics, it is uncertain whether more subtle NP-induced immune disorders can be detected on time.

The additional studies recommended in the ICH S8 guideline include T-cell–dependent antibody response, immunophenotyping, natural killer cell activity, host resistance studies, macrophage/neutrophil function and cell-mediated immunity performed in vivo, in vitro or ex vivo. However, as such, they cover only partially the immune effects frequently induced by nanomaterials, such as complement activation, inflammation or cytokine release (Table 1). Contrary to medicinal products, complement activation is routinely tested for medical devices. Besides the low predictivity of animal models, technical challenges such as the nanospecific optical and chemical properties that may interfere with the results of many conventional assays have to be taken into account. Therefore, many regulators are requesting that validated state-of-the-art in vitro methods containing specific nanosize-related precautions should be developed to assess the immunotoxic profile of NPs for medical use. The reflection papers issued by the European Medicines Agency in 2013 related to information needs for IV liposomal products, iron-based products and block copolymer micelle products recommend additional immunotoxicity studies including complement activation and macrophage activation, antigenicity and hypersensitivity reactions using validated in vitro and in vivo methods in sensitive models.

Up to now, there is only one standardized method for the evaluation of immunological response to nanoparticulate materials, developed by the American Society for Testing and Materials (ASTM). Another ASTM method refers

Table 1 Most relevant immune effects reported in the in vivo studies and corresponding regulatory guideline/guidance documents

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<th>Immune effect</th>
<th>Investigated endpoint</th>
<th>Model</th>
<th>Guideline/standard</th>
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<tr>
<td>Activation of innate</td>
<td>• Cytokine release&lt;sup&gt;16,28,37,88,108–110&lt;/sup&gt;</td>
<td>Rodents</td>
<td>Partially covered by ICH S8 as part of additional immunotoxicity studies</td>
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<td>immune cells</td>
<td>• Oxidative and nitrosative stress generation&lt;sup&gt;16,111&lt;/sup&gt;</td>
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<td>• NP uptake by macrophages&lt;sup&gt;26,53,108,112,113&lt;/sup&gt;</td>
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<td>Complement activation</td>
<td>• Complement detection in the plasma (C3, C3a, C5a, SC5b-9, hemolytic assays&lt;sup&gt;31,34,42&lt;/sup&gt;)</td>
<td>Pigs, mice, rats, patients, human blood</td>
<td>Not covered by the existing guidelines for medicinal products</td>
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<td></td>
<td>• Hypersensitivity reactions&lt;sup&gt;16,19,114&lt;/sup&gt;</td>
<td></td>
<td>Covered by ISO 10993-4 for medical devices</td>
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<tr>
<td>Adaptive immune response</td>
<td>• Immunophenotyping&lt;sup&gt;47,15–17&lt;/sup&gt;</td>
<td>Mice, rats, chicken, dogs</td>
<td>Partially covered by ICH S8 as a part of STS and by additional immunotoxicity studies</td>
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<tr>
<td></td>
<td>• Cytokine release&lt;sup&gt;47,118&lt;/sup&gt;</td>
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<td></td>
<td>• Antibodies production&lt;sup&gt;10,52,56,61&lt;/sup&gt;</td>
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<td></td>
<td>• Immune organ weight&lt;sup&gt;21,52,55,109&lt;/sup&gt;</td>
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<td>Inflammation</td>
<td>• Activation of inflammatory pathways&lt;sup&gt;17,24,27&lt;/sup&gt;</td>
<td>Rodents</td>
<td>Partially covered by ICH S8 as a part of STS</td>
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<tr>
<td></td>
<td>• Histopathology&lt;sup&gt;20,23,55&lt;/sup&gt;</td>
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<td></td>
<td>• Production of ROS and RNS&lt;sup&gt;22,19,120&lt;/sup&gt;</td>
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<td>Immunosuppression</td>
<td>• Cytotoxicity to immune cells&lt;sup&gt;48,67,120&lt;/sup&gt;</td>
<td>Rodents</td>
<td>Partially covered by ICH S8 as a part of additional immunotoxicity studies</td>
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<tr>
<td></td>
<td>• Lymphocyte proliferation&lt;sup&gt;16,78&lt;/sup&gt;</td>
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<td></td>
<td>• NK cell activity&lt;sup&gt;20,78&lt;/sup&gt;</td>
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<td>• Phagocytic function&lt;sup&gt;16,121&lt;/sup&gt;</td>
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<td></td>
<td>• Cytokine profiling&lt;sup&gt;42,71,122&lt;/sup&gt;</td>
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**Note:** ICH S8, ICH guideline for immunotoxicity studies for human pharmaceuticals, referring to STS and additional immunotoxicity studies. ISO 10993–4, ISO guidance “Biological evaluation of medical devices – Part 4: Selection of tests for interactions with blood”. **Abbreviations:** ICH, International Conference on Harmonization of Technical Requirements for Registration of Pharmaceuticals for Human Use; NK, natural killer; RNS, reactive nitrogen species; ROS, reactive oxygen species; STS, standard toxicity studies.
to assessment in vitro of the whole complement activation by solid materials used in medical devices. However, it does not address medicinal products or products based on nanotechnology. Complement activation, secretion of proinflammatory cytokines and leukocyte proliferation are part of the assay cascade provided by the European Nanomedicine Characterization Laboratory for the preclinical characterization of nanomaterials intended for medical use. More efforts are needed to develop and standardize test methods for assessment of the immunological response to nanotechnology-based pharmaceuticals.

**Conclusion**

Nanotechnology-based health products are essential for the implementation of personalized medicine. However, many of these products are triggering interaction with the immune system, leading to a compromised safety and efficacy. The activation of the immune system, including the activation of macrophages and the release of cytokines, is the most frequent effect of nanomaterials observed in vivo. Surface-related properties of the investigated nanomaterials were identified as the most critical parameter influencing the observed immune effect. Complement activation and related hypersensitivity reactions as well as antigenicity leading to the production of the specific antibodies and ABC were the main adverse effects described after the administration of lipid-based and polymer-based nanomaterials, the most relevant platforms currently used for the development of nanomedicines. Due to interspecies variations, the relevance of the animal studies used for this purpose remains limited. In consequence, relevant, suitable in vitro testing methods based on humanized cells should be developed for nanotechnology-based medicinal products.

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**Author contributions**

BHK performed the literature search, analyzed the results and drafted the manuscript. SBH conceived the study, helped in data interpretation and revised the manuscript. All authors contributed toward data analysis, drafting and revising the paper and agree to be accountable for all aspects of the work.

**Disclosure**

Any opinions expressed in this publication are those of the authors only, and this paper does not represent an official position of the European Commission. The authors report no conflicts of interest in this work.

**References**


111. Ilinskaya AN, Clogston JD, Mcneil SE, Dobrovolskaia MA. Induction of oxidative stress by Taxol® vehicle Cremophor-EL triggers production of interleukin-8 by peripheral blood mononuclear cells through the mechanism not requiring de novo synthesis of mRNA. Nanomedicine. 2015;11(8):1925–1938.


