Sirt1 protects neural stem cells from apoptosis by decreasing acetylation of histone 3K9

Chongdong Jian1,*
Cuihua Zou1,*
Ning Xu2
Guoying Chen2
Donghua Zou2
1Youjiang Medical University for Nationalities, Baise, Guangxi 533000, People’s Republic of China;
2Department of Neurology, The Fifth Affiliated Hospital of Guangxi Medical University, Nanning, Guangxi 530021, People’s Republic of China

*These authors contributed equally to this work

Objective: To explore the role and mechanism of Sirt1 in protecting neural stem cells (NSCs) from apoptosis.

Materials and methods: Transfection was used to overexpress Sirt1 in rat NSCs. The effect of Sirt1 overexpression on camptothecin-induced apoptosis of NSCs was evaluated. Western blotting was used to examine the expression of Sirt1, cleaved caspase-3, and acetylated histone 3K9.

Results: Overexpression of Sirt1 in NSCs decreased the cleavage of caspase-3 and acetylation of histone 3K9.

Conclusion: Sirt1 may protect NSCs from apoptosis by decreasing the acetylation of histone 3 on K9.

Keywords: neural stem cells, apoptosis, Sirt1, caspase-3, acetylated histone 3K9

Introduction
Our previous study found that proliferation of endogenous neural stem cells (NSCs) was temporarily enhanced in the p25 transgenic mouse model; however, the survival of NSCs was impaired.1 NSCs undergo apoptosis following temporary enhanced proliferation. Therefore, intervening strategies that can enhance the survival of NSCs may be beneficial for the treatment of neural injuries or degenerative diseases.

Originally identified in yeast, sirtuins are nicotinamide adenine dinucleotide (NAD+)-dependent deacetylases that play an important role in longevity.2 There are seven sirtuins in mammals. Sirt1 is involved in many biological functions, such as oxidative stress and inflammatory responses, glucose and lipid metabolism, autophagy, cell mitosis, apoptosis, cell cycle regulation, cell proliferation, cell senescence, and metabolism.3,4 In addition, Sirt1 plays important roles in the occurrence and development of neurons, the maintenance of normal neuronal functions, and the protection of neurons.5,6 However, its role in protecting NSCs from apoptosis has not been elucidated. In this study, we overexpressed Sirt1 in rat NSCs and induced cellular injury with camptothecin to determine the role and mechanism of Sirt1 in protecting NSCs during apoptosis.

Materials and methods
NSC culture, transfection, and Western blotting were performed as previously published.1 The rat NSC line was obtained from the Fred Gage Laboratory7 (The Salk Institute, La Jolla, CA, USA). Briefly, 1 µL of human simplex virus (MIT viral core facility, Cambridge, MA, USA) expressing flag or flag-Sirt1 was added to the medium to transfect
NSCs. Transfected NSCs were transferred to 6-well plates. Different concentrations of camptothecin (0, 0.5, 2, and 10 µM) were added to the medium for 16 hours and cells were collected for Western blot analysis. Total proteins were extracted from cultured NSCs using lysis buffer. The primary antibodies used in this study were anti-cleaved caspase-3 (1:500, Cell Signaling Technology; Denver, MA, USA), anti-acetyl-Histone3K9 (1:500, Cell Signaling Technology), anti-flag (1:1,000, Sigma-Aldrich Co., St. Louis, MO, USA), and anti-α-tubulin (1:2,000, Cell Signaling Technology). Protein bands were captured and analyzed using Quantity One software for OD values.

**Results**

Overexpression of flag-Sirt1 in rat NSCs decreased the level of cleaved caspase-3 protein when the cells were treated with camptothecin (Figure 1A). Similarly, the levels of acetylated histone 3K9 were reduced following camptothecin treatment (Figure 1B).

**Discussion**

Sirt1 plays important roles in many pathophysiological processes by deacetylating various substrates. Studies have reported that Sirt1 can inhibit apoptosis through various

**Figure 1** Overexpression of Sirt1 protects neural stem cells from camptothecin-induced cell death.

**Notes:** (A) Overexpression of flag-Sirt1 in neural stem cells reduced the elevated level of cleaved caspase-3 induced by camptothecin. IB: flag confirmed flag-Sirt1 expression by Western blotting. IB: cleaved caspase-3 detection of apoptosis in flag or flag-Sirt1 in different treatments of camptothecin. (B) Overexpression of flag-Sirt1 in neural stem cells reduced the level of acetylated histone 3K9. IB: flag confirmed flag-Sirt1 expression by Western blotting. IB: Sirt1: detection of endogenous Sirt1 in control and overexpression Sirt1 group by using anti-Sirt1 antibody. IB: H3K9 (A) and H3: representative Western blotting and summary statistics of acetylation of histone 3 at K9, normalized to total histone 3.

**Abbreviation:** IB, immunoblotting.
pathways in tissues. For example, Sirt1 reduced endoplasmic reticulum stress and apoptosis of brown adipocytes in vivo and in vitro by inhibiting Smad3/ATF4 signaling. Activation of Sirt1 promotes recovery of mitochondrial proteins and functions by increasing mitochondrial biogenesis and by reducing apoptosis following intracerebral hemorrhage via the PGC-1α mitochondrial pathway. Sirt1 may attenuate endoplasmic reticulum stress-induced cardiomyocyte apoptosis via PERK/eIF2α, ATF6/CHOP, and IRE1α/JNK-mediated pathways. Additionally, Sirt1 inhibits apoptosis by deacetylating p53.

There are limited reports on whether Sirt1 can protect NSCs from apoptosis. We found that overexpressing Sirt1 in rat NSCs resulted in reduced levels of cleaved caspase-3 and reduced apoptosis in response to camptothecin, suggesting that Sirt1 protects NSCs from apoptosis. In addition, the expression of acetylated histone 3K9 in NSCs was reduced. Acetylation of histone 3K9 is frequently associated with DNA damage. Sirt1 can remove acetyl groups from histone lysine residues, resulting in deacetylation of histone H3 on K9. Thus, our data suggest that Sirt1 could reduce DNA damage that triggers apoptosis.

Conclusion

In conclusion, Sirt1 may protect NSCs from apoptosis by decreasing the acetylation of histone 3 on K9.

Acknowledgments

This study was supported by the National Natural Science Foundation of China (grant number: 81860244), Guangxi Natural Science Foundation (grant number: 2016GXNSFA380012), the Basic Ability Enhancement Program for Young and Middle-age Teachers of Guangxi (grant number: 2017KY0516), the Project of Nanning Scientific Research and Technology Development Plan (grant number: 20163142), the Scientific Research Project of Guangxi Health and Family Planning Commission (grant number: Z20170001), and the China Scholarship Council (201708455059).

Disclosure

The authors report no conflicts of interest in this work.

References