Pooling analysis on prognostic value of PHH3 expression in cancer patients

Qian Hao^{1,*}
 Cong Dai^{1,*}
 Yujiao Deng^{1,*}
 Peng Xu¹
 Tian Tian¹
 Shuai Lin¹
 Meng Wang¹
 Kang Liu¹
 Dingli Song¹
 Ying Wu¹
 Yan Guo²
 Zhijun Dai¹

¹Department of Oncology, Second Affiliated Hospital of Xi'an Jiaotong University, Xi'an, People's Republic of China; ²School of Life Science and Technology, Xi'an Jiaotong University, Xi'an, People's Republic of China

*These authors contributed equally to this work

Correspondence: Zhijun Dai
 Department of Oncology, Second Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710004, Shaanxi, People's Republic of China
 Email dzj0911@126.com

Yan Guo
 School of Life Science and Technology, Xi'an Jiaotong University, Xi'an 710049, Shaanxi, People's Republic of China
 Tel +86 029 87679226
 Email guoyan253@xjtu.edu.cn

Background: Various studies have evaluated the significance of phospho-histone-H3 (PHH3) expression in cancer patients, but controversy over its reliability remains. We conducted a meta-analysis to summarize the prognostic relevance of PHH3 expression in cancer patients.

Patients and methods: Nineteen studies, including 4803 patients, were identified by searching PubMed, Web of Science, Embase, and Cochrane Library. The correlation of PHH3 expression level with overall survival (OS), disease-free survival, and recurrence-free survival was analyzed.

Results: Overall, the results suggest that high expression of PHH3 can predict a poor OS (HR=2.66, 95% CI=1.74–4.08, $P<0.001$), disease-free survival (HR=3.40, 95% CI=1.47–7.87, $P=0.004$), and recurrence-free survival (HR=2.80, 95% CI=1.61–4.85, $P<0.001$) in cancer patients. The subgroup analysis showed that highly expressed PHH3 was significantly related to breast cancer (HR=5.66, 95% CI=2.72–11.78, $P<0.001$) and urogenital tumors (HR=3.01, 95% CI=1.78–5.09, $P<0.001$). Furthermore, no significant difference was found between Asian (HR=1.98, 95% CI=1.08–3.63, $P=0.026$) and Caucasian populations (HR=3.01, 95% CI=1.87–4.85, $P<0.001$) regarding OS and PHH3 expression.

Conclusion: This meta-analysis indicates that high expression of PHH3 may serve as a biomarker for poor prognosis in patients with cancer.

Keywords: PHH3, prognosis, human cancer, meta-analysis, survival

Introduction

Cancer is still considered to be a major challenge for modern medicine and a major public health problem worldwide.¹ Tumor-specific markers can be useful for monitoring and evaluating tumor progression and therapeutic efficacy. However, there is still a shortage of highly sensitive and specific markers for assessing cancer progression and prognosis.² Therefore, it is necessary for decision-making concerning clinical therapy to figure out an effective pretreatment parameter that can be used to evaluate survival probability and prognosis of cancer patients.

Phospho-histone-H3 (PHH3) is a core histone protein, which in its phosphorylated state forms the principal constituents of eukaryotic chromatin, with histone H3 being phosphorylated at serine (Ser) 10 or Ser28 as well as its phosphorylation of Ser10 being strongly correlated with the late G2 to M-phase transition in mammalian mitotic cells.³ On the basis of previous research, a few cell line- and animal model-based researches have displayed an increase in phosphorylation of histone H3 at Ser10 (H3S10ph), the only histone marker that is involved in carcinogenesis and cellular transformation.^{4–7}

Also, earlier in vitro studies have suggested that a higher level of H3S10ph alone directly participated in cellular transformation.⁵ PHH3 has long been accepted as a cellular proliferation marker in a large variety of cancers, and it was revealed to be of prognostic value in several recent studies.^{8–10} For instance, PHH3 expression is elevated in gliomas tissues, and patients with high PHH3 expression have a shorter overall survival (OS) compared with patients with lower expression levels of PHH3.⁸ Elevated PHH3 expression is also detected in prostate carcinoma tissues and is strongly correlated with an advanced clinical stage.⁹ In breast cancer, increased PHH3 expression is correlated with an lymph vessel invasion.¹⁰ However, the association between elevated PHH3 expression and prognosis has not been identified in patients with gastrointestinal stromal tumors.¹¹ Therefore, we conducted this meta-analysis to summarize the global findings from analyses of PHH3 expression as a predictive indicator of clinical results in cancer patients.

Patients and methods

Search strategy

We performed a network search using PubMed, Web of Science, Embase, and Cochrane Library for original articles that analyzed the prognostic value of PHH3 in various cancers, with the last search update on February 1, 2018. The search strategy included the following keywords combined with “phospho-histone-H3:” “PHH3”, “neoplasm”, “cancer”, “prognostic”, and “prognosis”. Furthermore, additional eligible studies were collected by a manual search for relevant research from reference lists.

Selection criteria

The studies were considered eligible if they met all the following criteria: 1) studies that evaluated the relationship of PHH3 expression in cancer patients with detailed information about OS, disease-free survival (DFS), or recurrence-free survival (RFS); 2) HRs and 95% CIs, raw data, or survival curves were reported in the article; and 3) the articles were written in English or Chinese. Articles were excluded based on the following criteria: 1) meeting abstract, review or laboratory articles, as well as case studies or letters; 2) articles that described the survival outcome of other indicators; 3) duplicate publications; and 4) animal studies.

Quality assessment

To control the quality of the research, the Newcastle–Ottawa scale, ranging from 0 to 9 stars,¹² was used to assess the quality of the eligible papers. Studies with more than six stars were considered high quality. Two authors (Qian Hao and

Yujiao Deng) performed the assessment independently, and any disagreement was resolved by discussion.

Data extraction

Two independent reviewers (Qian Hao and Cong Dai) extracted the details of included studies using a standardized form, and any disagreements were resolved through discussion with a third reviewer (Zhijun Dai). The following information was recorded: the first author's name, year of publication, number of patients, patient source, tumor types, PHH3 assessment method, prognostic outcomes, analytical method, and HR with the corresponding 95% CI.

Statistical analysis

Statistical analyses were conducted using STATA version C14.0 (Stata Corporation, College Station, TX, USA). The HRs with their 95% CIs were used for evaluating the strength of the correlations between PHH3 and the survival outcomes. We evaluated the study heterogeneity (I^2) by chi-squared test. All statistical tests were two-sided, and the significance level was set at 0.05. The fixed effects model was used when no obvious heterogeneity was observed among the included studies ($I^2 < 50\%$), otherwise the random effects model was selected to perform the meta-analysis. Subgroup analyses were conducted to investigate the potential factors of heterogeneity. Publication bias was estimated by Begg's funnel plot and Egger's test. Sensitivity analysis was performed by sequential omission of each study.

Results

Characteristics of included studies

Based on the study design, we initially searched a total of 410 articles. After duplicates were removed, 216 articles remained. Hundred and seventy-four articles qualified for review after we screened titles and abstracts. Subsequently, 122 studies were assessed by screening the full text, of which 103 articles were excluded ([Table S1](#)). Finally, 19 studies, published between 2011 and 2017, were included in this meta-analysis as shown in Figure 1. The Newcastle–Ottawa scores of these studies ranged between 6 and 8. The total number of patients included were 4803, ranging between 27 and 637 patients per study. The main characteristics of the 19 articles were extracted and are summarized in Table 1. The patients were from People's Republic of China,^{8,13,14} Korea,¹⁵ Norway,^{10,16–18} India,¹⁹ Germany,^{9,20} the United Kingdom,^{21,22} the United States of America,^{11,23} New Zealand,²⁴ Sweden,²⁵ Australia,²⁶ and Italy.²⁷ The types of carcinoma included glioma, neuroblastoma, gastric cancer, gastroin-

Figure 1 Flow diagram of the study selection process.

testinal stromal tumor, Merkel cell carcinoma, cutaneous melanoma, uterine smooth muscle tumor, endometrial cancer, adrenocortical carcinoma, prostate cancer, and breast cancer. Immunohistochemical staining was used to determine the level of PHH3 expression.

Association of PHH3 expression with OS in human cancer

Twelve studies comprising 1259 cancer patients were included in the survival analysis. The pooled HR was 2.66 (95% CI=1.74–4.08, $P<0.001$; Figure 2 and Table 2). High expression of PHH3 was significantly related to poor OS of cancer patients. Due to the emergence of clear heterogeneity

in this study ($I^2=74.5\%$, $P<0.001$; Table 2), a subgroup analysis was conducted to explore the source of heterogeneity further. The subgroups included tumor types, ethnicity, and HR estimate (Table 2). High PHH3 expression was significantly associated with reduced OS in breast cancer (HR=5.66, 95% CI=2.72–11.78, $P<0.001$) and urogenital tumors (HR=3.01, 95% CI=1.78–5.09, $P<0.001$), but not with nervous system tumors (HR=1.58, 95% CI=0.76–3.28, $P=0.226$), digestive system tumors (HR=1.57, 95% CI=0.73–3.35, $P=0.247$), or cutaneous tumors (HR=2.99, 95% CI=0.85–10.56, $P=0.089$). There was clear heterogeneity in nervous system tumors ($I^2=69.4\%$, $P=0.071$) and digestive system tumors ($I^2=56.6\%$, $P=0.129$), while

Table 1 Main characteristics of all studies included in the meta-analysis

First author	Year	Patient source	Number of patient	Tumor types	Specimen	Method	PHH3 high expression (%)	Median follow-up (month)	Outcome	M/U	HR (95% CI)	NOS
Kim et al ¹⁵	2017	Korea	218	BC	Tissue	IHC	40.30	46	DFS	U	8.90 (1.07–74.0)	9
Chow et al ¹³	2017	People's Republic of China	55	USMT	Tissue	IHC	–	65.5	OS	M	2.71 (1.06–6.94)	7
Zhu et al ⁸	2016	People's Republic of China	61	Gliomas	Tissue	IHC	51.67	44.3	OS	M	1.19 (1.01–1.41)	8
Winther et al ¹⁶	2016	Norway	160	Meningiomas	Tissue	IHC	–	–	RFS	M	3.05 (1.46–6.58)	7
Khan et al ¹⁹	2016	India	101	GC	Tissue	IHC	58.42	–	OS	U	2.15 (1.07–4.28)	8
Goltz et al ⁹	2015	Germany	637	PC	Tissue	IHC	–	51	DFS	U	1.44 (0.61–3.41)	8
Iwasaki et al ²¹	2015	UK	27	MCC	Tissue	IHC	–	42	RFS	U	1.84 (1.09–3.11)	8
Ramani et al ²²	2015	UK	80	Neuroblastomas	Tissue	IHC	30.00	121.3	OS	M	7.47 (1.41–39.61)	6
Alkhasawneh et al ¹¹	2015	USA	50	GSTS	Tissue	IHC	34.00	56.4	OS	M	2.60 (1.10–5.80)	8
Gerring et al ²⁴	2015	New Zealand	108	BC	Tissue	IHC	–	85	OS	U	0.97 (0.36–2.62)	7
Olar et al ²³	2015	USA	363	Meningiomas	Tissue	IHC	15.42	–	OS	M	4.35 (2.17–8.71)	7
Duregon et al ²⁷	2014	Italy	48	AC	Tissue	IHC	–	40	RFS	U	1.03 (1.02–1.05)	8
Xu ¹⁴	2014	China	52	PC	Tissue	IHC	94.23	–	OS	U	2.22 (1.0–5.0)	6
Klintman et al ²⁵	2013	Sweden	221	BC	Tissue	IHC	30.77	–	OS	M	5.43 (1.24–23.80)	6
Gudlaugsson et al ¹⁷	2013	Norway	237	BC	Tissue	IHC	35.44	–	DFS	U	2.40 (1.40–4.30)	8
Ladstein et al ¹⁸	2012	Norway	338	CNM	Tissue	IHC	74.85	61.5	RFS	M	10.6 (4.4–25.6)	8
Brunner et al ²⁶	2012	Australia	99	EC	Tissue	IHC	48.48	72	DFS	M	8.9 (3.4–23.5)	9
Gudlaugsson et al ¹⁰	2011	Norway	240	BC	Tissue	IHC	35.00	117	OS	M	1.90 (1.04–3.36)	8
Kim et al ²⁰	2007	Germany	99	Meningiomas	Tissue	IHC	–	42	OS	M	5.9 (1.7–31.4)	9
									RFS	M	9.6 (3.1–29.9)	8
											1.07 (1.06–1.10)	8

Note: ‘–’ unobtainable data.

Abbreviations: AC, adrenocortical carcinoma; BC, breast cancer; CNM, cutaneous nodular melanoma; DFS, disease-free survival; EC, endometrial carcinoma; GC, gastric cancer; GSTS, gastrointestinal stromal tumors; IHC, immunohistochemistry; M, multivariate; MCC, Merkel cell carcinoma; NOS, Newcastle–Ottawa scale; OS, overall survival; PC, prostate carcinoma; PHH3, phospho-histone-H3; RFS, recurrence-free survival; U, univariate; USMT, uterine smooth muscle tumor.

Figure 2 Forest plot of HR for the association of PHH3 expression and OS.

Note: Weights are from random effects analysis.

Abbreviations: PHH3, phospho-histone-H3; OS, overall survival.

Table 2 Main meta-analysis results

Analysis	No. of studies	No. of patients	Model	HR (95% CI)	P-value	Heterogeneity		Publication bias	
						I^2 (%)	P-value	Begg's P	Egger's P
OS	12	1259	Random	2.66 (1.74–4.08)	0.000	74.5	0.000	0.047	0.000
Tumor type	–	–	–	–	–	–	–	–	–
Nervous system tumor	2	141	Random	1.58 (0.76–3.28)	0.226	69.4	0.071	–	–
Digestive system tumor	2	151	Random	1.57 (0.73–3.35)	0.247	39.4	0.199	–	–
Cutaneous tumor	2	365	Random	2.99 (0.85–10.56)	0.089	56.6	0.129	–	–
Breast cancer	2	348	Random	5.66 (2.72–11.78)	0.000	26.7	0.243	–	–
Urogenital tumor	4	254	Random	3.01 (1.78–5.09)	0.000	0.00	0.567	–	–
Ethnicity	–	–	–	–	–	–	–	–	–
Asian	4	269	Random	1.98 (1.08–3.63)	0.026	66.6	–	–	–
Caucasian	8	990	Random	3.01 (1.87–4.85)	0.000	51.0	–	–	–
HR estimate	–	–	–	–	–	–	–	–	–
Multivariate analysis	7	1033	Random	3.17 (1.66–6.04)	0.000	81.8	0.000	–	–
Univariate analysis	5	226	Random	1.93 (1.31–2.82)	0.001	74.5	0.000	–	–
DFS	4	777	Random	3.40 (1.47–7.87)	0.004	67.4	0.027	–	–
RFS	5	1496	Random	2.80 (1.61–4.85)	0.000	72.6	0.006	–	–

Note: '–' unobtainable data.

Abbreviations: DFS, disease-free survival; OS, overall survival; RFS, recurrence-free survival.

no obvious heterogeneity was found in other tumor types. Subgroup analysis showed that the correlation between OS and PHH3 expression did not differ between ethnicities (Asian: HR=1.98, 95% CI=1.08–3.63, $P=0.026$; Caucasian: HR=3.01, 95% CI=1.87–4.85, $P<0.001$) and HR estimates (multivariate analysis: HR=3.17, 95% CI=1.66–6.04; univariate analysis: HR=1.93, 95% CI=1.31–2.82).

Association of PHH3 expression with DFS and RFS

Four studies comprising 777 patients were included in determining the relationship between elevated PHH3 expression and DFS. Correspondingly, five studies involving 1496 patients were included for determining the association between PHH3 overexpression and RFS. The pooled outcome indicated that high expression levels of PHH3 might imply poor DFS (HR=3.40; 95% CI=1.47–7.87, $P=0.004$; Figure 3 and Table 2) and RFS (HR=2.80; 95% CI=1.61–4.85, $P<0.001$; Figure 4 and Table 2). Furthermore, significant heterogeneity was noted among the studies (DFS: $I^2=67.4\%$, $P=0.027$; RFS: $I^2=72.6\%$, $P=0.006$). However, we did not

conduct subgroup analyses to determine sources of heterogeneity because of the small number of studies.

Sensitivity analyses and publication bias

In order to determine whether an individual study had a significant influence on pooled HR and to confirm the stability and reliability of HR estimates, all the studies were sequentially removed. We found that the pooled HRs were not significantly influenced by any single study (Figure 5). Both the Begg's and Egger's tests were performed to assess if any publication bias existed in the included studies. The funnel plot (Figure 6) was asymmetrical. Begg's test ($P=0.193$) revealed no publication bias among the 12 eligible studies, while Egger's test ($P=0.002$) showed a clear bias.

Discussion

PHH3, a chief protein component of nucleosomes in eukaryotic cells, is a less extensively studied proliferation factor. The clear and contrast-rich staining of PHH3 is easily assessed, with high inter-observer reproducibility, making it the subject of intense research in recent years.^{28,29} PHH3 has been found

Figure 3 Forest plot of HR for the association of PHH3 expression and DFS.

Note: Weights are from random effects analysis.

Abbreviations: DFS, disease-free survival; PHH3, phospho-histone-H3.

Figure 4 Forest plot of HR for the association of PHH3 expression and RFS.
Note: Weights are from random effects analysis.
Abbreviations: PHH3, phospho-histone-H3; RFS, recurrence-free survival.

Figure 5 Sensitivity analysis of pooled HRs on the association between PHH3 expression and OS.
Abbreviations: OS, overall survival; PHH3, phospho-histone-H3.

Figure 6 Funnel plots of publication bias for all of the included studies reported with OS.
Abbreviations: log[HR], logarithm hazard ratio; OS, overall survival; s.e., standard error.

to be an independent prognostic factor in meningioma, cutaneous melanoma, and gastric cancer.^{18,20,30,31} We conducted this meta-analysis to mitigate sample size problems of individual studies and enhance their statistical power. Our study is the first meta-analysis to summarize research on PHH3 and its prognostic role in various types of cancer.

In this meta-analysis, we analyzed 19 studies including 4803 patients, with OS data from 12 studies, DFS data from 4 studies, and RFS data from 5 studies, which revealed that a high expression of PHH3 was correlated with poor OS, DFS, and RFS and significantly associated with a reduced OS in patients with breast cancer and urogenital tumors. Thus, PHH3 expression may be considered as a potential prognostic factor for cancer patients. It is well known that Ki-67 is commonly used to assess cell proliferation. However, according to Takahashi et al and Brenner et al, PHH3 expression was an independent factor for poor prognosis and an improved mitotic marker in gastric cancer and endometrial tissues compared to Ki-67.^{31,32} Likewise, some previous studies have noted that PHH3 is an excellent predictability marker of survival in long- and short-term follow-up.^{15,24,25} Therefore, we collected and analyzed the results of several studies and found that PHH3 is a novel, useful proliferation marker that can be used as a prognostic indicator in astrocytoma, melanoma, prostate carcinoma, pituitary adenoma, and breast cancer.^{25,28,29,33–36} PHH3 may also be a prognostic marker in various other types of cancer. To further analyze the role of PHH3 in different cancers, a subgroup analysis

was performed. PHH3 expression was found to be statistically associated with breast, gynecological, cutaneous, and urogenital tumors.

Through the subgroup analysis, we found that the heterogeneity of our study originated from tumors of the nervous and digestive systems. Additionally, PHH3 doesn't have standard testing methods and it's also not routinely utilized. Its practical utility is limited so far, different phosphorylation sites (Ser10 and Ser28) and measurement units were used in these studies in different labs, detection assays, antibodies, and thresholds for positivity, given that it may cause significant discrepancies and biases in our studies. Furthermore, we conducted subgroup analyses of study region and analysis method; however, no significant differences were detected. It is important to note that there was a publication bias, suggesting that large-scale investigations are required. An inadequate number of included studies analyzed the OS, DFS, and RFS rates, with controversial conclusions. Overall, our meta-analysis supports the conclusion that PHH3 may be a predictive biomarker of OS, DFS, and RFS. Thus, further research is necessary to evaluate the relationship between overexpression of PHH3 and outcomes in human cancers.

Moreover, we found that some studies have identified a close correlation between PHH3 and mitotic cell condensation.^{37,38} Immunohistochemical staining of PHH3 has suggested that modification occurs almost exclusively in actively proliferating cells during M-phase, not during apoptosis.³⁹ From a clinical standpoint, PHH3 immunostaining

has been validated in pulmonary melanoma, neuroendocrine carcinomas, and pancreatic cancer. Several previous studies have measured PHH3 levels among different types of cancers. Studies suggested that PHH3 index increased with higher grade of tumor, including cancers of breast, ovarian, melanoma, and meningioma.^{20,29,40–43} Therefore, PHH3 staining not only has prognostic value but also supports tumor grading by facilitating mitotic counting. PHH3 staining can be easily identified and can minimize inter-observer and inter-laboratory technical variations. In addition, one study suggested that the PHH3 index is a more sensitive measure of mitotic activity in Merkel cell carcinoma than the hematoxylin and eosin (H&E)-stained mitotic count index. Thus, it is believed to be a useful complementary diagnostic tool for standard H&E mitotic counts. Another study confirms that PHH3 was a better proliferation marker than Ki-67 due to its high reproductivity of immunohistochemistry, accuracy, and consistency among raters in breast cancer.¹⁵ Finally, the “true” negative resection margin in gastric cancer has reportedly been determined using the distance-dependent association of PHH3 with clinical parameters. Therefore, PHH3 could be helpful in limiting the extent of resection and preventing post-surgical locoregional recurrence of the disease.¹⁹

Generally, PHH3 expression levels are helpful in predicting the prognosis of different cancers, and PHH3 staining can be a useful complementary tool to routine methods in grading and determining “true” negative resection margins.

Conclusion

In summary, we found that increased expression of PHH3 indicated poor survival outcomes in patients with cancer, and therefore, PHH3 is a potential novel prognostic indicator in cancer patients.

Acknowledgment

This work was supported by the National Natural Science Foundation of China (No. 81471670) and the Key Research and Development Plan, Shaanxi Province, China (2017ZDXM-SF-066).

Author contributions

Qian Hao and Cong Dai designed the study. Qian Hao, Yujiao Deng, and Peng Xu wrote the manuscript. Tian Tian, Shuai Lin, Meng Wang, and Kang Liu extracted the relevant studies and data. Dingli Song, Ying Wu, and Yan Guo analyzed the data. All authors contributed toward data analysis, drafting, revision of and final approval of the paper, and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. *CA Cancer J Clin*. 2018;68(1):7–30.
2. Pritzker KP. Predictive and prognostic cancer biomarkers revisited. *Expert Rev Mol Diagn*. 2015;15(8):971–974.
3. Fukushima S, Terasaki M, Sakata K, et al. Sensitivity and usefulness of anti-phosphohistone-H3 antibody immunostaining for counting mitotic figures in meningioma cases. *Brain Tumor Pathol*. 2009;26(2):51–57.
4. Chadee DN, Hendzel MJ, Tyllipski CP, et al. Increased Ser-10 phosphorylation of histone H3 in mitogen-stimulated and oncogene-transformed mouse fibroblasts. *J Biol Chem*. 1999;274(35):24914–24920.
5. Choi HS, Choi BY, Cho YY, et al. Phosphorylation of histone H3 at serine 10 is indispensable for neoplastic cell transformation. *Cancer Res*. 2005;65(13):5818–5827.
6. Ibuki Y, Toyooka T, Zhao X, Yoshida I. Cigarette sidestream smoke induces histone H3 phosphorylation via JNK and PI3K/Akt pathways, leading to the expression of proto-oncogenes. *Carcinogenesis*. 2014;35(6):1228–1237.
7. Li B, Huang G, Zhang X, et al. Increased phosphorylation of histone H3 at serine 10 is involved in Epstein-Barr virus latent membrane protein-1-induced carcinogenesis of nasopharyngeal carcinoma. *BMC Cancer*. 2013;13:124.
8. Zhu P, Zhang CB, Yang P, et al. Phosphohistone H3 (pHH3) is a prognostic and epithelial to mesenchymal transition marker in diffuse gliomas. *Oncotarget*. 2016;7(29):45005–45014.
9. Goltz D, Montani M, Braun M, et al. Prognostic relevance of proliferation markers (Ki-67, PHH3) within the cross-relation of ERG translocation and androgen receptor expression in prostate cancer. *Pathology*. 2015;47(7):629–636.
10. Gudlaugsson E, Skaland I, Undersrud E, Janssen EA, Soiland H, Baak JP. D2-40/p63 defined lymph vessel invasion has additional prognostic value in highly proliferating operable node negative breast cancer patients. *Mod Pathol*. 2011;24(4):502–511.
11. Alkhasawneh A, Reith JD, Toro TZ, et al. Interobserver variability of mitotic index and utility of PHH3 for risk stratification in gastrointestinal stromal tumors. *Am J Clin Pathol*. 2015;143(3):385–392.
12. Stang A. Critical evaluation of the Newcastle–Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. *Eur J Epidemiol*. 2010;25(9):603–605.
13. Chow KL, Tse KY, Cheung CL, et al. The mitosis-specific marker phosphohistone-H3 (PHH3) is an independent prognosticator in uterine smooth muscle tumours: an outcome-based study. *Histopathology*. 2017;70(5):746–755.
14. Xu H. *Expression of EN2, PHH3 in Prostate Cancer and Prognostic Significance* [master's thesis]. Taiyuan: Shanxi Medical University; 2014.
15. Kim JY, Jeong HS, Chung T, et al. The value of phosphohistone H3 as a proliferation marker for evaluating invasive breast cancers: a comparative study with Ki67. *Oncotarget*. 2017;8(39):65064–65076.
16. Winther TL, Arnli MB, Salvesen Ø, Torp SH. Phosphohistone-H3 proliferation index is superior to mitotic index and MIB-1 expression as a predictor of recurrence in human meningiomas. *Am J Clin Pathol*. 2016;146(4):510–520.
17. Gudlaugsson E, Klos J, Skaland I, et al. Prognostic comparison of the proliferation markers (mitotic activity index, phosphohistone H3, Ki67), steroid receptors, HER2, high molecular weight cytokeratins and classical prognostic factors in T(1)(-)(2)N(0)M(0) breast cancer. *Pol J Pathol*. 2013;64(1):1–8.
18. Ladstein RG, Bachmann IM, Straume O, Akslen LA. Prognostic importance of the mitotic marker phosphohistone H3 in cutaneous nodular melanoma. *J Invest Dermatol*. 2012;132(4):1247–1252.

19. Khan SA, Amnekar R, Khade B, et al. p38-MAPK/MSK1-mediated overexpression of histone H3 serine 10 phosphorylation defines distance-dependent prognostic value of negative resection margin in gastric cancer. *Clin Epigenetics*. 2016;8:88.
20. Kim YJ, Ketter R, Steudel WI, Feiden W. Prognostic significance of the mitotic index using the mitosis marker anti-phosphohistone H3 in meningiomas. *Am J Clin Pathol*. 2007;128(1):118–125.
21. Iwasaki T, Matsushita M, Nonaka D, et al. Phosphohistone-H3 (PHH3) is prognostic relevant in Merkel cell carcinomas but Merkel cell polyomavirus is a more powerful prognostic factor than AJCC clinical stage, PHH3, Ki-67 or mitotic indices. *Pathol Int*. 2015;65(8):404–409.
22. Ramani P, Taylor S, Miller E, Sowa-Avugrah E, May MT. High phosphohistone H3 expression correlates with adverse clinical, biological, and pathological factors in neuroblastomas. *J Histochem Cytochem*. 2015;63(6):397–407.
23. Olar A, Wani KM, Sulman EP, et al. Mitotic index is an independent predictor of recurrence-free survival in meningioma. *Brain Pathol*. 2015;25(3):266–275.
24. Gerring Z, Pearson JF, Morrin HR, Robinson BA, Harris GC, Walker LC. Phosphohistone H3 outperforms Ki67 as a marker of outcome for breast cancer patients. *Histopathology*. 2015;67(4):538–547.
25. Klintman M, Strand C, Ahlin C, et al. The prognostic value of mitotic activity index (MAI), phosphohistone H3 (PPH3), cyclin B1, cyclin A, and Ki67, alone and in combinations, in node-negative premenopausal breast cancer. *PLoS One*. 2013;8(12):e81902.
26. Brunner A, Riss P, Heinze G, Brustmann H. pHH3 and survivin are co-expressed in high-risk endometrial cancer and are prognostic relevant. *Br J Cancer*. 2012;107(1):84–90.
27. Duregon E, Molinaro L, Volante M, et al. Comparative diagnostic and prognostic performances of the hematoxylin-eosin and phospho-histone H3 mitotic count and Ki-67 index in adrenocortical carcinoma. *Mod Pathol*. 2014;27(9):1246–1254.
28. Skaland I, Janssen EA, Gudlaugsson E, Hui Ru Guo L, Baak JP. The prognostic value of the proliferation marker phosphohistone H3 (PPH3) in luminal, basal-like and triple negative phenotype invasive lymph node-negative breast cancer. *Cell Oncol*. 2009;31(4):261–271.
29. Skaland I, Janssen EA, Gudlaugsson E, et al. Phosphohistone H3 expression has much stronger prognostic value than classical prognosticators in invasive lymph node-negative breast cancer patients <55 years of age. *Mod Pathol*. 2007;20(12):1307–1315.
30. Takei H, Bhattacharjee MB, Rivera A, Dancer Y, Powell SZ. New immunohistochemical markers in the evaluation of central nervous system tumors: a review of 7 selected adult and pediatric brain tumors. *Arch Pathol Lab Med*. 2007;131(2):234–241.
31. Takahashi H, Murai Y, Tsuneyama K, et al. Overexpression of phosphorylated histone H3 is an indicator of poor prognosis in gastric adenocarcinoma patients. *Appl Immunohistochem Mol Morphol*. 2006;14(3):296–302.
32. Brenner RM, Slayden OD, Rodgers WH, et al. Immunocytochemical assessment of mitotic activity with an antibody to phosphorylated histone H3 in the macaque and human endometrium. *Hum Reprod*. 2003;18(6):1185–1193.
33. Sambrook J, Russell DW. Dephosphorylation of plasmid DNA. *CSH Protoc*. 2006;2006(1).
34. Ali HR, Dawson SJ, Blows FM, Provenzano E, Pharoah PD, Caldas C. Aurora kinase A outperforms Ki67 as a prognostic marker in ER-positive breast cancer. *Br J Cancer*. 2012;106(11):1798–1806.
35. Jonsdottir K, Assmus J, Slewa A, et al. Prognostic value of gene signatures and proliferation in lymph-node-negative breast cancer. *PLoS One*. 2014;9(3):e90642.
36. Nowak M, Svensson MA, Carlsson J, et al. Prognostic significance of phospho-histone H3 in prostate carcinoma. *World J Urol*. 2014;32(3):703–707.
37. Goto H, Tomono Y, Ajiro K, et al. Identification of a novel phosphorylation site on histone H3 coupled with mitotic chromosome condensation. *J Biol Chem*. 1999;274(36):25543–25549.
38. Juan G, Traganos F, Darzynkiewicz Z. Histone H3 phosphorylation in human monocytes and during HL-60 cell differentiation. *Exp Cell Res*. 1999;246(1):212–220.
39. Sun A, Zhou W, Luncford J, Strack P, Dauffenbach LM, Kerfoot CA. Level of phosphohistone H3 among various types of human cancers. *BMJ Open*. 2012;2(5):e001071.
40. Bossard C, Jarry A, Colombeix C, et al. Phosphohistone H3 labeling for histoprognostic grading of breast adenocarcinomas and computer-assisted determination of mitotic index. *J Clin Pathol*. 2006;59(7):706–710.
41. Ribalta T, McCutcheon IE, Aldape KD, Bruner JM, Fuller GN. The mitosis-specific antibody anti-phosphohistone-H3 (PHH3) facilitates rapid reliable grading of meningiomas according to WHO 2000 criteria. *Am J Surg Pathol*. 2004;28(11):1532–1536.
42. Skaland I, Janssen EA, Gudlaugsson E, et al. Validating the prognostic value of proliferation measured by phosphohistone H3 (PPH3) in invasive lymph node-negative breast cancer patients <71 years of age. *Breast Cancer Res Treat*. 2009;114(1):39–45.
43. Scott IS, Heath TM, Morris LS, et al. A novel immunohistochemical method for estimating cell cycle phase distribution in ovarian serous neoplasms: implications for the histopathological assessment of paraffin-embedded specimens. *Br J Cancer*. 2004;90(8):1583–1590.

Cancer Management and Research

Publish your work in this journal

Cancer Management and Research is an international, peer-reviewed open access journal focusing on cancer research and the optimal use of preventative and integrated treatment interventions to achieve improved outcomes, enhanced survival and quality of life for the cancer patient. The manuscript management system is completely online and includes

Submit your manuscript here: <https://www.dovepress.com/cancer-management-and-research-journal>

a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Dovepress