Computer-aided drug design and inhibitive effect of a novel nitrogenous heterocyclic compound and its mechanism on glioma U251 cells and breast cancer MCF-7 cells

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Background: Glioma and breast cancer are severe malignant cancerous tumors that highlight the importance of developing new anti-cancer drugs. The aim of this study was to explore the effects of a novel nitrogenous heterocyclic compound on glioma and breast cancer cells and to determine its mechanism of action.

Methods: We designed and synthesized a novel nitrogenous heterocyclic compound, 3-(4-amino-1H-benzof[d]imidazole-2-carboxamido)-4-oxo-3,4-dihydropyrimidazo[5,1-d][1,2,3,5]tetrazine-8-carboxamide, based on alkylglycerone phosphate synthase (AGPS) using computer-aided drug design (CADD), and we measured its effect on the proliferation, invasion, cell cycle and apoptosis of U251 glioma and MCF-7 breast cancer cells. In addition, the compound’s effect on the expression of tumor-related mRNA, circular RNAs (circRNAs) and long non-coding RNAs (lncRNAs) was explored.

Results: It was found that the nitrogenous heterocyclic compound could induce cell cycle arrest at the G2/M phase of U251/MCF-7 cells and activate apoptosis. Real-time PCR showed that the expression levels of tumor-related mRNA, circRNAs and lncRNAs were impacted.

Conclusion: We concluded that the nitrogenous heterocyclic compound inhibits the proliferation and invasion of U251 glioma and MCF-7 breast cancer cells through the induction of apoptosis and cell cycle arrest by regulating tumor-related genes.

Keywords: nitrogenous heterocyclic compound, glioma, breast cancer, proliferation, invasion

Introduction
Glioma and breast cancer are both severe malignant cancerous tumors with suboptimal therapeutic treatments, for example, the 5-year survival rate of malignant glioma patients is about 5%1 and breast cancer is the most common invasive malignancy and the second leading cause of tumor-related death among women worldwide;2 there are excessive side-effects in current anti-cancer drugs, therefore, it is very important to develop new anti-cancer drugs. Previous studies have indicated that alkylglycerone phosphate synthase (AGPS) has a close association with tumor development. Overexpression of AGPS can increase the viability and migration potential of various tumor cells, including 231MFP, C8161 melanoma, PC3 prostate cancer and primary breast cancer cells, and advance the growth and migration of the tumor cells,3 demonstrating the important role of AGPS in tumor development. Meanwhile, in our previous study it was also reported that AGPS silencing suppresses the potential of proliferation and invasion.
invasion of U251 glioma cells; thus, we considered that AGPS may be a viable target for anti-cancer drugs.

Benzyl isothiocyanate is a natural compound that is extracted from cruciferase plants and shows anti-tumor activity in many tumor types. However, the potential application of this compound in cancer is limited due to the toxicity of the isothiocyanate moiety.

We designed and synthesized a novel nitrogenous heterocyclic compound, 3-(4-amino-1H-benzo[d]imidazole-2-carboxamido)-4-oxo-3,4-dihydroimidazo[5,1-d][1,2,3,5]tetrazine-8-carboxamide, with reduced toxicity based on AGPS using computer-aided drug design (CADD) to improve potency and decrease the side effects. The effect of the nitrogenous heterocyclic compound on the proliferation and invasion of U251 glioma and MCF-7 breast cancer cells was explored, and the molecular mechanism was investigated to understand the anti-tumor effects of this compound.

Methods

Computer-aided drug design and compound synthesis

The AGPS three-dimensional structure (PDBID: 2UUV) was downloaded from the Protein Data Bank, and the molecular structure was drawn using ChemBioOffice2010. "CDOCKER ENERGY" was measured using Discovery Studio 3.5.

The compound was synthesized as follows: A solution of 5-amino-1H-imidazole-4-carboxamide (100 mmol) and aminoisocyanate (100 mmol) in 300 mL of tetrahydrofuran was stirred at 60 °C for 5 h to give 3-amino-4-oxo-3,4-dihydroimidazo[5,1-d][1,2,3,5]tetrazine-8-carboxamide. This intermediate was then reacted with 4-amino-1H-benzo[d]imidazole-2-carboxylic acid (100 mmol) at 60 °C for 12 h to give the desired compound, 3-(4-amino-1H-benzo[d]imidazole-2-carboxamido)-4-oxo-3,4-dihydroimidazo[5,1-d][1,2,3,5]tetrazine-8-carboxamide (95% purity). All the major chemicals were purchased from Weiran Biotech (Jinan, China). 1H NMR (DMSO-d6 400 MHz) δ 8.70 (s, 1H), 8.37 (s, 2H), 8.03 (s, 1H), 6.97 (d, J=8.4 Hz, 1H), 6.89 (dd, J=6.4 Hz, 1H), 6.46 (d, J=8.2 Hz, 1H), 6.27 (s, 2H), 5.10 (s, 1H), 4.51 (s, 1H). MS (ESI) m/z 354.1 (M+1)⁺.

Cell lines and culture

Rat adrenal medulla PC12 (ATCC Number: CRL-1271), human glioma U251 (ATCC Number: CRL-1966), normal breast MCF10A (ATCC Number: CRL-10317) and breast cancer MCF-7 (ATCC Number: HTB-22) cell lines were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). The PC12 cell line was cultured in Ham’s F12K (with 10% fetal bovine serum), while the U251, MCF10A and MCF-7 cell lines were cultured in DMEM (with 10% fetal bovine serum). Both above culture media contained 100 U/mL of penicillin and 100 mg/mL of streptomycin and were maintained at 37°C with 5% CO₂. The heterocyclic or isothiocyanate compounds were dissolved by DMSO and diluted by DMEM without fetal bovine serum.

MTT assays

PC12/U251/MCF10A/MCF-7 cells were cultured in 96-well plates (3x10³/well) overnight. The cells were treated with the nitrogenous heterocyclic compound or benzyl isothiocyanate at various concentrations (0, 10, 20, 50, 100, 200, 500 μM) for 72 h. MTT (20 μL) was added, and the cells were cultured at 37°C for 4 h. The solution was removed, and then 150 μL of DMSO was added at 37°C for 10 min, after which time the OD was determined at 490 nm. The inhibitory rate of the drugs on the cells was calculated as follows: % Inhibition = (1 – OD value / OD value) x 100%.

Flow cytometry assay

U251/MCF-7 cells (2x10⁵/well) were cultured in 6-well plates in the presence of the nitrogenous heterocyclic compound (50 and 100 μM) at 37°C for 72 h. After the cells were digested, washed with cold PBS, and stained with Annexin V-FITC (Annexin V-FITC Apoptosis Detection Kit, Beyotime Biotechnology, Shanghai, China) at room temperature away from light, for 15 min, apoptosis was measured by flow cytometry at 488 nm. Likewise, after the cells were treated with 20 and 50 μM of the nitrogenous heterocyclic compound at 37°C for 72 h, fixed using 70% absolute alcohol at 4°C overnight, and stained by propidium iodide (Sigma-Aldrich, St. Louis, MO, USA) at room temperature away from light, for 30 min, the cell cycle was analyzed using flow cytometry at 488 nm.

Caspase-3/8 activity assays

U251/MCF-7 cells (2x10⁵/well) were cultured in 6-well plates in the presence of the nitrogenous heterocyclic compound (20 and 50 μM) at 37°C for 72 h. Caspase-3/8 activity was measured using caspase detection kits (ImmunoChemistry Technologies LLC, Bloomington, MN, USA) and a microplate reader at 488 nm.

Transwell assay

U251/MCF-7 cells (2x10⁵/well) were cultured in 6-well plates in the presence of the nitrogenous heterocyclic compound (20 and 50 μM) at 37°C for 72 h. After digestion by Trypsin-EDTA, DMEM was used to suspend the cells (5x10⁵/mL), and 100 μL of the suspension was added into the Transwell (Corning, NY, USA) upper chamber with 25 μL Matrigel (200 μg/mL,
BD Biosciences, San Jose, CA, USA). DMEM containing 10% fetal bovine serum (FBS) (600 µL) was added into the lower chamber, and the cells were cultured at 37°C for 24 h. The membrane was fixed using 4% methyl ester at room temperature for 10 min and stained by 0.01% crystal violet at room temperature for 20 min, and then the number of migrating cells was counted using a light microscope (×200).

Western blot assay
U251/MCF-7 cells (2×10⁵/well) were cultured in 6-well plates in the presence of the nitrogenous heterocyclic compound (20 and 50 µM) at 37°C for 72 h. The cells were lysed, and the total proteins were extracted by centrifugation at 12,000 g at 4°C for 10 min using protein extraction buffer (Bio Scientific, Austin, TX, USA). Protein was measured using the Bradford assay (Beyotime Biotechnology, Shanghai, China), and 50 ng of the protein was separated via SDS-PAGE and then transferred onto a polyvinylidene fluoride membrane. The membrane was blocked using 1% BSA at 37°C for 1 h. The membrane was incubated with AGPS (1:2,000, sc-374201), p21 (1:1,500; sc-166630), p27 (1:1,500; sc-71813), Bcl-2 (1:1,500; sc-23960), survivin (1:1,500; sc-101433) and Bim (1:1,500; sc-374358) antibodies at 4°C overnight and was then incubated at 37°C for 1 h with peroxidase-labeled anti-rabbit immunoglobulin G (1:2,000). The membrane was washed three times with PBS containing 0.05% Tween20. The membrane was visualized using Immobilion Western chemiluminescent horseradish peroxidase substrate (EMD Millipore, Billerica, MA, USA). β-Actin (1:5,000; sc-58673) was used as the control. All antibodies were purchased from Santa Cruz (Dallas, TX, USA).

Quantitative real time polymerase chain reaction (qRT-P) CR assay
U251/MCF-7 cells (2×10⁵/well) were cultured in 6-well plates in the presence of the nitrogenous heterocyclic compound (20 and 50 µM) at 37°C for 72 h. Trizol (Invitrogen, Thermo Fisher Scientific, Waltham, MA, USA) was used to extract the total RNA from each group, and an RT-PCR kit (Takara Bio Inc., Kusatsu, Shiga Prefecture, Japan) was used in reverse transcription of the RNA into DNA. A 2 µL aliquot of the RT product was used to perform the PCR reaction. Total RNA was then reverse transcribed and the expression of mRNAs, circRNAs and IncRNAs was detected using the real-time PCR assay (Applied Biosystems 7500 Real-Time PCR System, Thermo Fisher Scientific, Waltham, MA, USA). The PCR conditions were as follows: denaturation at 95°C for 10 min, followed by 40 cycles at 95°C for 15 s, 60°C for 60 s and a final elongation at 95°C for 15 s. The genes were normalized using β-actin as a control. The full details of the primers used in these experiments are shown in Table 1.

Table 1 Quantitative real time polymerase chain reaction primers in the experiments

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primers</th>
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| P21  | Forward: 5'-CATGGGTGTTGCTACGAGGACATC-3'  
Reverse: 5'-TTGGCAAATCATGCTTTGGT-3' |
| P27  | Forward: 5'-CATTCCATGAAATCTACGAGGT-3'  
Reverse: 5'-CGTCAAGCTTAACTGCTG-3' |
| Bcl-2 | Forward: 5'-CGTACAGTTCAACAAAGGCA-3' 
Reverse: 5'-ATGGTGTTGAGGATCGTCAA-3' |
| Survivin | Forward: 5'-TCCGCAAGTTTCTCATAAT-3' 
Reverse: 5'-GTTGCGGCTTCTTTGTCG-3' |
| Bim  | Forward: 5'-GATAGTTGTTAGGAGGCTGG-3'  
Reverse: 5'-CTCTCCCTACAGACAGCAGG-3' |
| DQ786243 | Forward: AATCGGCTCTGGAGAAGGGA 
Reverse: CGTCTGCTTCTGTGATCTTTTGA |
| HOXD-AS1 | Forward: GGCTCTCCTCTAGTGGTTG 
Reverse: CAGTGCACAGTACAGCAAG |
| CAST2 | Forward: GCACATTGAGGCTGTTCC 
Reverse: CCACTGCTCAGAGGTC |
| ANCR | Forward: GACATTCTCAGTGCTGTTGG 
Reverse: AAGCGAC |
| MEG3 | Forward: TTTTTGCGCAAGGCTTTGGG 
Reverse: AGGGGCACTAAGGACAGGTTA |
| circUBP2 | Forward: AGGCTTGAAGGCTTTGC 
Reverse: TCAGTTTGGAGTATGCAAGAT |
| circZNF292 | Forward: GCTCAAAGACTGGGGTGT 
Reverse: AGTTTGTGTTTGGGGCAAG |
| circTGF25 | Forward: CGGAATCTCCGTTAATAGCTATCTTA 
Reverse: CAGAGAGGCGGTCTAGCAGTGA |
| circHIPK3 | Forward: TATGTGTTGAGATCTCTGTGGCA 
Reverse: TGGTTGCTAGACAGAAGTGTGGA |
| circCdrl | Forward: GTGCTCAGTTGATCAGGG 
Reverse: TACCTGAGCAACCTTGGGAA |
| circZKSCAN1 | Forward: AGTCCCACTCAACACTGTGCT 
Reverse: CACCTCCATCTATGACATCAT |
| circTCH | Forward: GCAGAGGCCAACACTGGGA 
Reverse: TCCCTGAGTCTGACTGAGAG |
| circMTO1 | Forward: GAGCTGTAAGAAGTCTATTTC 
Reverse: CACAGGGCATCCAAGGGGTC |
| β-actin | Forward: AGGCACAGGGCGTGAT 
Reverse: GCCCACATGGAATCTTCTGAC |

Statistical analysis
Experimental data are represented by x±s. SPSS 11.0 statistical software was used to perform one-way analysis of variance (ANOVA) and a P-value ≤0.05 was considered of statistical significance.
Results
The expression of AGPS of PC12, U251, MCF10A and MCF-7 cells

The Western blotting showed an increased expression of AGPS in U251 and MCF-7 cells compared with PC12 and MCF10A cells, thus demonstrating that there was an overexpression of AGPS in tumor cells (Figure 1A). The result also showed that nitrogenous heterocyclic compound suppressed the expression of AGPS in U251 and MCF-7 cells, indicating its target for AGPS (Figure 1B).

The effect of the nitrogenous heterocyclic compound and benzyl isothiocyanate on the proliferation of U251 and MCF-7 cells

Using virtual screening, we designed and synthesized the nitrogenous heterocyclic compound 3-(4-amino-1H-benzo[d]imidazole-2-carboxamido)-4-oxo-3,4-dihydropyrimido[5,1-d][1,2,3,5]tetrazine-8-carboxamide (Figure 2A); the 2D and 3D combined mode with amino acid residues of AGPS is shown in Figure 2B and C. The synthetic procedure to prepare the nitrogenous heterocyclic compound is shown in Figure 2D. Furthermore, the MTT assay showed that the nitrogenous heterocyclic compound (Figure 2E) and benzyl isothiocyanate (Figure 2F) inhibit the proliferation of PC12, U251, MCF10A and MCF-7 cell lines in a dose-dependent manner. The results showed that after the nitrogenous heterocyclic compound and benzyl isothiocyanate acted on the PC12, U251, MCF10A and MCF-7 cells for 72h, nitrogenous heterocyclic compound was less toxic to the normal than non-cancerous cells due to a lower level of growth inhibition compared with benzyl isothiocyanate (Figure 2G), indicating increased safety and cancer-targeting ability of the nitrogenous heterocyclic compound compared to benzyl isothiocyanate.

Nitrogenous heterocyclic compound concentrations that were non-toxic (20 \( \mu \)M, inhibition rate <5%) or that had low toxicity (50 \( \mu \)M, inhibition rate <15%) in U251 and MCF-7 cells were used in subsequent experiments to minimize the effects on cell proliferation, with 0 \( \mu \)M serving as the control group.

The effect of the nitrogenous heterocyclic compound on the cell cycle of U251 and MCF-7 cells

Compared to the control group’s effect on U251 and MCF-7 cells, the nitrogenous heterocyclic compound at 20 and 50 \( \mu \)M resulted in a significant decrease in the number of cells in the G1 phase and an increase in the number of cells in the G2/M phase after 72 h (Figure 3), thus demonstrating that the nitrogenous heterocyclic compound arrests the cell cycle of U251 and MCF-7 cells in G2/M phase.

The effect of the nitrogenous heterocyclic compound on the apoptosis of U251 and MCF-7 cells

Flow cytometry showed an increase in apoptosis after treating U251 and MCF-7 cells with the nitrogenous heterocyclic compound at 50 and 100 \( \mu \)M for 72 h, which demonstrates that the nitrogenous heterocyclic compound could cause the apoptosis of U251/MCF-7 cells. Furthermore, we found that the nitrogenous heterocyclic compound induced increased caspase-3/8 activity in U251 and MCF-7 cells (Figure 4).

The effect of the nitrogenous heterocyclic compound on the invasion of U251 and MCF-7 cells

To study the effect of the nitrogenous heterocyclic compound on the invasion of U251 and MCF-7 cells, we examined the
invasion ability of U251 and MCF-7 cells using a transwell assay. Cell invasion significantly decreased after treatment with the nitrogenous heterocyclic compound compared to that of the control group, thus demonstrating that the nitrogenous heterocyclic compound affects the invasion ability of U251 and MCF-7 cells (Figure 5).

The effect of the nitrogenous heterocyclic compound on the expression of tumor-related genes
The real time PCR assay showed that treatment of U251 and MCF-7 cells with the nitrogenous heterocyclic compound at 20 and 50 μM for 72 h resulted in significantly increased protein and mRNA expression of p27, p21 and Bim and decreased expression of Bcl-2 and survivin. The nitrogenous heterocyclic compound was also found to suppress the expression of circRNAs circUBAP2, circZNF292, circHIPK3, circTCF25, and circCdr1, as well as lncRNAs H19, DQ786243, HOXD-AS1, CCAT1, and HULC, while it increased the expression of circRNAs circZKSCAN1, circMTO1, and circITCH, as well as lncRNAs GAS5, CASC2, ANCR, and MEG3 (Figure 6).

Discussion
Both glioma and breast cancer are the most common types of cancer, and it remains a huge challenge to public health not just in China but throughout the world; thus, it is important to develop new cancer-targeting drugs with decreased toxicity.

Figure 2. The design of the nitrogenous heterocyclic compound and its inhibition in the proliferation of glioma and breast cancer cell lines.
Notes: The structure (A), 2D combined mode with amino acid residues of AGPS (B), where the green circle and green line represent a hydrophobic bond, the pink circle and blue line represent a hydrogen bond, and the orange line represents a pi bond (a covalent bond) in the interactions of the compounds, and 3D combined mode (C) with AGPS and the nitrogenous heterocyclic compound. (D) The synthetic procedure to prepare the nitrogenous heterocyclic compound. (E) The inhibitive effect of the nitrogenous heterocyclic compound on proliferation of PC12, U251, MCF10A, and MCF-7 cell lines as determined by the MTT assay. (F) The inhibitive effect of benzyl isothiocyanate on proliferation of PC12, U251, MCF10A, and MCF-7 cell lines as determined by the MTT assay. (G) The IC_{50} of the nitrogenous heterocyclic compound and benzyl isothiocyanate in PC12, U251, MCF10A, and MCF-7 cell lines.
Abbreviation: AGPS, alkylglycerone phosphate synthase.
It has been reported that benzyl isothiocyanate demonstrates anti-tumor activity; however, the potential application of this compound is limited due to its toxicity.

Increased expression of AGPS in human U251 glioma and MCF-7 breast cancer cell lines was observed compared to non-cancer PC12 and MCF10A cell lines, indicating that AGPS may be a suitable target for anti-tumor drug therapy. Therefore, we hoped to modify the structure of benzyl isothiocyanate to improve its effect on cell cycle and apoptosis.

Figure 3 Effect of the nitrogenous heterocyclic compound on the cell cycle of U251 glioma and MCF-7 breast cancer cell lines.

Notes: An increased number of cells in the G2/M phase of U251 glioma (A) and MCF-7 breast cancer (B) cell lines was observed after treatment with the nitrogenous heterocyclic compound. *compared to the 0 μM group, P < 0.05.

Figure 4 Effect of the nitrogenous heterocyclic compound on the apoptosis of U251 glioma and MCF-7 breast cancer cell lines.

Notes: Increased apoptosis and caspase-3/8 activity was observed in U251 glioma (A) and MCF-7 breast cancer (B) cell lines after treatment with the nitrogenous heterocyclic compound. *compared to the 0 μM group, P < 0.05.
selectivity for cancer cells and develop a potentially new drug by CADD.

In a previous study, we designed and synthesized an isothiocyanate derivative by CADD targeting AGPS, predicted the ADME (absorption, distribution, metabolism, excretion) and toxicity, and investigated the inhibitory effect of the compounds on the proliferation and invasion of glioma U251 cells, providing the basis for further development and clinical application of a nitrogenous heterocyclic compound.9

In order to obtain a comprehensive understanding of the anti-tumor activity associations among the candidate compounds and then further optimize the lead compound structure, we investigated the inhibitive effect of nitrogenous heterocyclic compound in U251 and MCF-7 cells.

We found that the nitrogenous heterocyclic compound had increased safety and cancer-targeting ability compared to that of benzyl isothiocyanate via affecting the proliferation and invasion of glioma U251 cells, providing the basis for further development and clinical application of a nitrogenous heterocyclic compound.9

Anti-oncogenes p27 and p21 and cyclin-CDK are important mechanisms that are known to regulate the cell cycle.10–12 The combination of p21 with cyclin and CDK suppresses kinase activity and causes cell cycle arrest; p27 is also known to prevent the transition of cells from the G2 to the M phase and inhibit the cell cycle of tumor cells.13,14

Our study found that the nitrogenous heterocyclic compound increased the expression of p27 and p21, which suppressed the proliferation of U251 and MCF-7 cells.

Bcl-2 and survivin are known apoptosis-associated genes that inhibit cell apoptosis by suppressing the activation of caspase,15–17 while Bim is an apoptosis promoting gene that causes tumor cell apoptosis.18 In this study, the nitrogenous heterocyclic compound was found to decrease the expression of Bcl-2 and survivin and increase the expression of Bim, thus providing a mechanism for the induction of apoptosis in U251 and MCF-7 cells.

All of the Wnt/b-catenin, PI3K-Akt, JAK/STAT, and MAPK/ERK signal pathways play an important role in the function of p27, p21, Bcl-2, survivin and Bim, thus we considered that they may also play an important role in the anti-tumor activity of nitrogenous heterocyclic compound.

Although their mechanism in the tumor process is not well understood, circRNAs and lncRNAs are considered important targets for anti-tumor drug therapy. Our study showed that the nitrogenous heterocyclic compound reduced the expression of circRNAs circUBAP2, circZNF292, circHIPK3, circTGF25 and circCdr1 and lncRNAs H19, DQ786243, HOXD-AS1, CCAT1 and HULC, which are known oncogenes. Meanwhile, the compound increased the expression of circRNAs circZKSCAN1, circMT01 and circITCH and lncRNAs GAS5, CASC2, ANCR and MEG3, which are known tumor suppressor genes. These results suggest a potential role for circRNAs and lncRNAs in the nitrogenous heterocyclic compound. The involved signal pathways of lncRNAs and circRNAs are shown in Table 2, and suggest that there is a possibility that the nitrogenous heterocyclic compound affects tumor cells via
Figure 6 Effect of the nitrogenous heterocyclic compound on the expression of tumor-related genes in U251 glioma and McF-7 breast cancer cell lines.

Notes: Western blot and real-time PCR assays showed increased expression of p27, p21, Bim, circZKSCAN1, circMTO1, and circITCH, as well as lncRNAs GAS5, CASC2, ANCR, and MEG3, and decreased expression of Bcl-2, survivin, circUBaP2, cZNF292, circHIPK3, circTCF25, and Cdr1, as well as lncRNAs H19, DQ786243, HOXD-AS1, CCAT1, and HULC in U251 glioma (A) and MCF-7 breast cancer (B) cell lines. *compared to the 0 μM group, P<0.05.

Abbreviation: PCR, polymerase chain reaction.
the above-mentioned signal pathways. We will continue to explore this further.

**Conclusion**

This study shows that the nitrogenous heterocyclic compound inhibits the proliferation of tumor cells and causes their apoptosis and cell cycle arrest. The nitrogenous heterocyclic compound may be a new and potential anti-tumor drug.

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**Disclosure**

The authors report no conflicts of interest in this work.

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