Factors correlated with the improvement of endothelial dysfunction during Abatacept therapy in patients with rheumatoid arthritis

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Background: Rheumatoid arthritis patients are exposed to a high risk of cardiovascular morbidity and mortality even in the early phases of the disease.

Methods: We evaluated carotid common carotid intimal media thickness (ccIMT) intimal thickness and brachial flow-mediated dilation (FMD) of 45 rheumatoid arthritis patients without known cardiovascular risk factors or heart disease on a stable dose of prednisone 5.2±1.2 mg/day and Methotrexate 11.5±2.1 mg at baseline (T0) and after 12 months (T1) of treatment with Abatacept 125 mg/week. The comparison between T0 and T1 (t- and Mann–Whitney test), correlation (Spearman r), and predictivity (linear regression) of FMD, ccIMT vs clinical and laboratory parameters (disease activity 28 score, tumor necrosis factor alpha [TNFα], interleukin-6, erythrocyte sedimentation rate, C-reactive protein (CRP), CD3+, CD3+/CD4+, CD3+/CD8+, CD19+(B), CD20+(B), NK CD3-CD56+CD16+, CD14+ HLA DR+, CD4+CD28+, CD4+CD28, rheumatoid factor IgM, IgA, RF IgG, anti-citrullinated peptide antibodies) were also evaluated.

Results: During Abatacept treatment, ccIMT and FMD remained stable and disease activity 28 score, CRP, erythrocyte sedimentation rate, and interleukin-6 decreased significantly (p=0.0001, 0.002, 0.0002, 0.0001 respectively). At T0, only ccIMT resulted as correlated with baseline TNFα values (p=0.0245) and in an inverse proportion. At T1, ccIMT correlated with CD3/CD8+ lymphocytes number (p=0.0351) and FMD with CRP (p=0.0075). In regression analysis, baseline ccIMT and FMD had a low predictivity for TNFα (p=0.011) and CRP (p=0.049) at T1, respectively.

Conclusion: This study shows that the endothelial function remained stable during Abatacept treatment.

Keywords: Abatacept, rheumatoid arthritis, cardiovascular disease

Introduction

We know from several studies that rheumatoid arthritis (RA) is an important cardiovascular (CV) risk factor in terms of morbidity as well of mortality, and this risk is present even at the beginning of the disease’s course.1-2 Epidemiological studies have also shown that RA is associated with an increased risk of premature CV diseases (CVD), including acute myocardial infarction and CV-related mortality.3-7 Inflammation plays the main role in this risk assessment, and its effects are involved not only in patients suffering from arthritis but also in the atherogenic process of otherwise healthy people.8-10 Endothelial dysfunction (ED) characterizes every step of the atherosclerosis progression, up to the latest stages of vascular disease. Mechanisms leading to ED are pleiotropic, but most of them can be found more expressed in RA patients rather than...
in healthy controls: alterations of nitric oxide’s metabolism, increased synthesis of VEGF, chemokines, adhesion cytokines, and ROS production, only to cite the most representative.11 Notably, these processes are the same as involved in inflammatory proliferation of synovial vasculature during RA. In order to assess the state of coronary artery endothelium, it is possible to explore its correlation with the brachial artery, which can be investigated in a noninvasive way with the help of vascular ultrasonography. This technique allows to evaluate the endothelium-dependent flow-mediated vasodilation (FMD).12 ED is a feature shared by patients with long-standing RA treated with Methotrexate,13 but an incredibly rapid improvement in FMD can be seen in patients when a treatment with tumor necrosis factor (TNF) blockers is performed.14 Conversely, not many data are available regarding treatment with anti-TNF in terms of ED improvement. Two papers (one by Mathieu et al and the other by Sandoo and Kitas)15,16 show neither exciting nor comforting results in terms of arterial stiffness in the course of therapy with Abatacept, a soluble cytotoxic T-Lymphocyte Antigen (CTLA)-4Ig fusion protein that blocks CD28–CD80/86 costimulatory T-cell activation, describing a worsening of this parameter.

**Materials and methods**

We evaluated in 45 RA (2010 ACR/EULAR classification criteria) patients (mean: 62.75 years old patients [standard error of the mean {SEM}: 1 and 7], lower and higher 95% interval confidence values IC: 61.68–18.32). FMD on brachial artery using a probe with 10 MHz transducer (HP Sonos 5500, Hewlett-Packard Development Co., Palo Alto, CA, USA), and common carotid intima media thickness (ccIMT) (10 MHz linear array transducer, HP Sonos 5500) intimal thickness at baseline and after 12 months of treatment with Abatacept 125 mg/week. Patients were on a stable dose of prednisone 5.2±1.2 mg/day and Methotrexate 11.5±2.1 mg/week, did not have significant changes in CV factors (no cerebral and CV events), and had stable arterial pressure during the follow-up, and did not have diabetes, smoking habit, and obesity at baseline. Furthermore, demographics (gender and illness duration) were considered in the statistical analysis.

We did not analyze patients switched to other treatments (DMARDs and bDMARDs) because the time of washout of Abatacept because in the short term it could influence the parameters evaluated. Because of the retrospective nature of the study and possible other CV confounding factors in the control population, we could not compare this data with another cohort.

Patients have also been evaluated for disease activity 28 score (DAS28) and for the following laboratory parameters: TNFα (Human TNF-alpha Quantikine Immunoassay; R&D Systems Inc, Minneapolis, MN, USA); interleukin (IL)-6 (Human IL-6 Instant Enzyme-linked Immunosorbent assay; eBioscience, Bender MedSystem GmbH, Vienna, Austria); the erythrocyte sedimentation rate (ESR) and/or C-reactive protein (CRP) levels (Unicel Coulter DxC 800 Synchron Central System; Beckman Coulter Inc, Brea, CA, USA); CD3+, CD3+/CD4+, CD3+/CD8+, CD19+(B), CD20+(B), NK CD3-CD56+CD16+, CD14+ HLA DR+, CD4+CD28+, CD4+CD28 null peripheral mononuclear cells (BD FACSCanto II flow cytometer; Biosciences, San Jose, CA, USA); rheumatoid factor (RF) IgM (N Latex RF, Siemens AG, Munich, Germany); RF IgA and RF IgG (Enzyme Immuno Assay Orgentec Diagnostika GmbH, Mainz, Germany); anti-citrullinated peptide antibodies (ACPA; Anti-CCP Axis Shield Diagnostics Ltd, Luna Place, The Technology Park, Dundee, DD2 1XA, UK). All patients gave their written informed consent according to the retrospective nature of the study according to the Declaration of Helsinki and Italian legislation (Authorization of the Privacy Guarantor No. 9, December 12, 2013). The Institutional Review Board, the Health Director of San Giovanni di Dio Hospital in Florence, reviewed and approved this research and the use of clinical and laboratory data of common clinical practice, in the respect of the Privacy Law, for clinical and scientific studies and publications.

**Assessment of brachial artery FMD and ccIMT**

A single trained sonographer estimated, with a 10-MHz linear ultrasound probe (HP Sonos 5500; Hewlett-Packard Development Co.), common carotid artery for ccIMT and brachial artery for FMD, at baseline (before the first infusion) and after 12 months. ccIMT measurements were obtained with patients lying supine, the neck extended, and the chin directed to the side opposite to the examined one. Considering that we indicate as IMT the distance between the leading edges of the lumen interfaces and the media-adventitia interface of the far wall and knowing that in ultrasonography this distance can be assessed measuring the space between the first and the second echogenic lines from the lumen, both carotid arteries were delaminated three times, 1 cm distal to the carotid bifurcation, at the level of the posterior wall, expressed in millimeters. The average of these measures was taken into account as an indicator of carotid vascular stiffness. Although
any normal range has already been established, an increase of >1 mm was considered abnormal.\(^{17}\)

For FMD, patients had rested previously for 30 minutes in a temperature-controlled room. Then, a B-mode longitudinal section of the right arm brachial artery was evaluated above the antecubital fossa. A pneumatic cuff was positioned on the forearm, inflated to an over-systolic pressure for 4.5 minutes and then released with the aim of inducing reactive hyperemia. After deflation, the maximal flow velocity and arterial diameter were recorded for 90 minutes, ECG-gated, and detected offline. The FMD measurement was performed at baseline (before the first infusion) and repeated after 12 months.

**Statistical analysis**

Descriptive statistics were expressed as mean ± SEM, 95% lower and upper confidence intervals of mean, and as number and percentage for categorical variables. A \(p\)-value <0.05 was considered statistically significant. Normal distribution of parameters was verified by “KS normality test” and “Wilcoxon Signed Rank Test”.

T0 and T1 parameters were compared with parametric \(t\)-test or nonparametric Mann–Whitney, when indicated. The correlation of FMD and ccIMT with other parameters at T0 and T1 was evaluated with nonparametric Pearson and parametric Spearman \(r\) test, respectively, when indicated. Finally, predictive values of T0 FMD and ccIMT for other parameters at T1 were analyzed with linear regression, for the absence of other CV bias factors as illustrated in methods.

**Results**

In the 45 RA patients examined, ccIMT and FMD T0 and T1 were independent by gender (32 female vs 13 male) and RA disease duration (5.7 years [0.53 SEM], 4.6–6.8 95% CI) at statistical analysis (Mann–Whitney and Spearman \(r\) test, respectively). During treatment, ccIMT and FMD remained stable, with a low increase in FMD from T0 to T1 (+1.35%). Following Abatacept treatment, DAS28, CRP, ESR, and IL-6 significantly decreased by 23.55%, 38.90%, 29.4%, and 60.66%, respectively (percentage expressed over T0 values) with significant values shown in Table 1. At T0, only ccIMT resulted as correlated with baseline TNF\(\alpha\) values \((p=0.0245)\) in an inverse proportion (data not shown) and not with other evaluated parameters. Baseline FMD does not correlate with any of the parameters considered. At T1, after 12 months of Abatacept 125 mg/week treatment, FMD correlated with CRP \((p=0.0075)\), while ccIMT shows correlation with the number of CD3+/CD8+lymphocytes \((p=0.0351)\) and TNF\(\alpha\) \((p=0.0299)\), both evaluated after treatment (Table 2). In regression analysis the baseline levels of ccIMT and FMD correlated with TNF\(\alpha\) levels \((p=0.011)\), while at the time T1 with CRP levels \((p=0.049)\).

**Table 1** Significant comparison between baseline and follow-up parameters analyzed before and after treatment with Abatacept in RA patients

<table>
<thead>
<tr>
<th>Parameter</th>
<th>T0</th>
<th>T1</th>
<th>(p)-statistics</th>
</tr>
</thead>
<tbody>
<tr>
<td>ESR</td>
<td>29.82 (1.9), 25.99–33.65</td>
<td>21.07 (2.2), 16.51–25.63</td>
<td>0.0002 (MW)</td>
</tr>
<tr>
<td>CRP</td>
<td>1.37 (0.1), 1.04–1.70</td>
<td>0.83 (0.12), 0.57–1.1</td>
<td>0.0023 (MW)</td>
</tr>
<tr>
<td>DAS28</td>
<td>3.79 (0.10), 3.57–4</td>
<td>2.89 (0.15), 2.59–3.20</td>
<td>&lt;0.0001 (TT)</td>
</tr>
<tr>
<td>IL-6</td>
<td>10.22 (2.08), 6.01–14.43</td>
<td>4.02 (0.37), 3.25–4.78</td>
<td>&lt;0.0001 (TT)</td>
</tr>
<tr>
<td>CRP (mg/dL)</td>
<td>&lt;4.02 (0.37), 3.25–4.78</td>
<td>0.83 (0.12), 0.57–1.1</td>
<td>&lt;0.0001 (TT)</td>
</tr>
<tr>
<td>CRP</td>
<td>0.0351 Not significant</td>
<td></td>
<td></td>
</tr>
<tr>
<td>TNF(\alpha)</td>
<td>0.0245 Not significant</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IL-6</td>
<td>0.0075 Not significant</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Table 2** Significant correlation of FMD and ccIMT at baseline (T0) and at 1 year follow-up (T1) with other parameters at T0 and T1, respectively

<table>
<thead>
<tr>
<th>Parameter</th>
<th>FMD T0</th>
<th>ccIMT T0</th>
<th>FMD T1</th>
<th>ccIMT T1</th>
</tr>
</thead>
<tbody>
<tr>
<td>CRP (mg/dL)</td>
<td>5.52 (0.04), 5.42–5.61</td>
<td>0.77 (0.01), 0.74–0.80</td>
<td>5.604 (0.04), 5.50–5.70</td>
<td>0.74 (0.001), 0.71–0.77</td>
</tr>
<tr>
<td>CRP T1</td>
<td>0.83 (0.12), 0.57–1.1</td>
<td>Not significant</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>TNF(\alpha) T0</td>
<td>21.49 (3.6), 14.14–28.84</td>
<td>Not significant</td>
<td>(p=0.02)</td>
<td>–</td>
</tr>
<tr>
<td>TNF(\alpha) T1</td>
<td>25.80 (6.09), 6.01–14.43</td>
<td>Not significant</td>
<td>–</td>
<td>(p=0.02)</td>
</tr>
<tr>
<td>CD3/CD8+T0</td>
<td>514.3 (42.6), 428.5–600.2</td>
<td>Not significant</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>CD3/CD8+T1</td>
<td>500.2 (35.53), 428.6–571.9</td>
<td>–</td>
<td>Not significant</td>
<td>(p=0.03)</td>
</tr>
</tbody>
</table>

**Note:** Values are expressed in mean, SEM, and 95% CI.

**Abbreviations:** CRP (mg/dL), C-reactive protein; FMD, flow-mediated dilation; SEM, standard error mean; TNF\(\alpha\) (pg/mL), tumor necrosis factor alpha; ccIMT, common carotid intima media thickness.
Discussion

Although our study has important limitations, such as its retrospective nature, that do not allow the availability of a control group and the possibility of a bias in assessing CV risk driven by the elevated age of patients examined, data in our possession represent, in the light of our knowledge, the first evidence of Abatacept’s safety in terms of ED. In fact, a previous study by Mathieu et al found a worsening in arterial stiffness evaluated by pulse wave velocity (PWV) during the treatment with this drug, rising concerns about its role in the fragile equilibrium of RA patients’ vasculature. Also Sandoo et al, in a letter to the editor, described a negative effect on both micro and large vessels’ endothelial function in a 32-year-old woman affected by rheumatoid arthritis and treated with Abatacept. By contrast, Abatacept’s pharmacodynamical activity suggests stimulating speculations regarding its possible incisive role in controlling both the rheumatic disease and the CV-associated comorbidity. In fact, it is known that rheumatoid arthritis can lead to accelerated atherogenesis by promoting endothelial dysfunction, following the activation of the cytokine system. A healthy endothelium lies in a state that can be defined as antiinflammatory, antithrombotic, anticoagulant, profibrinolytic, and anti-proliferative. Also, this homeostatic condition depends on refined and sensitive regulatory pathways, and it can easily be perturbed and transformed into the opposite-featured condition identified as “endothelial dysfunction”. Atherosclerosis is actually a chronic inflammatory disease in which endothelial alteration leads to retention of oxidized low-density lipoprotein cholesterol particles into the intimal artery layer, attracting leukocytes and causing a localized inflammatory reaction. Once oxidized, LDL cholesterol particles become neoantigens whose T-cells are able to respond thanks to the signals provided by peptide-major histocompatibility complex (MHC) antigen complexes on antigen-presenting cells. MHC complexes bind the T-cell antigen receptor, and, along with costimulatory signals, this leads to an inflammatory activation. Resting T-cells constitutively express the dominant costimulatory receptor CD28, whereas activated T-cells are characterized by the expression of the coinhibitory CTLA-4 receptor. In light of the substantial intravascular CTLA-4+ T-cell infiltration found in samples while investigating the course of the postinterventional remodeling process, Ewing et al used a femoral artery cuff mouse model to elucidate the role of T-cells and the CD28–CD80/86 costimulatory and CTLA-4 coinhibitory pathways in this context. CD4−/− and CD80−/−CD86−/− development of mice intimal lesions was reduced in comparison to normal C57Bl/6J controls. Data are available showing that systemic Abatacept treatment, a soluble CTLA-4Ig fusion protein that blocks CD28–CD80/86 costimulatory T-cell activation, can prevent intimal thickening by 58.5%. Being an inflammatory disease, atherosclerosis accounts for several cytokines involved in its pathogenesis. The role of IL-6 in the development of CV disease has been deeply investigated. There is evidence that genetic variations in the IL-6 gene and its receptor gene (IL-6R) lead to different immune responses, and they are associated with predisposition to CVD. In brief, the IL-6 polymorphism genotypes are mainly associated with proinflammatory cytokines, whereas the IL-6R polymorphism genotypes are associated with antiinflammatory cytokines. Moreover, people with established CAD can host different cytokines subsets, confirming the importance of the genetic contribution in CAD. CTLA4-Ig is able to modulate macrophagic cytokine expression (TNFα and IL-6) probably acting on NF-kB pathway. This effect, which can also be encountered in vitro on activated human macrophages, is stronger using CTLA4-Ig-DEX and CTLA4-Ig-DEX-MTX combined treatments rather than monotherapy. In particular, a decrease of cytoplasmic NF-kB expression can be observed along with an increase of NF-kB inhibitor, IκBα. In our attempt to discover correlation between markers of inflammation (CRP, ESR, IL-6, and TNFα) and measures of arterial stiffness (FMD and ccIMT), we did not have statistically significative data, except for the unexpected negative correlation between TNFα levels and ccIMT. When we reassessed these parameters after 12 months of treatment with Abatacept, we detected a significant reduction in both laboratory (CRP, ESR, IL-6, and TNFα) and clinimetric indexes (DAS28) of inflammation. A nonsignificant reduction was found in ccIMT, along with a slightly increased FMD. Surprisingly, the improvement of FMD positively correlated with follow-up levels of CRP while ccIMT showed correlation with the number of CD3+/CD8+ lymphocytes and TNFα, both evaluated after treatment. Moreover, in regression analysis, we detected that T0 value of ccIMT can be predictive of TNFα at follow–up (p=0.011), maybe indicating a higher resistance against therapy in patients with more severe arterial stiffness at baseline.

Also, the high average age of our population should be taken into account in the evaluation of TNFα levels both at baseline and at follow-up. In fact, this parameter tends to increase with aging, independently from any other morbidity, and its amount can fluctuate without true clinical significance. On the other hand, FMD value at T0 was predictive of CRP levels at T1 (p=0.049), confirming the controversial
relationship between these two parameters, emerging at follow-up. The lack of a clear relationship between routine laboratory parameters of inflammation and arterial stiffness, in the face of their parallel comprehensive improvement at T1, may suggest the role of other unexplored inflammation mediators in this complex dialogue.

Our findings are in contrast with the ones noted by Mathieu et al., although in their case, analysis of arterial stiffness was made through PWV and not with FMD; neither measurement of ccIMT was performed; authors of that study assume the lack of total control on inflammation as a possible reason for their results, without excluding the role of an increase of total cholesterol levels. Indeed, PWV got worse only in patients who did not reach disease remission, while it remained stable in the group in which the treatment achieved success. Elevation of levels of lipids is largely described in the course of bDMARDS therapies, although their effect on the progression of atherogenesis remains controversial. Tocilizumab, a monoclonal antibody against IL-6 receptor available in RA treatment, accounts for the most debated concerns in this field, but latest studies seemed to be reassuring about CV safety of this drug.

Data are also available about anti-TNFα being discordant as to its effect on lipidic profile, whereas the improvement of CV risk with this class of drug is well known. We did not assess lipidic profile in our study, whereas Sidiropoulos et al. analyzing the variation of FMD and ccIMT of 21 RA patients undergoing 18 months of anti-TNFα treatment described a significant improvement of FMD without changes to the ccIMT – agreeing with our findings – with no correlation with lipidic levels. From what we observed, FMD seems to be the most variable and modifiable parameter regarding vascular stiffness, while ccIMT appears to be less plastic. This could reflect a more chronic and irreversible process on the basis of increasing intimal thickness. Furthermore, Spinelli et al., in a study conducted to evaluate the role of classic and novel RA autoantibodies in determining CV risk, noted that RA patients have an FMD lower than healthy controls but they lack divergence in terms of ccIMT. So FMD could be burdened by higher pathogenic power.

In conclusion, this study confirms that T-cell costimulation through the CD28–CD80/86 pathway may play a role in accelerated atherosclerosis development observed in patients with chronic inflammatory diseases, and it could be negatively regulated by CTLA-4 coinhibition. Data on CD3+CD8+ lymphocytes’ correlation with ccIMT at T1 could also deserve investigation to assess the role of this cellular population in vascular damage. Overall, our results may also dispel concerns about using Abatacept in RA patients suffering from CV disease: endothelial parameters do not worsen in the course of treatment, while the improvement in joint and systemic inflammation could help in reducing atherogenesis and CV risk. Further studies could be helpful in better understanding which mediators have the deepest involvement in being a molecular bridge between RA and atherosclerosis.

Disclosure
The authors report no conflicts of interest in this work.

References


