In silico and in vivo characterization of cabralealactone, solasodin and salvadorin in a rat model: potential anti-inflammatory agents

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Background: The present study investigates the hepato- and DNA-protective effects of standardized extracts of Cleome brachycarpa (cabralealactone), Solanum incanum (solasodin), and Salvadora oleoides (salvadorin) in rats.

Materials and methods: Hepatotoxicity was induced with intraperitoneal injection of carbon tetrachloride (CCl4) (1 mL/kg b.wt.) once a week for 12 weeks. The hepato- and DNA protective effects of the extracts in different combinations were compared with that of a standard drug Clavazin (200 mg/kg b.wt.). Tissue alanine aminotransferase, alpha-fetoprotein, tumor necrosis factor alpha (TNF-α), isoprostanes-2α, malondialdehyde, and 8-hydroxydeoxyguanosine, the significant hallmarks of oxidative stress, were studied.

Results: Histopathological findings of the liver sections from the rat group which received CCl4+cabralealactone, solasodin, and salvadorin demonstrated improved centrlobular hepatocyte regeneration with moderate areas of congestion and infiltration comparable with Clavazin. For in silico study, the identified compounds were subjected to molecular docking with cyclooxygenase-2 and TNF-α followed by a molecular dynamics study, which indicated their potential as anti-inflammatory agents.

Conclusion: Cabralealactone, solasodin, and salvadorin confer some hepatoprotective and DNA-damage protective effects against CCl4-induced toxicity. They successfully restored the normal architecture of hepatocytes and have the potential to be used as inhibitor to main culprits, that is, cyclooxygenase-2 and TNF-α. They can combat oxidative stress and liver injuries both as mono and combinational therapies. However, combination therapy has more ameliorating effects.

Keywords: cabralealactone, solasodin, salvadorin, antioxidant, COX-2 inhibitor, anti-inflammatory

Introduction

Currently, more than 10% of humans are affected by continuous exposure to hepatotoxic mediators like alcohol, viruses, toxic substances, and biotransformed metabolites, which may contribute to the development of liver cirrhosis and hepatocellular carcinoma.1 Liver is an organ, which is at higher risk for toxic metabolic reactions that lead to chronic liver inflammation, mainly associated with the innate defense system of the biological organization for elimination of toxic metabolites. Elevation of liver enzymes (alanine aminotransferase [ALT], aspartate aminotransferase, gamma-glutamyl transferase, and alkaline phosphatase) is a useful indicator of an abnormal hepatic functioning in addition to the clinical manifestations seen.2 Since hepatotoxicity is associated with inflammation and oxidative stress, herbal extracts with potent anti-inflammatory
and anti-oxidative properties are known to have therapeutic values in the treatment of fibrosis and toxic injuries. Therefore, the herbal extracts may be potential sources of new chemical metabolites with good therapeutic potential in xenobiotic-induced liver injuries.1,2

Cleome brachycarpa belongs to the Cleomaceae family and is locally known as Ponwar. It grows widely in sandy regions of Africa (especially in North Africa) and Asia (particularly in Egypt, Saudi Arabia, Afghanistan, Pakistan, India, and Iran). The plant is also known as spider flower because of its flower shape.3 C. brachycarpa is reported to have some amazing biochemical activities in the biological system such as antimicrobial, hepatoprotective, anticancer, antiviral, analgesic, antiimmune, immuno-modulatory, and antioxidant properties.5 It is also used in the Iranian traditional medicine in the treatment of jaundice, diarrhea, bronchitis, scabies, malaria, heart failure, immune system deficiency, seizure and rheumatoid arthritis, fever, chills, and stomach disorders, although in high doses, it has been reported to induce stomach swelling.5,6 Moreover, in a recent study, the ethanolic extract of its stems, roots, and leaves shows some antioxidant effects.7 The total phenolic and flavonoid contents of these parts are also reported to be active against 1,1-diphenyl-2-picyrylhydrazyl (DPPH) radical scavenging and ferric reducing ability of plasma activities.8 Another research established in vitro antibacterial effects of aerial parts,9 memory-boosting effects in Alzheimer,10 and anxiolytic effects in the central nervous system.7 Aerial parts of plant may show cytotoxic effects because of its constituents i.e., flavonoid and sesquiterpene.11 Spectroscopic analysis reveals that 90% of its ethanolic extract contains active phytochemical constituents, including cabralealactone (5.1%), trinortriterpenoid dilactone (0.6%), deacetoxylbrachycarpone (1.1%), and ursoic acid (0.2%).5,7 In 2010, the anticancer activity of ethanolic extract of C. brachycarpa was evaluated by using albino mice against Ehrlich Ascites Carcinoma cell line in 2 doses (200 and 400 mg/kg) for 9 consecutive days. The anticancer effect was evaluated by observing the tumor volume, viable tumor cell count, and hematological parameters. All the above-mentioned parameters showed a significant (p<0.01) decrease and the life span of the tumor-bearing host showed an increase, indicating that the plant extract has a potential anticancer activity.11

Solanum incanum is locally known as thorn apple or bitter apple, an important plant utilized in Pakistan. It grows abundantly in Africa and is also found in the Middle East and India. All the parts of the plant are valued since they are mainly used to combat liver pain, ulcer, and benign tumors.13 Isolation of solasodin metabolite from S. incanum berries indicates the presence of a chemical structure related to that of a steroidal glycol-alkaloid, which can confer a protective effect against carbon tetrachloride (CCL4)-induced hepatic insult. Cytotoxic investigation of solasodin indicates the suppressive and apoptotic characteristic against hepatitis (specifically HepG2 and Hep3B) cell lines.14 The anti-cancer effect of the herb is mainly attributed due to its glycosidal alkaloids.15–17 It has been reported that solasodin causes necrosis of cancer cells via upregulation of tumour necrosis factor (TNF) receptor and activation of intrinsic mitochondrial apoptotic signaling pathways.18,19 Studies have reported the hepatoprotective effect of fresh tuber roots of S. incanum on CCL4-induced liver damage.20

Salvadora oleoides or locally known as “Mitha Jall or Peelu” is an evergreen plant found in India and Pakistan, as well as in Southern Iran. It is traditionally used in the treatment of enlarged spleen, rheumatism, low fever, snake bites, and cough, alongside its widespread use as a purgative.21 Two compounds, octadecanoyl heptanoate and δ-lactone of 3 isobutyl-5 hydroxy-19-oxononadeconic acid have been isolated from its stem bark, and salvadorin, a new dimeric dihydroisocoumarin has been isolated from chloroform fraction of S. oleoides.

Gas chromatography and mass spectrometry analysis of essential oils based on analysis of hydro-distilled extracts indicated the presence of 35 (94.0%) and 25 (91.1%) compounds in the leaf and stem, respectively. The leaf was found to contain high concentrations of 2-methoxy-4-vinylphenol (25.4%), cis-3-hexenyl benzoate (16.8%), phytol (13.9%), n-hexadecanoic acid (6.9%), and trans-β-damascenone (2.1%). On the other hand, the stem oil contains high concentrations of 2-methoxy-4-vinylphenol (21.6%), phytol (12.9%), n-hexadecanoic acid (3.6%), octacosane (7.9%), nonacosane (7.3%), 1-octadecene (5.8%), heptacosane (5.9%), hexacosane (4.5%), pentacosane (3.4%), squalene (3.9%), and trans-β damascenone (2.3%).22

A study was conducted to evaluate the hepatoprotective activity induced in albino rats following administration of S. oleoides aqueous extract. The extract significantly elevated the high-density lipoprotein (HDL) cholesterol level and decreased serum glucose, total cholesterol (TC), triglycerides, low-density lipoprotein (LDL) cholesterol, TC/HDL, and LDL/HDL ratios while the body weight, liver function indices, and selected tumor markers were approximately comparable with the control animals. These findings suggest that S. oleoides extract is very potent in ameliorating liver injury.23 New technological advances have
been employed to evaluate the pharmacological properties of natural products using state-of-the-art computer-based approaches, the use of which has become inevitable in drug discovery pipelines due to molecular docking and molecular dynamics simulations. The combination of such molecular modeling techniques alongside investigation of biological activity is an important approach in elucidating novel plant-based drugs. Additionally, although many natural products are widely utilized in folk medicine, their toxic potentials have not been reported. Considering these factors, the aim of the current study is to evaluate the protective effects of *C. brachycarpa* (cabralealactone), *S. incanum* (salvadorin), and *S. oleioides* (solasodin) against DNA damage and confirm their hepatoprotective effects.

**Materials and methods**

All chemicals and reagents used were of analytical grades and were purchased from Sigma Chemical Co. (St Louis, MO, USA). Standardized 90% ethanolic extracts of cabralealactone, 80% ethanolic extracts of both salvadorin and solasodin were purchased from Sigma Chemical Co. Ethical approval was obtained from the Institutional Review Board of The University of Lahore (Approval No: USM/Animal Ethics approval/2009/45 [140]) and all animal experiments were performed in accordance with the Institutional Guidelines for the Care and Use of Animals for Scientific Purposes. A *p*-value of <0.05 was considered as statistically significant.

The animals were divided into several groups and received different treatments (Table 1). Subcutaneous injection of CCl₄ (1:1 v/v) was administered with saline (1 mL/kg b.wt. per week) to albino rats (*Rattus norvegicus*) every week for 12 consecutive weeks to induce hepatocellular injury as confirmed by the liver function tests. At the end of the experiment, the animals were sacrificed by an intraperitoneal injection of ketamine (Arevipharma GmbH, Radebeul, Germany) (1 mL/g).

Following sacrifice, the liver lobes were separated and collected in glass tubes containing 5% formalin for histopathology examination. For homogenate preparation, freshly dissected liver tissue was homogenized in phosphate buffer (50 mM, pH 7.2), centrifuged at 3,000 rpm for 10 min and the supernatant separated to yield 25% (w/v) homogenate. It was stored at −80°C until biochemical analysis. The estimation of alanine aminotransferase (ALT) was conducted by using commercially available Bio Merux and Randox kit. The levels of α-fetoprotein (AFP), 8-hydroxy-2-deoxyguanosine (8-OHdG), tumor necrosis factor alpha (TNF-α), and isoprostanes-2 alpha (isoP-2α) were determined by using commercially available ELISA kits (Abcam, Cambridge, MA, USA).

Lipid peroxidation in the blood samples was estimated calorimetrically by measuring thiobarbituric acid reactive substances (TBARS). Briefly, sample (0.2 mL), sodium dodecyl sulfate (8.1%, 0.2 mL), acetic acid (20%, 1.5 mL), and thiobarbituric acid (0.8%, 1.5 mL) were mixed, followed by centrifugation (3,000 rpm for 10 min). The upper organic layer was aspirated, and the optical density was read at 532 nm using a spectrophotometer (Echelle; LTB Lasertechnik Berlin GmbH, Berlin, Germany) against the blank. The levels of lipid peroxides were expressed as millimoles of TBARS/g of liver tissue based on a standard curve. DPPH scavenging activity of plant extracts was measured to investigate their antioxidant status. Ethanol DPPH was added to the prepared stirred utions with the plant extracts and the mixture was kept in the dark for 30 min. Absorbance was measured at 517 nm against a blank whereas, ascorbic acid was used as a positive control.

**Statistical analysis**

The data was subjected to analysis of variance (ANOVA) using COSTAT computer package (Cohort software, Berkeley, CA, USA). The mean values were compared with the least significance difference (at <0.05) test. One-way ANOVA were performed followed by Duncan’s Multiple Range Test. The values are mean ± SD and *p*-values (<0.05) were considered as significant.

**In silico studies**

**Dataset collection and molecular docking**

The 3 active compounds (cabralealactone, solasodine, and salvadorin) were retrieved from ChemSpider (IDs: 23281688, 391288, and 28282881 respectively) and then converted to

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**Table 1** Experimental design

<table>
<thead>
<tr>
<th>Groups (n=5)</th>
<th>Treatments</th>
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<tbody>
<tr>
<td>A</td>
<td>Control</td>
</tr>
<tr>
<td>B</td>
<td>CCl₄ alone</td>
</tr>
<tr>
<td>C</td>
<td>CCl₄+cabralealactone</td>
</tr>
<tr>
<td>D</td>
<td>CCl₄+solasodin</td>
</tr>
<tr>
<td>E</td>
<td>CCl₄+solvadorin</td>
</tr>
<tr>
<td>F</td>
<td>CCl₄+cabralealactone+solasodin</td>
</tr>
<tr>
<td>G</td>
<td>CCl₄+cabralealactone+solvadorin</td>
</tr>
<tr>
<td>H</td>
<td>CCl₄+solasodin+solvadorin</td>
</tr>
<tr>
<td>I</td>
<td>CCl₄+cabralealactone+solasodin+solvadorin</td>
</tr>
<tr>
<td>J</td>
<td>CCl₄+Clavazin</td>
</tr>
</tbody>
</table>

**Notes:** Dose of CCl₄ (1 mL/kg b.wt. per week). Dose of cabralealactone, solasodin, salvadorin (100 mg/kg b.wt.). Dose of Clavazin (200 mg/kg b.wt.).

**Abbreviation:** CCl₄, carbon tetrachloride.
3D Mol2 format using an Open Babel.\textsuperscript{31,32} Since the crystal structures of \textit{Rattus norvegicus} cyclooxygenase-2 (COX2) and TNF-\(\alpha\) were not present in the protein data bank (PDB) repository, a homology modeling has to be conducted by employing a SwissModel.\textsuperscript{33} The crystal structures of murine COX2 (PDB ID: 4FM5) and TNF-\(\alpha\) (PDB ID: 2TNF) were selected to model the rat proteins.\textsuperscript{34,35} To generate a dimeric structure of TNF-\(\alpha\), individual modeled chains were later recombined as a single PDB coordinate file.

The predicted models were further evaluated based on Procheck, ERRAT, Z-score, and VERIFY 3D.\textsuperscript{36–38} The structural alignment of COX-2 and TNF-\(\alpha\) homology models on their templates are presented in Figure 1. Subsequently, the AutoDock Vina standard molecular docking procedure was executed as previously documented and analysis of the binding site was performed before docking.\textsuperscript{39–41} COX2 showed high homology 96\% with the template, considering a 100\% query cover and an E-value of 0.0, while TNF-\(\alpha\) was predicted to have a 95\% homology, 87\% query cover, and an E-value of 6e\(-10\). Superimposition of the co-crystallized structure with inhibitor led to the identification of binding pocket residues. For each protein, docking grid of 30Å \(\times\) 30Å \(\times\) 30Å was made covering the binding site residues. Details of the predocking procedure, including protein and ligand preparation, optimization, and minimization were elaborated elsewhere.\textsuperscript{42}

**Molecular dynamics simulations**

The molecular dynamics simulation is performed using an AMBER12 package to gain insight into the conformational

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**Figure 1** Three-dimensional structural representations of COX-2 and TNF-\(\alpha\), and Ramachandran plot assessment.

**Notes**: Structural alignment of homology models (in red) of TNF-\(\alpha\) (**A**) and COX-2 (**B**) with templates (green) are presented in ribbon representation. Ramachandran plots are shown below for TNF-\(\alpha\) and COX-2 in the left and right panels, respectively.

**Abbreviations**: COX-2, cyclooxygenase-2; TNF-\(\alpha\), tumor necrosis factor alpha.
dynamics of the protein and its binding site during a particular
and, evaluate the validity of the obtained binding
mode of ligands from docking calculations. In addition,
molecular dynamics simulations were used to rid unfavor-
able forces on the protein to reach equilibrium. The ligand
geometry obtained from the docking calculations was used as
a starting conformation for molecular dynamics study.
Partial charges for ligands were calculated and the force field
parameters (force constants for angles and bonds) were used
based on the proposed parameters of ANTECHAMBER and
General Amber Force Field. Amber ff99SB was also used
to provide protein parameters for the modeling. Hydrogen
atoms were added by using the t-leap module in Amber16.

The system was solvated to a 10 Å octagonal box of
TIP3P water molecules and neutralized by adding an appro-
nate number of counter ions. The resultant simulation setup
was subjected to the MD simulations in the ligand-bound
modes of ligands from docking calculations. In addition,
before the MD simulation of 20 ns, the system was relaxed
by performing an energy minimization to remove the largest
strains in the system. Then, after heating the system from
0 to 300 K, isotropic position scaling was used to keep the
system at a constant pressure (1 atm) while the Berendsen
thermostat method was used to maintain a constant tempera-
ture (300 K) during the MD simulation. The PME method
was used to investigate the contribution of long-range elec-
 trostatic interactions. A 10 Å cut-off was used for both PME
and van der Waals interactions. To examine the convergence
of the system to equilibrium, the temperature, density, and
the total energy was checked to ensure post-MD analysis.
MD production run of 20 ns was executed for each complex
while the Combined Cluster Analysis (CPTRAJ) module of
Ambert 16 was used for post-MD analysis.

Results
Significant restoration of the investigated biochemical vari-
ables in rat groups that received the herb treatments were
seen (Table 2). The biochemical parameters of rats that
received monotherapy of salvadorin and solasodin extracts
(group H) were significantly ameliorated (41.55±4.34 IU/L,
17.71±1.20 ng/mL, 18.01±2.38 pg/mL, 15.20±2.58 ng/mL,
15.41±3.03 pg/mL, 3.65±0.53 mmol/mL, and 1.55±0.386 ng/mL)
compared with the control group. In addition, the levels of
ALT, AFP, 8-OHdG, TNF-α, IsoP-2α, malondialdehyde
(MDA), and COX-2 were significantly improved compared
with the control. There remained a significant change in the
variables of groups treated with plant extracts. Also, combi-
nation therapies of these plant extracts caused significant and
remarkable restoration of all parameters compared with the
standard drug Clavazin. It is plausible that the antioxidant
effect of the plant extracts activate the proteolytic enzymes
involved in DNA repair that supports its traditional use as
anti-cancer agent.

Histopathological findings
The histopathological findings of the liver sections in the
control group (Figure 2A) indicated the presence of visible
central veins, a terminal hepatic venule, and an interlobular
portal vein. The hepatocytes were in contact with the

Table 2 Biochemical response of cabralealactone, solasodin, salvadorin following CCl4-induced liver insult in rats

<table>
<thead>
<tr>
<th>Groups</th>
<th>Mean ± SD (n=10)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>ALT (IU/L)</td>
</tr>
<tr>
<td>A</td>
<td>25.1±4.32</td>
</tr>
<tr>
<td>B</td>
<td>179.82±20.03</td>
</tr>
<tr>
<td>C</td>
<td>74.24±11.97</td>
</tr>
<tr>
<td>D</td>
<td>63.50±5.58</td>
</tr>
<tr>
<td>E</td>
<td>53.07±17.48</td>
</tr>
<tr>
<td>F</td>
<td>42.07±13.25</td>
</tr>
<tr>
<td>G</td>
<td>40.13±2.75</td>
</tr>
<tr>
<td>H</td>
<td>41.55±4.34</td>
</tr>
<tr>
<td>I</td>
<td>20.45±5.05</td>
</tr>
<tr>
<td>J</td>
<td>35.10±8.12</td>
</tr>
<tr>
<td>LSD (0.05)</td>
<td>11.70</td>
</tr>
<tr>
<td>p-value</td>
<td>0.0413</td>
</tr>
</tbody>
</table>

Notes: CCl4, @ 1 mL/kg b.w. per week. Cabralealactone, solasodin, salvadorin @ 100 mg/kg b.w. Clavazin @ 200 mg/kg b.w. Superscript letters indicate that values not sharing a common letter differ significantly.

Abbreviations: 8-OHdG, 8-hydroxy-2-deoxyguanosine; ALT, alanine aminotransferase; AFP, α-fetoprotein; CCl4, carbon tetrachloride; COX-2, cyclooxygenase-2; IsoP-2α, isoprostanes-2 α; LSD, least significance difference; MDA, malondialdehyde; TNF-α, tumor necrosis factor alpha; A, Control; B, CC14; C, cabralealactone; D, solasodin; E, salvadorin; F, cabralealactone+solasodin; G, cabralealactone+salvadorin; H, solasodin+salvadorin; I, cabralealactone+solasodin+salvadorin; J, Clavazin.
sinusoids, which appear as distensible vascular channels in connection with highly fenestrated endothelial cells surrounded by a population of phagocytic Kupffer cells. However, CCl₄-treated liver sections indicated the presence of prominent necrosis, vacuolization, central vein congestion, and fat vesicles (Figure 2B). The sinusoids, which are in dilated form, are characteristic of widened hepatic capillaries suggestive of hepatic tumors occurring due to sinusoidal infiltration by malignant or benign cells. The prominent architectural alterations in the CCl₄-treated toxic liver are karyomegaly (enlargement of the nucleus) and populated activated Kupffer cells. Examination of the liver sections in animals treated with standardized extracts of cabralealactone (100 mg/kg) revealed that the liver almost regained up to 30% of its normal architecture with reduced intensity of liver fibrosis (Figure 2C). Also, some liver recovery was observed in animals exposed to solasodin (Figure 2D) and salvadorin (Figure 2E) monotherapies as characterized by the lower extent of vacuolization seen.

Nevertheless, compared with the monotherapies, a significantly better protective effect was observed in the liver sections of rats that received combined treatment of standardized extracts of cabralealactone and salvadorin (Figure 2F), cabralealactone and solasodin (Figure 2G), and solasodin and salvadorin (Figure 2H), which were characterized by the presence of normal hepatic cords, absence of fatty infiltration, reduced intensity of sinusoidal dilation, and central vein congestion. However, the best recovery was seen in rat groups exposed to the 3 polyherbal treatment of cabralealactone, salvadorin, and solasodin (Figure 2I), indicating the potentials of these plants as anti-inflammatory agents.

**DPPH scavenging activity**
Significant DPPH scavenging activity was observed with salvadorn (94.26%) followed by cabralealactone (89.26%) and solasodin (87.06%) compared with ascorbic acid (91.26%) as standards as in Figure 3.
In silico studies
Molecular docking and MD simulations
The 3 active compounds were used for molecular docking with COX-2 and TNF-α. The chemical 2D structure representations of the active compounds are shown in Figure 4. The binding energy along with the pharmacokinetics properties of each compound is tabulated in Table 3 while the molecular interactions are diagrammatically represented in Figures 5 and 6. Relative to the COX-2 enzyme, salvadorin exhibited a higher binding energy of $-9.8 \text{ kcal/mol}$ compared with solasodine and cabralealactone, which showed $-7.9$ and $-6.8 \text{ kcal/mol}$, respectively. In relation to TNF-α, salvadorin exhibited a potentially higher binding energy of $-11.4 \text{ kcal/mol}$, while the binding energies corresponding to solasodine and cabralealactone were $-8.1$ and $-7.4 \text{ kcal/mol}$, respectively. Moreover, all ligands were docked in the lowest energy conformation inside the binding pocket as shown in Figures 5 and 6.

MD simulation of 20 nanosecond (ns) was performed on each lowest docking energy conformation that was chosen as a starting complex for COX-2 and TNF-α. The root mean square deviation (RMSD) of all backbone atoms of each target protein on the bound ligand was computed as a function of time in 1 ns as displayed in Figure 7. Relative to COX-2-salvadorin complex, the RMSD initially increased until 6 ns and then gradually became stable between the mean value of 1.75Å and 2.51Å until the end of the simulation (red), while the RMSD continued to fluctuate between 1.1Å and 2.2Å in the final 10 ns and continued to increase in both cases where COX-2 complexed with solasodine (yellow) and cabralealactone (green). In relation to the TNF-α-salvadorin complex, the backbone atoms RMSD remained stable throughout the 20 ns simulations between 1.15Å and 1.76Å and oscillated around the mean value of 0.51Å. On the other hand, TNF-α-solasodine showed stability in RMSD with small fluctuations while the backbone atoms of TNF-α complexed with cabralealactone showed higher fluctuations and continued to rise.

Figure 3 DPPH scavenging activity (%). Abbreviation: DPPH, 1,1-diphenyl-2-picrylhydrazyl.
Table 3  Molecular docking energies and drug-likeness properties of the active compounds

<table>
<thead>
<tr>
<th>Properties</th>
<th>Cabralealactone</th>
<th>Solasodine</th>
<th>Salvadorin</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mass</td>
<td>552.70</td>
<td>429.68</td>
<td>414.62</td>
</tr>
<tr>
<td>Log-p-value</td>
<td>8.69</td>
<td>5.57</td>
<td>6.34</td>
</tr>
<tr>
<td>H-bond acceptors</td>
<td>3.00</td>
<td>3.00</td>
<td>3.00</td>
</tr>
<tr>
<td>H-bonds donors</td>
<td>0.00</td>
<td>1.00</td>
<td>0.00</td>
</tr>
<tr>
<td>Rotatable bonds</td>
<td>5.00</td>
<td>0.00</td>
<td>1.00</td>
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<tr>
<td>PSA</td>
<td>35.53</td>
<td>32.70</td>
<td>43.37</td>
</tr>
<tr>
<td>ROS violations</td>
<td>2.00</td>
<td>1.00</td>
<td>1.00</td>
</tr>
<tr>
<td>Atoms</td>
<td>78.00</td>
<td>78.00</td>
<td>72.00</td>
</tr>
<tr>
<td>Rings</td>
<td>7.00</td>
<td>6.00</td>
<td>5.00</td>
</tr>
<tr>
<td>Binding affinity of compounds (kcal/mol) with COX-2</td>
<td>-6.80</td>
<td>-7.90</td>
<td>-9.80</td>
</tr>
<tr>
<td>Binding affinity of compounds (kcal/mol) with TNF-α</td>
<td>-7.40</td>
<td>-8.10</td>
<td>-11.40</td>
</tr>
</tbody>
</table>

Abbreviations: COX-2, cyclooxygenase-2; PSA, polar surface area; ROS, Lipinski’s rule of 5; TNF-α, tumor necrosis factor alpha.

Discussion

To our knowledge, this is the first study to confirm that cabralealactone, solasodin, and salvadorin have DNA-damage protective effects against CCl₄-induced genotoxicity and may have the potential to be investigated as a COX-2 inhibitor and anti-inflammatory agent. The emergence of oxidative stress as a central player in chronic metabolic diseases has opened new avenues for research in the pathology of a wide range of diseases and their treatments to increase the survival rate. The mechanism of action of CCl₄-induced liver damage is a complex mechanism that indicates its absorption in parenchymal hepatocytes and its metabolism results in the formation of free radicals in the presence of cytochrome P₄₅₀-dependent monooxygenases. The resultant free radicals form a covalent bond with biological macromolecules preferably with fatty acids and phospholipids of the plasma membrane. Once free radicals are produced, lipid peroxidation is initiated, resulting in cellular membrane peroxidation, chloromethylation reaction, polyunsaturated fatty acids (PUFA) deterioration, and ultimately disturbance of the structural integrity. The destruction of phospholipid-associated PUFA leads to physiological alterations in the mitochondria, endoplasmic reticulum, and is mostly prevalent in cellular plasma membrane. In addition to this, the metabolism of calcium-associated homeostasis is disturbed, leading to the loss or leakage of cellular calcium. The resulting end products of PUFA form a covalent association with proteins and mimic the catalytic activity of enzymes.

Figure 5  Structure representation of cross-sectional area of COX-2 with docked compounds.

Notes: Cross-sectional area of COX-2 with docked compounds, cabralealactone (pink), solasodin (light brown), and salvadorin (green) in stick representation inside the binding pocket. Molecular interactions in different colors are displayed alongside using a DS visualizer (Accelrys Inc., San Diego, CA, USA).

Abbreviation: COX-2, cyclooxygenase-2.
CCL4 disrupts the cellular integrity by activating inflammatory cytokines such as TNF-α, interleukin (IL)-6, ALT, 8-OHdG (the hallmark of DNA-based oxidative damage, MDA, and isoP-2α). A study reported that patients with raised levels of AFP, histologically increased steatosis, and inflammatory infiltration point to DNA-based oxidative stress. Another research demonstrated that high levels of oxidative stress contribute to dysfunctioning of adipokines, leading to dysregulated production of reactive oxygen species from the adipose tissues that caused increased oxidative damage in the blood. This ultimately affected the normal functioning of the liver, skeletal muscles, and aorta of the patients with chronic liver disease. The response of cells to the DNA damage is demonstrated via the production of tumor suppressor gene p53 that allows genetic fidelity by inducing cell cycle arrest, ultimately ameliorating the cell damage due to apoptosis as shown in Figure 8.

TNF-α is a cytokine with a molecular mass of 55 kDa, which mediates its cytotoxic activities through the expression of 2 receptors (p55 and p75). The induction of reactive oxygen species production by TNF-α occurs via the endothelial mitochondria and nicotinamide adenine dinucleotide phosphate in the cell membrane. Thus, the cytotoxic activity of TNF-α induced by CCL4 disrupts the hepatocyte mitochondrial function, which stimulates the production of reactive oxygen species in the mitochondria, particularly at the ubiquinone site while TNF-α disrupts the mitochondrial electron transport chain at the complex III region, ultimately resulting in the accumulation of free oxygen radicals inside the mitochondrion. In addition, the increased production of TNF-α also stimulates the nuclear factor-kappa beta activation and upregulated IL-6 expression.

The antioxidant-rich standardized extracts of cabralea lactone, salvadorin, and solasodin are useful in repairing the DNA damage and ameliorating the TNF-α-induced cytotoxicity in hepatocytes and malignant transformation of hepatocytes as evidenced by the histological findings. IsoP-2α is recognized as potent indicator of increased oxidative stress in hepatic cirrhosis conditions. The formation of isoprostanes (the isomer of prostaglandin F-2α) is non-enzymatic when free oxygen radicals attack arachidonic acid in the lipid component of the cell membrane. Therefore, isoP-2α is extensively employed as a gold standard index of oxidative stress and DNA damage in liver tissue (Table 4). In our study, high significant correlation exists between some of the aforesaid parameters, which reveals that increased release of ALT from the hepatocytes is responsible for elevated AFP levels in the tissue due to increased production of reactive oxygen species associated with cellular damage, ultimately leading to the DNA damage. Elevated levels of

**Figure 6** Molecular surface representation of dimeric TNF-α.

**Notes:** Molecular surface representation of dimeric TNF-α (in 2 different colors for each chain) with docked compounds, cabralea lactone (pink), solasodine (light brown), and salvadorin (green) in stick representation inside the binding pocket. Molecular interactions in different colors are displayed alongside using a DS visualizer (Accelrys Inc., San Diego, CA, USA).

**Abbreviation:** TNF-α, tumor necrosis factor alpha.
AFP may result in the induction of cycle of inflammation; necrosis and regeneration due to CCl₄-induced liver injury. Upregulated cellular turnover in the inflammation cycle and oxidative DNA damage results in the appearance of genetic and epigenetic changes, which may be due to the increased expression of cellular oncogenes and growth and angiogenic factors; activation of telomerase activity; and downregulation of tumor suppressor genes. Nevertheless, the result should be interpreted with caution since AFP has been reported to yield some false-positive findings (20%). Nevertheless, both in silico and in vivo studies to determine the binding potential and stability of protein’s all-backbone atoms via molecular docking, targeting TNF-α and COX2 enzyme followed by 20 ns MD simulation to analyze the atomistic behavior of protein’s all-backbone atoms with ligand bound inside the binding pocket. Between the 3 compounds, salvadorin yielded the most stable RMSD with minimum fluctuation seen in the final 10–12 ns, which inferred better stability during the formation of the complex (Figure 7).
studies suggested that all 3 active compounds have good and desirable affinity parameters, yielding stable molecular interactions.59–61 Thus, these compounds can be configured as promising agents for the development of COX-2 and TNF-α antagonists.

**Limitations**

The limitation of the current study is the relatively low number of samples and variables due to financial constraints. Efficacy of the results can be improved by increasing the number of samples and taken into consideration its molecular level.

**Table 4** Pearson correlation coefficients of different variables in liver tissues of rats under CCL₄ stress receiving cabralealactone, solasodin, salvadorin, and when used in combinations

<table>
<thead>
<tr>
<th>Variables</th>
<th>ALT (IU/L)</th>
<th>AFP (ng/L)</th>
<th>8-OHdG (pg/mL)</th>
<th>TNF-α (ng/mL)</th>
<th>IsoP-2α (pg/mL)</th>
<th>MDA (nmol/mL)</th>
<th>COX-2 (ng/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ALT (IU/L)</td>
<td>1.000</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AFP (ng/L)</td>
<td></td>
<td>0.913**</td>
<td>0.751***</td>
<td>0.953**</td>
<td>0.905**</td>
<td>0.912**</td>
<td>0.594***</td>
</tr>
<tr>
<td>8-OHdG (pg/mL)</td>
<td>0.817**</td>
<td>0.909**</td>
<td>0.889**</td>
<td>0.824**</td>
<td>0.815**</td>
<td>0.679**</td>
<td></td>
</tr>
<tr>
<td>TNF-α (ng/mL)</td>
<td>1.000</td>
<td>0.912**</td>
<td>0.884**</td>
<td>0.749**</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IsoP-2α (pg/mL)</td>
<td>1.000</td>
<td>0.912**</td>
<td>0.884**</td>
<td>0.749**</td>
<td>0.679**</td>
<td></td>
<td></td>
</tr>
<tr>
<td>MDA (nmol/mL)</td>
<td>1.000</td>
<td>1.000</td>
<td>0.669**</td>
<td>0.648**</td>
<td>0.679**</td>
<td></td>
<td></td>
</tr>
<tr>
<td>COX-2 (ng/mL)</td>
<td>1.000</td>
<td>1.000</td>
<td>1.000</td>
<td>1.000</td>
<td>1.000</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Note:** **Significant at the 0.01 level (two-tailed).**

**Abbreviations:** 8-OHdG, 8-hydroxy-2-deoxyguanosine; ALT, alanine aminotransferase; AFP, α-fetoprotein; COX-2, cyclooxygenase-2; isoP-2α, isoprostanes-2-alpha; MDA, malondialdehyde; TNF-α, tumor necrosis factor alpha.
Hence, further studies in these areas would be helpful in understanding and raising new sets of ideas in the field.52–64

Conclusion
It is concluded that cabralealactone, solasodin, and salvadorin showed hepatoprotective and DNA-damage protective effects against CCl₄-induced toxicity. The herbs successfully restored the normal architecture of hepatocytes and had the potential to be used as inhibitor to COX-2 and TNF-α. They can combat oxidative stress and liver injuries when used both as mono and combination therapies, with the later showing better findings perhaps due to synergistic effects.

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Disclosure
The authors report no conflicts of interest in this work.

References


