

Fat infiltration in the gluteus minimus muscle in older adults

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Purpose: Fat infiltration (FI) in the gluteus minimus muscle (GMin) has been reported as being restricted to the anterior part. The aim of this study was to verify whether FI is localized only to the anterior part of the GMin through the dissection of large number of elderly cadavers and to clarify the underlying mechanism of FI localized to a specific region.

Materials and methods: Thirty-eight hips from 20 embalmed elder cadavers were dissected for verification of FI localization. The samples of three regions – the fat infiltrated region of the anterior part (FI), the region without FI (non-FI), and the posterior region – were harvested from nine hips for the histological fiber type assessment.

Results: The FI area was distributed at 2.6°–38° from the anterior border within the anterior part, of which function is responsible for hip extension and internal rotation and is likely reduced by disuse following aging. The number of type I fibers was lowest in FI, followed by non-FI, while the number of type I fibers was highest in the posterior. Meanwhile, the number of type II fibers was equivalent across the three regions. Atrophic type II fibers were observed in overall samples of the three regions, and grouped atrophy, which is evidence of denervation, was observed in FI and non-FI samples. Type grouping, which suggests to be reinnervated, was found in the non-FI and posterior samples but very little in the FI samples.

Conclusion: FI in the GMin in older adults restricted to deep within the anterior part is secondary to disuse and is strongly related to impaired denervation–reinnervation process.

Keywords: gluteus minimus, fat infiltration, disuse, denervation, fiber type

Introduction

There have been many reports on the decline in physical activity and the risk of falls associated with the atrophy of the gluteus medius and minimus muscles (GMed and GMin, respectively), including fat infiltration (FI).^{1–6} FI in the GMin is localized deep within the anterior part, and its content increases with increasing age.^{2,4} Only one study has investigated the degree of FI in the GMin using cadavers, revealing that out of 12 cadavers, mild FI and extensive FI were found in 6 and 2 cadavers, respectively. Furthermore, FI was restricted to the anterior part, suggesting that FI is associated with aging.

Muscle fiber type studies assessing aging process that results in muscle atrophy, especially reduction in the cross-sectional areas (CSAs) of type II fibers, have been investigated.^{7–12} Type II fiber has a short contraction time and an ability to produce high tension. With aging and being inactive, the muscles lose their activity for rapid force production as a result of preferential atrophy of type II fibers. The strength decline in old age was correlated with the CSAs of type II fibers.⁷ The aim of this study was to verify whether FI is localized only into the anterior part of the GMin through the dissection of a large number of elderly cadavers and to clarify the underlying mechanism

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of FI localized to a specific region of the GMin through a muscle fiber-type assessment.

Materials and methods

Macroscopic observation

We dissected 38 hips from 20 embalmed cadavers of elderly individuals (14 males and 6 females) to expose the GMin. Their mean age at death was 82.8 (ranged 64–95) years. Two hips were excluded due to previous surgery invading the tendon of the GMin.

According to the previous radiological studies that FI localized to the bony surface of muscle, the dissection was performed using a technique described by Walters et al.¹³ The muscle belly was elevated from its attachment to the outer aspect of the pelvis. The capsular attachment and the tendinous insertion to the greater trochanter were sharply dissected. The specimens formed a fan with an apex assuming the tendinous insertion. The fat content in the muscle, which was greater than or equal to the muscle tissue content (corresponding to stages III and IV of Goutallier's classification on computer tomography¹⁴), was observed on the deep surface and measured as the angle from the anterior border (Figure 1).

Microscopic observation

Tissue sampling and immunohistochemistry

We assessed randomly selected 9 hips from 38 cadavers and harvested the samples from three regions: the fat infiltrated

region of the anterior part by gross inspection (FI), the region without FI (non-FI), and the posterior (C) region.

Samples of muscle tissue, each ~10–20 mm in length and 5–10 mm in depth, were collected from three regions of the GMin of each specimen. Each sample was post-fixed overnight in Bouin's solution at 4°C. Tissues were then dehydrated and embedded in Paraplast embedding media (Sigma, St Louis, MO, USA). Cross sections of each sample were cut at a thickness of 5 µm.

The sections were deparaffinized and hydrophilized. H&E staining for GMin was performed according to conventional methods. Nearly adjacent sections were incubated with an antigen retrieval agent (pH 9.0; Nichirei Co., Tokyo, Japan) at 95°C for 20 minutes. A blocking solution consisting of 2% bovine serum albumin (Sigma) in PBS was applied at 37°C for 30 minutes. Two primary antibodies, anti-fast myosin skeletal heavy chain (1:1,000, #ab91506; Abcam, Cambridge, UK) and anti-slow myosin skeletal heavy chain (1:10,000, #ab11083; Abcam), were used to label the myosin heavy chains of type I and type II fibers, respectively. Sections were incubated overnight at 37°C with the primary antibody. The secondary antibodies, anti-mouse IgG Alexa Fluor 488 (1:400, #A-11029; Thermo Fisher Scientific Inc., Cambridge, MA, USA) and anti-rabbit IgG Alexa Fluor 568 (1:400, #A-11036; Thermo Fisher Scientific Inc.), were used for an hour at 37°C. As a negative control of immunohistochemistry, nearly adjacent sections were incubated without the primary antibody and then incubated with the secondary antibody. No labeling was confirmed in these control sections.

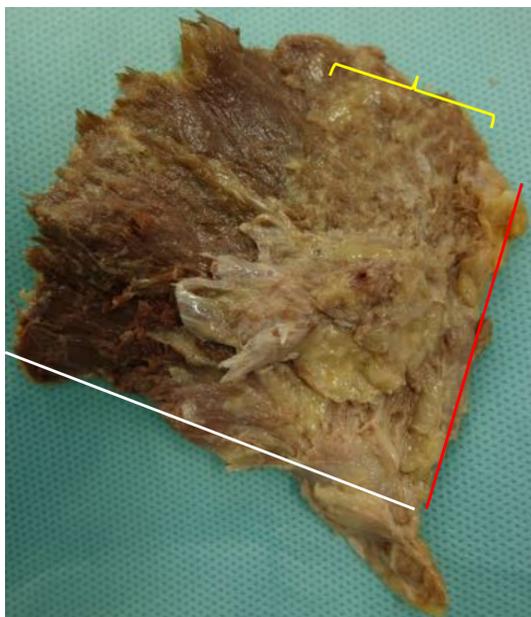


Figure 1 Appearance of the deeper surface of gluteus minimus.

Notes: Red line: the anterior border; white line: the posterior border; yellow bracket: FI area.

Abbreviation: FI, fat infiltration.

Imaging, counting the fiber types, and determining the fiber size

A total of five images were obtained for each specimen. Each fiber was counted on the Adobe Photoshop (Adobe Systems Inc., San Jose, CA, USA); the number of fibers was then expressed in terms of the number of fibers/mm² (Figure 2). The hybrid fibers were excluded because they were so few in this study. The CSA of each fiber was determined using ImageJ (National Institutes of Health, Bethesda, MD, USA). Over 50 sections coming from nine specimens (six sections from a single specimen) were examined. In adult male, the diameter of muscle fibers of the quadriceps is regarded as between 40 and 80 µm. We defined the fiber with more than 80 µm of diameter as enlarged fiber or hypertrophy and the fiber <40 µm as small fiber or atrophy.

Statistical analysis

Free statistical software “R” version 2.8.1 (The R Project for Statistical Computing; open source) was used for statistical

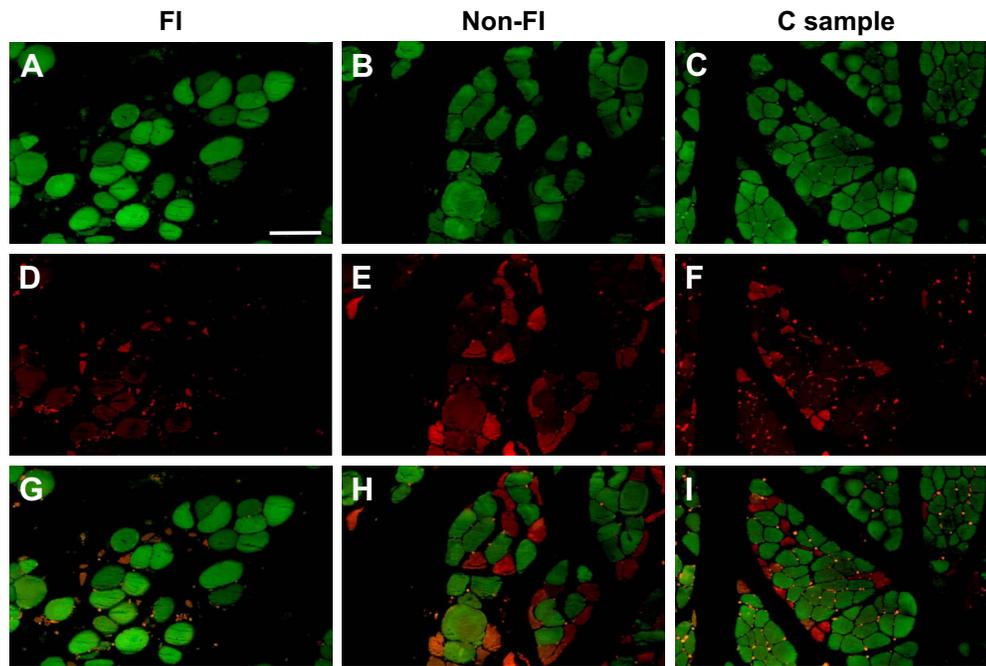


Figure 2 Complementary immunohistochemical staining for each fiber type.

Notes: Complementary immunohistochemical staining for type I fibers (A–C), type II fibers (D–F), and a merging (G–I) of the FI, non-FI, and C samples, respectively. Magnification: $\times 200$. Scale bar indicates 100 μm .

Abbreviations: C, posterior; FI, fat infiltration.

analysis. The data were analyzed using repeated measures ANOVAs. The threshold used for statistical significance was $p=0.01$.

Ethics

The cadavers used in this study were donated for the dissection training for medical students and the related studies for researchers. Written informed consent was obtained from all of them before their death. This study was approved by the Ethics Committee of Yamagata University School of Medicine (#315), Yamagata, Japan, and was also approved by the Ethics Review Board of the Yamagata Prefectural University of Health Sciences (#1701-22), Yamagata, Japan.

Results

Macroscopic observation

On the superficial surface of the GMin, the muscle fibers ran fan-shaped with a mean angle of 78.9° (60° – 110°). The muscle could be divided into two parts because the fibers of the anterior and posterior parts were oriented craniocaudally and horizontally, respectively. The anterior part was larger and amounted to two-thirds of the muscle. The fibers of the posterior part converged on the caudal area of the aponeurosis, while the cranial area centralized the fibers of the anterior part. Then, the aponeurosis gradually turned into tendinous portion (Figure 3). Superficial infiltration of fatty tissue was

rarely found other than in one hip in which all muscle fibers became fatty tissue.

The deep surface of the GMin below the inferior gluteal line was covered by the aponeurotic tissue, where the muscle fibers of both parts blended. Therefore, the observation

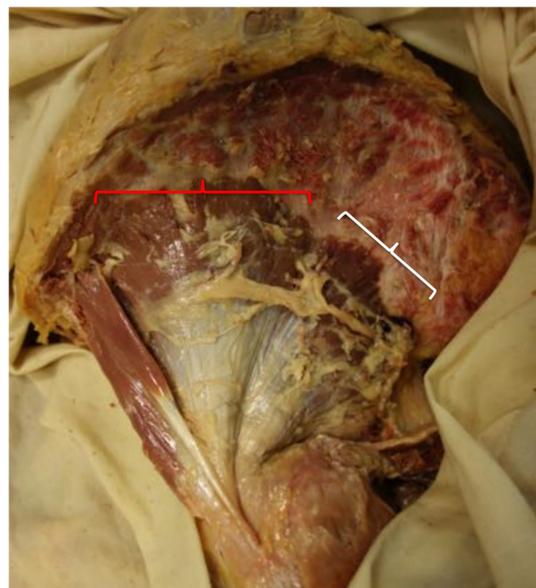


Figure 3 Appearance of the superficial surface of gluteus minimus.

Notes: The muscle fibers ran fan-shaped. The muscle could be divided into the anterior (red bracket) and posterior parts (white bracket). The anterior part was larger and amounted to two-thirds of the muscle.

of the FI area was performed on the deep surface between the anterior and inferior gluteal lines.

The macroscopic FI was deeply localized in the bony origin in all specimens. The FI area was distributed at a mean angle of 33.4°, between 2.8° (minimum 0°) and 36.2° (maximum 65°) from the anterior border, within the anterior part. The fatty fibers were hardly found in the posterior part. These findings are summarized in Table 1.

Microscopic observation

Predominance of type I fiber was observed in all the three regions (62%–74%). The total number of type I and type II

Table 1 Demographics of all specimens

Sex	Age	Hip	Fan-shaped angle (°)	FI distribution (°)
M	90	R	90	10–40
		L	80	10–35
M	74	R	90	0–60
		L	60	0–15
M	89	R	90	5–35
		L	110	0–30
M	75	R	70	10–30
		L	70	0–20
F	93	R	80	15–50
		L	70	0–35
M	92	R	75	0–30
		L	90	0–35
F	94	R	80	10–45
		L	65	5–20
F	77	L	90	0–40
		F	90	R
L	80			0–30
F	95	R	90	15–55
		L	70	15–30
M	89	R	90	10–45
		L	110	0–65
M	76	R	80	0–30
		L	80	0–30
M	67	L	80	0–40
M	82	R	80	0–40
		L	80	0–40
M	77	R	80	0–50
		L	80	0–35
F	76	R	80	0–35
		L	70	0–30
M	77	R	80	0–40
		L	70	0–40
M	64	R	70	0–40
		L	70	0–35
M	88	R	60	0–20
		L	70	0–30
M	90	R	70	0–40
		L	70	0–30
Mean			78.9	2.8–36.2 (33.4)

Abbreviations: F, female; FI, fat infiltration; M, male; L, left; R, right.

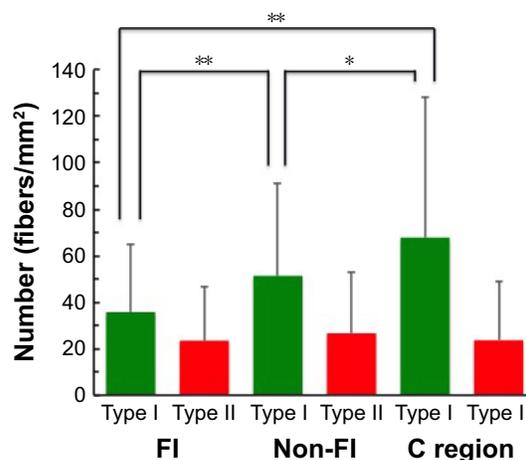


Figure 4 The number of each fiber type. **Notes:** The number of type I fibers decreased progressively from the FI to the non-FI to the C regions, while that of type II fibers was equivalent across the three regions. Each column shows the mean, and bar represents the SD in each group. * $p < 0.01$; ** $p < 0.001$. **Abbreviations:** C, posterior; FI, fat infiltration.

fibers and the number of type I fibers were lowest in the FI, second lowest in the non-FI, and highest in the C regions, while that of type II fibers was equivalent across the three regions. There was a significant difference between the number of type I fibers in the FI and non-FI and that in the C region ($p = 1.2e-06$ and 0.0026 , respectively; Figure 4). The CSAs of type I and type II fibers among the three regions were $1,520.18 \pm 604.39$ and $374.62 \pm 170.58 \mu\text{m}^2$, respectively. The CSAs of type I and type II fibers of each region are shown in Figure 5. In the statistical analysis of the sizes of each fiber type, there were significant differences between the CSAs

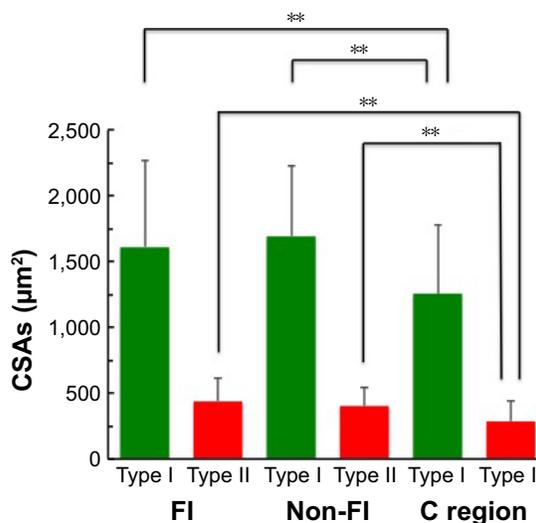


Figure 5 The CSAs of type I and type II fibers in the three regions. **Notes:** The CSAs of type I and type II fibers in the FI region were equal to those in the non-FI region. The ones in the C region were significantly smaller than those in the other regions. Each column shows the mean, and bar represents the SD in each group. ** $p < 0.001$. **Abbreviations:** C, posterior; CSAs, cross-sectional areas; FI, fat infiltration.

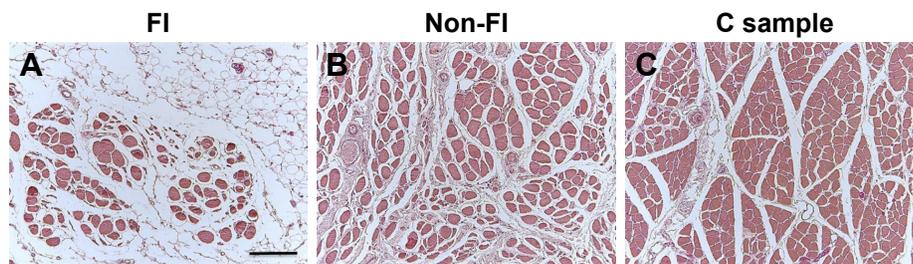


Figure 6 Cross section of samples.

Notes: In the FI sample, the rounded and enlarged fibers and atrophied fibers were scattered (A). The interstitial spaces were slightly widened and closed in the samples of the non-FI and C, respectively. The form of these fibers was ensiform (B, C). H&E staining, magnification: $\times 100$. Scale bar indicates 200 μm .

Abbreviations: C, posterior; FI, fat infiltration.

of type I fibers in the FI and non-FI and that in the C region ($p=1.1\text{e-}05$ and $1.1\text{e-}7$, respectively). The CSAs of type II fibers in the FI and non-FI were statistically different from that in the C region ($p=5.6\text{e-}10$ and $4.2\text{e-}10$, respectively). The CSAs of type I and type II fibers in the FI region were equal to those in the non-FI region.

From the results of CSA measurement, the diameters of type I and type II fibers were estimated at 60–90 and 28–46 μm , respectively. There were more than a few enlarged type I fibers and small type II fibers. The histologic appearance of the FI samples showed markedly widespread muscle tissue loss and fatty tissue infiltration. The rounded and enlarged type I fibers and atrophied type II fibers were scattered. In the non-FI samples, the interstitial space was slightly widened. The form of type I fibers was varied; some were small, and others were rounded and enlarged. Type II fibers atrophied. In the samples of the C group, small fibers were observed. Type II fibers atrophied as well (Figure 6). Type grouping was found in the non-FI and C samples, but very little was found in the FI samples (Figure 7).

Discussion

In macroscopic observation, we found that the FI in the GMin of the elderly cadavers was localized within the angle

of 2.8° – 36.2° from the anterior border of the GMin. Flack et al² found that the GMin was composed of four compartments, namely, the anterior, mid-anterior, mid-posterior, and posterior subregions. They also found FI in 8 of 12 embalmed cadaver specimens that were predominantly restricted within the anterior subregion and throughout the mid-anterior subregion. The finding in this study coincides with Flack's results.

Pfirmsmann et al¹ previously noted in an MRI study that the FI of the GMin was evident in the anterior two-thirds and uncommon in the posterior part. We previously reported that the FI of the GMin was localized deep within the anterior region, and the fat content increased with increasing age.⁴

Beck et al¹⁵ artificially divided the GMin into four sectors, and sectors I and II, which correspond to the anterior and mid-anterior subregions of Flack's divisions, worked well in abduction and acted as internal rotators with hip extension. Semciw et al¹⁶ demonstrated different roles between the anterior and posterior parts of the GMin electromyographically. The posterior part is more active in the early stance phase, both parts co-contract in the late mid-stance phase, and the anterior part has a large internal rotational torque and counteracts the hip extension moment. Flack et al² also reported that the physiological CSAs of the anterior and mid-anterior

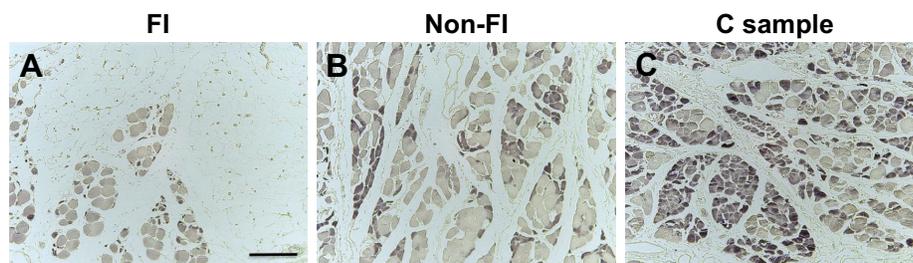


Figure 7 Vigorous regenerative areas in the three regions.

Notes: A cluster consisting of several atrophied type II fibers (stained in dark) was barely found in the FI sample (A). A small-type grouping of type II fibers was found in the non-FI sample (B), while reinnervated fibers occupied a large area in the C sample (C). DAB staining, Magnification: $\times 100$. Scale bar indicates 200 μm .

Abbreviations: C, posterior; FI, fat infiltration.

subregions were smaller than those of the mid-posterior and posterior subregions. It is well known that with increasing age, stride length is reduced and hip extension capacity decreases, followed by a decline in stance phase. Using MRI, Kumagai et al¹⁷ reported that the GMin worked more actively in hip abduction because the increase in signal intensity of the GMin was greater than that of the GMed immediately after a one-legged stance, and even the GMed has a larger CSA. In an electromyographical study, Ganderton et al¹⁸ suggested that a lack of terminal hip extension and internal rotation reduces the stimulus for the anterior part to contract. A reduction of gait performance with increasing age, such as reduced stance phase duration and a loss of hip extension and subsequent loss of internal rotation, may be exacerbated predominantly in the thinner anterior part of the GMin.

The details of individual activities of the specimens during lifetime were unidentified because the specimens were donated elder cadavers. However, to a greater or lesser extent, all specimens showed FI in the deep anterior part of the GMin. Therefore, aging or disuse secondary to aging might influence the FI localization in the anterior part of the GMin.

In microscopic assessment of the muscular fiber type, the number of type I fibers was lowest in the FI, second lowest in the non-FI, and highest in the C regions, while the number of type II fibers was equivalent across all the three regions. The CSAs of type I and type II fibers in the C region were significantly lower than those in the FI and non-FI regions. The CSAs of type I and type II fibers were 1,520.18 and 374.62 μm^2 , respectively. These values were relatively low, especially those of type II. Lexell et al⁸ found that the total number of fibers in the vastus lateralis significantly decreases with increasing age. Nilwik et al¹² reported that age-related reduction in muscle mass was mainly attributed to specific type II fiber atrophy with no substantial decline in the number of fibers, namely, a decline in the CSAs of type II fibers. Lexell and Tayler found a selective reduction in the size of type II fibers in the elders compared with the youngsters. They concluded that such a finding was very common in the elders due to inactivity and immobilization.⁹ Larsson et al revealed that type II fiber atrophy was responsible for age-related strength in the quadriceps muscle.⁷ The type II fiber size in the GMin was very small in this study compared with those of the vastus lateralis in previous studies. The reason for small size of the type II fiber was not only the subjects' age (mean age at death was 82.8 years) but may be characteristic of the GMin. In an 11-year study, Aniansson et al¹⁰ established that there was a significant increase in both type I and type II fiber areas

instead of significant reductions in the proportion of type II fibers. Frontera et al¹¹ also reported that the number of both fiber types declined, while their diameter increased with age due to compensatory hypertrophy, as determined through an 8.9-year longitudinal study with a sample of participants with a mean age of 71 years. In the FI and non-FI regions, the number of type I fibers was lower, while the CSAs of both types were larger in the present study. These fiber characteristics suggest an influence of inactivity and aging in the FI and non-FI regions.

In the histological analysis of the FI samples, the residual fibers with rounded and/or enlarged type I and atrophied and/or rounded type II fibers were scattered. Fiber hypertrophy in aged muscle is considered to be a compensatory mechanism to correct for a muscle size deficit resulting from motor unit loss.⁹⁻¹¹ The rounded and enlarged forms seen in the FI and non-FI samples seemed to result from fiber splitting and myogenic compensation.

Neurogenic changes in histologic appearance are characterized as atrophic fibers in acutely denervated muscle and as grouped atrophy in progressively denervated one. Type grouping occupied by type II, which is evidence of muscle reinnervation by sprouting, was found in the non-FI and C samples but very little in the FI samples. This finding suggests that the fibers in the non-FI and C regions have a capacity to regenerate. In the FI region, denervation outpaces reinnervation because the fibers were replaced by fatty tissue subsequent to fiber loss without any alteration of fiber type. In other words, the fibers in the FI region no longer had any regenerative capacity. These facts suggest that the denervation–reinnervation process was impaired in the FI region.

The number of motor neurons in the spinal anterior horn decreases with increasing age.¹⁹ Aging also brings about changes in the motor unit and neuromuscular junction in the context of the impaired denervation–reinnervation process. Campbell et al²⁰ found motor unit loss apparent over the age of 60 years. Hourigan et al²¹ demonstrated that neuromuscular junction instability (increased near fiber jiggle) is associated with aging. In a study using mice, Butikofer et al²² indicated that aging is associated with neuromuscular junction fragmentation and that neuromuscular junction fragmentation and muscle fiber loss are not secondary to the loss of motor neurons. Chai et al²³ found that the denervation of muscles is not due to the loss of motor neurons but rather results from the denervation of neuromuscular junctions. These findings suggest that for muscle fiber loss to occur, motor neuron loss is not essential, but neuromuscular junction instability is necessary. Baehr et al²⁴ reported that poor muscle recovery

following disuse was associated with neuromuscular junction instability in a rat hind limb unloading model. That is, prolonged disuse induces neuromuscular junction instability and delays the reinnervation process.

When denervation outpaces reinnervation, denervated fibers will degenerate, and much space will consequently be created. Beeler et al²⁵ proposed that with chronic muscle denervation, the muscle fiber atrophy and fat fills the space created by the reduction in fiber size. In the present study, fat filled the space that was created by the loss of fibers resulting from an impaired denervation–reinnervation process. Therefore, we advocate another hypothesis: the interstitial spaces created by muscle tissue loss followed by an impaired denervation–reinnervation process can lead to a crude FI-related implicitly to disuse but directly to denervation in the anterior GMin.

Conclusion

FI in the GMin in older adults restricted to deep within the anterior part is secondary to disuse and strongly related to impairment of denervation–reinnervation process.

Disclosure

The authors report no conflicts of interest in this work.

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