The role of HIF-1α in chemo-/radioresistant tumors

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Abstract: Chemo-/radioresistance is a major obstacle in clinical oncology. The precise failure mechanisms of chemo-/radioresistance are multifactorial failures. It is now widely accepted that a tumor hypoxia microenvironment contributes significantly to chemo-/radioresistance. Hypoxia is the most common and obvious neoplastic microenvironment and is due to the rapid proliferation of tumor cells. HIF-1α is a principal molecular mediator of adaptability to hypoxia in tumor cells. HIF-1α activation leads to the transcription of a plethora of target genes that promote physiological changes associated with chemo-/radioresistance, including increasing the ability of DNA repair, the inhibition of apoptosis, and alterations of the cellular metabolism. Moreover, recent findings suggest that HIF-1α-activated autophagy is a crucial factor in the promotion of cell survival under the distressed microenvironment, thereby leading to the chemo-/radioresistance of tumor cells.

Keywords: HIF-1α, chemo-/radioresistance, cancer

Introduction

In recent years, cancer has become a major threat to public health due to the increase in both morbidity and mortality of malignancies.1 Torre et al identified the mortality rates. In 2012, 14.1 million new cancer cases occurred and 8.2 million cancer deaths occurred. Breast cancer is the leading killer at 25% among females, and lung cancer is both the leading killer at 19.6% among males and 13% of all cancer deaths for both males and females.

The main methods of antitumor therapy currently are surgery, chemotherapy, radiotherapy, and targeted therapy. Normally, patients with a malignant tumor need to have the tumor removed first by surgical resection, followed by chemotherapy, and radiotherapy at different times;2 however, cancer treatments with either drugs or radiation are seriously limited by chemo-/radioresistance of tumors.3 Both Rueff and Rodrigues4 and Gatti and Zunino5 reported that chemo-/radioresistance is the primary cause for treatment failure in the majority of patients with malignant carcinoma. Thus, both how to overcome the resistance of chemo-/radiotherapy and how to improve the curative effect of cancer treatment are pressing problems to be solved.

The precise failure mechanisms of chemo-/radioresistance are multifactorial failures and can be classified broadly into the following two categories: tumor cell intrinsic resistance, which is largely due to the germinal genetic makeup according to Kartal-Yandim et al,6 and due to tumor microenvironment-related factors according to Gottesman.7 In recent years, Prasad et al identified three tumor cell intrinsic resistance mechanisms, the high expression of MDRI (multidrug resistance gene)
in tumor cells, alterations in drug metabolism, and drug transport. Moreover, Verduzco et al, Muz et al, and others now widely accept that the alteration in tumor microenvironment during chemo-/radiotherapy leads to the expression of tumor-microenvironment-related factors, which contribute significantly to chemo-/radioresistance.8–11

Hypoxia is the most common and obvious neoplastic microenvironment in solid tumors and is due to the rapid proliferation of tumor cells.12 Moreover, Verduzco et al13 and Muz et al14 reported that the hypoxia tumor microenvironment is an independent prognostic factor in a diverse range of solid human tumors; HIF-1α, a nuclear transcription factor, is a biomarker of the hypoxia microenvironment.12 Both Wu et al15 and Lixia et al16 reported that HIF-1α protein expression has been detected in most types of solid tumors and is associated with increased tumor growth, vascularization, and metastasis. In addition, HIF-1α activation has an emerging role in increasing resistance to current cancer therapies.10,11 Moreover, both Dong et al17 and Greco and Scott18 reported that many mechanisms are involved in the roles of HIF-1α during the promotion of the survival of tumor cells under chemo-/radiotherapy and chemo-/radioresistance.

Constitutive activation of HIF-1α is observed in a broad spectrum of solid tumors, and HIF-1α has an emerging role in tumorigenesis and chemo-/radioresistance. As a result, inhibiting the HIF-1α activity may represent a valuable anticancer therapeutic strategy. This review gives an overview of recent observations regarding the effects of HIF-1α on chemo-/radioresistance with special emphasis on the underlying molecular mechanisms.

**HIF-1α promotes chemo-/radioresistance in tumor cells**

A hypoxia microenvironment forms easily in the process of the rapid proliferation of cells in a tumor, and the hypoxia microenvironment is a common feature of malignant solid tumors. Both Verduzco et al13 and Muz et al14 identified that hypoxia has an emerging role in chemo-/radioresistance of solid tumors because the activity of chemotherapeutic agents decreases or even disappears in the hypoxia microenvironment. HIF-1α is a biomarker of hypoxia microenvironment. Muz et al15 identified HIF-1α as a principal molecular mediator of adaptability to hypoxia in tumor cells, and HIF-1α mediates cell proliferation, apoptosis, metabolism, immune responses, genomic instability, vascularization, invasion, and metastasis. Li et al17 and others demonstrated that HIF-1α is both highly expressed in most of the solid tumors such as in cervical cancer,18 breast cancer,19 colorectal cancer,20 and gastric cancer21 and is tightly associated with the occurrence and development of tumors. Readers interested in the detailed information on the role of HIF-1α in tumor tumorigenesis should read two recent reviews on this topic, because this article mainly focuses on the chemo-/radioresistance roles of HIF-1α.22,23

Recent findings by Li et al and others demonstrated that the high expression of HIF-1α is not only related to malignant progression17,18 but also has an important effect on the treatment outcome of tumors.24 Moreover, Zhao et al and others noted that a contribution of HIF-1α to drug resistance has been observed in a wide spectrum of clinical tumor samples such as gastric,24 pancreatic,25 and gall bladder types26 and HIF-1α expression is associated with both poor prognoses and relapses during treatment. Furthermore, Zhao et al and others noted that HIF-1α appears to be a crucial molecular target that can be exploited to improve on the current treatment of metastatic and treatment-resistant tumors of the stomach, pancreas, and gall bladder.24–26 These data suggested that HIF-1α plays important roles in treatment-resistant tumors.

The mechanisms of chemo-/radioresistance are complex and may change during different stages in tumors. According to Takasaki et al,27 Meijer et al,28 and Unruh et al,29 at least three mechanisms are involved in the roles of HIF-1α in promotion of chemo-/radioresistance: HIF-1α-mediated apoptosis, increased ability of DNA-repair, and induced alterations of cellular metabolism. Moreover, Feng et al30 suggested that HIF-1α-activated autophagy is a crucial factor in the promotion of cell survival under the distressed microenvironment, thereby leading to the therapy resistance. The overall conclusion of Takasaki et al’s,27 Meijer et al’s,28 Unruh et al’s,29 and Feng et al’s30 observations is that HIF-1α promotes chemo-/radioresistance of cancer cells and mechanisms are complex and varied. The following section outlines general molecular mechanisms that have been shown in the roles of HIF-1α in chemo-/radioresistance (Table 1).

**HIF-1α-mediated activation of DNA repair pathway**

DNA repair is a collection of processes by which a cell identifies and corrects damage to the DNA molecules that encode its genome. DNA damage is the mainstay of cancer treatment. For instance, chemo-/radiotherapy could lead to tumor cell death by inducing DNA damage. However, Wang et al31 reported that tumor cells such as glioblastoma cells could initiate DNA damage repair, thereby causing the inhibition...
of DNA damage induced by chemo-/radiotherapy. Also, Yang et al reported that the hepatocarcinoma cells exhibited higher activity of DNA damage repair pathway than normal cells and Stover et al reported that activated DNA repair pathway is a significant cause of chemo-/radioresistance in tumor cells. Thus, after exposure to chemotherapeutic drugs and radiation, many cancer cells could avoid death through the activation of DNA repair pathway.

Fortini et al reported that chemo-/radiotherapy may induce single-stranded DNA breaks (SSBs) or double-stranded DNA breaks (DSBs), which must be repaired or must start apoptosis. The repair of SSBs is carried out by major pathways: base excision repair (BER), poly-ADP-ribose polymerase (PARP-1), and both XPA and XPD. These three proteins could have an effect on the BER process, and Li et al identified that HIF-1α contributed to cisplatin resistance in lung cancer through regulation of DNA repair pathway (Table 1). Wran et al and Logsdon et al concluded that generally HIF-1α increases the ability of DNA damage repair through the regulation of DNA repair system in liver cancer, breast cancer, osteosarcoma, and pancreatic ductal adenocarcinoma cells.

Most of molecules in DNA repair pathway are regulated by HIF-1α. For example, HIF-1α mediates the overexpression of PARP-1, XPA, and XPD. These three proteins could have an effect on the BER process, and Li et al identified that BER is associated with resistance to some chemotherapeutic drugs in non-small-cell lung cancer (NSCLC) cells. In addition, Stover et al identified that the activities of ATM, DNA-PK, and H2AX in the DSBs repair pathway are also regulated by HIF-1α. Earlier, Wirchner et al suggested that an increased number of DSBs occurred in etoposide-treated HIF-1α-deficient mouse embryonic fibroblasts (MEFs). When Wirchner et al studied

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Abbreviations: DNA-PK, DNA-dependent protein kinase; RIP, receptor-interacting protein.

A large number of studies showed that HIF-1α could increase the ability of DNA damage repair through the regulation of DNA repair pathway, thus leading to the chemo-/radioresistance in tumor cells. For instance, Um et al identified that HIF-1α contributed to resistance to ionizing radiation and anticancer drug therapy through the upregulation of DNA-PK in hypoxia tumor cells. Liu et al identified that HIF-1α contributed to cisplatin resistance in lung cancer through regulation of DNA repair pathway (Table 1). Wran et al and Logsdon et al concluded that generally HIF-1α increases the ability of DNA damage repair through the regulation of DNA repair system in liver cancer, breast cancer, osteosarcoma, and pancreatic ductal adenocarcinoma cells.
the potential molecular mechanism, markedly reduced protein expression of DNA-PK was found in HIF-1α-deficient MEFs. This study demonstrated that etoposide treatment in HIF-1α-deficient MEFs both reduced the protein expression of DNA-PK and increased the susceptibility to DNA repair (Table 1). Shenoy et al.1 showed that HIF-1α enhanced DNA repair through upregulating XPA, which leads to cisplatin resistance in testicular germ cell tumors (Table 1). In a study about the mechanism of chemo-/radioresistance in hepatocellular carcinoma, Jin et al.2 demonstrated that HIF-1α inhibited the formation of both radiotherapy-induced DSBs and SSBs. Klein et al.3 (Table 1) suggested that the HIF-1α-activated DNA damage repair pathway also has an emerging role in chemo-/radioresistance in gastric cancer. In addition, Sugrue et al.4 suggested that the expressions of both DNA-PK and H2AX were positively correlated with the expression of HIF-1α in radiation-treated mouse mesenchymal stromal cells (MSCs) and showed that after knockdown of HIF-1α in MSCs, the MSC’s ability to repair DNA was impaired and that radiation-induced apoptosis in MSCs was increased. Consistent with previous outcomes, the study of Segrue et al.4 suggested that HIF-1α promoted radioresistance in MSC’s through enhancing the ability of DNA repair (Table 1). The collective research supported HIF-1α’s emerging role in DNA repair and HIF-1α’s emerging role in chemo-/radioresistance in a variety of tumor cells.

**HIF-1α-mediated alterations in cellular metabolism**

Reprogramming of energy metabolism is another hallmark of cancer. Tan et al.5 summarized that targeting metabolic pathways may increase sensitivity to either standard chemotherapy or radiotherapy. In addition, Gatenby and Gillies6 reported that the upregulation of enzymes involved in glycolysis has an emerging role in chemo-/radioresistance in various malignant tumors such as esophageal, gastric, breast, and colorectal malignant tumors. The first rate-limiting step of glucose metabolism is the transport of glucose across the plasma membrane, and GLUT1 is the transport membrane protein in this process. Using the xenograft model, Liu et al.7 demonstrated that the inhibition of GLUT1 increased cisplatin-induced lung cancer cell death (Table 1). Pyruvate dehydrogenase kinase (PDK) 3 is responsible for the conversion of pyruvate to acetyl-coenzyme A, which enters the tricarboxylic acid cycle to produce ATP. Lu et al.8 reported that knockdown of PDK3 both inhibited hypoxia-induced glycolysis and increased the sensitivity of colon cancer cell lines to chemotherapeutic agents such as cisplatin, paclitaxel, and oxaliplatin. Zhou et al.9 reported the following two observations: first, LDHA catalyzes the final three steps in the glycolytic pathway, such as the conversion of pyruvate, the reduction of nicotinamide adenine dinucleotide (NAD) to lactate, and the oxidation of NAD, and second, LDHA has a critical role in tumor maintenance. A further study by Zhou et al.10 reported that the knockdown of LDHA reduced survival under hypoxic conditions in breast cancer cell lines. Luo and Semenza11 reported the following three observations: first, PKM2 is the last rate-limiting enzyme in the glycolytic pathway, second, PKM2 is expressed predominantly in tumor cells, and third, PKM2 is important for both cancer metabolism and tumor growth. Moreover, the study suggested that the chemical inhibition of PKM2 could sensitize hypoxic tumors to radio-/chemotherapy. All these data indicated that the alterations in PKM2 metabolism and LDHA metabolism have a critical role in the therapy resistance of tumors, and targeting metabolic reprogramming represents promising novel anticancer strategies.

HIF-1α affects chemo-/radioresistance via regulation of genes related to metabolic pathways. For example, Meijer et al.12 showed that HIF-1α inhibition results in the following metabolic changes: decreased rate of glucose uptake, decreased lactate production, increased oxygen consumption, and increased production of reactive oxygen species (ROS), which could enhance the therapeutic efficacy of radiotherapy. Meijer et al.13 hypothesized that HIF-1α is also a critical regulator of many of the genes responsible for alterations in glycolysis of the tumor, which drives therapeutic resistance. For example, Meijer et al.12 observed that HIF-1α-mediated upregulation of GLUT-1 increased intracellular ATP, pyruvate, and lactate levels and, thus, induced glycolysis. Moreover, a study of Huang et al.14 reported that this metabolic shift enhanced both the production of ATP through mechanisms that are independent of the mitochondria and confers resistance to receptor-interacting protein-dependent necroptosis in colorectal cancer cells (Table 1). Kim et al.15 reported that HIF-1α has been shown to both bind to the promoter of PDK3, the most active isoform of the PDK family, and to induce PDK3 expression levels, resulting in a switch from mitochondrial respiration to glycolysis. Furthermore, Lu et al.16 reported that HIF-1α-mediated PDK3 upregulation both significantly inhibited cell apoptosis and increased resistance to either cisplatin or paclitaxel. According to previous studies, switching from mitochondrial respiration to glycolysis promotes tumor cells' survival; thus, these studies demonstrated that HIF-1α could promote chemoresistance via the upregulation of PDK3. Maiso et al.17...
recently demonstrated that HIF-1α increased the expression of LDHA and glucose uptake and that specific inhibition of LDHA and HIF-1α can restore sensitivity to therapeutic agents such as bortezomib in multiple myeloma cells (Table 1). This study confirmed that HIF-1α reduced the efficacy of chemotherapeutic drugs by increasing the expression of LDHA. Luo et al. reported that PKM2 is also a transcriptional target of HIF-1α and attaches directly with the HIF-1α subunit and proposed that the inhibition of PKM2 could be used to sensitize hypoxic tumors to radio-/chemotherapy. In addition, Mazure et al. reported recently, HIF-1α either blocked mitochondrial respiration or destroyed mitochondria via activation of PDK1, thus enhancing the cellular glycolytic metabolism and inducing cellular resistance to apoptosis. In conclusion, these findings indicated that HIF-1α-mediated alterations in cellular metabolism via regulating the activity of enzymes in the metabolic pathway, which plays a critical role in radio-/chemoresistance of tumor cells.

**HIF-1α-mediated inhibition of apoptosis in tumor cells under chemo-/radiotherapy**

In regard to therapeutic resistance, Zhao et al. reported that chemo-/radiotherapy can induce cell apoptosis, which is considered a major mechanism in chemo-/radiotherapy’s induced cell death. Thus, Zhao et al. argues both that apoptosis impairment represents a key cause of chemo-/radioresistance and that apoptosis activation relies on distinct signaling pathways, which mainly refer to the extrinsic pathway and the intrinsic pathway. Krakstad and Chekenya added that the extrinsic apoptosis pathway is activated upon ligand binding to death receptors (DR4/5, DcR2, and Fas), but the intrinsic pathway is triggered by signals such as DNA damage, oxidative stress, and growth factor deprivation, which are mainly regulated by the tissue trauma interactions by both proapoptotic and antiapoptotic proteins. Mohammad et al. proposed that both these pathways, extrinsic and intrinsic, are always highly deregulated in cancers and pathway deregulation could allow cancer cells to escape apoptosis resulting in both tumor survival and chemo-/radioresistance. These above observations confirmed that either defective apoptosis or changes in cell cycle regulation have a crucial role in chemo-/radioresistance in tumor cells.

HIF-1α’s activation can elicit both pro- and antiapoptotic effects depending on the cellular context. Takasaki et al., in regard to therapeutic resistance, demonstrated that HIF-1α both inhibited proapoptotic proteins and activated antiapoptotic proteins to inhibit the intrinsic cell death pathway. The inhibited proapoptotic proteins and activated antiapoptotic proteins promote the survival of tumor cells under the chemo-/radiotherapy. For example, Takasaki et al. reported that HIF-1α induced the expression of both c-myc and survivin, which are two of many antiapoptotic proteins. Takasaki et al. reported that both c-myc and survivin, thereby, inhibited the apoptosis of lung cancer cells. Nishimoto et al. suggested about colon cancer that HIF-1α inhibited the chemo-/radiotherapy-induced apoptosis of tumor cells through the promotion of antiapoptotic proteins (STAT3 and TCF4; Table 1). Rohwer et al. in a gastric cancer study, showed that HIF-1α-mediated suppression of p53 activation occurred in response to the chemotherapeutic agent 5-flourouracil. HIF-1α-mediated suppression of p53 provides an interesting new angle as the suppressive effect of HIF-1α on chemotherapy-induced apoptosis was dependent on a functional p53 pathway (Table 1). Zhao et al. recently, in a gastric cancer study, showed that after knockdown of HIF-1α in gastric cancer cells, both the decreased expression of survivin and the increased expression of proapoptotic proteins (notably, Bax and caspase 3/8) were found. Furthermore, Zhao et al. proposed that chemo-/radiotherapy-induced apoptosis of tumor cells was significantly increased (Table 1). In addition, in human colon cancer cells, Pei et al. reported that HIF-1α decreased proapoptotic signaling by inhibiting the extrinsic cell death pathway, which allows cells to tolerate higher levels of chemotherapeutic injury before activating cellular death pathways. For example, Pei et al. reported that HIF-1α reduced the expression of proapoptotic signaling factors, such as tumor necrosis factor-related apoptosis-inducing ligand (TRAIL). Taken together, the overall conclusion of these observations demonstrated that HIF-1α is an important mediator of chemo-/radioresistance in solid tumors through regulating the cell apoptosis and indicated the function of HIF-1α as an antagonist of apoptosis.

**HIF-1α-mediated activation of autophagy**

Besides apoptosis, the process of autophagy is increasingly recognized as an important regulator in cell death. According to both Dalby et al. and Song et al., autophagy is a metabolic process in response to both hypoxia and metabolic stress, can produce ATP to avoid necrotic or apoptotic cell death, and has an emerging role in promoting the survival of tumor cells. Both Fang et al. and Li et al. wrote that similar to apoptosis, autophagy is also highly regulated and that multiple proteins are involved in the regulation of autophagy. For example, autophagy-related protein (ATG) is an important family of proteins.
proteins involved in autophagy regulation, the activation of autophagy is induced mainly by ATG1, ULK1, ATG13, and Beclin-1, and the formation of the autophagosome is induced by some other ATG proteins such as ATG6, ATG9, and LC3. Furthermore, Paggetti et al reported that the activation of the mTOR/PI3K pair participates during the inhibition of autophagy. Sannigrahi et al and Lei et al reported that the activation of autophagy has an emerging role in inducing the therapy resistance in tumors. For example, Liu et al reported that the functional inactivation of autophagy pathways results in significantly enhanced efficacy of chemosensitivity in tumors. A large number of studies demonstrated that HIF-1α-mediated activation of autophagy is a crucial cause for the chemosensitivity in tumor cells. For example, as BCL2 inhibits autophagy through interacting with Beclin-1. Sun et al’s research suggested that HIF-1α inhibited the expression of BCL2 through upregulating the expression of miRNA210 in colon cancer cells. The miRNA210 upregulation induced the activation of autophagy resulting in radioresistance (Table 1). Recently, Wu et al wrote about the mechanism of resistance to cisplatin in lung cancer. Wu et al showed that HIF-1α activated autophagy through increasing the expression of BNIP3 and Beclin-1 in lung cancer cells. Furthermore, Wu et al reported that when HIF-1α inhibited the autophagy by using autophagy inhibitor 3-MA, the cisplatin-induced apoptosis of tumor cells was significantly increased. Wu et al’s results suggested that HIF-1α-mediated activation of autophagy can induce the resistance to cisplatin in lung cancer (Table 1). Consistent with previous outcomes, Zou et al reported that HIF-1α also increased the expression of Beclin-1 through activation of the c-Jun pathway and suggested that the activated autophagy inhibits the radiotherapy-induced ROS in lung cancer cells (Table 1). Feng et al reported that the activity of autophagy was markedly enhanced in hypoxic osteosarcoma cells and reported that both the significantly increased expression of both HIF-1α and LC3II were found and that a significant positive correlation between HIF-1α and LC3II was observed (Table 1). Moreover, Feng et al reported that HIF-1α-activated autophagy inhibited radiotherapy-produced ROS, which inhibited apoptosis of osteosarcoma cells. Jeong et al reported about the treatment of lung cancer with silver nanoparticles and drew similar conclusions. Jeong et al reported that HIF-1α-induced autophagy had a role in the removal of ROS produced by silver nanoparticles in lung cancer cells. Jeong et al reported the inhibited silver nanoparticle-induced death of tumor cells (Table 1). In addition, Lv et al showed that the overexpression of the HIF-1α activated autophagy through inhibiting mTOR/PI3K pathway in MSCs promotes survival of the MSCs under radiotherapy (Table 1). Most of the data above suggested that HIF-1α-induced autophagy may play a crucially important role in promotion of tumor cells’ survival under radiotherapy. Collectively, all these studies supported that HIF-1α activates autophagy through the regulation of a variety of autophagy-related proteins, which has an important role in chemosensitivity in tumors.

Table 1 shows that multiple mechanisms participate in HIF-1α-induced chemosensitivity in tumor cells; chemosensitivity is a crucial problem in antineoplastic therapy. Many references in this review showed that HIF-1α induced chemosensitivity, not only through the regulation of DNA repair pathway, cellular metabolism, apoptosis, and autophagy, but also through enhanced expression of MDR1, thereby promoting multiple-drug resistance in tumors. Therefore, inhibiting the expression of HIF-1α by targeted therapeutics is of great significance for improving chemosensitivity in tumors.

**Conclusion and prospects**

Despite significant advances in recent years, therapy resistance and the consequential need for both superior medicines and novel therapeutic approaches remain the main obstacles in clinical antineoplastic therapy. Studies showed that microenvironment-related factors of tumor cells are the main cause for chemosensitivity. Hypoxia is the most common and the most obvious feature in a solid tumor; HIF-1α is a biomarker of the hypoxia microenvironment in a solid tumor. HIF-1α protein expression has been detected in most types of solid tumors and is associated with increased tumor growth, vascularization, and metastasis. Furthermore, a large number studies showed that HIF-1α protein expression has been increasingly recognized as an important cause for chemosensitivity in tumors. The research about autophagy enriches the cognition of the role of HIF-1α in chemosensitivity.
As HIF-1α plays different roles in different organs and cancers under various stimuli, most of the studies demonstrated that the mechanisms of the roles of HIF-1α in chemo-/radioresistance are different upon different treatments. However, some studies about lung cancer suggested that the mechanisms of HIF-1α in the promotion of cells’ survival under the same treatment (cisplatin) are also different. This observation illustrates that the chemo-/radioresistance–HIF-1α-related network is complex and not quite clear, which require in-depth study. At least, it is now extensively accepted that HIF-1α plays a central role in chemo-/radioresistance and HIF-1α inhibition provides effective anticancer advantages that can reverse chemo-/radioresistance. Therefore, pharmacological HIF-1α inhibition is necessary for reversing chemo-/radioresistance in tumors. However, whether the inhibition of HIF-1α in tumor cells turns out to be beneficial for tumor therapy has not been reported. Future research and more tumor models in immunocompetent animals are needed for testing the inhibition of HIF-1α in clinical antitumor therapeutics.

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References


