DNA- and RNA-binding ability of oligoDapT, a nucleobase-decorated peptide, for biomedical applications

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Background: Nucleobase-bearing peptides and their interaction with DNA and RNA are an important topic in the development of therapeutic approaches. On one hand, they are highly effective for modulating the nucleic-acid-based biological processes. On the other hand, they permit to overcome some of the main factors limiting the therapeutic efficacy of natural oligonucleotides, such as their rapid degradation by nucleases.

Methods and results: This article describes the synthesis and characterization of a novel thymine-bearing nucleoamino acid based on the L-diaminopropionic acid (L-Dap) and its solid phase oligomerization to α-peptides (oligoDapT), characterized using mass spectrometry, spectroscopic techniques, and scanning electron microscopy (SEM) analysis. The interaction of the obtained nucleopeptide with DNA and RNA model systems as both single strands (dA-dT, rA-rT, and poly(rA)) and duplex structures (dA-dT, poly(rA)/poly(rU)) was investigated by means of circular dichroism (CD) and ultraviolet (UV) experiments. From the analysis of our data, a clear ability of the nucleopeptide to bind nucleic acids emerged, with oligoDapT being able to form stable complexes with both unpaired and double-stranded DNA and RNA. In particular, dramatic changes in the dA-dT and poly(rA)/poly(rU) structures were observed as a consequence of the nucleopeptide binding. CD titrations revealed that multiple peptide units bound all the examined nucleic acid targets, with T2θ/A or T2θ/A:T(U) ratios >4 in case of oligoDapT/DNA and ~2 in oligoDapT/RNA complexes.

Conclusion: Our findings seem to indicate that Dap-based nucleopeptides are interesting nucleic acid binding-tools to be further explored with the aim to efficiently modulate DNA- and RNA-based biological processes.

Keywords: nucleopeptides, nucleic acid interaction, poly(rA) binding, circular dichroism

Introduction

A large number of chimeric compounds that bear nucleobases or other types of heteroaromatic rings connected to amino acid-containing moieties were discovered in various natural sources or are the result of synthetic efforts.1-5 Among these, of great interest are nucleoamino acids or nucleobase amino acids (NBAs). The natural (S)-willardine and its analogs,6-7 for example, act as potent agonists of α-amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA) receptors or kainate receptors, whereas a large number of these synthetic monomers constitute the building blocks for the assembly of synthetic nucleobase-containing peptides (nucleopeptides), investigated for their potential applications in biomedicine and biomaterial fields.8-13

The range of applications of nucleopeptides recalls that of peptide nucleic acids (PNAs), the known DNA analogs, which are achiral, non-charged oligomers with a pseudo-peptide backbone, largely used in numerous diagnostic, antigen, and antisense strategies.14-20
The presence of both nucleobases and peptide backbone in nucleopeptides offers the possibility of binding nucleic acids or protein targets, modulating important cell regulatory processes, such as the viral reverse transcriptase inhibition, similar to other oligonucleotide (ODN) systems.

NBA-containing peptides were used to select nucleic acid-related proteins derived from Saccharomyces cerevisiae and to detect the peptide/protein interactions on T7 phages displaying proteins. Moreover, the ability of nucleopeptide to deliver ODNs into cells was recently demonstrated treating HeLa cells with NBA-containing peptides incubated in the presence of a fluorescent Cy5-labeled hairpin DNA. Fluorescence microscopy studies evidenced how nucleopeptides deliver hairpin DNA to the cytosol of live cells, while cell viability assays showed their biocompatibility even at high dosages after 72 h of treatment.

Despite all their favorable applications, one of the existing problems in the development of synthetic ODN analogs is the nonspecific interaction with DNA and RNA. A previous study reported that inserting short (R)-diethylene glycol (miniPEG) moieties in the PNA backbone can improve the specificity of the interaction. Alternatively, the insertion of chiral amino acids, possibly positively charged, into the PNA backbone provided both an improved water solubility and chirality, which has proven crucial in conferring high-sequence selectivity to artificial ODNs.

Overall, the important role that NBA-containing structures play in biomedicine, as well as the possibility to realize new DNA- and RNA-binding nucleopeptides, prompted us to realize and investigate a new nucleobase-decorated artificial ODNs, as well as the evaluation of its biomolecular recognition properties toward both single- and double-stranded DNA and RNA targets assessed by circular dichroism (CD) and ultraviolet (UV) spectroscopies.

Materials and methods

Materials

O-(7-aza-1H-Benzo triazol-1-yl)-N,N,N’,N’-tetramethyluronium hexafluorophosphate (HATU) and benzo triazol-1-yl-oxytrityl phosphonium hexafluorophosphate (PyBOP) were purchased from Novabiochem (Merck S.p.a., Milan, Italy), 9-fluorenylethoxycarbonyl (Fmoc)-L-Dap (tert-butoxycarbonyl [Boc]-OH) was purchased from Bachem (Weil am Rhein, Germany). Anhydrous N,N-dimethylformamide (DMF) was obtained from Labscan (Gliwice, Poland); piperidine from Biosolve ( Valkenswaard, The Netherlands); high-performance liquid chromatography (HPLC) solvents and acetic anhydride (AcO) from Riedel-de Haën (Seelze, Germany); polyl(rA), thimin-1-yl acetic acid (TCH, COOH), trifluoroacetic acid (TFA), 2,4,6-trimethylpyridine (TMP), and Rink-amide resin from Fluka (München, Germany); dichloromethane (DCM) and TFA (for HPLC) from Romil (Cambridge, UK); dA12 from Biomers.net GmbH (Ulm, Germany); and deuterated dimethyl sulf oxide (DMSO) from Sigma-Aldrich Co. (Milan, Italy).

1H NMR (nuclear magnetic resonance) and 13C NMR spectra were recorded at 25°C on a Varian 600 MHz (150 MHz) unit. Chemical shifts (δ) are given in parts per million (ppm), and all coupling constants (J) are given in hertz. 1H NMR chemical shifts are referenced to residual CH2SOCD3 (δ=2.49, quin). 13C NMR chemical shifts are referenced to the solvent (CD2SOCD; δ=39.5). Samples were centrifuged at 4,000 rpm on a Z 200 A centrifuge (Hermle from Del Chimica, Napoli, Italy).

Samples of both monomer and nucleopeptides underwent liquid chromatography–mass spectrometry (LC-MS) analysis (Figures S3 and S4, respectively) on an MSQ mass spectrometer (ThermoElectron, Milan, Italy) equipped with an electrospray ionization (ESI) source operating at 3 kV needle voltage and 320°C and with a complete Surveyor HPLC System, comprising an MS pump, an autosampler, and a photodiode array (PDA) detector, by using a Phenomenex (Castel Maggiore, Italy) Jupiter C18 300 Å (5 μm, 4.6 × 150 mm)
column. We performed the gradient elution at 25°C (monitoring at 260 nm) building up a gradient starting with buffer A (0.05% TFA in water) and applying buffer B (0.05% TFA in acetonitrile) with a flow rate of 0.8 mL·min⁻¹. We performed the semipreparative purifications on a Hewlett-Packard/Agilent 1100 series HPLC system (Agilent Technologies, Santa Clara, CA, USA), equipped with a PDA detector, and used a Phenomenex Jupiter C18 300 Å (10 µm, 4.6 × 250 mm) column. Gradient elution was performed at 25°C (monitoring at 260 nm) building up a gradient starting with buffer A (0.05% TFA in water) and applying buffer B (0.05% TFA in acetonitrile) with a flow rate of 0.8 mL·min⁻¹. Samples were lyophilized from water in an FD4 freeze dryer (Heto Lab Equipment, Birkerød, Denmark) for 16 h.

**Synthetic procedures**

**Synthesis of monomer 3**

Commercial Fmoc/Boc-protected 2,4-diaminopropionic acid 1 (Fmoc-L-Dap(Boc)-OH: 50 mg, 0.12 mmol; Figure 2) was treated with a 1:1 TFA/DCM solution (2 mL) at 45°C, and the mixture was stirred for 1.5 h. Afterward, the solvent was removed in vacuo, and the crude mixture was treated with cold diethyl ether. After centrifugation, a white precipitate was recovered by filtration and repeated washings with diethyl ether, and was dried in vacuo. This product was dissolved in anhydrous DMF (1 mL) treated with N,N-diisopropylethylamine (DIEA; 0.9 equiv., 19 µL, 0.11 mmol) and TMP (0.6 equiv., 9 µL, 0.07 mmol), and then reacted with TCH₂COOH (2: 2.2 equiv., 49 mg, 0.26 mmol), previously preactivated with HATU (2.0 equiv., 91 mg, 0.24 mmol) and DIEA (2.0 equiv., 42 µL, 0.24 mmol)/TMP (2.0 equiv., 32 µL, 0.24 mmol) in DMF (1 mL) for 2 min. After 1.5 h, the solvent was removed in vacuo. The crude material was treated with water, sonicated, and filtered in order to remove the aqueous solution from the white precipitate. This solid was purified by semipreparative HPLC and the desired product (3) was obtained as a pure compound in good yield (47 mg, 0.25 mmol, 81% yield): LC-ESI-MS.

**Figure 2** Schematic representation of the synthesis of the nucleobase-decorated peptides.

**Abbreviations:** Boc, tert-butoxycarbonyl; Fmoc, 9-fluorenylmethoxycarbonyl; TFA, trifluoroacetic acid; DCM, dichloromethane; HATU, O-(7-aza-1H-benzotriazol-1-yl)-N,N,N′,N′-tetramethyluronium hexafluorophosphate; DIEA, N,N-diisopropylethylamine; DMF, N,N-dimethylformamide; PyBOP, benzotriazol-1-yl-oxytripyrrolidinophosphonium hexafluorophosphate; Ac₂O, acetic anhydride; TIS, triisopropylsilane.
were dissolved in a known amount of milliQ water and quantified by UV ($T = 85^\circ C$, absorbance measured at $\lambda = 260\, nm$). The used molar extinction coefficients of 43,000 cm$^{-1}\cdot$M$^{-1}$ (4) and 51,600 cm$^{-1}\cdot$M$^{-1}$ (5) were calculated starting from that corresponding to the thymine-containing PNA monomer (ie, 8,600 cm$^{-1}\cdot$M$^{-1}$). ESI-MS characterization of oligoDapT 4 (Figure S4A): $m/z$: 1,795.59 (found), 1,795.76 (expected for [C$_{38}$H$_{59}$N$_{27}$O$_{17}$H$_2$]+); 899.72 (found), 898.85 (expected for [C$_{28}$H$_{48}$N$_{12}$O$_{15}$]+); and 600.20 (found), 599.26 (expected for [C$_{19}$H$_{50}$N$_{15}$O$_{13}$]+) and 5 (Figure S4B): $m/z$: 1,025.23 (found), 1,024.50 (expected for [C$_{28}$H$_{59}$N$_{27}$O$_{17}$]+) and 683.34 (found), 683.24 (expected for [C$_{19}$H$_{50}$N$_{15}$O$_{13}$]+).

Scanning electron microscopy (SEM) analysis

Samples for microscopic observations were obtained using compounds in solution. After slow solvent evaporation of a droplet taken from a 167 µM solution in H$_2$O, the sample of oligoDapT was coated with a conductive layer of Au–Pd and examined in Nova NanoSem 450 SEM from FEI Technologies Inc. (Hillsboro, OR, USA) at 2 and 3 kV in a high-vacuum mode. DNA (dA$_{16}$) and RNA (poly(rA))/nucleotide complex solutions were obtained from the CD titrations with oligoDapT.

CD and UV studies

CD and UV studies were carried out in analogy with other literature studies. In particular, we obtained the CD spectra in a 200–320 nm wavelength range at 5°C on a J-810 spectropolarimeter (Jasco Europe S.R.L., Cremella, Italy) equipped with a Peltier PTC-423S/15, using a Hellma (Milan, Italy) tandem quartz cell (2 × 0.4375 cm). UV spectra were recorded on a UV–Vis Jasco model V-550 spectrophotometer equipped with a Peltier ETC-505T temperature controller.

Results and discussion

Synthesis of the thymine-containing peptides

The artificial peptides were obtained following the synthetic procedure described in Figure 2.

First, we synthesized monomer 3 by reacting commercial Fmoc/Boc-protected L-Dap monomer 1 with TCH$_2$COOH (2) and characterized it by ESI-MS (Figure S3) and NMR spectroscopy (Figures S5 and S6). Subsequently, we used in a sequential manner 3 and 1 in order to synthesize in the solid phase, using Fmoc-chemistry protocol, two oligomers, ie, pentathymine 4 and hexathymine 5 (Figure 2). Fmoc UV measurements during the synthesis showed general high coupling efficiencies and, thus, overall good yields of the final nucleopeptides.

Nucleopeptide synthesis

Solid-phase synthesis was performed in analogy with other literature reports on a Rink-amide-NH$_2$ resin (0.55 mmol/g, 18 mg). After resin Fmoc cleavage by treatment with 25% piperidine in DMF over 30 min and repeated DMF washings of the resin, a mixture of Fmoc-L-Dap(Boc)-OH 1 or monomer 3 (300 µL of a 0.1 M solution in DMF, 30 µmol, 3 equiv.), PyBOP (50 µL of a 0.6 M solution in DMF, 30 µmol, 3 equiv.), and DIEA (10 µL, 60 µmol, 6 equiv.) was introduced into the reactor containing the free-NH$_2$ resin and stirred for 20 min. Then, the coupling mixture was removed from the resin, and the solid support was washed repeatedly with DMF. Unreacted amino functions were capped by treating the resin with Ac$_2$O (20%)/DIEA (5%) in DMF over 15 min. Finally, Fmoc was removed by treatment with 25% piperidine in DMF over 15 min. This procedure was repeated for the sequential insertion of L-Dap and L-DapT on the solid support. The overall yield, as estimated by UV Fmoc test, was 65% in case of the hexathymine derivative 5, whereas pentathymine nucleopeptide 4 was obtained in 89% yield.

The deprotection and detachment of the nucleopeptides from the resin were realized by treatment with TFA/triisopropylsilane/H$_2$O ($95\%$/$2.5\%$/$2.5\%$) over 2 h. After nucleopeptide precipitation with cold diethyl ether, the samples were treated with milliQ H$_2$O and purified by reversed-phase (RP)-HPLC using a gradient (10% [for 5 min] to 30%) of B’ in A’ over 30 min, collecting the peak at t$_R = 16.8$ min.

After HPLC purification, the nucleopeptide samples were dissolved in a known amount of milliQ water and
After detachment from the resin and deprotection under acidic conditions, the constructs 4 and 5 were purified by RP-HPLC and characterized by ESI-MS (Figures S4). The obtained oligomers presented peptide chains in which half of the residues carried the DNA nucleobases, while the others were base free, thus ensuring positive charges to the overall structures under physiological conditions. This led to cationic nucleopeptides, highly soluble in aqueous solution, that had both base–base and ionic (−NH₄⁺/−PO₄⁻) binding possibilities toward natural nucleic acids.

**SEM analysis**

SEM imaging of dried oligoDapT led to the micrographs shown in Figure 3.

Surface morphology of oligoDapT appears with a low roughness and some thread structures (Figure 3A). Several dense and long thread like structures, fiber-like, clearly appear (Figure 3B). At high magnifications, we found a fiber thickness ranging between 10 and 20 nm (Figure 3C) similar to that obtained by SEM, for self-assembling peptides rich in ionic and hydrophobic residues, or by TEM for aromatic peptides. In analogy with other nucleopeptide systems, here the interaction between aromatic moieties (nucleobases) has a role in the formation of the observed fibril-like structures.

Moreover, we analyzed by SEM the peptide/nucleic acid complexes to elucidate both morphology and size of the resulting structures. SEM of oligoDapT/DNA

![Figure 3 SEM micrographs of oligoDapT (A–C), oligoDapT/DNA complex (D and E), and oligoDapT/RNA complex (F). Notes: (A, B, D, E, and F) 100,000×, 1 µm; (C) 200,000×, 500 nm. Abbreviation: SEM, scanning electron microscopy.](image-url)
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(Figure 3D and E) and oligoDapT/RNA (Figure 3F) revealed the absence of thread-like structures and the formation of spherical morphologies (width 160–200 nm), in analogy with other literature reports on nucleic acid–peptide complexes or hybrids,\textsuperscript{34,35} arising from the nucleic acid/nucleopeptide binding, which induces the clustering of several nanofibers into larger aggregates.\textsuperscript{26}

**Nucleic acid-binding studies**

Afterward, we studied the interaction of oligoDapT with nucleic acids, in both single- and double-stranded forms, by means of CD and UV spectroscopies. We recorded in a two-chamber quartz cuvette the sum of CD and UV spectra relative to the separated solutions of the nucleopeptide (4 µM concentration in thymine) and complementary single-stranded nucleic acids (dA\textsubscript{12} DNA, rA\textsubscript{12} RNA, and poly(rA) RNA, all at 4 µM concentration in adenine) in 10 mM phosphate buffer (pH 7.5) at 5°C and compared them with those obtained after the abovementioned solutions were mixed. Extensive changes in DNA and RNA CD spectra upon mixing the solutions of the two-cell reservoirs reflected strong oligoDapT/DNA and oligoDapT/RNA interactions (Figures 4A, 5A, and 6A).

We have found only minor UV changes (Figure 4B) for oligoDapT/dA\textsubscript{12} mixed in a 1:1 ratio, suggesting a minor role of the base stacking in the formation of the T\textsubscript{Ldap}/dA complex. On the other hand, the analogous oligoDapT/rA\textsubscript{12} system largely involved nucleobase stacking as evidenced by the significant UV hypochromic effect (Figure 5B). An even more pronounced hypochromic effect was evidenced when poly(rA) was used instead of rA\textsubscript{12} (Figure 6B). This RNA vs DNA selectivity, previously evidenced also for other L-diamino acid-based nucleopeptides by CD and surface plasma resonance (SPR) experiments,\textsuperscript{36} can be associated with additional H-bonds with RNA structures due to ribose 2′-OH moieties (absent in DNA) that reinforce the overall ODN/nucleopeptide structuring.

By titrating the oligoDapT/DNA solution with further amounts of the nucleopeptide, we observed that the CD profiles underwent further modifications in correspondence

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**Figure 4** Nucleopeptide/DNA binding assays. Sum (black) and mixture (red) CD (A and C) and UV (B) spectra of oligoDapT (4 µM in T) and dA\textsubscript{12} DNA (4 µM in A) in 10 mM phosphate buffer (pH = 7.5; T = 5°C; optical path = 0.875 cm). (C) CD titration relative to the addition of oligoDapT (4) to dA\textsubscript{12} DNA at the following T\textsubscript{Ldap}/dA ratios: 2, 3, 4, and 5.

**Abbreviations:** CD, circular dichroism; UV, ultraviolet.
of T_{Ldap}/dA ratios higher than 2:1, which resulted in dramatic changes in both shape and intensity of the CD bands (Figure 4C). More in detail, a negative band at 274 nm and a positive one at 260 nm appeared, with the 260 nm band showing also a hypsochromic shift of its maximum upon titration (from 260 nm at T_{Ldap}/dA = 2 to 256 nm at T_{Ldap}/dA = 5). A substantial stabilization of the CD signal was observed at a 5:1 (T_{Ldap}/dA) ratio, indicating a multimeric oligoDapT/DNA complex.

Analogously, we performed a titration experiment also with rA_{12} and poly(rA); the latter is an RNA suggested as an important anticancer target.\textsuperscript{37,38} The stoichiometry of the obtained oligoDapT/RNA complexes was investigated by adding oligoDapT aliquots to the 1:1 (T_{Ldap}/rA) complex solutions until stabilization of the CD signal: for both rA_{12} and poly(rA), the formation of complexes with a 2/1 = T_{Ldap}/rA stoichiometry was evidenced (Figures 5A and 6A). However, upon titration with the nucleopeptide,
more dramatic conformational changes in the poly(rA) with respect to rA, reflected in strong CD profile changes, were observed.

To get more insight into the nature of the nucleopeptide/nucleic acid recognition, and in particular to establish the relative contribution of the electrostatic forces and sequence-specific recognition to the interaction, a binding experiment of oligoDapT with a noncomplementary poly(rU) strand was performed, and the obtained spectra were compared with those shown in Figure 6A for the poly(rA). CD experiments showed that the nucleic acid structure of the homouracil RNA underwent only negligible changes upon oligoDapT addition (Figure 6C), in contrast to the dramatic RNA structure perturbation observed for the oligoDapT/poly(rA) binding (Figure 6A). These findings indicate that the interaction is mainly driven by complementary base pairing, with the NH/phosphate ionic recognition being only of secondary importance, ie, useful to reinforce the overall interaction but not the major driving force.

The importance of new drugs able to bind double-stranded DNA, as well as to interfere with double-stranded RNA-driven biological processes or act as stabilizers and carriers of double-stranded RNA drugs prompted us to study the interaction of oligoDapT with dA/dT and poly(rA)/poly(rU), chosen as double-stranded DNA and RNA model systems, respectively. First, the target complexes were prepared by mixing equimolar amounts of DNA and RNA single strands, heating the resulting solutions at 90°C for 10 minutes and then slowly cooling overnight. Subsequently, by using a two-chamber quartz cuvette, we recorded and compared the CD spectra relative to the sum of the separated solutions of oligoDapT and double-stranded nucleic acids with those recorded after mixing the solutions and also after further oligoDapT additions. CD changes in the DNA/DNA mixture spectrum were evident when a 1/1 T/D ratio was achieved, with a stabilization of the signal only after further oligoDapT addition, corresponding to a 4/1 T/D ratio. Moreover, a bathochromic shift (4 nm) for the positive band at ~280 nm was observed in the CD titration (Figure 7).

More dramatic changes were observed studying the interaction of oligoDapT with the RNA:RNA duplex, with a significant decrease in the overall CD signal intensity and a hypsochromic shift of the positive band at 266 nm, relative to the poly(rA):poly(rU) complex, to 259 nm in the spectrum of the ternary complex oligoDapT/RNA:RNA when a 2/1 T/D ratio was reached (Figure 8). CD titration experiments suggested the formation of a complex with a 2/1 stoichiometry relative to the T/D and rA:rU bases, since a certain CD signal stabilization was obtained for this mixture.

Taken together, the abovementioned findings demonstrated the ability of oligoDapT to bind all the nucleic acid systems selected for this study. Furthermore, CD titration experiments evidenced some differences in the binding
Properties exploitable in the context of molecular devices for potential biomedical applications; indeed, this is a nucleic acid binder able to target complementary DNA and RNA of interest in antigen, antisense, and double-stranded RNA targeting antiviral strategies, as well as to be used for the delivery of therapeutic ODNs, a recently evidenced property of nucleopeptides.26

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Disclosure
The authors report no conflicts of interest in this work.

References


**Supplementary materials**

This section provides

1. a structural representation of a hexathymine oligoDapT;
2. a structural representation of the interaction between oligoDapT and nucleic acids;
3. liquid chromatography-electrospray ionization-mass spectrometry (LC-ESI-MS) characterization of the thymine-based monomer 3 and nucleopeptides 4 and 5;
4. $^1$H and $^{13}$C NMR spectra of the nucleoamino acid 3; and
5. ultraviolet (UV) melting curves relative to oligoDapT/DNA, oligoDapT/RNA, and controls (DNA/DNA and RNA/RNA).

![Figure S1](https://www.dovepress.com/)

*Figure S1* Structural representation of the hexathymine oligo(Dap).

![Figure S2](https://www.dovepress.com/)

*Figure S2* H-bonding and ionic-based interactions occurring between oligoDapT and complementary nucleic acids.
Figure S3 LC-ESI-MS (positive mode) analysis of the thymine-based monomer (3) used in this study (HPLC method on RP C18: 15%–70% of CH₃CN [0.05% TFA] in H₂O [0.05% TFA]). (A) ESI-MS spectrum of the chromatographic peak at 12 min; (B) extracted TIC chromatogram from 10.55 to 13.12 min; (C) extracted PDA chromatogram from 10.55 to 13.12 min in micro absorbance units.

Abbreviations: LC-ESI-MS, liquid chromatography-electrospray ionization-mass spectrometry; HPLC, high-performance liquid chromatography; RP, reversed phase; TFA, trifluoroacetic acid; ESI-MS, electrospray ionization-mass spectrometry; PDA, photodiode array; RT, retention time; TIC, total ion current.
Figure S4 ESI-MS (positive mode) analyses of the thymine-based nucleopeptides 4 (A) and 5 (B).

Abbreviation: ESI-MS, electrospray ionization-mass spectrometry.
Figure S5 1H NMR spectrum of the nucleoamino acid 3 recorded at 25°C in deuterated DMF on a Varian 600 MHz unit. 
Abbreviations: NMR, nuclear magnetic resonance; DMF, N,N-dimethylformamide.

Figure S6 13C NMR spectrum of the nucleoamino acid 3 recorded at 25°C in deuterated DMF on a Varian 600 MHz (150 MHz) unit. 
Abbreviations: NMR, nuclear magnetic resonance; DMF, N,N-dimethylformamide.
Figure S7 UV melting profiles for the complexes formed in 10 mM phosphate buffer (pH = 7.5) between da₁₂ (4 µM) and (A) 1 equiv. dT₁₂, (B) 3.5 equiv. oligoDapT 4, and (C) 3.5 equiv. oligoDapT 5.

Abbreviation: UV, ultraviolet.

Figure S8 Normalized (0–1) UV melting profiles for the complexes formed in 10 mM phosphate buffer (pH = 7.5) between da₁₂ (4 µM) and 1 equiv. dT₁₂ (black line), 3.5 equiv. oligoDapT 4 (blue), and 3.5 equiv. oligoDapT 5 (red).

Abbreviation: UV, ultraviolet.

Figure S9 (A) UV melting profile for the complex formed between rA₁₂ (4 µM) and 2.5 equiv. oligoDapT 4 in 10 mM phosphate buffer (pH = 7.5). (B) Normalized (0–1) UV melting profiles for the complexes oligoDapT/dA₁₂ (blue) and oligoDapT/rA₁₂ (red) formed in 10 mM phosphate buffer (pH = 7.5).

Abbreviation: UV, ultraviolet.
Figure S10 UV melting profiles for the complexes formed in 10 mM phosphate buffer (pH = 7.5) between poly(rA) (4 µM) and (A) 1 equiv. poly(rU) and (B) 3.5 equiv. oligoDapT 4.

Abbreviation: UV, ultraviolet.

Figure S11 Normalized (0–1) UV melting profiles for the complexes formed in 10 mM phosphate buffer (pH = 7.5) between poly(rA) (4 µM) and 1 equiv. poly(rU) (black line) or 3.5 equiv. oligoDapT 4 (red).

Abbreviation: UV, ultraviolet.