Inhibitory effect of the Kampo medicinal formula Yokukansan on acute stress-induced defecation in rats

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Objectives: Irritable bowel syndrome (IBS) is a functional gastrointestinal disorder with symptoms of abnormal defecation and abdominal discomfort. Psychological factors are well known to be involved in onset and exacerbation of IBS. A few studies have reported effectiveness of traditional herbal (Kampo) medicines in IBS treatment. Yokukansan (YKS) has been shown to have anti-stress and anxiolytic effects. We investigated the effect of YKS on defecation induced by stress and involvement of oxytocin (OT), a peptide hormone produced by the hypothalamus, in order to elucidate the mechanism of YKS action.

Methods and results: Male Wistar rats were divided into four groups; control, YKS (300 mg/kg PO)-treated non-stress (YKS), acute stress (Stress), and YKS (300 mg/kg PO)-treated acute stress (Stress+YKS) groups. Rats in the Stress and Stress+YKS groups were exposed to a 15-min psychological stress procedure involving novel environmental stress. Levels of plasma OT in the YKS group were significantly higher compared with those in the Control group (P < 0.05), and OT levels in the Stress+YKS group were remarkably higher than those in the other groups (P < 0.01). Next, rats were divided into four groups; Stress, Stress+YKS, Atosiban (OT receptor antagonist; 1 mg/kg IP)-treated Stress+YKS (Stress+YKS+B), and OT (0.04 mg/kg IP)-treated acute stress (Stress+OT) groups. Rats were exposed to acute stress as in the previous experiment, and defecation during the stress load was measured. Administration of YKS or OT significantly inhibited defecation; however, administration of Atosiban partially abolished the inhibitory effect of YKS. Finally, direct action of YKS on motility of isolated colon was assessed. YKS (1 mg/mL, 5 mg/mL) did not inhibit spontaneous contraction.

Conclusion: These results suggested that YKS influences stress-induced defecation and that increased OT secretion may be a mechanism underlying this phenomenon.

Keywords: Yokukansan, oxytocin, irritable bowel syndrome, acute stress, corticosterone, Kampo medicine

Introduction
Irritable Bowel Syndrome (IBS) is a disorder with an increasing number of cases in recent years. The prevalence rate in Japan is 10%–15% and the number of patients is estimated to be 12 million.1,2 IBS is a functional gastrointestinal disorder characterized by the presence of recurrent abdominal pain with abnormalities in stool frequency and form. No physical or biochemical abnormalities underlying the symptoms have been found. IBS is thought to be caused by diverse etiologies, and psychological factors are often related to its onset and deterioration.3,4 Psychological abnormalities affect intestinal motility through the central nervous system, for example, colon movement is accelerated by negative emotions such as anxiety, fear, and stress.5,6 Intestinal function
and central nervous system are known to be closely related, referred to as “brain-gut interaction”. This interaction is suggested to be influenced by the hypothalamic-pituitary-adrenal (HPA) axis, the autonomous nervous system, and the immune system.\(^6\)\(^7\)

IBS patients are categorized into subgroups including IBS with diarrhea (IBS-D), IBS with constipation (IBS-C), IBS with mixed diarrhea and constipation (IBS-M), and unclassified IBS. Drug therapy used for IBS-D primarily includes serotonin-3 (5-HT\(_3\)) receptor antagonists, anticholinergic drugs, anti-diarrheal drugs, and selective serotonin reuptake inhibitors.\(^22\) Recent studies have demonstrated that intraperitoneal administration (IP) of OT inhibited stress-induced colonic contraction in rats.\(^25\) In the current study, the effects of YKS on increased intestinal motility induced by stress, and involvement of OT in YKS action were evaluated.

Materials and methods

Animals

Male Wistar rats (7–8 weeks old), purchased from Nippon Bio-Supp. Center (Tokyo, Japan), were used. During the experimental period, the animals were housed in standard plastic cages in our animal facilities at 25°C ± 2°C with 55% ± 5% humidity under a light/dark cycle of 12 h/12 h.

Food (CLEA Japan, CE-2, Tokyo, Japan) and water were provided ad libitum. Experiments were performed as per the guidelines of the Committee of Animal Care and Welfare of Showa University. All experimental procedures were approved by the Committee of Animal Care and Welfare of Showa University (certificate number: 07061).

Drugs

Dry powdered extracts of YKS (Lot No 2110054010) used in the present study were supplied by Tsumura & Co. (Tokyo, Japan). The seven herbs comprising YKS (Table 1) were mixed and extracted with purified water at 95.1°C for 1 h, a soluble extract was separated from insoluble waste and concentrated by removing water under reduced pressure. YKS was dissolved in distilled water and orally administered. OT (Peptide Institute, Osaka, Japan) and Atosiban, an OT-receptor antagonist (Sigma-Aldrich, St Louis, MO, USA), were dissolved in saline and intraperitoneally administered.

Influence of YKS and acute stress on OT secretion

To evaluate effects of YKS and acute stress on OT secretion, 28 rats were randomly divided into four groups (seven per group) as follows: (1) Control group; (2) YKS-treated non-stress (YKS) group; (3) acute stress (Stress) group, and (4) YKS-treated acute stress (Stress+YKS) group. YKS (300 mg/kg/day) was administered daily for 3 days to the YKS and Stress+YKS groups, and water was administered orally to the Control and Stress groups. In preliminary experiments, single administration of YKS did not show inhibitory

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**Table 1 Component galenicals of YKS**

<table>
<thead>
<tr>
<th>Plant name</th>
<th>Weight</th>
<th>Major active components</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Uncaria rhynchophylla</em></td>
<td>3.0 g</td>
<td>Rhynchophylline, Geissoschizine methyl ether</td>
</tr>
<tr>
<td><em>Cnidii rhizoma</em></td>
<td>3.0 g</td>
<td>Ligustilide, Senkyunolide</td>
</tr>
<tr>
<td><em>Bupleuri radix</em></td>
<td>2.0 g</td>
<td>Saikosaponin a, c, d, e, f</td>
</tr>
<tr>
<td><em>Atractylodis Lanceae rhizoma</em></td>
<td>4.0 g</td>
<td>Asaractylodin, β-eudesmol</td>
</tr>
<tr>
<td><em>Poria</em></td>
<td>4.0 g</td>
<td>Eburicoic acid, Pachymic acid</td>
</tr>
<tr>
<td><em>Angelicae radix</em></td>
<td>3.0 g</td>
<td>Ligustilide, n-butyldiphenalide</td>
</tr>
<tr>
<td><em>Glycyrrhizae radix</em></td>
<td>1.5 g</td>
<td>Glycyrrhizin, Glycyrrhizic acid</td>
</tr>
</tbody>
</table>

**Note:** Weights show the amounts mixed.

**Abbreviation:** YKS, Yokukansan.
effects on increased defecation (data not shown), and YKS was therefore pre-administered for 3 days. YKS dose was chosen based on the results of published studies. It has been reported that OT is involved in stress responses. In our study, we investigated the influence of acute psychological stress on OT secretion. On day 4, 1 h after administration of YKS or water, rats in the Stress and the Stress+YKS groups were exposed to 15 min of an acute stress procedure, involving novel environmental stress. Each rat was transferred from group-housed cages and placed in an opaque box (60 × 60 × 60 cm³) individually for 15 min. Next, all rats were anesthetized with intraperitoneal pentobarbital sodium (50 mg/kg; Kyoritsu Seiyaku, Somnopentyl, Tokyo, Japan), and blood samples were obtained from the inferior vena cava. To avoid influences of fluctuation in routine, all blood sampling was performed between 13:00 h and 15:00 h. Blood samples were centrifuged at 4°C and 3,000 rpm for 10 min, and supernatants collected. The plasma was stored at −80°C until measurements. Acetonitrile was used for the precipitation of plasma proteins. The thawed plasma was mixed with acetonitrile (Wako Pure Chemical Industries, Osaka, Japan) and centrifuged. Subsequently, the supernatant was evaporated and reconstituted with the assay buffer. Plasma OT level was measured using an enzyme-linked immunosorbent assay (ELISA) kit (Enzo Life Sciences, Farmingdale, NY, USA), according to the manufacturer’s instructions.

Intraperitoneal administration of OT
To determine the dose of OT to be used in subsequent experiments, OT was intraperitoneally administered to rats and plasma OT concentrations were measured. Doses of OT were 0.02 mg/kg (OT0.02; n = 6), 0.04 mg/kg (OT0.04; n = 6), and 0.20 mg/kg (OT0.20; n = 6). These doses were determined by referring to a previous study. It was reported that intraperitoneal administration of OT (500 pmol per rat) inhibited water-avoidance stress-induced colonic contraction in rats. Control rats (n = 6) were intraperitoneally administered with saline. One hour after administration, plasma OT concentrations were measured using the method described above.

Acute stress-induced defecation
It is known that defecation in rats is increased by an acute stress load, and this model is used for studies on IBS-D. Thirty-six rats were randomly divided into four groups (nine per group) as follows: (1) acute stress (Stress); (2) YKS-treated acute stress (Stress+YKS); (3) Atosiban-treated acute stress (Stress+YKS+B); and (4) OT-treated acute stress (Stress+OT). YKS (300 mg/kg/day) or water was orally administered daily for 3 days. On day 4, rats in the Stress+OT group were administered OT (0.04 mg/kg, IP) 1 h before the stress load, and those in the Stress+YKS+B group were administered Atosiban (1 mg/kg, IP) 10 min before YKS administration. In the Stress and Stress+YKS groups, saline was administered instead of Atosiban or OT (Table 2). One hour after administration of YKS or water, rats were exposed to acute stress, and defecation during the stress load was measured using a balance. Atosiban dose was determined based on a previous study.

Influence of YKS and acute stress on corticosterone secretion
Levels of plasma corticosterone, which reflects activation of the HPA axis, were measured. Twenty-four rats were randomly divided into four groups (six per group) as follows: (1) Control; (2) acute stress (Stress); (3) YKS-treated acute stress (Stress+YKS); and (4) OT-treated acute stress (Stress+OT). Drug administration, stress load, and blood sampling were conducted as described above. Corticosterone levels were measured using an ELISA kit (Enzo Life Sciences).

Recording of colon contraction
We investigated whether the inhibitory effect of YKS on increased defecation was mediated via direct action on the bowel. Segments of the distal colon were isolated from five intact rats and were longitudinally cut into 15 × 5 mm² strips, and then transferred to a 20-mL organ bath. The organ baths contained aerated (5% CO₂, 95% O₂) Kreb’s solution (composed of NaCl 6.9 g/L, KCl 0.36 g/L, KH₂PO₄ 0.16 g/L, O₂).
MgSO$_4$ 0.14 g/L, CaCl$_2$ 0.28 g/L, NaHCO$_3$ 2.1 g/L, and glucose 1.8 g/L) maintained at 37°C. The colon segments were suspended in Kreb’s solution with an initial tension of 2.0 g for recording. Then, YKS dissolved in Kreb’s solution was perfused into the bath at 5 mL/min after stable colonic contractions were established. The YKS concentrations used were 1 mg/mL and 5 mg/mL based on previous reports. Colon contractions were recorded with a force displacement transducer (FD Pickup TB-611T; Nihon Koden, Tokyo, Japan) and a PowerLab data acquisition system (ADI Instruments, Dunedin, New Zealand).

Statistical analysis
All experimental data are presented as mean ± standard error of mean (SEM). Statistical significance of differences between groups was evaluated using paired t-test for comparisons between two groups, and one-way analysis of variance (ANOVA) for comparisons among >2 groups. Post hoc comparisons between the four groups were performed using the Tukey’s post hoc test. All P-values < 0.05 were considered to be statistically significant.

Results
Influence of YKS and acute stress on OT secretion
The influence of YKS and acute psychological stress on OT secretion was investigated. Administration of 300 mg/kg/day YKS significantly increased plasma OT levels compared with those in the control group (Control, 14.47 ± 0.96 pg/mL; YKS, 19.43 ± 0.86 pg/mL; P < 0.05). No significant difference was observed between the Control and Stress groups (Stress, 15.74 ± 0.83 pg/mL); however, OT level in the Stress+YKS group was markedly increased compared with the other groups (Stress+YKS, 27.47 ± 1.47 pg/mL; P < 0.01) (Figure 1).

Intraperitoneal administration of OT
Plasma OT concentrations were measured 1 h after intraperitoneal administration of OT (0, 0.02, 0.04, and 0.20 mg/kg). The results are shown in Table 3. The plasma OT level in the OT (0.04) group was almost identical to that in the Stress+YKS group (27.47 ± 1.47 pg/mL; Figure 1); therefore, OT dose in subsequent experiments was set to 0.04 mg/kg.

Acute stress-induced defecation
Defecation during the 15-min stress load was measured by weighing. Acute stress load-induced defecation (1.31 ± 0.23 g), and defecation in the absence of stress were significantly suppressed by pre-administrations of YKS (0.14 ± 0.09 g) and OT (0.18 ± 0.11 g) (P < 0.01). However, administration of Atosiban decreased the effect of YKS (0.71 ± 0.23 g) (Figure 2).

Influence of YKS and acute stress on corticosterone secretion
Influence of YKS and acute psychological stress on plasma corticosterone levels were also investigated. Plasma corticosterone level was significantly increased in the Stress and Stress-OT groups compared with that in the Control group (Control, 71.08 ± 17.43 ng/mL; Stress, 192.61 ± 14.72 ng/mL; Stress+OT, 210.33 ± 32.40 ng/mL; P < 0.01); however, there was no significant difference between the Control and Stress+YKS groups (128.51 ± 17.06 ng/mL) (P = 0.266) (Figure 3).

Influence of YKS on isolated colon contraction
Spontaneous muscle contractions vary in their tension and frequency among muscle preparations. Thus, baseline recordings were obtained as controls during 10 min prior to administration of YKS and compared with a 10-min period

Table 3 Plasma OT concentrations

<table>
<thead>
<tr>
<th>(pg/mL)</th>
<th>Control (n = 6)</th>
<th>OT (0.02) (n = 6)</th>
<th>OT (0.04) (n = 6)</th>
<th>OT (0.20) (n = 6)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Average</td>
<td>15.36</td>
<td>15.13</td>
<td>27.70</td>
<td>270.45</td>
</tr>
<tr>
<td>SEM</td>
<td>1.58</td>
<td>3.41</td>
<td>9.12</td>
<td>10.25</td>
</tr>
</tbody>
</table>

Note: Plasma OT concentrations (pg/mL) following the intraperitoneal administration of OT (0, 0.02, 0.04 and 0.20 mg/kg).

Abbreviations: OT, oxytocin; SEM, standard error of mean.
after YKS administration to assess response. Representative contraction waves are shown in Figure 4A (YKS 1 mg/mL) and 4B (YKS 5 mg/mL). Contractile amplitude (g) was obtained by integrating area under the contractile wave above baseline. The amplitudes before and after administration of 1 mg/kg YKS did not show a significant difference (before, 49.15 ± 3.23 g; after, 54.63 ± 6.18 g) (Figure 4C). The value after administration of 5 mg/kg YKS was higher compared with that before administration (before, 50.85 ± 5.04 g; after, 76.57 ± 12.32 g) (P = 0.052) (Figure 4D). Overall, YKS did not appear to directly inhibit colonic motility.

**Discussion**

This study addressed the use of Kampo medicines as potential treatment for patients with IBS-D, by evaluating effectiveness and mechanism of YKS in a rat model with increased defecation induced by stress.25

It has been reported that YKS shows 5-HT_{1A} receptor agonist action, 18 5-HT_{2A} receptor downregulation, 40 inhibition of glutamate secretion, 41 enhancement of glutamate clearance from extracellular fluid by astrocytes, 42 and N-methyl-D-aspartate (NMDA) receptor antagonism. 43 However, no reports concerning its action on OT secretion are present. Therefore, in the present study, we examined the influence of YKS on OT secretion and whether peripheral OT is involved in the control of defecation induced by acute stress.

First, influences of pre-administration of YKS and acute psychological stress on OT secretion in rats were investigated. Administration of YKS promoted peripheral secretion of OT, which was further increased under the 15-min novel environmental stress conditions, although OT level did not change under the stress alone (Figure 1). Previous studies showed that some acute stressors, 44 45 such as immobilization, ether exposure, and forced swimming, increase the plasma OT levels but not after novel stress load. 44 46

The level was unchanged in the present study as well. The trends in the secretion of OT seem to be different depending on the types of stressors. These results suggested that administration of YKS might enhance sensitivity of OT neurons to stress. Zheng et al 44 reported that adaptation to stress may involve upregulation of OT expression in the hypothalamus, which in turn attenuates CRF expression. YKS is thus expected to enhance resistance against stress.

Next, the inhibitory effect of YKS on defecation induced by stress was examined, and the amount of feces in the Stress+YKS group significantly decreased compared with that in the Stress group (Figure 2). Mizoguchi et al 47 also reported that pre-administration of YKS in aged rats for 3 months inhibited the anxiety-related defecation under a novel environment and that the increases of serotonin and dopamine in the prefrontal cortex were also involved. Therefore, we investigated whether this action is a result of OT secretion. Atosiban, an OT receptor antagonist, was intraperitoneally administered before stress load. The result showed that the inhibitory effect of YKS was partially obstructed by Atosiban. Moreover, OT was intraperitoneally injected to result in a plasma OT level similar to that due to YKS administration (Figure 1, Table 2), and defecation was significantly decreased similar to that caused by YKS. Intraperitoneal injection of OT and Atosiban is not expected to result in brain entry due to the blood–brain barrier. 48 49 Plasma corticosterone concentration, which reflects activation of the HPA axis, was also measured in this study. The level

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**Figure 2** Amount of defecation (g) following the acute stress load.

**Notes:** Data are presented as mean ± SEM. Statistical analysis: one-way ANOVA followed by Tukey’s post hoc test. **P < 0.01 (vs Stress group).

**Abbreviations:** ANOVA, analysis of variance; B, blocker of oxytocin receptor; OT, oxytocin; SEM, standard error of mean; YKS, Yokukansan.

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**Figure 3** Plasma corticosterone (ng/mL) levels following administration of YKS and the acute stress procedure.

**Notes:** Data are presented as mean ± SEM. Statistical analysis: one-way ANOVA followed by Tukey’s post hoc test. **P < 0.01 (vs Control group).

**Abbreviations:** ANOVA, analysis of variance; OT, oxytocin; SEM, standard error of mean; YKS, Yokukansan.
was significantly increased by the stress load; however, this significant increase was prevented by pre-administration of YKS (ns, comparing Stress+YKS and Control groups), but not OT. Central administration of OT inhibits the release of CRF, adrenocorticotropic hormone (ACTH), and corticosterone,24,25 also suggesting that OT was not delivered into the brain. Finally, the lack of a direct inhibitory effect of YKS on colonic motility was confirmed with studies on isolated colons (Figure 4). As described above, OT has a regulatory effect on bowel movement, acting on the OT receptor in the gastrointestinal tract.25,27–30 Based on the above results, YKS may control stress-induced defecation at least partially via peripheral OT secretion.

Paraventricular nuclei in the hypothalamus also deliver OT to several forebrain nuclei. YKS administration has a high probability of increasing OT secretion in the brain because the peripheral OT level increased. Intraventricular administration of OT has been shown to inhibit accelerated colonic motility induced by acute stress,25 highlighting the need for more detailed studies to clarify the central mechanism of YKS.

Almost all OT neurons express both 5-HT1A and 5-HT2A receptors,56 and administration of 5-HT1A receptor agonists increases secretion of OT. However, administration of a 5-HT2A receptor agonist was shown to induce functional desensitization of 5-HT1A receptors, and pre-administration of a 5-HT2A receptor antagonist prevented desensitization of 5-HT1A receptors.51 As mentioned above, YKS has been shown to have 5-HT1A receptor agonist action18 and a 5-HT2A receptor downregulation effect.53 In this study, YKS was pre-administered for 3 days before the stress load because single administration was not effective in preliminary experiments. Antagonist action on 5-HT2A receptors for 3 days might enhance sensitivity of 5-HT1A receptors in OT neurons. In the neuropharmacological studies of YKS, geissoschizine methyl ether, an alkaloid synthesized by Uncariae cum Uncis ramulus, has been identified as an active compound and to have a partial 5-HT1A receptor agonistic action52 and 5-HT2A receptor antagonistic action.53 Accordingly, the actions of Uncariae cum Uncis ramulus via the 5-HT1A and 5-HT2A receptors may represent one of the mechanisms via which YKS affects OT neurons. Therefore, future studies are required to identify crude medicinals that regulate OT secretion as well as to identify their mechanisms. This is the first report to show that YKS increases plasma OT level and the possibility of using YKS to treat increased defecation induced by stress, similar to symptoms of abnormal defecation in IBS-D. OT is also reported to downregulate mesenteric afferent sensitivity through the nNOS-NO-KATP pathway,54 and therefore action of YKS via OT is also expected to play an analgesic role in modulating the decline of the pain threshold in IBS.

Conclusion

Our findings suggest that administration of YKS enhances sensitivity of OT neuron to stress, elevates secretion of OT, and controls stress-induced defecation. YKS is expected to be useful in treating abnormal defecation induced by stress.
Acknowledgments

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Author contributions

All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; gave final approval of the version to be published; and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

References