Antifungal susceptibility testing of vulvovaginal Candida species among women attending antenatal clinic in tertiary care hospitals of Peshawar

Maria Khan1
Jawad Ahmed2
Amina Gul3
Aamer Ikram1
Farida Khurram Lalani1

1Department of Microbiology, Armed Forces Institute of Pathology, National University of Management Sciences, CMH Rawalpindi, Rawalpindi, Pakistan; 1Department of Microbiology, Institute of Basic Medical Sciences, Khyber Medical University, Peshawar, Pakistan; 1Department of Microbiology, Khyber Medical College, Khyber Medical University, Peshawar, Pakistan

Background: Vulvovaginal candidiasis (VVC) is considered as a pervasive gynecological problem among women worldwide. Owing to this fact, in the current study, we aimed at assessing the prevalence rate of Candida spp. causing VVC in symptomatic pregnant women and their antifungal susceptibility pattern.

Methods: This study was carried out in the tertiary care hospitals of Peshawar during the period of July 1, 2016 to December 31, 2016. The study group included 450 pregnant women in the age group of 17–44 years with symptoms of excessive vaginal discharge, pain and pruritis. In all, 108 pregnant women were culture positive for Candida. Antimicrobial susceptibility testing (AST) was conducted on specimens against various azoles and polyene F group of antifungals.

Results: Out of 108 Candida spp. isolated from vaginal swabs, there were 45 (41.7%) Candida albicans, 18 (16.7%) Candida tropicalis, 18 (16.7%) Candida krusei, 16 (14.8%) Candida glabrata and 11 (10.2%) Candida dubliniensis. According to age distribution, 27 years was the mean age. Pregnancy trimester distribution among patients was as follows: 21 (19.4%) patients were in their first trimester, 65 (60.2%) patients were in their second trimester and 22 (20.4%) patients were in the third trimester. Susceptibility of fluconazole was determined as follows: 33.3% of the Candida isolates were sensitive, 4.6% were susceptible dose dependent (SDD) and 62% were resistant. Susceptibility of Candida spp. with respect to nystatin in patients with VVC was as follows: 25% were sensitive, 16.7% were SDD and 58.3% were resistant. Susceptibility of clotrimazole was analyzed, and it was sensitive in 21.3% of patients, SDD in 19.4% of patients and resistant in 59.3% of patients. Voriconazole susceptibility was recorded to be sensitive in 85.2% of patients, SDD in 4.6% of patients and resistant in 10.2% of patients suffering from VVC. Susceptibility results for itraconazole in patients with VVC were as follows: 42.6% of patients were sensitive, 16.7% of patients were SDD, and 40.7% of patients were resistant.

Conclusion: In this study, frequency of VVC was noted to be high in the second trimester of pregnancy, with the highest frequency of C. albicans isolated, followed by C. tropicalis and C. krusei. Antifungal susceptibility testing revealed that fluconazole was exceedingly resistant against Candida species (62%), followed by clotrimazole (59.3%) and nystatin (58.3%). On the contrary, voriconazole had the highest antimicrobial activity against Candida species (85.2%).

Keywords: vulvovaginal candidiasis, fluconazole, voriconazole, itraconazole, ketoconazole, nystatin

Plain language summary

Problem: Resistance to antifungal agents has represented a major challenge for the clinic and a major public health problem, especially if the patient has been treated previously with antifungal agents.
What is already known? Vulvovaginal candidiasis (VVC) is the most frequent gynecological ruling among women of childbearing age. Among sexually active women, 75% have at least once experienced symptomatic VVC, which usually presents as soreness, burning, itching and abnormal curd-like vaginal discharge.

What this paper adds? With multiple antifungals and varying susceptibility patterns of Candida, the study was conducted to determine the frequency of common Candida species among women with VVC and also the antifungal susceptibility pattern of identified Candida species causing VVC and make information accessible to the clinicians for effectual therapeutic outcome.

Introduction
Globally amid the fungal infections in human beings, candidal infections are predominantly reported. More than 200 species of Candida have been reported.1 In healthy human beings, Candida is considered as a commensal, and its scope to produce either superficial or systemic infections depends on the host immune system and various risk factors.2 Vulvovaginal candidiasis (VVC) is the most frequent gynecological ruling among women of childbearing age.3 Among sexually active women, 75% have at least once experienced symptomatic VVC, which usually presents as soreness, burning, itching and abnormal curd-like vaginal discharge.4 In pregnant women, VVC complications include abortion, chorioamnionitis and subsequent preterm delivery.5 Transmission of Candida can take place from the infected mother’s vagina to the newborn, leading to Candida congenital infection.6 It is perceived that Candida albicans presented for VVC cases of 70–90%, with a recent surfacing of non-albicans species.7

The improvement of standardized antimicrobial susceptibility testing (AST) has been the area under discussion of several studies in the past few decades. Reference scheme for yeast susceptibility testing (National Committee for Clinical Laboratory Standards [NCCLS]) is available. Agar-based susceptibility testing procedure has been a focus of interest for many researchers and comprises the classical disk diffusion method along with novel E-test method.8,9 Antifungal susceptibility testing may specify clinical response, predict treatment failure and develop local antibiograms, aiding in empirical selection of antifungals.10 Resistance to antifungal agents has represented a major challenge in public health problems, particularly if the patient has been in the past treated with an azole group of antifungal agent; the likelihood of microbiological resistance should likely be considered. With a diverse range of antifungals use and anecdotal susceptibility patterns of Candida, it has at the present turn out to be indispensable to perform AST and make information accessible to the clinicians for effectual therapeutic outcome.

Methods
The study was conducted at antenatal units of tertiary care hospitals in Peshawar. Samples were processed at the Department of Microbiology, Institute of Basic Medical Sciences (IBMS), Khyber Medical University, Peshawar, from July 2016 to December 2016. Permission from the institutional ethical committee of Khyber Medical University was obtained before commencement of the study. An informed consent form was obtained from the participants after the contents of the form were clearly explained. Furthermore, they were assessed on the basis of age, pregnancy trimester and signs/symptoms during the gynecological examination. The participants younger than 18 years were able to sign informed consent on their own behalf, and this was acceptable to the institutional ethical committee of Khyber Medical University.

Both symptomatic and non-symptomatic pregnant women’s samples that were colonized or presenting with VVC, i.e., pruritus, leukorrhea, edema and vulvovaginal erythema presenting with whitish plaques on the mucosa were used for mycological cultivation. Samples were collected upon inserting a sterile vaginal speculum into the vagina; two high vaginal swabs, one after the other, were taken by a sterile cotton wool swab into the posterior vaginal fornix and rotated gently. The swabs were then inserted into its outer casing and were labeled with the patient’s case number, name and date. One of the swabs was used for direct smear examination, and the second one was inoculated on Sabouraud dextrose agar (SDA) and incubated aerobically at 37°C.

Examination of high vaginal swabs was done by the wet preparation method, i.e., 10% potassium hydroxide (KOH) preparation and Gram staining. Sterile normal saline drops were added to one of the tubes and shaken to extricate materials from the swab. Then, a wet film was prepared by putting a drop of the saline deposit onto a clean glass slide, which was then covered with a cover slip and examined under the microscope for budding yeast cells and pseudohyphae using 40× objective lens. Thereafter, a sterile inoculating loop was used to transfer small amount of the deposit onto a clean glass slide to form a smear. The smear was air-dried, and then, Gram staining was performed. The slide was then examined under the microscope using the oil immersion objective lens for yeast cells.
For phenotypic identification, the second swab was streaked onto culture plates of SDA (Oxoid, Basingstoke, UK) and Sabouraud dextrose agar with chloramphenicol (SC; Oxoid), which was then incubated for 48 hours at 37°C. The isolated pure colonies were confirmed on Gram staining for yeast cells.

*Candida* spp. was isolated on CHROMagar Candida (Oxoid) which is a differential and selective medium. Isolated *Candida* colonies on SDA were subcultured onto CHROMagar using an inoculating loop and incubated at 37°C for 24 hours. Presumptive identification was done based on colony color of the growing *Candida* strains. Identification of *Candida* was based on the color of individual colony. API 20C AUX (bioMérieux, Marcy-l’Étoile, France) was further used for species identification and confirmation, which is based on 19 carbohydrate assimilation tests plus a negative control, read by assessing cupules for turbidity. The kit was used in accordance with the guidelines given by the manufacturer. Reading of the strips was done after 48 and 72 hours of incubation at 30°C. Using this method, the following *Candida* species were identified: *C. albicans* (green colonies), *Candida krusei* (pink colonies) *Candida glabrata* (purple colonies), *Candida tropicalis* (blue colonies) and *C. dublinensis* (darker green colonies).

**Antifungal susceptibility testing**

A suspension was prepared by picking five to six colonies from the SDA culture plate of ~1 mm diameter from a 24-hour old culture of *Candida* species. Colonies were then inoculated in 5 mL of sterile saline, and its turbidity was adjusted to 0.5 McFarland standards visually. A sterile cotton wool swab was moistened in the adjusted inoculum suspension, and then, excess fluid was rinsed by rolling the swab on the inside surface of the tube above the fluid surface. Müller-Hinton agar (MHA) surface was streaked to make a lawn of the isolate.

Antifungal susceptibility testing was undertaken by the disk diffusion method. Using disk dispenser (Oxoid™), fluconazole disk (10 μg), itraconazole (10 μg), voriconazole (10 μg), clotrimazole (10 μg) and nystatin (100 IU) antifungal discs (Thermo Scientific™ Oxoid™) were applied on MHA (Thermo Scientific™ Oxoid™) as recommended by the Clinical Laboratory Standard Institute (CLSI) M44A document.

The plates were incubated in ambient air at 35°C and read at 24 hours. The diameters of zones of inhibition were measured in millimeters using a ruler for each antifungal disk. Interpretation of all antifungal susceptibility (susceptible S, susceptible dose dependent [SDD], and resistant R) was done according to CLSI standards (Table 1). Quality control was undertaken by using quality control strains, American Type Culture Collection (ATCC) 90028.

### Results

A total of 108 patients were observed to determine frequency of in vitro susceptibility of *Candida* spp., and the results were analyzed.

Age distribution among 108 patients was analyzed as follows: 11 (10.1%) patients were in age range 17–20 years, 28 (25.9%) patients were in age range 21–24 years, 31 (28.7%) patients were in age range 25–28 years, 20 (18.5%) patients were in age range 29–32 years, 12 (11.1%) patients were in age range 33–36 years, three (2.7%) patients were in age range 37–40 years and three (2.7%) patients were in age range 41–44 years. The minimum age calculated was 17 years, and the maximum age was 44 years. Mean age was 27 years with standard deviation (SD) ±5.61012 (Figure 1).

Pregnancy trimester distribution among patients was analyzed as follows: 21 (19.4%) patients were in their first trimester, 65 (60.2%) patients were in their second trimester and 22 (20.4%) patients were in their third trimester (Figure 2).

Frequency of *Candida* spp. distribution among patients with VVC was analyzed as follows: 45 (41.7%) patients had isolated *C. albicans*, 18 (16.7%) patients had *C. tropicalis*, 18 (16.7%) patients had *C. krusei*, 16 (14.8%) patients had *C. glabrata* and 11 (10.2%) patients had isolated *C. dublinensis*. In all, 65% of the patients had a past history of antifungal use.

### Table 1 Interpretative breakpoints of antifungal agents

<table>
<thead>
<tr>
<th>Antifungals</th>
<th>Disk concentration</th>
<th>Zone of activity (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Sensitive</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Nystatin</td>
<td>100 U</td>
<td>≥15</td>
</tr>
<tr>
<td>Clotrimazole</td>
<td>10 μg</td>
<td>≥20</td>
</tr>
<tr>
<td>Fluconazole</td>
<td>10 μg</td>
<td>≥19</td>
</tr>
<tr>
<td>Voriconazole</td>
<td>10 μg</td>
<td>≥19</td>
</tr>
<tr>
<td>Itraconazole</td>
<td>10 μg</td>
<td>≥15</td>
</tr>
</tbody>
</table>

Abbreviation: SDD, susceptible dose dependent.
Susceptibility of Candida spp. with respect to nystatin was analyzed as follows: nystatin was sensitive in 27 (25%) of Candida isolates, SDD in 18 (16.7%) of Candida isolates and resistant in 63 (58.3%) of Candida isolates causing VVC (Table 2). Mean zone of inhibition for nystatin disk was 10 mm with SD ±4.58981.

Susceptibility of Candida spp. with respect to clotrimazole was analyzed as follows: clotrimazole was sensitive in 23 (21.3%) of Candida isolates, SDD in 18 (16.7%) of Candida isolates and resistant in 64 (59.3%) of Candida isolates in patients with VVC (Table 2). Mean zone of inhibition for clotrimazole disk was 12 mm with SD ±6.56882.

Susceptibility of Candida spp. with respect to voriconazole was analyzed as follows: voriconazole was sensitive in 92 (85.2%) of Candida isolates, SDD in five (4.6%) of Candida isolates and resistant in 11 (10.2%) of Candida isolates (Table 2). Mean zone of inhibition for voriconazole disk was 21 mm with SD ±4.23074.

Susceptibility of Candida spp. with respect to itraconazole was analyzed as follows: itraconazole was sensitive in 46 (42.6%) of Candida isolates, SDD in 18 (16.7%) of Candida isolates and resistant in 44 (40.7%) of Candida isolates (Table 2). Mean zone of inhibition for itraconazole disk was 13 mm with SD ±5.51711.

Susceptibility of 45 C. albicans isolates with respect to each of the antifungal was analyzed as follows: fluconazole was sensitive in 17 (37.7%) of Candida isolates, intermediate isolates and resistant in 64 (59.3%) of Candida isolates in patients with VVC (Table 2). Mean zone of inhibition for clotrimazole disk was 12 mm with SD ±6.56882.

Table 2 Antifungal susceptibility pattern of isolated Candida spp.

<table>
<thead>
<tr>
<th>Drugs</th>
<th>Sensitive (%)</th>
<th>SDD (%)</th>
<th>Resistant (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fluconazole</td>
<td>33.3</td>
<td>4.6</td>
<td>62</td>
</tr>
<tr>
<td>Nystatin</td>
<td>25</td>
<td>16.7</td>
<td>58.3</td>
</tr>
<tr>
<td>Clotrimazole</td>
<td>21.3</td>
<td>19.4</td>
<td>59.3</td>
</tr>
<tr>
<td>Voriconazole</td>
<td>85.2</td>
<td>4.6</td>
<td>10.2</td>
</tr>
<tr>
<td>Itraconazole</td>
<td>42.6</td>
<td>16.7</td>
<td>40.7</td>
</tr>
</tbody>
</table>

Abbreviation: SDD, susceptible dose dependent.
in four (8.8%) of Candida isolates and resistant in 24 (53.3%) of C. albicans isolates. Nystatin was sensitive in 12 (26.6%) of C. albicans isolates, intermediate in eight (17.7%) of C. albicans isolates and resistant in 25 (55.5%) of C. albicans isolates. Clotrimazole was sensitive in eight (17.7%) of the isolated C. albicans, SDD in 10 (22.2%) of the isolated C. albicans and resistant in 27 (60%) of the isolated C. albicans. Voriconazole was sensitive in 39 (86.6%) of the isolated C. albicans, SDD in three (6.6%) of the isolated C. albicans and resistant in three (6.6%) of the isolated C. albicans. Itraconazole was sensitive in 11 (61.1%) of the isolated C. albicans, SDD in two (11.1%) of the isolated C. albicans. Clotrimazole was sensitive in 11 (61.1%) of the isolated C. albicans and resistant in seven (38.8%) of the isolated C. albicans (Figure 3).

Susceptibility of 18 C. tropicalis isolates with respect to each of the antifungal was analyzed as follows: fluconazole was sensitive in 11 (61.1%) of Candida isolates, SDD in none of the isolated C. tropicalis and resistant in seven (38.8%) of the isolated C. tropicalis. Nystatin was sensitive in seven (38.8%) of the isolated C. tropicalis, SDD in four (22.2%) of the isolated C. tropicalis and resistant in seven (38.8%) of the isolated C. tropicalis. Voriconazole was sensitive in five (27.7%) of the isolated C. tropicalis, SDD in six (33.3%) of the isolated C. tropicalis and resistant in seven (38.8%) of the isolated C. tropicalis. Itraconazole was sensitive in 15 (83.3%) of the isolated C. tropicalis, SDD in two (11.1%) of the isolated C. tropicalis and resistant in one (5.5%) of the isolated C. tropicalis. Clotrimazole was sensitive in 11 (61.1%) of the isolated C. tropicalis, SDD in three (16.6%) of the isolated C. tropicalis and resistant in four (22.2%) of the isolated C. tropicalis (Figure 4).

Susceptibility of 18 C. krusei isolated with respect to each of the antifungal was analyzed as follows: fluconazole was sensitive in none of the patients, SDD in none of the patients and resistant in 18 (100%) of the patients isolated with C. krusei. Nystatin was sensitive in five (27.7%) of the isolated C. krusei, SDD in four (22.2%) of the isolated C. krusei and resistant in nine (50%) of the isolated C. krusei. Voriconazole was sensitive in five (27.7%) of the isolated C. krusei, SDD in three (16.7%) of the isolated C. krusei and resistant in 10 (55.5%) of the isolated C. krusei. Itraconazole was sensitive in 16 (88.8%) of the isolated C. krusei, SDD in none of the isolated C. krusei and resistant in two (11.1%) of the isolated C. krusei. Itraconazole was sensitive in five (27.7%) of the isolated C. krusei, SDD in one (5.5%) of the isolated C. krusei and resistant in 12 (66.6%) of the isolated C. krusei (Figure 5).

Susceptibility of 16 isolates of C. glabrata with respect to each of the antifungal was analyzed as follows: fluconazole was sensitive in six (37.5%) of the Candida isolates, SDD in none of the Candida isolates and resistant in 10 (62.5%) of the Candida isolates isolated with C. glabrata. Nystatin was sensitive in three (18.7%) of the isolated C. glabrata, SDD in two (12.5%) of the isolated C. glabrata and resistant in 11 (68.7%) of the isolated C. glabrata. Clotrimazole was sensitive in five (31.25%) of the isolated C. glabrata, SDD in one (6.25%) of the isolated C. glabrata and resistant in 10 (62.5%) of the isolated C. glabrata. Voriconazole was sensitive in 13 (81.25%) of the isolated C. glabrata, SDD in none of the isolated C. glabrata and resistant in three (18.7%) of the isolated C. glabrata. Itraconazole was sensitive in four

Figure 3 Antifungal susceptibility pattern in Candida albicans.
Abbreviation: SDD, susceptible dose dependent.
(25%) of the isolated C. glabrata, SDD in five (31.25%) of the isolated C. glabrata and resistant in seven (43.7%) of the isolated C. glabrata (Figure 6).

Susceptibility of 11 isolates of C. dubliniensis with respect to each of the antifungal was analyzed as follows: fluconazole was sensitive in two (18.2%) of the patients, SDD in one (9.1%) of the patient and resistant in eight (72.7%) of the patients isolated with C. dubliniensis. Nystatin was sensitive in none of the isolated C. dubliniensis, SDD in none of the isolated C. dubliniensis and resistant in 11 (100%) of the isolated C. dubliniensis. Clotrimazole was sensitive in none of the isolated C. dubliniensis, SDD in one (9.1%) of the isolated C. dubliniensis and resistant in 10 (90.9%) of the isolated C. dubliniensis. Voriconazole was sensitive in nine (81.8%) of the isolated C. dubliniensis, SDD in none of the isolated C. dubliniensis and resistant in two (18.2%) of the isolated C. dubliniensis. Itraconazole was sensitive in two (18.2%) of the isolated C. dubliniensis, SDD in one (9.1%) of the isolated C. dubliniensis and resistant in eight (72.7%) of the isolated C. dubliniensis (Figure 7).
Discussion

Worldwide, many studies have revealed that Candida spp. can switch from a commensal state into a pathogen causing infections, in reaction to transformation in the host leading to infections of oral mucosa, gastrointestinal lining and genital tract epithelium. One of the studies demonstrated that prevalence of candidiasis was more (33.8%) in women in the age range of 20–29 years, followed by those (24.3%) in the age range of 30–39 years, and the overall prevalence of VVC was 25%.13 The prevalence rate is parallel to 26% recorded in Ibadan14 and is more or less twofold to that reported in Burkina Faso (14%) and approximately half to that in (55.4%) Cameroon.15 In another study, the prevalence reported was 30.7% in Jamaica16 and 30% in Nigeria.17 A range of studies have revealed asymptomatic vaginal colonization of Candida as 5–30%.18 Many investigators noted a high infection rate in the 20–29 age group, which was consistent with our study, as many of the patients were in the age range of 25–28 years.
most probably due to indiscriminate drug usage, especially contraceptives. Various researchers declared women of age group 15–45 years to have the highest incidence, which is almost the same in the present study in women in the age range of 17–44 years. The cause for this high incidence could be due to decreased levels of protective cervical antibodies in the reproductive tract.

Spinillo et al reported that a high rate of Candida infections occurs in married women between 30 and 45 years with a reproductive history, repeated sexual activity and taking oral contraceptive pills. Rural women had an elevated rate of infectious vaginitis, owing mostly to conditions of poor medical care, lack of health education, scarce economic resources and difficulty in timely medical treatment.

The presenting complaints in 31% patients were itching, 29.4% suffered from vaginal discharge as the primary complaint, 15.6% of women presented with pain only and 13.3% presented with a triad of discharge, itching and pain. Among other complaints described were vaginal/vulvar, erythema, dysuria and dyspareunia. In another study, females (55%) who reported to clinics with VVC had pregnancy. This most likely owes to raised level of hormones during pregnancy, which is an energy source for Candida growth. An increased propensity to infection is due to both an elevated level of vaginal colonization and a higher prevalence of symptomatic vaginitis. Our findings were comparable with this study; 19.67% of C. tropicalis was isolated from vaginal swab, which was second to urine.

Oyewole et al also detected the highest occurrence of VVC in the second trimester (61%) of pregnancy, which is almost similar to our study with the highest frequency (60.2%) in the second trimester. A Brazilian study reported C. albicans isolated in 92.3%, C. krusei isolated in 3.3%, C. glabrata isolated in 2.2%, Candida parapsilosis isolated in 1.1% and C. tropicalis isolated in 1.1% of pregnant women with VVC. A study in Kenya showed that C. albicans was the most frequent isolated (73.7%) followed by C. glabrata (13%), Candida famata (5%), C. krusei (3%) and C. parapsilosis (1%).

Our findings were somewhat similar to another study publishing the highest occurrence of C. albicans (54.3%), followed by C. glabrata (25.7%), C. tropicalis (5.7%) and C. dublinensis (14.3%), identified by the CHROMagar test. Nelson et al also reported C. albicans as the commonest vaginal species followed by C. glabrata in VVC cases in pregnancy. Another report by Oyewole et al showed the maximum rate of C. albicans as 50%, followed by C. glabrata 21.4%, C. tropicalis 14.3%, C. krusei 11.9% and Candida pseudotropicalis 2.4%. In colonized women, the predominant species isolated was C. albicans, corresponding to 62.9% from the women with VVC. Other colonizing species in the vaginal mucosa of asymptomatic women were C. glabrata 14.3%, followed by Candida sphaeria 8.6% and C. parapsilosis 2.9%. In different countries, studies reported C. albicans being most widespread species in VVC (76–89%), followed by C. glabrata (7–16%). The proportion of non-albicans spp. related to VVC among these countries varied from 11% to 24.

In India, five Candida species were isolated and identified in a study, where a minor proportion of C. albicans (35.5%) was noted, followed by increased occurrence of non-albicans species, with C. tropicalis (26.4%), C. glabrata (20.6%), C. krusei (15.7%) and C. dubliniensis (1.6%). One study has described a higher level of non-C. albicans species occurrence over time. As stated by Ferrara et al, almost 25% of VVC cases were caused by non-albicans species. Comparing the results of our study with those other studies which demonstrated, C. glabrata to be the second most significant species among VVC cases because of its higher frequency and increased rate of clinical resistance to antifungals. In the present study, 41.7% of VVC cases were due to C. albicans, followed by C. tropicalis (16.7%), C. krusei (16.7%), C. glabrata (14.8%) and C. dubliniensis (10.2%).

A study in Uganda showed species frequency as follows: C. albicans 78.9%, C. glabrata 14.3%, C. krusei 3.3%, C. tropicalis 1.4%, C. famata 0.96%, Candida lusitaniae 0.4% and C. parapsilosis 0.4%. Resistance among antifungals was noted as nystatin 0.61% of C. albicans. C. krusei showed fluconazole resistance of 71.43%. C. lusitaniae, C. krusei, C. famata and C. glabrata were observed to be 100% resistant to itraconazole. C. famata and C. glabrata showed 50% and 36.7% resistance to clotrimazole correspondingly. C. albicans displayed resistance of 20.6% to itraconazole and 6.6% to voriconazole. In an 8-year study, fluconazole resistance increased from 2.4% to 55.4% (2006–2012), but the rate dropped to 8.9% in 2013. Considering miconazole and itraconazole, resistance increased from 2.4% and 7.1% to 59.8% and 58.9% (2006–2013), respectively. Among 100 Candida isolates, C. albicans (30%), C. parapsilosis (10%), C. tropicalis (21%), C. glabrata (8%), Candida parakruusei (8%) and C. krusei (3%) were isolated by workers. In vitro antifungal activity points to clotrimazole minimum inhibitory concentration of 16 and 8 µg/mL to be effective against 70% of Candida spp., fluconazole MIC of 64 and 32 µg/mL to be effective against 36.2% of isolates and nystatin disk to be 63.5% effective. In a study conducted in India,
Candida isolates showed 97.2% sensitivity to fluconazole, 80% sensitivity to clotrimazole, 57% sensitivity to itraconazole and 37% sensitivity to miconazole. In most reports, regarding fluconazole and itraconazole, resistance was noted in 42% and 48%, respectively, of the Candida isolates. In the current study, C. krusei showed 100% resistance to fluconazole, followed by 66.6% resistance to itraconazole. Prostitutes with vulvovaginitis were evaluated in a study that reported an increased susceptibility of C. albicans to fluconazole compared to C. glabrata. One resistant strain and four SDD strains of C. glabrata were isolated. On the other hand, when isolates of Candida were tested in a study by Mishra et al all C. glabrata, 50% C. tropicalis and 12% C. albicans isolates were found to be resistant to fluconazole. Recently, some authors reported that among all C. tropicalis isolates, amphotericin B was 100% sensitive, whereas 37.7% were found to be resistant to fluconazole, which is similar to the finding in our study with fluconazole resistance of 38.8%. Our study indicated that most Candida spp. tested were resistant to fluconazole (62%), followed by clotrimazole (59.3%), nystatin (58.3%) and itraconazole (40.7%); the lowest was recorded for voriconazole (10.2%). C. glabrata in our study showed resistance to nystatin (68.7%), followed by fluconazole (62.5%), itraconazole (43.7%) and voriconazole (18.7%). The most resistant C. dubliniensis isolate in the current study was nystatin (100%), followed by clotrimazole (90.9%), itraconazole (72.7%) and voriconazole (18.2%).

Conclusion

The present study demonstrated the importance of species identification and susceptibility testing for antifungals in pregnant women attending antenatal units. The predominant cause of vulvovaginal candidiasis in this study was C. albicans. An escalating number of Candida spp. from clinical isolates were resistant to antifungal agents that are routinely used for the treatment of VVC due to the fact that majority of the women had previous history of antifungal use. A significant number of non-albicans Candida were recognized, which demonstrated decreased susceptibility to all drugs, particularly the azoles, which are generally used for the management of vaginal candidiasis. Isolation of non-albicans yeasts may have clinical implication due to their reduced susceptibility to various antifungals. Antifungal susceptibility testing may possibly be used to calculate clinical response, to forecast malfunction in management, and accordingly, local antibiograms can aid in empirical assortment of antifungals, guiding options for long-term therapy, and are meant for alternative regimens in testing of isolates from recurrent infections.

Disclosure

The authors report no conflicts of interest in this work.

References


