

Liposome encapsulation of doxorubicin and celecoxib in combination inhibits progression of human skin cancer cells

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Abstract: Therapeutic agents aimed at inhibiting a single molecular target have not been successful in cancer therapy, but rather they impart resistance. However, multi-target inhibitors have shown promising results in circumventing the development of resistance and inducing apoptosis in cancer cells/tissues. In this study, we encapsulated doxorubicin and celecoxib in a single liposome at a ratio of 1:10. These dual drug-encapsulated liposomes showed excellent anticancer activity compared to individually encapsulated liposomes. The expression of key proteins such as AKT and COX-2 was suppressed, which suggests that doxorubicin and celecoxib synergistically inhibit multiple key signaling pathways.

Keywords: drug delivery, nanoliposomes, nanomedicines, cancer nanotechnology

Introduction

During the past few decades, there has been significant development in cancer treatment using chemotherapy, radiation therapy and surgery.¹ Traditional methods of cancer treatment have limited success due to systemic side-effects, development of drug resistance and sub-optimal drug concentration at the tumor site.² To increase the local drug concentration in tumor region, several strategies have been developed and are centered on targeting the cancer cells/tissues and oncogenes involved in controlling the proliferation and key survival pathways of cancer types. Several targeted monotherapies such as vemurafenib, trastuzumab, imatinib, gefitinib and erlotinib are successful in treating various cancer types.³ However, prolonged monotherapy results in the development of resistance due to multi-genetic abnormalities present in cancer cells. It has been reported that single-target inhibitors (STIs) cannot combat cancer; therefore, multi-target inhibitors (MTIs) are an attractive alternative as they have shown more efficacy and do not impart resistance compared to STIs. MTIs synergistically inhibit multiple pathways that are essential for the growth of cancer cells. Therefore, liposome encapsulation of such MTIs may offer several benefits such as improved solubility of hydrophobic drugs, natural retention of drugs at tumor site by enhanced permeability and retention (EPR) effect, enhanced circulating half-life and favorable pharmacokinetic behavior.⁴ In this study, we have synthesized liposomal formulation of two anti-cancer drugs, doxorubicin and celecoxib, which inhibit the protein kinase B (AKT) and cyclooxygenase-2 (COX-2) pathway respectively, that are overexpressed in human skin cancer cells/tissues.^{5,6}

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Materials and methods

Synthesis of liposomes

Blank liposomes (BLs) were synthesized using phosphatidylcholine (PC) and 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[methoxy(polyethylene glycol)]-2000 (m-DSPEG) film hydration under N₂ flow followed by redispersion of film in 1% saline solution. To synthesize doxorubicin encapsulating liposomes (Dox L) and celecoxib encapsulating liposomes (Cele L), the dried lipid film was rehydrated with phosphate buffer solution containing either doxorubicin or celecoxib was used. The doxorubicin and celecoxib encapsulating liposomes (Dox-Cele L) were synthesized by the aforementioned method except that the drugs were added in the ratio of 1:10.

Characterization of liposomes

Blank and Dox L and Cele L were characterized by dynamic light scattering, UV-visible spectrophotometer and transmission electron microscope.

Determination of anticancer activity

The anticancer activity of Dox L and Cele L was evaluated by 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) and 5'-bromo-2'-deoxyuridine (BrdU) assay. Different concentrations of Dox L and Cele L were exposed to human skin carcinoma (A431) cells (5,000 cells seeded in a 96-well plate) followed by the addition of MTS dye. The resultant water-soluble formazan color was read at 450 nm. A431 cells were commercially purchased from National Centre for Cell Sciences, Pune, India.

Results and discussion

The BLs were ~80 nm in diameter, whereas Dox L, Cele L and a combination of Dox L and Cele L (Dox-Cele L) (1:10) were ~87 nm, ~86 nm and ~88 nm in diameter, respectively (Table 1). It has been shown that nanoparticles of ~80 nm

Table 1 Size and zeta potential measurement of liposomes

Liposome type	Size (nm)	Zeta potential (mV)
Blank liposomes (BLs)	80.61±2.9	-46.5±3.2
Doxorubicin-encapsulated liposomes (Dox L)	87.66±3.4	-47.0±2.3
Celecoxib-encapsulated liposomes (Cele L)	86.34±4.5	-50.6±4.6
Doxorubicin and celecoxib-encapsulated liposomes (Dox-Cele L)	88.81±2.1	-50.1±5.1

Notes: Data presented as mean ± SD.

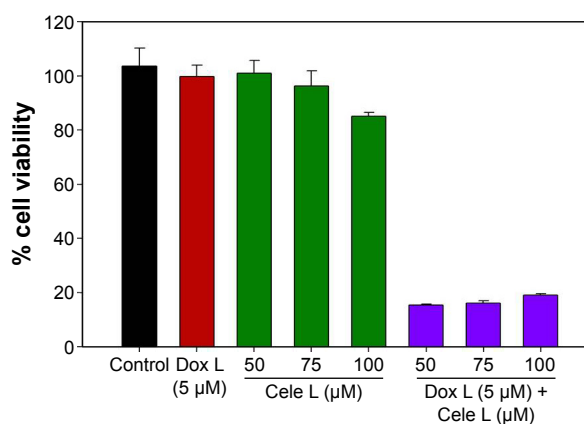


Figure 1 MTS assay showing significant decrease in A431 cell viability when exposed to Dox-Cele L than Cele L or Dox L alone.

Abbreviations: Cele L, celecoxib-encapsulated liposomes; Dox-Cele L, combination of doxorubicin- and celecoxib-encapsulated liposomes; Dox L, doxorubicin-encapsulated liposomes; MTS, 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium.

diameter are internalized the most by cancerous cells/tissues and produce EPR effect.⁴

Compared to BL, the increase in diameter of Dox L, Cele L and Dox-Cele L demonstrate the successful encapsulation of drugs. Furthermore, the high negative zeta potential values imply high stability of these liposomes in aqueous suspension.

Encapsulation of drugs did not alter the zeta potential values considerably, which indicates that drugs are present in the internal cavity of liposomes and not physically adsorbed on the surface of the liposomes. We estimated cell viability on A431 cells by two methods: MTS (Figure 1) and BrdU (Figure 2) assay. MTS assay using A431 cells clearly demonstrated that Dox L at 5 µM concentration did not cause any decrease in cell viability, whereas Cele L at 100 µM,

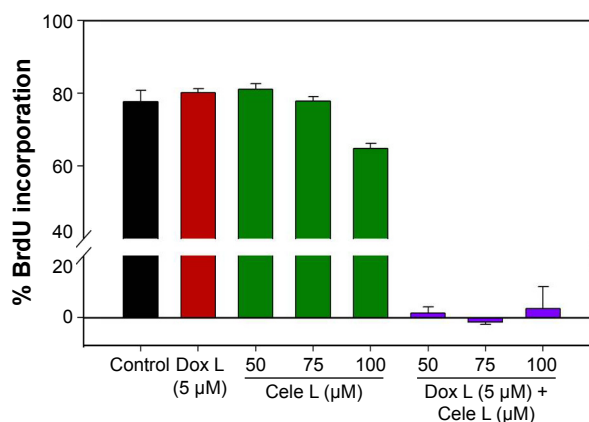


Figure 2 BrdU assay showing exposure of Dox-Cele L greatly reduced the cell proliferation in A431 cells than Cele L or Dox L alone.

Abbreviations: BrdU, 5'-bromo-2'-deoxyuridine; Cele L, celecoxib-encapsulated liposomes; Dox L, doxorubicin-encapsulated liposomes.

75 μM and 50 μM concentrations induced 18%, 7% and 3% decrease in cell viability, respectively.

Interestingly, Dox–Cele L that consisted of doxorubicin (5 μM) and celecoxib (ranging from 100 μM to 50 μM) induced significant (>90%) decrease in cell viability. This observation was further confirmed by BrdU incorporation assay (Figure 2). The results of BrdU assay showed that when used alone, 5 μM concentration of Dox L and 75 μM and 50 μM of Cele L did not induce any cytotoxicity against A431 cells; however, 100 μM of Cele L showed ~20% decrease in cell viability. Conversely, a combination of doxorubicin and celecoxib in single liposome (Dox–Cele L) showed >99% decrease in cell viability. This observation clearly suggests that the co-exposure of doxorubicin and celecoxib is much effective than either of the drugs used alone. It has been well documented that AKT and COX-2 pathways are key regulators of cancer cell progression, metastatic spread and even tumor initiation. Doxorubicin is a known inhibitor of AKT pathway and celecoxib inhibits COX-2 pathway; therefore, co-exposure of these two drugs possibly might have suppressed the expression of AKT and COX-2 simultaneously, leading to cell apoptosis.

Conclusion

Liposomes ratiometrically loaded with combinations of doxorubicin and celecoxib were successfully synthesized in this study. These dual drug-loaded liposomes were able to inhibit the cancer cell viability up to >99% even at lower

concentrations, which were ineffective when used alone. The co-exposure of doxorubicin and celecoxib might have synergistically inhibited the AKT and COX-2 pathways leading to cell apoptosis.

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Disclosure

The author reports no conflicts of interest in this work.

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