

Measurement of serum PODXL concentration for detection of pancreatic cancer

Keisuke Taniuchi^{1,2}

Makiko Tsuboi²

Masahiko Sakaguchi^{3,4}

Toshiji Saibara^{1,2}

¹Department of Endoscopic Diagnostics and Therapeutics, Kochi Medical School, Kochi University, Nankoku, Kochi, Japan; ²Department of Gastroenterology and Hepatology, Kochi Medical School, Kochi University, Nankoku, Kochi, Japan; ³Integrated Center for Advanced Medical Technologies, Kochi Medical School, Kochi University, Nankoku, Kochi, Japan; ⁴Cancer Prevention and Control Division, Kanagawa Cancer Center Research Institute, Yokohama, Kanagawa, Japan

Background: The aim of this study was to investigate the use of podocalyxin (PODXL) and secretoglobulin family 1D, member 2 (SCGB1D2) expressions in whole blood as diagnostic biomarkers to distinguish between patients with pancreatic cancer and control participants, in comparison with serum cancer antigen 19-9 (CA19-9), which is the current clinical standard.

Patients and methods: Flow cytometric analysis was performed to determine the expressions of PODXL and SCGB1D2 on the surface of cultured pancreatic cancer cells. Immunoblotting was performed to determine whether PODXL and SCGB1D2 were detectable in the media of cultured pancreatic cancer cells. A discovery-stage clinical study was performed in a cohort of 23 patients with pancreatic cancer and 51 control individuals without pancreatic disease who had been treated in the Department of Gastroenterology and Hepatology at Kochi Medical School Hospital from April 2014 to January 2016. Serum PODXL and SCGB1D2 levels were measured by enzyme-linked immunosorbent assay (ELISA).

Results: PODXL and SCGB1D2 accumulated in the protrusions of cultured pancreatic cancer cells, and they were detectable both on the cell surface and in the cultured media from these cells. The discovery-stage clinical study showed that the area under the receiver-operating characteristic curve (AUC) was 0.96 (95% confidence interval [CI] 0.91–1.000) for PODXL, 0.80 (95% CI 0.67–0.94) for SCGB1D2, and 0.78 (95% CI 0.66–0.90) for CA19-9. The AUC for PODXL was thus significantly higher than that for CA19-9 ($P = 0.006$). The combination of SCGB1D2 with CA19-9 did not significantly increase the AUC (0.83; 95% CI 0.70–0.96) compared with the AUC for either SCGB1D2 or CA19-9 alone ($P = 0.563$).

Conclusion: PODXL may be a novel, non-invasive diagnostic biomarker for the detection of pancreatic cancer.

Keywords: podocalyxin-like protein, secretoglobulin, pancreatic cancer, diagnostic marker

Introduction

Pancreatic ductal adenocarcinoma (PDAC) is the fourth most common cause of cancer death in the Western world.¹ The prognosis is poor, with 1- and 5-year survival rates of only 20% and 6%, respectively.² Early diagnosis of PDAC is difficult, and there are no blood biomarkers that can assist in the identification of such patients at an early stage.³ Therefore, the discovery of blood biomarkers that can improve the diagnosis of PDAC and provide prognostic information for these individuals would greatly assist patient management.

We have previously described two glycoproteins, podocalyxin (PODXL) and secretoglobulin family 1D, member 2 (SCGB1D2), which promote the motility and invasiveness of PDAC cells.^{4,5} PODXL is a highly glycosylated and sialylated trans-membrane protein.⁶ High PODXL expression is associated with a significantly higher risk of death from PDAC,^{4,7} as well as poor differentiation, perineural invasion, and

Correspondence: Keisuke Taniuchi
Department of Endoscopic Diagnostics and Therapeutics, Kochi Medical School, Kochi University, Kohasu, Oko-cho, Nankoku, Kochi 783-8505, Japan
Tel/fax +81 88 880 2338
Email ktaniuchi@kochi-u.ac.jp

perivascular invasion.⁷ PODXL contributes to the formation of additional membrane protrusions in PDAC cells through the recruitment of gelsolin to filamentous actin, resulting in increased motility and invasiveness.⁴ PODXL is thus an important marker of poor prognosis in PDAC.⁴

SCGB1D2, also termed lipophilin B, is a member of the secretoglobulin (SCGB) superfamily, a group of small, secretory, rarely glycosylated, dimeric proteins with unclear physiological functions, mainly expressed in mucosal tissues.⁸ Mature SCGBs are secreted proteins composed of homo- and heterodimers of SCGB polypeptides.⁹ SCGB1D2 is expressed in a high proportion of breast cancers,¹⁰ and it is more frequently expressed in estrogen receptor-positive tumors.¹¹ We recently reported that, like PODXL, SCGB1D2 is also a biomarker of poor prognosis in PDAC and plays a role in the formation of cell protrusions important for cancer cell migration.⁵

Although it was discovered >30 years ago, serum cancer antigen 19-9 (CA19-9) remains the gold standard serum marker for patients with PDAC. In a cohort where approximately half of the patients had Stage IV disease, CA19-9 was found to have a median sensitivity of 79% (70%–90%) and a median specificity of 82% (68%–91%) for PDAC diagnosis; however, the marker is not useful in the mass screening of asymptomatic subjects.¹² Moreover, inadequate sensitivity and specificity limit the use of CA19-9 in the early diagnosis of PDAC.¹³ Several other potential serum or tissue markers for PDAC, such as TIMP-1 and ICAM-1, are currently being evaluated.¹⁴ Overall, however, in current clinical practice, biomarkers have a limited role to play in diagnosing PDAC.

The aim of the present study was to assess the utility of serum PODXL and SCGB1D2 as diagnostic markers for differentiating PDAC from control participants without pancreatic disease. The differential expressions of PODXL and SCGB1D2 in serologic samples of patients with early-stage (0, I, and II) and late-stage (III and IV) PDAC are also reported.

Patients and methods

Cell culture

The human PDAC cell line S2-013, a subline of SUIT-2, was obtained from Dr T Iwamura (Miyazaki Medical College, Miyazaki, Japan).¹⁵ The human PDAC cell line, PANC-1, was purchased from the American Type Culture Collection (Manassas, VA, USA). HPNE immortalized normal pancreatic epithelial cells were a kind gift from Dr Ouellette.¹⁶ The ethics review board of Kochi Medical School approved

the use of the gifted cell lines. All cells were grown in Dulbecco's Modified Eagle's Medium (DMEM; Thermo Fisher Scientific, Waltham, MA, USA) supplemented with 10% heat-inactivated fetal calf serum at 37°C in a humidified atmosphere saturated with 5% CO₂.

Confocal immunofluorescence microscopy

Immunocytochemistry was carried out, as previously described.¹⁷ Briefly, coverslips were coated with 10 µg/mL of fibronectin (Sigma-Aldrich Co., St Louis, MO, USA) for 1 h at room temperature. Cells were seeded on fibronectin-coated glass coverslips, incubated for 5 h, and then fixed with 4% paraformaldehyde and permeabilized with 0.1% Triton X-100. Blocking solution (3% bovine serum albumin/PBS) was added, followed by incubation with anti-PODXL antibody (PAB20111; Abnova, Taipei, Taiwan) or anti-SCGB1D2 antibody (sc-48327; Santa Cruz Biotechnology Inc., Dallas, TX, USA) for 1 h. After washing, Alexa488- and Alexa594-conjugated secondary antibody (Molecular Probes, Carlsbad, CA, USA) was applied. Each specimen was visualized using a Zeiss LSM 510 META microscope (Carl Zeiss Meditec AG, Jena, Germany).

Flow cytometric analysis

S2-013 and HPNE cells (1×10^6 cells) were incubated with anti-PODXL antibody (NBP2-27219; Novus Biologicals, Littleton, CO, USA) or anti-SCGB1D2 antibody (sc-48327) at 4°C for 1 h (0.34 mg/mL). Mouse IgM for PODXL and goat IgG for SCGB1D2 were used as isotype controls (0.34 mg/mL, Santa Cruz Biotechnology Inc.). The cells were washed in PBS and then incubated with Alexa488-conjugated secondary antibodies (Molecular Probes) at 4°C for 30 min. The cells were washed in PBS and analyzed using a FACScan flow cytometer (BD Biosciences, San Jose, CA, USA) and ModFit (Verity Software House Inc., Topsham, ME, USA). Mean fluorescence intensity was calculated as relative signal intensity, ie, cells treated with anti-PODXL/cells treated with mouse IgM isotype control or cells treated with anti-SCGB1D2/cells treated with goat IgG isotype control.

Immunoblot analysis of cell lysates

Cell pellets were resuspended in 20 mM HEPES (pH 7.4), 100 mM KCl, 2 mM MgCl₂, 0.5% Triton X-100, protease inhibitor cocktail tablets (Hoffman-La Roche Ltd., Basel, Switzerland), and a phosphatase inhibitor cocktail (Nacalai Tesque, Kyoto, Japan). The bicinchoninic acid (BCA) assay was used to determine the protein concentration of each lysate;

an aliquot of each lysate was then diluted with sample buffer (50 mM Tris, 2% sodium dodecyl sulfate [SDS], 0.1% bromophenol blue, and 10% glycerol) to a final concentration of 1–2 µg/µL, and it was then analyzed by SDS–polyacrylamide gel electrophoresis (PAGE) and Western blotting using anti-PODXL (PAB20111), anti-SCGB1D2 (sc-48327), and anti-GAPDH (014-25524; Wako Pure Chemical Industries, Ltd., Osaka, Japan) antibodies.

Immunoblot analysis of culture conditioned media

S2-013 and HPNE cells were grown to 70% confluency, washed three times with PBS, and incubated for 48 h in the presence of conditioned media supplemented with 1% heat-inactivated fetal calf serum. Culture-conditioned media were collected, and cell cultures were trypsinized and analyzed for the presence of necrotic cells using trypan blue staining. Culture-conditioned media harvested from S2-013 and HPNE cells were centrifuged using a Beckman Coulter Allegra X-15R centrifuge at 300× *g* at 4°C for 10 min to remove detached cells. Supernatant was collected and filtered through 0.22 µm filters (Merck Millipore, Billerica, MA, USA) to remove contaminating apoptotic bodies, microvesicles, and cell debris. Next, the samples were concentrated using an Amicon Ultra 10K Centrifugal Filter (Merck Millipore), according to the manufacturer's instructions. The concentrations of PODXL and SCGB1D2 in these concentrated conditioned media were determined by immunoblotting. An aliquot of each conditioned medium was diluted with sample buffer (50 mM Tris, 2% SDS, 0.1% bromophenol blue, and 10% glycerol) and analyzed by SDS-PAGE and Western blotting using anti-PODXL (PAB20111), anti-SCGB1D2 (sc-48327), anti-GAPDH (014-25524), and anti-α-tubulin (017-25031; Wako Pure Chemical Industries, Ltd.) antibodies.

Human serum PDAC samples for enzyme-linked immunosorbent assay (ELISA)

The serum PODXL and SCGB1D2 levels and the clinicopathological data were analyzed retrospectively. All serum samples analyzed by ELISA were collected in the Kochi Medical School Hospital from April 2014 to January 2016. Serum samples from PDAC patients were selected for the study on the basis of the following criteria: 1) patients who were newly diagnosed and previously untreated, and 2) those with tumors pathologically diagnosed as PDAC. Tumors were classified (Stages I–IV) according to the classification of the International Union against Cancer (Table 1).¹⁸

Table 1 Summary of characteristics

Characteristics	PDAC, % (n)	Control, % (n)
Age, years		
≤39	0 (0)	7.8 (4)
40–49	4.3 (1)	11.7 (6)
50–59	4.3 (1)	9.8 (5)
60–69	26.0 (6)	41.2 (21)
70–79	56.5 (13)	13.7 (7)
≥80	8.7 (2)	15.7 (8)
Gender		
Male	52.2 (12)	43.1 (22)
Female	47.8 (11)	56.9 (29)
Diagnosed with diabetes		
Yes	21.7 (5)	13.7 (7)
No	78.3 (18)	86.3 (44)
Diagnosed with hypertension		
Yes	43.5 (10)	37.3 (19)
No	56.5 (13)	62.7 (32)
Serum uric acid		
Upregulated	13.0 (3)	17.6 (9)
Normal range	87.0 (20)	82.4 (42)
Serum triglyceride		
Upregulated	8.7 (2)	31.4 (16)
Normal range	91.3 (21)	68.6 (35)
Stage ^a		
0	4.3 (1)	
IA	0 (0)	
IB	0 (0)	
IIA	8.7 (2)	
IIB	17.4 (4)	
III	26.1 (6)	
IV	43.5 (10)	
Serum CA19-9, median (IQR)	75.6 (13.4–306.5)	12.7 (0.93–28.4)

Note: ^aClassified according to the classification of the International Union against Cancer.
Abbreviations: CA19-9, cancer antigen 19-9; IQR, interquartile range; PDAC, pancreatic ductal adenocarcinoma.

Clinicopathological parameters were classified according to the pancreatic carcinoma criteria of the Japan Pancreas Society.¹⁹ All individuals classified as controls were diagnosed with benign gastrointestinal diseases such as gastric and colonic polyps while being evaluated for non-pancreatic diseases. The study was approved by the ethics review board of Kochi Medical School (approval number: ERB-101891), and the methods were carried out in accordance with the approved guidelines. Written informed consent was obtained from each patient. All serum samples were obtained at the time of diagnosis and stored at –80°C until use.

ELISA

Serum levels of PODXL and SCGB1D2 were measured by ELISA using commercially available enzyme test kits (SEA768Hu [CLOUD-CLONE, Katy, TX, USA] for PODXL and CSB-EL020814HU [CUSABIO, Hubei Province, China] for SCGB1D2). All assays used a

quantitative sandwich enzyme immunoassay technique. Serum samples were diluted 1 in 10 in assay buffer, and then 100 μ L of the diluted sample was incubated in pre-coated ELISA 96-well plates, along with standards, for 2 h at 37°C. After washing the wells, 100 μ L of biotin-labeled polyclonal secondary antibody (detection reagent A) was added and incubated for 1 h at 37°C. After washing, 100 μ L of avidin-conjugated horseradish peroxidase (detection reagent B) was added and incubated for 30 min at 37°C. After a final washing step, 90 μ L of tetramethylbenzidine (TMB) substrate was added to each well and incubated for ~15 min in the dark at 37°C, until the second lowest standard could be distinguished from the blank by a change in color. Stopping solution (50 μ L of sulfuric acid) was then added, and the absorbance at 490 nm was analyzed using the SpectraMax 190 Microplate Reader (Molecular Devices LLC, Sunnyvale, CA, USA). Levels of CA19-9 in serum were measured by ELISA using a commercially available enzyme test kit (TM-CA-19-9 ELISA KIT; DRG International Inc., Springfield, NJ, USA), following the same protocol as mentioned earlier.

Statistical analysis

For the in vitro experiments, StarFlex software (version 6; YUMIT, Osaka, Japan) and SAS software (version 9.1.3; SAS Institute Inc., Cary, NC, USA) were used for statistical analysis. Student's *t*-test was used for the comparison of continuous variables. *P*-values < 0.05 were considered significant, and all tests were two-tailed.

In the discovery stage clinical study, statistical analysis was performed using R (version 3.3.3; The R Foundation, Vienna, Austria). The pROC package (The R Foundation) was used for receiver operating characteristic (ROC) curve analysis. Patient sample characteristics, including PODXL, SCGB1D2, and CA19-9 expressions, were compared using the Wilcoxon rank sum test. Pearson's correlation coefficients were calculated for PODXL, SCGB1D2, and CA19-9. The predictive performances of PODXL and SCGB1D2 were evaluated using ROC analysis, the area under the receiver-operating characteristic curve (AUC), and the corresponding 95% confidence intervals (CIs). DeLong's test was used for differences in AUC values.²⁰ Multivariate logistic regression was used to establish the diagnostic mathematical model. The variance inflation factor (VIF) was used to check for multicollinearity using the DAAG package (The R Foundation). Diagnostic values of PODXL and SCGB1D2 were evaluated based on AUCs, with sensitivity and specificity evaluated at various cutoff values. The three cutoff values were calculated as: 1) the highest

Youden index,²¹ 2) the highest sensitivity with specificity not <80%, and 3) the highest specificity with sensitivity not <80%. The logit model was used to determine cutoff values for combinations of multiple biomarkers.²² In all analyses, *P*-values < 0.05 were considered significant.

Results

Extracellular localization of PODXL and SCGB1D2 from cultured PDAC cells

Immunocytochemistry was used to determine the subcellular localization of PODXL and SCGB1D2 in the moderately differentiated PDAC cell line (S2-013), the poorly differentiated PDAC cell line (PANC-1), and the immortalized normal pancreatic epithelial cell line (HPNE). Western blotting showed that S2-013 and PANC-1 cells expressed higher levels of PODXL and SCGB1D2 than HPNE cells (Figure 1A). Confocal microscopy showed that PODXL and SCGB1D2 were present in the cytoplasm of the cell bodies and in membrane protrusions in S2-013 and PANC-1 cells, which had many peripheral actin structures (Figure 1B). In contrast, there was no obvious staining of PODXL and SCGB1D2 in HPNE cells (Figure 1B).

The cell surface expressions of PODXL and SCGB1D2 were assessed in S2-013 and HPNE cells by flow cytometry. This demonstrated that PODXL and SCGB1D2 antibodies bound to the surface of S2-013 cells and that PODXL and SCGB1D2 were expressed on the surface of S2-013 cells (Figure 1C). PODXL and SCGB1D2 were not detectable in HPNE cells. Because PODXL is a transmembrane glycoprotein whose extracellular domain is modified heavily by *O*-linked glycosylation and the addition of highly charged sialic acid residues,²³ and the extracellular domain of SCGB1D2 is thought to be secreted,⁸ immunoblotting was next used to examine the presence of these proteins in the culture media of S2-013 cells. The anti-PODXL antibody (PAB20111) recognized the extracellular domain of PODXL. Western blotting showed that PODXL and SCGB1D2 were both detected in media from S2-013 cultures (Figure 1D). In contrast, PODXL and SCGB1D2 were not detectable in media from HPNE cells (Figure 1D). We used two loading control markers, glyceraldehyde-3-phosphate dehydrogenase (GAPDH) and α -tubulin. GAPDH with the same molecular mass as the intracellular GAPDH is detected in the conditioned medium of mammalian cultured cell lines,²⁴ and α -tubulin is an intracellular protein marker. GAPDH was abundantly present in media from S2-013 cultures, and α -tubulin was not detectable in media from S2-013 and HPNE cultures (Figure 1D).

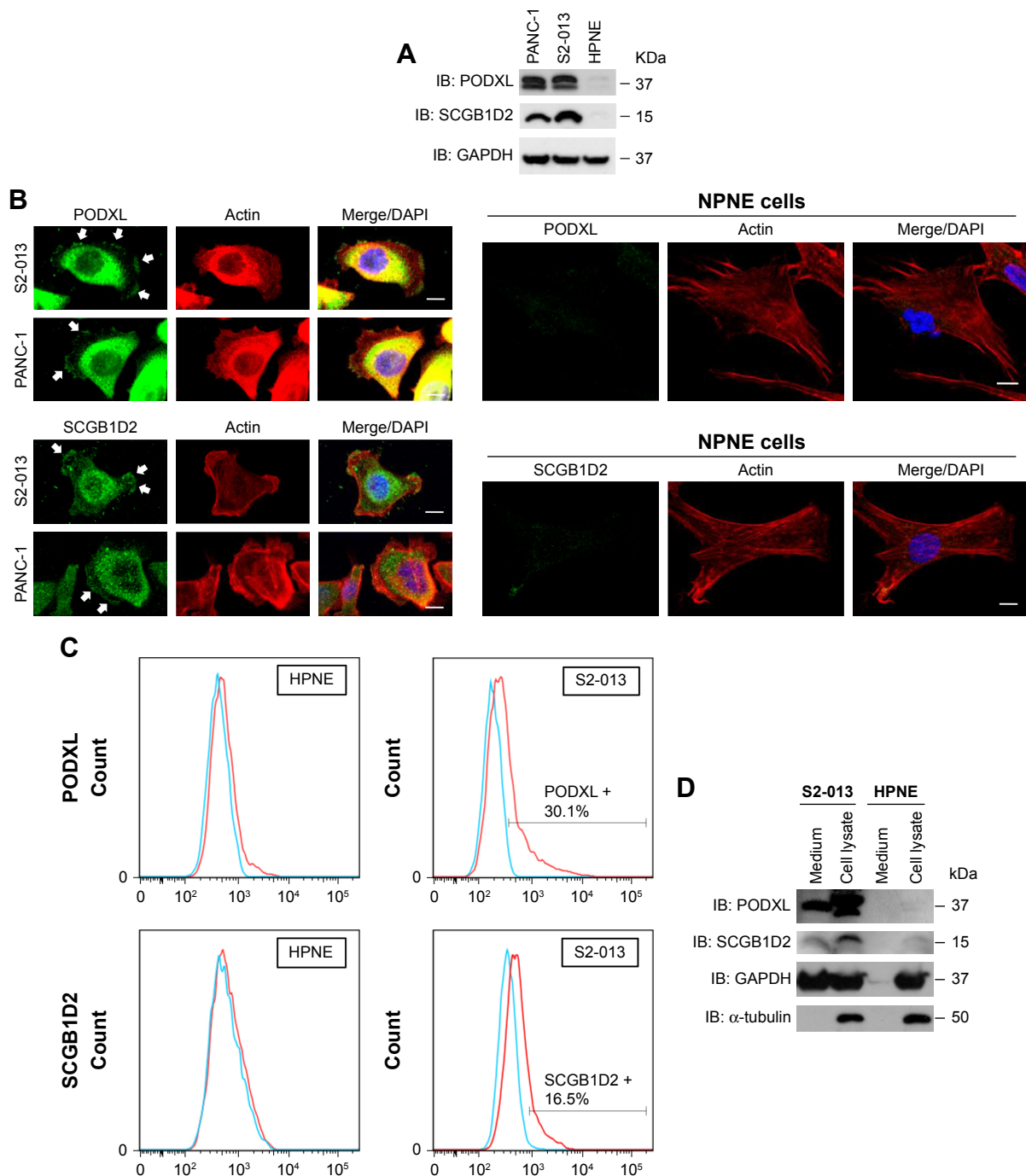


Figure 1 Subcellular localization of PODXL and SCGB1D2 in PDAC and immortalized normal pancreatic epithelial cells.

Notes: (A) Western blotting of total cell lysates of S2-013, PANC-1, and HPNE cells was performed using anti-PODXL and anti-SCGB1D2 antibodies. Loading control: GAPDH. (B) Confocal immunofluorescence microscopic images of S2-013, PANC-1, and HPNE cells labeled with anti-PODXL antibody (green) or anti-SCGB1D2 (green), and phalloidin (red; actin filaments); arrows, PODXL and SCGB1D2 localized to cell protrusions; blue, DAPI (nuclear) staining; bars, 10 μ m. (C) Expressions of PODXL and SCGB1D2 on the surface of S2-013 and HPNE cells, as evaluated by flow cytometry. Cells treated with anti-PODXL or anti-SCGB1D2 antibody (red) vs cells treated with the corresponding isotype control (blue). (D) Western blotting of total cell lysates and culture media of S2-013 and HPNE cells was performed using anti-PODXL and anti-SCGB1D2 antibodies. Loading control: α -tubulin (an intracellular protein marker) and GAPDH.

Abbreviations: GAPDH, glyceraldehyde-3-phosphate dehydrogenase; PDAC, pancreatic ductal adenocarcinoma; PODXL, podocalyxin; SCGB1D2, secretoglobulin family 1D, member 2.

Serum levels of PODXL

Because the in vitro findings suggested the potential use of PODXL as a novel tumor marker for PDAC, whether PODXL is present in the serum of patients with PDAC

was investigated. The discovery-stage clinical study included a total of 74 serologic samples from patients with PDAC and control individuals without pancreatic disease. For individuals with no prior therapy for PDAC,

disease staging (0–IV) for PDAC was performed at the time of specimen procurement. Clinical characteristics at the time of specimen procurement are summarized in Table 1. Of the individuals with PDAC, seven (30.4%) had early-stage disease (0/I/II) at the time of specimen procurement.

The PODXL levels in serum samples from both PDAC patients and control individuals were detected by ELISA experiments; the median serum level of PODXL in PDAC patients was 8.74 ng/mL (interquartile range [IQR] 7.1–12.4), while in control individuals it was significantly lower (0.15 ng/mL; IQR –1.01 to 0.86) (Figure 2A, $P < 0.001$). The detection sensitivity of the PODXL ELISA kit was 0.156 ng/mL. The relationships between serum PODXL concentration and clinicopathological features were analyzed by the Wilcoxon rank sum test (Table 2). However, there were no significant associations between serum PODXL concentration and the clinical characteristics such as age, gender, tumor size, clinical stage, and serum CA19-9 levels in PDAC patients.

Serum levels of SCGB1D2

The SCGB1D2 levels in serum samples of both PDAC patients and control individuals were detected by ELISA experiments; the median serum level of SCGB1D2 in PDAC patients was 238.87 ng/mL (IQR 115.11–683.74), while in control individuals it was 71.09 ng/mL (IQR 13.08–104.51) (Figure 2A, $P < 0.001$). The relationship between SCGB1D2 levels and clinicopathological features is shown in Table 3. Serum SCGB1D2 level was significantly correlated with gender and serum triglyceride ($P < 0.05$). However, there were no significant associations between serum SCGB1D2 levels and other clinicopathological features such as tumor size, clinical stage, and serum CA19-9 levels in PDAC patients.

ROC curve analyses

ROC curves were used to evaluate the performance of PODXL and SCGB1D2 in distinguishing patients with PDAC from control individuals (Figure 2B). For the detection of PODXL and SCGB1D2 in PDAC and control individuals

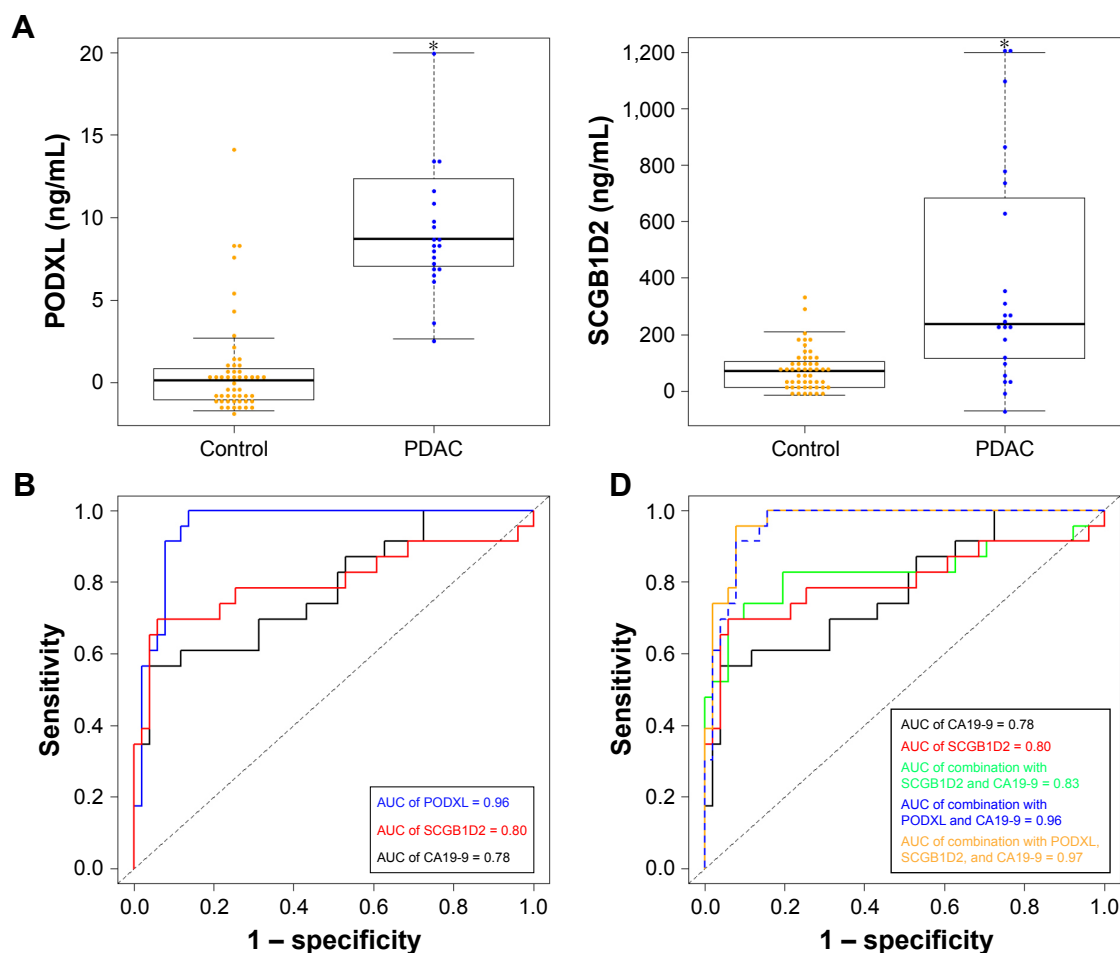


Figure 2 (Continued)

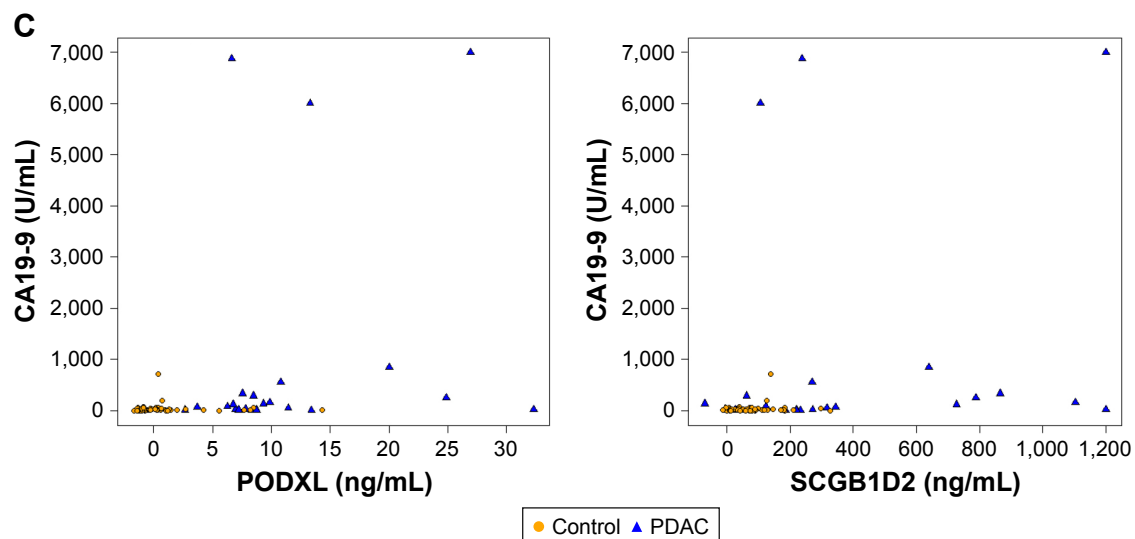


Figure 2 The performance of PODXL, SCGB1D2, CA19-9, and the combination of SCGB1D2 with CA19-9 determined by ELISA for distinguishing PDAC patients from control individuals.

Notes: (A) Distribution of PODXL and SCGB1D2 in sera from PDAC patients and control individuals. The horizontal line in the middle of each box indicates the median, whereas the top and bottom borders of the box mark the 75th and 25th percentiles, respectively. The upper whisker is the 75th percentile + $(1.5 \times \text{IQR})$. The lower whisker is the 25th percentile - $(1.5 \times \text{IQR})$. Differences are significant between PDAC patients and control individuals ($*P = 0.001$, Mann-Whitney U test). (B) ROC curves of the levels of PODXL, SCGB1D2, and CA19-9 in sera from PDAC patients and control individuals; X-axis, $1 - \text{specificity}$; Y-axis, sensitivity. ($*P = 0.001$, Mann-Whitney U test) (C) Relationships between serum levels of PODXL, SCGB1D2, and CA19-9 (X-axis, CA19-9 concentration; Y-axis, PODXL or SCGB1D2 concentration). (D) ROC curves for the performance of SCGB1D2 and CA19-9 alone, the combination of SCGB1D2 with CA19-9, the combination of PODXL with CA19-9, and the combination of PODXL and SCGB1D2 with CA19-9.

Abbreviations: CA19-9, cancer antigen 19-9; ELISA, enzyme-linked immunosorbent assay; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; IQR, interquartile range; PDAC, pancreatic ductal adenocarcinoma; PODXL, podocalyxin; ROC, receiver operating characteristic; SCGB1D2, secretoglobin family 1D, member 2.

Table 2 Correlation between serum PODXL level and clinicopathological parameters in PDAC

Characteristics	Serum PODXL level (ng/mL)		P-value
	Median	IQR	
Age, years			0.699
<70 (n=8)	8.6	7.4–10.3	
≥70 (n=15)	8.7	7.1–15.7	
Gender			0.310
Male (n=12)	9.3	7.4–15.0	
Female (n=11)	8.4	6.7–10.4	
Diagnosed with diabetes			0.766
Yes (n=5)	8.7	6.7–11.4	
No (n=18)	8.6	7.3–12.7	
Diagnosed with hypertension			0.321
Yes (n=10)	7.7	6.8–9.6	
No (n=13)	9.3	8.4–13.3	
Serum uric acid			0.553
Upregulated (n=3)	10.8	9.3–12.0	
Normal range (n=20)	8.6	6.9–11.9	
Serum triglyceride			0.785
Upregulated (n=2)	8.3	7.7–8.8	
Normal range (n=21)	8.7	6.9–13.3	
Stage ^a			0.973
0, I, II (n=7)	8.5	7.0–18.2	
III, IV (n=16)	8.7	7.1–11.4	
Tumor diameter			0.137
0–3 cm (n=13)	9.3	8.4–13.3	
>3 cm (n=10)	7.1	6.7–10.3	

Note: ^aClassified according to the classification of the International Union against Cancer.

Abbreviations: IQR, interquartile range; PDAC, pancreatic ductal adenocarcinoma; PODXL, podocalyxin.

Table 3 Correlation between serum SCGB1D2 level and clinicopathological parameters in PDAC

Characteristics	Serum SCGB1D2 level (ng/mL)		P-value
	Median	IQR	
Age, years			0.438
<70 (n=8)	203.0	94.9–440.0	
≥70 (n=15)	269.7	172.4–714.3	
Gender			0.023
Male (n=12)	455.6	234.2–808.1	
Female (n=11)	124.5	33.5–274.6	
Diagnosed with diabetes			0.502
Yes (n=5)	317.1	238.9–344.1	
No (n=18)	226.2	73.4–773.6	
Diagnosed with hypertension			0.457
Yes (n=10)	291.5	85.6–1,044.5	
No (n=13)	232.1	124.5–317.1	
Serum uric acid			0.185
Upregulated (n=3)	105.7	49.8–187.7	
Normal range (n=20)	255.1	170.3–743.0	
Serum triglyceride			0.043
Upregulated (n=2)	–15.0	–42.8–12.8	
Normal range (n=21)	269.7	185.6–727.7	
Stage ^a			0.504
0, I, II (n=7)	124.5	44.5–553.0	
III, IV (n=16)	254.3	211.7–661.7	
Tumor diameter			0.598
0–3 cm (n=13)	220.3	105.7–788.9	
>3 cm (n=10)	270.5	225.0–565.8	

Note: ^aClassified according to the classification of the International Union against Cancer.

Abbreviations: IQR, interquartile range; PDAC, pancreatic ductal adenocarcinoma; SCGB1D2, secretoglobin family 1D, member 2.

without pancreatic disorders, the AUC was 0.96 (95% CI 0.91–0.99) and 0.80 (95% CI 0.67–0.94), respectively. The levels of CA19-9 were determined using a commercial ELISA kit, and the AUC of CA19-9 was 0.78 (95% CI 0.66–0.90). The performance of PODXL in distinguishing PDAC from control individuals was found to be superior to that of CA19-9 ($P = 0.006$). However, SCGB1D2 was not superior to CA19-9 ($P = 0.802$).

The sensitivity and specificity of PODXL for PDAC diagnosis were analyzed using different cutoff values (Table 4). Three cutoff values for each marker were obtained by minimizing the total prediction error, as described in the “Patients and methods” section. The sensitivity of PODXL ranged from 91.3% to 100%, and the specificity varied from 86.3% to 92.2%, for the different cutoff values. Specifically, when a threshold of 6.27 ng/mL was used, a sensitivity of 91.3%, a specificity of 92.2%, and an accuracy of 91.9% were achieved (Table 4). The sensitivity of SCGB1D2 ranged from 69.6% to 82.6%, and the specificity varied from 86.3% to 94.1% for the different cutoff values. Specifically, when a threshold of 185.58 ng/mL was used, a sensitivity of 69.6%, a specificity of 94.1%, and an accuracy of 86.5% were achieved (Table 4). Using the same method for CA19-9, instead of using the standard cutoff value (37.0 U/mL), when a threshold of 66.0 U/mL was used, a sensitivity of 56.5%, a specificity of 96.1%, and an accuracy of 83.8% were achieved (Table 4). Decreasing the cutoff value to 50.0 U/mL improved the sensitivity for the detection of PDAC to 60.9%, but lowered the specificity to 88.2% and the accuracy to 79.7%.

Combination with SCGB1D2 and CA19-9

There were significant correlations between the serum levels of PODXL and CA19-9 (Pearson's correlation coefficient $[R] = 0.38$, 95% CI 0.17–0.56, $P = 0.007$), between SCGB1D2 and CA19-9 ($R = 0.31$, 95% CI 0.09–0.50, $P = 0.007$), and between PODXL and SCGB1D2 ($R = 0.72$, 95% CI 0.588–0.81, $P < 0.001$) (Figure 2C). To take account of multicollinearity, VIF was employed. None of the VIF values for the two models reached a value of 10, meaning that there was no collinearity in the model. VIF values for CA19-9, PODXL, gender, and age were 1.02, 1.08, 1.07, and 1.01, respectively. VIF values for CA19-9, SCGB1D2, gender, and age were 1.05, 1.25, 1.24, and 1.04, respectively.

SCGB1D2 and CA19-9 were combined in an attempt to improve the sensitivity and specificity for the detection of PDAC. ROC analysis showed that the AUC for the

Table 4 Sensitivity and specificity for PODXL, SCGB1D2, and CA19-9 at specified cutoff values

Marker	Cutoff P-value	Sensitivity (95% CI)	Specificity (95% CI)	Accuracy (95% CI)	AUC (95% CI)	True positives (n)	True negatives (n)	False positives (n)	False negatives (n)	LR+ (95% CI)	LR- (95% CI)
PODXL											
A	2.68	100 (78.9–100)	86.3 (73.7–94.3)	90.5 (81.5–96.1)	0.96 (0.91–0.99)	23	44	7	0	7.3 (3.7–14.5)	0 –
B	2.68	100 (78.9–100)	86.3 (73.7–94.3)	90.5 (81.5–96.1)		23	44	7	0	7.3 (3.7–14.5)	0 –
C	6.27	91.3 (72.0–98.9)	92.2 (81.1–97.8)	91.9 (83.2–97.0)		21	47	4	2	11.6 (4.5–30.1)	0.1 (0.0–0.4)
SCGB1D2											
A	185.58	69.6 (47.1–86.8)	94.1 (83.8–98.8)	86.5 (76.5–93.3)	0.80 (0.67–0.94)	16	48	3	7	11.8 (3.8–36.6)	0.3 (0.2–0.6)
B	185.58	69.6 (47.1–86.8)	94.1 (83.8–98.8)	86.5 (76.5–93.3)		16	48	3	7	11.8 (3.8–36.6)	0.3 (0.2–0.6)
C	62.58	82.6 (61.2–95.0)	47.1 (32.9–61.5)	58.1 (46.1–69.5)		19	24	27	4	1.6 (1.1–2.1)	0.3 (0.1–0.9)
CA19-9											
A	66	56.5 (34.5–76.8)	96.1 (86.5–99.5)	83.8 (73.4–91.3)	0.78 (0.66–0.90)	13	49	2	10	14.4 (3.5–58.7)	0.5 (0.3–0.7)
B	50	60.9 (38.5–80.3)	88.2 (76.1–95.6)	79.7 (68.8–88.2)		14	45	6	9	5.2 (2.3–11.7)	0.4 (0.3–0.7)
C	9.7	82.6 (61.2–95.0)	49.0 (34.8–63.4)	59.5 (47.4–70.7)		19	25	26	4	1.6 (1.2–2.2)	0.4 (0.1–0.9)

Abbreviations: AUC, area under the curve; CA19-9, cancer antigen 19-9; CI, confidence interval; LR, likelihood ratio; PODXL, podocalyxin; SCGB1D2, secretoglobulin family 1D, member 2.

combination of SCGB1D2 and CA19-9 was 0.83 (95% CI 0.70–0.96), which was not significantly different from the AUC of SCGB1D2 alone ($P = 0.213$) or CA19-9 alone ($P = 0.563$) (Figure 2D). The sensitivity of the combination of SCGB1D2 and CA19-9 ranged from 73.9% to 82.6%, the specificity ranged from 80.4% to 90.2%, and the accuracy ranged from 81.1% to 85.1% for the different cutoff values (Table 5). When a threshold of 0.306 was used, a sensitivity of 73.9%, a specificity of 90.2%, and an accuracy of 85.1% were achieved (Table 5). With the combination of PODXL and CA19-9, the AUC of this combination was not significantly different from the AUC of PODXL alone (Figure 2D), indicating that PODXL was independently able to distinguish PDAC patients from control individuals.

Association of serum tumor marker levels with the risk of PDAC

Next, the significance of serum tumor marker levels for the diagnosis of PDAC was evaluated using logistic regression to obtain crude odds ratios (ORs) (Table 6). To exclude the possible effects of age and gender, crude ORs were then adjusted for the effects of age and gender (adjusted OR [aOR] values). The results showed that among PODXL, SCGB1D2, and CA19-9, the serum level of PODXL was the most highly correlated with the risk of PDAC. The combination of PODXL with CA19-9 did not improve PDAC detection compared to the use of PODXL alone (aOR of CA19-9 = 1.00, 95% CI 0.99–1.01, $P = 0.227$, for the multivariate regression model of PODXL and CA19-9).

Detection of early-stage PDACs using PODXL, SCGB1D2, or CA19-9

The level of PODXL in serum samples from patients in the early stages of PDAC (Stages 0, I, and II) was significantly higher than the level in controls, as well as the level of CA19-9 in these patients (Figure 3A). ROC curves for PODXL, SCGB1D2, and CA19-9 are shown in Figure 3B. The AUC values of PODXL used to distinguish patients with Stage 0/I/II and Stage III/IV PDAC from controls were 0.96 and 0.96, respectively (Table 7). For SCGB1D2, the AUC values used to distinguish patients with Stage 0/I/II and Stage III/IV PDAC from controls were 0.66 and 0.87, respectively (Table 7). For CA19-9, the AUC values to distinguish patients with Stage 0/I/II and Stage III/IV PDAC from controls were 0.79 and 0.77, respectively (Table 7). The performance of PODXL in distinguishing PDAC at Stage 0/I/II from control individuals was found to be superior to that of CA19-9 ($P = 0.048$).

Table 5 Sensitivity and specificity for combination with SCGB1D2 and CA19-9 at specified cutoff values

Marker	Cutoff	P-value	Sensitivity (95% CI)	Specificity (95% CI)	Accuracy (95% CI)	AUC (95% CI)	True positives (n)	True negatives (n)	False positives (n)	False negatives (n)	LR ⁺ (95% CI)	LR [–] (95% CI)
PODXL + CA19-9												
A	2.68		100 (78.9–100)	84.3 (71.4–93.0)	89.2 (79.8–95.2)	0.97 (0.92–0.99)	23	43	8	0	6.4 (3.4–12.0)	0
B	2.68		100 (78.9–100)	84.3 (71.4–93.0)	89.2 (79.8–95.2)		23	43	8	0	6.4 (3.4–12.0)	0
C	6.27		91.3 (72.0–98.9)	92.2 (81.1–97.8)	91.9 (83.2–97.0)		21	47	4	2	11.6 (4.5–30.0)	0.0 (0.0–0.3)
SCGB1D2 + CA19-9												
A	0.306		73.9 (51.6–89.8)	90.2 (78.6–96.7)	85.1 (75.0–92.3)	0.83 (0.70–0.96)	17	46	5	6	7.5 (3.1–17.9)	0.3 (0.1–0.6)
B	0.227		82.6 (61.2–95.0)	80.4 (66.9–90.2)	81.1 (70.3–89.3)		19	41	10	4	4.2 (2.3–7.6)	0.2 (0.1–0.5)
C	0.227		82.6 (61.2–95.0)	80.4 (66.9–90.2)	81.1 (70.3–89.3)		19	41	10	4	4.2 (2.3–7.6)	0.2 (0.1–0.5)

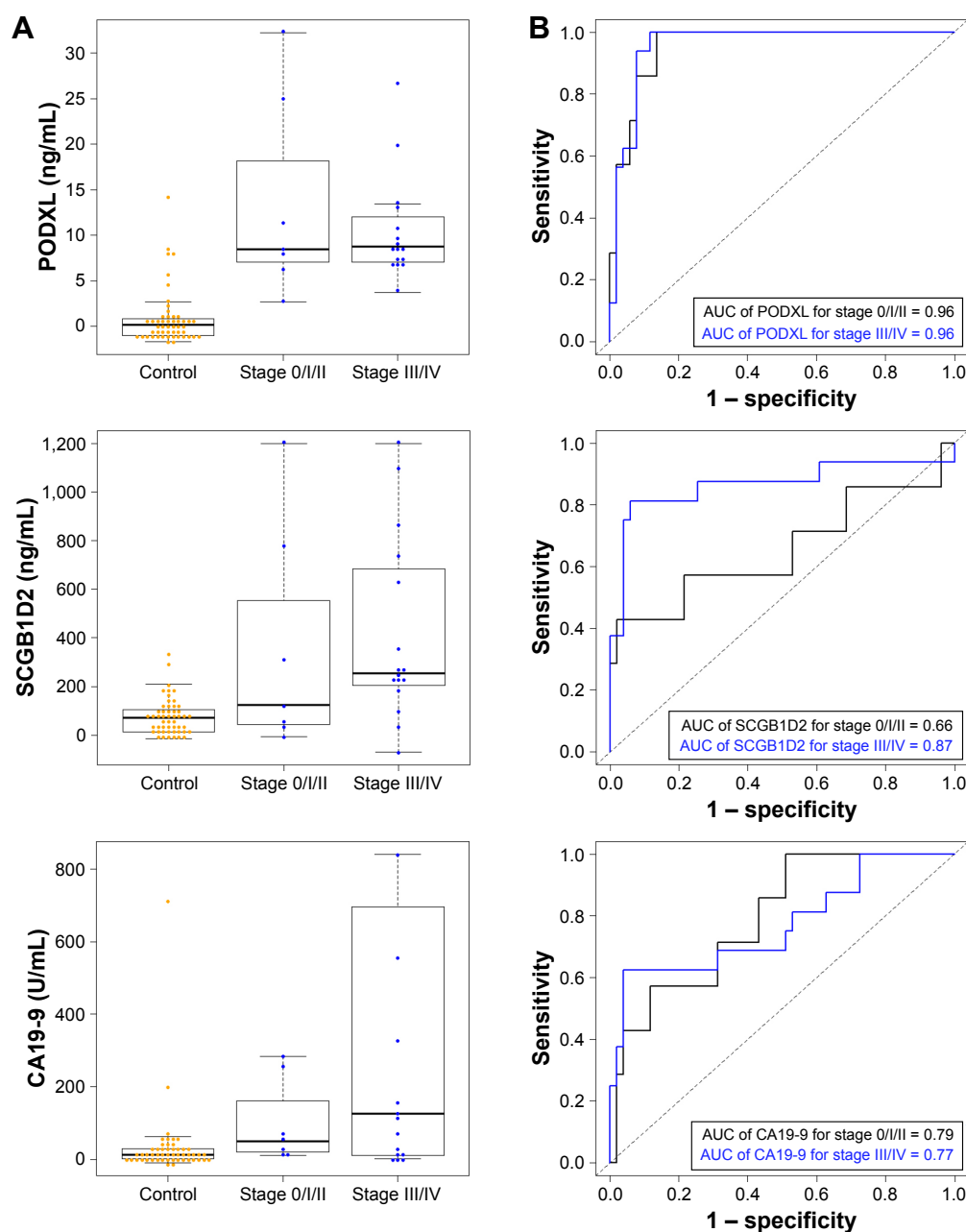
Abbreviations: AUC, area under the curve; CA19-9, cancer antigen 19-9; CI, confidence interval; LR, likelihood ratio; PODXL, podocalyxin; SCGB1D2, secretoglobulin family 1D, member 2.

Table 6 Univariate and multivariate logistic regression analyses

Indicators	Crude OR	P-value	Adjusted OR of Model 1	P-value	Adjusted OR of Model 2	P-value
PODXL	1.69 (95% CI 1.35–2.12)	<0.001	PODXL 1.66 (95% CI 1.32–2.10)	<0.001	PODXL 1.67 (95% CI 1.31–2.13)	<0.001
			CA19-9 1.00 (95% CI 0.99–1.01)	0.227	CA19-9 1.00 (95% CI 0.98–1.01)	0.225
SCGB1D2	1.01 (95% CI 1.00–1.02)	0.002	SCGB1D2 1.01 (95% CI 1.00–1.01)	0.006	SCGB1D2 1.01 (95% CI 1.00–1.02)	0.005
			CA19-9 1.00 (95% CI 0.99–1.01)	0.308	CA19-9 1.00 (95% CI 0.99–1.01)	0.460
CA19-9	1.01 (95% CI 1.00–1.01)	0.033	–	–	1.01 (95% CI 1.00–1.01)	0.058

Notes: Model 1: Odds ratio adjusted for CA19-9. Model 2: Odds ratio adjusted for CA19-9, age and gender.

Abbreviations: AUC, area under the curve; CA19-9, cancer antigen 19-9; CI, confidence interval; OR, odds ratio; PODXL, podocalyxin; SCGB1D2, secretoglobin family 1D, member 2.

**Figure 3** Determination of PODXL, SCGB1D2, and CA19-9 by ELISA in Stage 0/I/II and Stage III/IV PDACs.

Notes: (A) PODXL, SCGB1D2, and CA19-9 levels in sera from Stage 0/I/II PDACs, Stage III/IV PDACs, and control individuals. The horizontal line in the middle of each box indicates the median, whereas the top and bottom borders of the box mark the 75th and 25th percentiles, respectively. The upper whisker is the 75th percentile + (1.5 × IQR). The lower whisker is the 25th percentile – (1.5 × IQR). (B) ROC curves for the performance of PODXL, SCGB1D2, and CA19-9 from Stage 0/I/II PDACs, Stage III/IV PDACs, and control individuals.

Abbreviations: CA19-9, cancer antigen 19-9; ELISA, enzyme-linked immunosorbent assay; IQR, interquartile range; PDAC, pancreatic ductal adenocarcinoma; PODXL, podocalyxin; ROC, receiver operating characteristic; SCGB1D2, secretoglobin family 1D, member 2.

Table 7 Concentration and AUC of PODXL, SCGB1D2, and CA19-9 at each clinical stage of PDAC

Indicators	State	PDAC stage ^a	Number	Median (IQR)	AUC (95% CI)
PODXL	Control		51	0.15 (–1.01–0.86)	Reference
	PDAC	0/I/II	7	8.48 (7.05–18.2)	0.96 (0.90–1)
		III/IV	16	8.74 (7.14–11.42)	0.96 (0.91–1)
SCGB1D2	Control		51	71.09 (13.08–104.50)	Reference
	PDAC	0/I/II	7	124.50 (44.52–553.00)	0.66 (0.37–0.95)
		III/IV	16	254.30 (211.70–661.70)	0.87 (0.73–1)
CA19-9	Control		51	12.70 (0.93–28.41)	Reference
	PDAC	0/I/II	7	50.00 (19.00–161.80)	0.79 (0.62–0.96)
		III/IV	16	125.40 (11.02–624.20)	0.77 (0.62–0.92)

Note: ^aClassified according to the classification of the International Union against Cancer.

Abbreviations: AUC, area under the curve; CA19-9, cancer antigen 19-9; CI, confidence interval; IQR, interquartile range; PDAC, pancreatic ductal adenocarcinoma; PODXL, podocalyxin; SCGB1D2, secretoglobin family 1D, member 2.

Discussion

Kaplan–Meier plots and univariate and multivariate analyses of pathological findings have previously shown that high expressions of PODXL and SCGB1D2 are significant prognostic factors associated with worse survival after PDAC resection.^{4,5} Expression levels of PODXL and SCGB1D2 were evaluable in all 102 PDAC cases, and these cases were classified into low-expressing (70.6% for PODXL and 63.7% for SCGB1D2) and high-expressing (29.4% for PODXL and 36.3% for SCGB1D2) groups.^{4,5} Therefore, an increase in PODXL and SCGB1D2 within tumors would be expected to affect signaling pathways relevant to cell proliferation, invasiveness, and metastasis in PDAC. Since sensitive and specific biomarkers to identify patients with PDAC at an early stage are needed, it is important to investigate whether serum PODXL and SCGB1D2 are useful in the diagnosis of PDAC, as was done in the present study.

We previously reported that intracellular PODXL promotes PDAC cell motility and invasion by physically binding to the cytoskeletal protein gelsolin, and PODXL is a significant prognostic factor that predicts the overall survival of patients with PDAC.⁴ Since membranous PODXL is rarely expressed in migrating PDAC cells, intracellular PODXL bound to gelsolin may be more important for the promotion of cell motility and invasion than membranous PODXL.⁴ The main function of PODXL is to regulate cell morphology and adhesion by mediating the connection between intracellular proteins and extracellular ligands.²⁵ The present study demonstrated that PODXL accumulates in the protrusions of PDAC cells and localizes to the cell surface. In addition, it was shown that PODXL is present in the media of PDAC cell cultures; however, the molecular differences (both in terms of amino acid sequence and post-translational modifications) between membrane-bound PODXL and extracellular PODXL, as well as the functions of extracellular PODXL, are currently unknown. Future

studies should thus evaluate the structure of the soluble form of PODXL that can be detected in culture-conditioned media from PDAC cells.

Tumor-specific transmembrane or secretory proteins have significant advantages as biomarkers because they are present on the cell surface, within the extracellular space, or in the blood and are thus easily accessible.²⁶ Like PODXL, SCGB1D2 was also found to accumulate in the protrusions of migrating PDAC cells, and it was detected both on the cell surface and in the culture media of PDAC cells. Western blotting using anti-PODXL antibody that recognized the extracellular domain of PODXL showed two bands of PODXL (30 kDa and 45 kDa) in the cell lysate of PDAC cells and a single band of PODXL (35 kDa) in the culture media of PDAC cells (Figure 1D). PODXL contains a potential NH₂-terminal signal peptide, a single COOH-terminal membrane-spanning region, and the extracellular domain containing multiple potential sites for *N*- and *O*-linked glycosylation.²³ It is possible that 30 kDa and 40 kDa PODXLs were present due to post-translational modifications, most likely glycosylation, in PDAC cells. The molecular weight of a portion of the intracellular domain of PODXL is ~10 kDa,²³ and, thus, a portion of the extracellular domain of PODXL could be detected in the culture media of PDAC cells. There were also high levels of PODXL and SCGB1D2 in serum samples from PDAC patients. It is possible that the extracellular domain of PODXL was present in the serum obtained from PDAC patients. Serum SCGB1D2 was correlated with gender and serum triglyceride levels, but other clinicopathological factors were not significantly correlated with SCGB1D2 serum levels. The reasons for the observed associations between serum SCGB1D2, gender, and serum triglyceride levels are unknown; however, this should be further examined in future clinical studies.

To validate the feasibility of using serum levels of PODXL and SCGB1D2 as diagnostic tools, the sensitivity and

specificity of these markers were compared with those of the current standard serum marker, CA19-9. PODXL performed better than serum CA19-9 when using AUC to discriminate between PDAC and control individuals; however, the increase in AUC was not significant for SCGB1D2. It was also found that the combination of SCGB1D2 and CA19-9 did not significantly increase the AUC (0.83; 95% CI 0.70–0.96) compared with either SCGB1D2 (0.80; 95% CI 0.67–0.94) or CA19-9 alone (0.78; 95% CI 0.66–0.90). Importantly, these results suggest that increased expression of serum PODXL is more accurate for the diagnosis of PDAC than serum CA19-9. However, the combination of SCGB1D2 and CA19-9 does not appear to be particularly useful as a serum biomarker.

A multicenter trial comparing resection and chemoradiotherapy for locally invasive resectable PDAC (Stages IIA and IIB) showed significantly better outcomes with surgery, despite the premature termination of the trial.^{27,28} The circumferential encasement of the celiac axis, the hepatic artery, or the superior mesenteric artery with suspected arterial tumor infiltration (T4 or Stage III) is categorized as non-resectable in most cases.^{28,29} Not only could better markers improve the early diagnosis of PDAC and allow more patients to undergo curative surgical resection but they could also potentially be used for patients at high risk of developing pancreatic cancer, to identify precancerous lesions while they are amenable to cure.³⁰ The present study included seven cases of Stage 0/I/II PDAC, and the AUC value for PODXL in the detection of these tumors was significantly higher than that for CA19-9 ($P = 0.048$). This study included only one patient in Stage 0-IB; therefore, additional validation with a larger set of serum samples covering various clinical stages, especially Stage 0-IB, is necessary to validate that PODXL is a potential biomarker for the screening of early-stage PDAC.

Conclusion

The use of serum PODXL as a biomarker achieved a sensitivity of 94.6% and a specificity of 84.3% for the detection of PDAC, at a cutoff value of 2.68 U/mL. All 23 patients with PDAC (100%) and all seven patients with Stage 0/I/II PDAC (100%) were correctly diagnosed as positive, whereas 13.7% of the 51 control individuals were falsely diagnosed as positive. Although it was quantitatively demonstrated that the detection of PODXL in serum may provide a novel, non-invasive approach for distinguishing PDAC from control participants, the present findings are preliminary until more elaborate studies clarify that PODXL is superior to CA19-9. Further studies are required to elucidate whether PODXL is useful as a diagnostic marker for the detection of

Stage 0/I/II PDACs, as well as how much additional benefit it may add in comparison to the use of serum CA19-9. To this end, we have started a prospective clinical validation study (UMIN000021938) in the Department of Gastroenterology and Hepatology at Kochi Medical School Hospital to assess the accuracy of increased expression of PODXL in the diagnosis of PDAC and the benefit of PODXL as a potential diagnostic marker for the early detection of PDACs in comparison to CA19-9. This prospective clinical study also includes patients with chronic pancreatitis and a non-cancerous pancreatic lesion, such as intraductal papillary mucinous neoplasm, to assess the utility of serum PODXL as a diagnostic marker for differentiating PDAC from other pancreatic disorders.

Acknowledgments

The authors would like to thank Miki Nishigawa and Rieko Takahashi for their excellent technical assistance. This study was supported by a Grant-in-Aid for Scientific Research (KAKENHI; 15K14396).

Author contributions

All authors contributed toward data analysis, drafting and critically revising the paper and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

References

1. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. *CA Cancer J Clin*. 2013;63(1):11–30.
2. Hidalgo M. Pancreatic cancer. *N Engl J Med*. 2010;362(17):1605–1617.
3. Locker GY, Hamilton S, Harris J, et al. ASCO 2006 update of recommendations for the use of tumor markers in gastrointestinal cancer. *J Clin Oncol*. 2006;24(33):5313–5327.
4. Taniuchi K, Furihata M, Naganuma S, Dabanaka K, Hanazaki K, Saibara T. Podocalyxin-like protein, linked to poor prognosis of pancreatic cancers, promotes cell invasion by binding to gelsolin. *Cancer Sci*. 2016;107(10):1430–1442.
5. Taniuchi K, Furihata M, Naganuma S, et al. Elevated expression of SCGB1D2 predicts unfavorable prognosis in patients with pancreatic ductal adenocarcinoma. *J Pancreas*. 2017;18:216.
6. Nielsen JS, McNagny KM. The role of podocalyxin in health and disease. *J Am Soc Nephrol*. 2009;20(8):1669–1676.
7. Saukkonen K, Hagström J, Mustonen H, et al. Podocalyxin is a marker of poor prognosis in pancreatic ductal adenocarcinoma. *PLoS One*. 2015;10(6):e0129012.
8. Klug J, Beier HM, Bernard A, et al. Uteroglobin/Clara cell 10 kDa family of proteins: nomenclature committee report. *Ann NY Acad Sci*. 2000;923:348–354.
9. Ni J, Kalfi-Suske M, Gentz R, Schageman J, Beato M, Klug J. All human genes of the uteroglobin family are localized on chromosome 11q12.2 and form a dense cluster. *Ann NY Acad Sci*. 2000;923:25–42.

10. Dillon DC, Reynolds LD, Retter MW, et al. Serum antibodies to lipophilin B detected in late stage breast cancer patients. *Clin Cancer Res*. 2003;9(2):749–754.
11. O'Brien N, Maguire TM, O'Donovan N, et al. Mammaglobin a: a promising marker for breast cancer. *Clin Chem*. 2002;48(8):1362–1364.
12. Goonnetilleke KS, Siriwardena AK. Systematic review of carbohydrate antigen (CA19-9) as a biochemical marker in the diagnosis of pancreatic cancer. *Eur J Surg Oncol*. 2007;33(3):266–270.
13. Duffy MJ, Sturgeon C, Lamerz R, et al. Tumor markers in pancreatic cancer: a European Group on Tumor Markers (EGTM) status report. *Ann Oncol*. 2010;21(3):441–447.
14. Brand RE, Nolen BM, Zeh HJ, et al. Serum biomarker panels for the detection of pancreatic cancer. *Clin Cancer Res*. 2011;17(4):805–816.
15. Iwamura T, Katsuki T, Ide K. Establishment and characterization of a human pancreatic cancer cell line (SUIT-2) producing carcinoembryonic antigen and carbohydrate antigen 19-9. *Jpn J Cancer Res*. 1987;78(1):54–62.
16. Lee KM, Yasuda H, Hollingsworth MA, Ouellette MM. Notch 2-positive progenitors with the intrinsic ability to give rise to pancreatic ductal cells. *Lab Invest*. 2005;85(8):1003–1012.
17. Taniuchi K, Nishimori I, Hollingsworth MA. Intracellular CD24 inhibits cell invasion by posttranscriptional regulation of BART through interaction with G3BP. *Cancer Res*. 2011;71(3):895–905.
18. Sobin LH, Gospodarowicz MK, Wittekind C. *TNM Classification of Malignant Tumors*. 7th ed. New York: Wiley-Blackwell; 2009: 132–135.
19. Japan Pancreatic Society. *Classification of Pancreatic Carcinoma*. 2nd ed. Tokyo: Kanehara & Co; 2003.
20. DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics*. 1988;44(3):837–845.
21. Youden W. Index for rating diagnostic tests. *Cancer*. 1950;3(1): 32–35.
22. Soureshjani MH, Kimiagari AM. Calculating the best cut off point using logistic regression and neural network on credit scoring problem – a case study of a commercial bank. *Afr J Bus Manag*. 2013;7:1414–1421.
23. Kershaw DB, Beck SG, Wharram BL, et al. Molecular cloning and characterization of human podocalyxin-like protein: orthologous relationship to rabbit PCLP1 and rat podocalyxin. *J Biol Chem*. 1997; 272(25):15708–15714.
24. Yamaji R, Chatani E, Harada N, Sugimoto K, Inui H, Nakano Y. Glyceraldehyde-3-phosphate dehydrogenase in the extracellular space inhibits cell spreading. *Biochim Biophys Acta*. 2005;1726(3): 261–271.
25. Thomas SN, Schnaar RL, Konstantopoulos K. Podocalyxin-like protein is an E-/L-selectin ligand on colon carcinoma cells: comparative biochemical properties of selectin ligands in host and tumor cells. *Am J Physiol Cell Physiol*. 2009;296(3):C505–C513.
26. Ishikawa N, Daigo Y, Yasui W, et al. ADAM8 as a novel serological and histochemical marker for lung cancer. *Clin Cancer Res*. 2004;10(24):8363–8370.
27. Imamura M, Doi R, Imaizumi T, et al. A randomized multicenter trial comparing resection and radiochemotherapy for resectable locally invasive pancreatic cancer. *Surgery*. 2004;136(5):1003–1011.
28. Hartwig W, Werner J, Jäger D, Debus J, Büchler MW. Improvement of surgical results for pancreatic cancer. *Lancet Oncol*. 2013;14(11): e476–e485.
29. Callery MP, Chang KJ, Fishman EK, et al. Pretreatment assessment of resectable and borderline resectable pancreatic cancer: expert consensus statement. *Ann Surg Oncol*. 2009;16(7):1727–1733.
30. Goggins M. Molecular markers of early pancreatic cancer. *J Clin Oncol*. 2005;23(20):4524–4531.

OncoTargets and Therapy

Publish your work in this journal

OncoTargets and Therapy is an international, peer-reviewed, open access journal focusing on the pathological basis of all cancers, potential targets for therapy and treatment protocols employed to improve the management of cancer patients. The journal also focuses on the impact of management programs and new therapeutic agents and protocols on

Submit your manuscript here: <http://www.dovepress.com/oncotargets-and-therapy-journal>

patient perspectives such as quality of life, adherence and satisfaction. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Dovepress