Exercise-induced changes in stress hormones and cell adhesion molecules in obese men

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Purpose: The current study examined the relationship between exercise-induced changes in stress hormones (epinephrine, norepinephrine, and cortisol) and vascular inflammatory markers (soluble intracellular adhesion molecule-1 [sICAM-1], soluble endothelial selectin [sE-selectin], and soluble vascular adhesion molecule-1 [sVCAM-1]) in obese men over a 24-hour period following exercise at lower and higher intensity.

Patients and methods: Fifteen physically inactive, obese, college-aged men performed a single bout of cycling exercise at lower and higher intensities (lower intensity: 50% of maximal heart rate, and higher intensity: 80% of maximal heart rate) in random order. Overnight fasting blood samples were collected at baseline, immediately postexercise (IPE), 1-hour PE (1-h PE), and 24-hour PE. Changes in stress hormones and inflammatory markers were analyzed with a repeated-measures analysis of variance using Bonferroni multiple comparisons and a linear regression analysis (p<0.05).

Results: sICAM-1, sVCAM-1, epinephrine, and norepinephrine did not change over time, while sE-selectin was significantly lower at 1-h PE (10.25±1.07 ng/mL, p=0.04) than at baseline (12.22±1.39 ng/mL). Cortisol and sICAM-1 were negatively related at 1-h PE following lower-intensity exercise (r²=0.34, p=0.02), whereas cortisol and sVCAM-1 were positively related at IPE following higher-intensity exercise (r²=0.36, p=0.02).

Conclusion: Regardless of intensity, an acute bout of aerobic exercise may lower sE-selectin in sedentary obese men. Responses of cortisol are dependent on exercise intensity, and cortisol may be a key stress hormone playing a major role in regulating sICAM-1 and sVCAM-1.

Keywords: vascular inflammation, ICAM-1, VACM-1, E-selectin, stress hormones, exercise intensity

Introduction

Atherosclerosis is the most common type of cardiovascular disease (CVD) and is strongly associated with elevated vascular inflammatory markers, such as cell adhesion molecules (CAMs). CAMs are glycoproteins expressed on the surface of various cells and play a crucial role in many cellular processes. In particular, intracellular adhesion molecules-1 (ICAM-1), vascular cell adhesion molecules-1 (VCAM-1), and E-selectin are considered the key molecules that are responsible for transmigrating leukocytes into the vascular intima, which in turn initiates the inflammatory process. However, a direct measurement of the actual expression of CAMs on endothelia is quite difficult in humans, whereas a soluble form of CAMs in the circulation, known

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as soluble CAMs (sCAMs), can be easily obtained from blood samples and indirectly reflects the rate of endothelial expression of CAMs.

Stress hormones, such as catecholamines (epinephrine [E] and norepinephrine [NE]) and cortisol, can modulate the CAM metabolism. When catecholamine receptors on leukocytes are activated in response to physical activity or stress, catecholamines and cortisol stimulate the expression of CAMs, leading to an increase of sCAMs in circulation. Therefore, it is presumed that the magnitude of changes in catecholamines and cortisol induced by physical activity can lead to a change in sCAMs, which in turn may be dependent upon the intensity of exercise performed.

Exercise can be of great benefit for preventing CVD and possibly provide a positive impact on vascular health. However, there is controversy regarding the effects of different intensities of exercise on the responses of CAMs. Despite substantial evidence suggesting that exercise may influence CAMs, it is still unclear how exercise at different intensities (lower vs higher) and exercise-induced stress hormones affect CAMs. Moreover, the majority of the previous studies focused on investigating the effect of exercise on CAMs in recreational athletes and trained individuals rather than in sedentary overweight or obese individuals. In obesity, excessive adipose tissue produces a number of inflammatory cytokines that contribute to acute and chronic inflammation. In addition, the major risk factors for CVD include male sex, physical inactivity, and obesity. Therefore, the purpose of the current study was to examine the response of sCAMs (sICAM-1, sVCAM-1, and sE-selectin) in sedentary obese men following exercise performed at different intensities and to determine whether there was any relationship between exercise-induced stress hormones and sCAMs.

Patients and methods

Participants

Fifteen physically inactive obese men (body mass index ≥30 kg/m²) between the ages of 18 and 30 years, who had not engaged in any form of physical activity (<2 days per week) for at least 6 months prior to the study, volunteered for this study. All participants were at low risk for developing CVD and had no contraindications to exercise as outlined by the American College of Sports Medicine. The study protocol was reviewed and approved by the Baylor University Institutional Review Board for human subjects, and all participants signed written informed consent documents prior to performing any study protocols.

Maximal heart rate test protocol

The maximal heart rate (HRmax) test was conducted using a Monark cycle ergometer (Medgraphics, Ergomedic, Model No. 828E; Monark, St Paul, MN, USA) based on the protocol used by Keytel et al²⁴ to estimate the participants’ HRmax, which was defined as maximal heart rate achieved at the point of maximal exhaustion. During the HRmax test, the participants warmed up on the bicycle for 2 minutes with a workload of 2 W/kg of body weight. After the warm-up, the participants began cycling at 70 revolutions per minute (rpm) with an exercise intensity of 3.33 W/kg of body weight for 150 seconds (stage 1). Upon completion of stage 1, the workload increased by an additional 50 W for another 150 seconds (stage 2). Thereafter, the workload increased by 25 W every 150 seconds until the participants could no longer maintain 70 rpm or volitionally stopped the test. The participants wore a heart rate monitor (Polar Electro Inc., Lake Success, NY, USA) around the chest and heart rate was recorded every minute throughout the test.

Study design and exercise trials

In a randomized, crossover design, each participant performed two exercise trials consisting of the lower- (50% of HRmax) and higher-intensity exercise (80% of HRmax) trials. Each exercise trial was randomly assigned and performed at least 7 days apart to allow the participants to fully recover and return to basal conditions. During each exercise trial, the participants exercised on the cycle ergometer until they expended 300 kcal, which was calculated using the following equation by Keytel et al²⁴ [gender × (-55.0969 + 0.6309 \times \text{heart rate} – 0.1988 \times \text{body weight (kg)} + 0.2017 \times \text{age}) + (1–gender) × (-20.4022 + 0.4472 \times \text{heart rate} – 0.1263 \times \text{body weight (kg)} + 0.074 \times \text{age})], where gender = 1 for males and 0 for females]. The participants refrained from any types of physical activity until completing the last blood draw at 24-hour following each exercise trial.

Blood sample collection

Overnight fasting blood samples were collected at baseline (PRE), immediately postexercise (IPE), 1-hour PE (1-h PE), and 24-hour PE (24-h PE) for each exercise trial. After 10 minutes of resting in a chair, venous blood from the antecubital vein was collected into a serum separator tube. Immediately after blood draws, blood samples remained at room temperature for 20 minutes to be clotted and then were centrifuged (1,000 g) for 20 minutes to separate the serum. Aliquots of serum samples were pipetted into 1.5-mL polypropylene tubes and immediately frozen at −80°C for later analyses.
Analysis of sCAMs and stress hormones

Serum samples were analyzed in duplicate for sICAM-1 and sVCAM-1 (Cat# ZF000000A; Bio-Rad Laboratories, Hercules, CA, USA) by a multiplex flow immunoassay. The concentrations of sICAM-1 and sVCAM-1 were determined by median fluorescent intensity using a five-parameter logistic curve-fit. Serum samples in duplicate were analyzed for E-selectin (Kit# EK0501; BOSTER, Fermont, CA, USA) by an enzyme-linked immunosorbent assay. The optical density was read at 450 nm using a spectrophotometer (SmartSpec Plus; Bio-Rad Laboratories) and the concentration was determined by a semi-log fit. Serum samples were analyzed in duplicate for E and NE (Kit# EA613/192; DLD Diagnostika GmbH, Adlerhost, Hamburg, Germany) and cortisol (Kit# DKO001; DiaMetra, Milan, Italy) by enzyme-linked immunosorbent assay. The optical density was read at 450 nm by the spectrophotometer (SmartSpec Plus; Bio-Rad Laboratories) and the concentrations were determined by a four-parameter logistic fit. Each assay was performed as instructed by the manufacturer’s assay procedure. Intra- and interassay %CV for each marker was as follows: sICAM-1 (5.91% and 7.25%), sVCAM-1 (6.32% and 7.97%), E-selectin (4.82% and 4.54%), E (8.12% and 9.01%), NE (5.85% and 6.67%), and cortisol (5.80% and 4.05%).

Statistical analyses

All statistical analyses were performed using the IBM Statistical Package for the Social Sciences 23.0 (IBM SPSS, Armonk, NY, USA) and reported as a mean ± standard error (SE) unless stated otherwise. The Shapiro–Wilk test with the normal Q–Q plots was used to examine normality, and all data were log-transformed if normality did not exist. Effect sizes were calculated for each outcome variable using Cohen’s d. A 2 × 4 (intensity × time) factorial analysis of variance (ANOVA) with repeated measures was used to examine the effects of exercise intensity (lower vs. higher) on sCAMs (sICAM-1, sVCAM-1, E-selectin) over the 24-hour period (PRE, IPE, 1-h PE, and 24-h PE). Since it has been well documented that exercise typically yields an acute increase in stress hormones during exercise, which in turn return to the pre-exercise condition within a couple of hours upon completion of exercise,25 the current study only included the blood samples of PRE, IPE, and 1-h PE to examine the responses of stress hormones (E, NE, and cortisol). A 2 × 3 (intensity × time) ANOVA with repeated measures was used to examine the responses of E, NE, and cortisol at PRE, IPE, and 1-h PE. If there were significant main effects on time, Bonferroni pairwise comparisons were conducted as post hoc tests to locate the significant mean differences. If a significant interaction between intensity and time was found, a follow-up simple effects test with Bonferroni pairwise comparisons was conducted as a post hoc test. Additionally, linear regression analysis was conducted to examine the relationship between sCAMs and stress hormones. A p-value <0.05 was set for the statistical significance.

Results

The anthropometric data of the participants are shown in Table 1. The average HR and duration for each lower- and higher-intensity exercise trial were 98.80 ± 1.82 bpm (42.00 ± 1.77 minutes) and 154.20 ± 2.76 bpm (19.00 ± 0.53 minutes), respectively. One of the major findings of the present study was that either lower- or higher-intensity of aerobic exercise did not significantly change sICAM-1 or sVCAM-1 (Table 2), while E-selectin was significantly (p = 0.04) reduced following exercise, regardless of intensity. The post hoc test indicated that E-selectin at 1-h PE (10.25 ± 1.07 ng/mL) was significantly lower than PRE (12.22 ± 1.39 ng/mL, p = 0.01) and IPE (12.20 ± 1.55 ng/mL, p = 0.02) (Figure 1). There were no significant changes in E or NE, whereas cortisol had a significant interaction effect between exercise intensity and time (Table 3). The further analysis post hoc revealed that cortisol at IPE (187.52 ± 31.09 ng/mL, p = 0.01) and at 1-h PE (156.24 ± 31.11 ng/mL, p = 0.01) were significantly lower than PRE (259.74 ± 23.07 ng/mL), respectively, during the lower-intensity exercise trial. In addition, cortisol at 1-h PE (189.35 ± 31.11 ng/mL) significantly decreased by 27.76% (p = 0.01) from IPE (262.12 ± 31.09 ng/mL) following higher-intensity exercise (Figure 2).

Table 1 displays the relationship between sCAMs and exercise-induced stress hormones following the lower- and higher-intensity exercise trials.
Table 2 Changes in sCAMs over 24 hours

<table>
<thead>
<tr>
<th>Intensity</th>
<th>Variables</th>
<th>PRE</th>
<th>IPE</th>
<th>1-hour PE</th>
<th>24-hour PE</th>
</tr>
</thead>
<tbody>
<tr>
<td>LI</td>
<td>sICAM-1 (ng/mL)</td>
<td>126.0±17.27</td>
<td>120.4±8.86</td>
<td>118.6±7.99</td>
<td>126.1±8.25</td>
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<td></td>
<td>sVCAM-1 (ng/mL)</td>
<td>119.0±7.00</td>
<td>112.5±8.25</td>
<td>107.0±6.21</td>
<td>116.1±5.83</td>
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<tr>
<td></td>
<td>sE-selectin (ng/mL)</td>
<td>12.99±8.85</td>
<td>11.36±6.55</td>
<td>9.60±4.81</td>
<td>10.48±6.67</td>
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<tr>
<td>HI</td>
<td>sICAM-1 (ng/mL)</td>
<td>127.5±7.27</td>
<td>137.9±8.86</td>
<td>129.5±7.99</td>
<td>127.8±8.25</td>
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<tr>
<td></td>
<td>sVCAM-1 (ng/mL)</td>
<td>119.8±7.00</td>
<td>130.8±8.25</td>
<td>122.7±6.21</td>
<td>121.3±5.83</td>
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<tr>
<td></td>
<td>sE-selectin (ng/mL)</td>
<td>11.45±6.13</td>
<td>13.04±10.10</td>
<td>10.90±6.76</td>
<td>12.14±5.83</td>
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</tbody>
</table>

Note: All data are presented as mean ± SE. Abbreviations: sCAM, soluble cell adhesion molecule; LI, lower-intensity; HI, higher-intensity; sICAM-1, soluble intracellular adhesion molecule-1; sVCAM-1, soluble vascular cellular adhesion molecule-1; sE-selectin, soluble endothelial selectin; PRE, pre-exercise; IPE, immediately postexercise; PE, postexercise; SE, standard error.

Figure 1 sE-selectin concentration at each time point.
Notes: *A significant difference between IPE and 1-hour PE (p=0.029); **a significant difference between baseline and IPE (p=0.001).
Abbreviations: sE-selectin, soluble endothelial selectin; IPE, immediately postexercise; PE, postexercise.

Table 3 Changes in stress hormones at each time point by exercise intensity

<table>
<thead>
<tr>
<th>Intensity</th>
<th>Variables</th>
<th>PRE</th>
<th>IPE</th>
<th>1-hour PE</th>
<th>24-hour PE</th>
</tr>
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<tr>
<td>LI</td>
<td>E (pg/mL)</td>
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<td></td>
<td>NE (pg/mL)</td>
<td>257.97±34.99</td>
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<td>299.87±26.46</td>
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<td></td>
<td>COR (ng/mL)</td>
<td>259.74±23.07</td>
<td>187.52±31.09</td>
<td>156.24±31.11</td>
<td></td>
</tr>
<tr>
<td>HI</td>
<td>E (pg/mL)</td>
<td>44.29±8.02</td>
<td>48.35±10.72</td>
<td>38.05±9.29</td>
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<tr>
<td></td>
<td>NE (pg/mL)</td>
<td>214.71±41.40</td>
<td>335.67±59.08</td>
<td>257.39±31.31</td>
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</tr>
<tr>
<td></td>
<td>COR (ng/mL)</td>
<td>243.75±23.07</td>
<td>262.12±31.09</td>
<td>189.35±31.11</td>
<td></td>
</tr>
</tbody>
</table>

Note: All data are presented as mean ± SE. *Significantly different from PRE (p=0.001); †Significantly different from IPE (p=0.01).
Abbreviations: LI, lower-intensity; HI, higher-intensity; E, epinephrine; NE, norepinephrine; COR, cortisol; PRE, pre-exercise; IPE, immediately postexercise; PE, postexercise; SE, standard error.

Discussion
The present study examined the acute responses of sCAMs and their association with exercise-induced stress hormones in sedentary obese men following a single bout of aerobic exercise at two different intensities (lower: 50% and higher: 80% HRmax). In the current study, either lower- or higher-intensity exercise did not significantly alter sICAM-1 and sVCAM-1. A similar result has been reported by several previous studies that utilized a variety of subject populations including physically active and sedentary men along with patients with type-2 diabetes. Several studies, however, have reported an acute increase in sICAM-1 and sVCAM-1 immediately following exercise, which in turn returned to the pre-exercise levels within several hours postexercise. For instance, healthy men showed increased sICAM-1 by higher-intensity exercise trials. Cortisol and sICAM-1 had a weak, but significant, negative relationship (r²=0.34, p=0.02) at 1-h PE during the lower-intensity exercise trial, while cortisol and sVCAM-1 had a positive relationship (r²=0.36, p=0.02) at IPE during the higher-intensity exercise trial. Cortisol and sE-selectin had a negative relationship at PRE during the lower- (r²=0.31, p=0.03) and higher-intensity (r²=0.70, p=0.01) exercise trials. In addition, E and sE-selectin had a positive relationship (r=0.56, p=0.01) at 1-h PE during the higher-intensity exercise trial.
Exercise on cell adhesion molecules

24% immediately following a maximal exercise test, which then returned to the pre-exercise level 1-h PE.28 Untrained healthy males demonstrated increased both sICAM-1 and sVCAM-1 by 10% immediately following maximal cycling exercise, whereas only a minor elevation (sICAM-1: 5% and sVCAM-1: 1%) was observed following moderate-intensity exercise (60% of maximal cycling exercise).17 Most of the studies including the current study that have reported no significant changes in sICAM-1 or sVCAM-1 utilized a moderate to submaximal exercise intensity protocol with relatively short duration,16,17,21 whereas it seems that maximal-intensity exercise causes a transitory elevation of sICAM-1 and sVCAM-1. Thus, aerobic exercise at any intensity, except for maximal intensity, may not negatively influence sICAM-1 or sVCAM-1.

Only a few studies have previously examined the responses of sE-selectin to exercise and most of these studies used untrained nonobese men and reported no changes in sE-selectin.16,17,26 However, it should be noted that sedentary obese men in the current study significantly decreased sE-selectin following exercise, regardless of intensity. This result was similar to the study by Roberts and associates that reported a reduction in sE-selectin following moderate-intensity aerobic exercise accompanied with a dietary intervention in overweight/obese male patients with type 2 diabetes.27 sE-selectin mediates the initial step of leukocyte transmigration and rolling,28 and inflammatory stimuli are known to induce maximum expression of E-selectin on the surface of endothelial cells.29 Thus, it seems that aerobic exercise in obese individuals may improve endothelial health by lowering sE-selectin, which may further downregulate the activity of leukocyte transmigration and rolling.

The current literature is generally agreed that stress including physical exercise stimulates a secretion of stress hormones, such as catecholamines and cortisol, which could also influence sCAMs.7 However, we found no significant changes in catecholamines following either lower- or high-intensity exercise. A possible explanation for the lack of changes in catecholamines in response to exercise may be related to our participants’ characteristic – obese men. Indeed, obese individuals had significantly lower responses of catecholamines than the nonobese, both at rest and during exercise, possibly due to obesity-mediated decreases in adrenergic receptor sensitivity.30 Additionally, the current study observed a positive relation between E and sE-selectin at 1-h PE during higher-intensity exercise. This result may be associated with exercise-induced beta-adrenergic stimulation since it has been reported that the expression of sE-selectin is mainly related to a response of E to exercise. It has been reported that exercise may increase the recruitment of

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### Table 4 Relationships between sCAMs and stress hormones

<table>
<thead>
<tr>
<th>Intensity</th>
<th>Time</th>
<th>Predictor</th>
<th>sICAM-1</th>
<th>p-value</th>
<th>R² (%)</th>
<th>sVCAM-1</th>
<th>p-value</th>
<th>R² (%)</th>
<th>sE-selectin</th>
<th>p-value</th>
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<tbody>
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<td>LI</td>
<td>PRE</td>
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<td></td>
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<td></td>
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<td>COR</td>
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<td>–31.5</td>
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<td>1-hour PE</td>
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<td>–4.6</td>
<td>0.44</td>
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Note: *p<0.05.

Abbreviations: sCAMs, soluble cell adhesion molecules; LI, lower-intensity; HI, higher-intensity; sICAM-1, soluble intracellular adhesion molecule-1; sVCAM-1, soluble vascular adhesion molecule-1; sE-selectin, soluble endothelial selectin; E, epinephrine; NE, norepinephrine; COR, cortisol; PRE, pre-exercise; IPE, immediately postexercise; PE, postexercise.
leukocytes through beta-adrenergic stimulation that interacts with E in endothelial tissue and induces an increase in the expression of E-selectin. One previous study also demonstrated that beta-adrenergic stimulation was required to upregulate E-selectin, suggesting that E is directly related to changes in sE-selectin.31

Conversely, cortisol was significantly decreased at IPE and 1-h PE from baseline during lower-intensity exercise, while cortisol at IPE was significantly higher than at 1-h PE during higher-intensity exercise. This result was similar to the previous study, which suggested that an increase in cortisol following a brief exercise was influenced by the intensity of exercise, since exercise intensity above 60% VO2max increases cortisol, while exercising below 50% VO2max seems to reduce cortisol mainly due to an enhanced elimination and suppressed secretion of cortisol.32

We found a statistically significant relationship between sCAMs and cortisol. However, only sVCAM-1 at IPE showed a positive relationship with cortisol at IPE during higher-intensity exercise. An exercise-induced increase in cortisol during higher-intensity exercise may be related to an increase in proinflammatory cytokines, which consequently increases in sVCAM-1,33 while, interestingly, anti-inflammatory cytokines modulate only sVCAM-1.34 Thus, the increase in cortisol immediately following higher-intensity exercise may cause an increase in sVCAM-1, primarily due to an exercise-induced increase in proinflammatory cytokines. Additionally, cortisol has an inhibitory effect on sICAM-1 by switching off the activated ICAM-1 gene through glucocorticoid receptors, and a low level of cortisol is also related to a decrease in inflammatory gene transcription.35 Thus, a low level of cortisol for a certain condition, such as low-intensity exercise, may suppress sICAM-1, and this may explain our result showing a negative relationship between cortisol and sICAM-1 at 1-h PE during lower-intensity exercise. One study has proposed a concentration-dependent, biphasic (both stimulatory and suppressive) effect of cortisol on inflammation. Effects of cortisol on a stimulus-induced inflammatory response can be either suppressive or stimulatory in a concentration- and time-dependent manner.36 Thus, our current results showing a positive relation with sVCAM-1 at IPE following higher-intensity exercise and the negative relation with sICAM-1 at 1-h PE following lower-intensity suggest that exercise-induced cortisol may have a biphasic effect on vascular inflammation and may play a central role in the regulation of sICAM-1 and sVCAM-1.

Conclusion
The findings of the current study suggest that an acute bout of exercise, regardless of intensity, lowers sE-selectin in sedentary obese men. It seems that higher volume and duration of exercise training may be necessary to significantly change both sICAM-1 and sVCAM-1. One of the important findings of the current study is that cortisol may be a key stress hormone playing an important role in regulating sICAM-1 and sVCAM-1. Future studies examining the mechanisms among pro- and anti-inflammatory cytokines, exercise-induced stress hormones, and CAMs are recommended to better understand the role of exercise in endothelial function.

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References


