ISL1 is upregulated in breast cancer and promotes cell proliferation, invasion, and angiogenesis

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Abstract: ISL1 plays a key role in several cancers, including pheochromocytoma, gastrointestinal, pancreatic, and lung tumors and bile duct carcinoma. In order to elucidate the role of ISL1 in breast cancer, we performed quantitative real-time polymerase chain reaction and Western blotting analysis, and we found that ISL1 was upregulated in breast cancer cells and tissues. Moreover, high expression of ISL1 was correlated with tumor size, metastasis, and poor prognosis. Colony formation analysis and CCK-8 analysis revealed that ISL1 facilitated breast cancer cell proliferation. In addition, wound healing analysis and transwell invasion analysis demonstrated that ISL1 played a role in cell migration and invasion. Interestingly, the expression of ISL1 was also associated with the expression of vascular endothelial growth factor (VEGF) in breast cancer, and ISL1 promoted angiogenesis in breast cancer. In conclusion, reducing the expression of ISL1 suppresses proliferation, migration, invasion, and angiogenesis in breast cancer, suggesting that ISL1 might serve as a novel molecular therapy target in breast cancer.

Keywords: ISL1, proliferation, metastasis, angiogenesis, breast cancer

Introduction

Breast cancer is a global problem; it has become one of the most common cancers among women in the world.¹ Treatment of breast cancer is associated with the diagnosis and specific characteristics of the breast carcinoma, including genetic mutations and hormone production.² Although there have been a lot of advances in medicine, early diagnosis and treatment of breast cancer is infrequent, and therefore, breast cancer is usually diagnosed in the advanced stages.³

ISL1, also known as Islet-1, is a LIM-homeodomain transcription factor.⁴ ISL1 has been found to play crucial roles in multiple tissue types, such as heart, kidneys, skeletal muscle, nervous system, and endocrine organs.⁵ Moreover, several studies also reveal that ISL1 plays a key role in differentiation, cell specification, and maintenance of phenotypes of the horizontal cells, cholinergic amacrine and ganglion in the retina.⁶ In addition, previous studies have suggested that ISL1 is highly expressed in rhabdomyosarcoma⁷ and gastric cancer,⁸ and associates with advanced tumor stage, tumor size, metastasis, and poorer overall survival.^{8,9} Recently, ISL1 has been reported to promote angiogenesis in human umbilical vein endothelial cells (HUVECs) through regulating the expression of vascular endothelial growth factor (VEGF).¹⁰ However, the detailed function of ISL1 in breast cancer remains unknown.

Angiogenesis takes part in the pathophysiology of ischemic cardiomyopathy, wound healing, atherosclerosis, stroke, and cancer. Many studies have suggested that the angiogenic factor VEGF is a new molecular treatment for diseases.¹¹ In addition, anti-VEGF therapy is a novel treatment of cancer through inhibiting angiogenesis.¹²

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To date, the main anti-VEGF antibodies are Macugen,¹³ Lucentis,¹⁴ and Avastin.¹⁵

Here, our study indicates that ISL1 is upregulated in breast cancer cell lines and tissue samples. Moreover, a high expression of ISL1 is corrected with tumor size, metastasis, and poor prognosis. Several functional experiments, including colony formation analysis, CCK-8 assay, wound healing assay, and transwell invasion analysis, demonstrate that ISL1 facilitates cell proliferation, migration, and invasion in breast cancer cells. In breast cancer, ISL1 also promotes angiogenesis. Blood vessels can supply nutrition for cancer cells, promoting cancer cell proliferation and metastasis, which might explain the reason why high expression of ISL1 is corrected with tumor size and metastasis.

Materials and methods

Specimens and ethics statement

A total of 108 pairs of adjacent normal tissue samples and breast cancer tissue samples were obtained from the Affiliated Hospital of Shandong Medical College from 2010 to 2015. This study was approved by the ethical committee of the Affiliated Hospital of Shandong Medical College. All patients provided written informed consent.

Cell culture

Human breast cancer cell lines MDA-MB-231 and MCF-7, human normal breast tissue cells MCF-10A, and HUVECs were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). MDA-MB-231 and MCF-7 cells were cultured in RPMI 1640 (GIBCO, Grand Island, NY, USA) or Dulbecco's Modified Eagle's Medium (DMEM; GIBCO), respectively, containing 1% Penicillin-Streptomycin and 10% fetal bovine serum (FBS; HyClone, Logan, UT, USA) at 37°C with 5% CO₂. MCF-10A cells were cultured in the Mammary Epithelial Cell Medium containing Mammary Epithelial Cell Growth Supplement and 1% Penicillin-Streptomycin (Sciencell, Carlsbad, CA, USA). The HUVECs were incubated in Endothelial Cell Medium (ECM; Sciencell) containing 1% Penicillin-Streptomycin, 1% Endothelial Cell Growth Supplement (ECGS), and 5% FBS.

Cell transfection

Cells were transfected with pCMV-Tag2B (vector) or pCMV-Tag2B-ISL1 using lipo2000 reagent (Thermo Fisher Scientific, Waltham, MA, USA) when cell confluence was 70%–80%. For knockdown ISL1, cells were transfected with scramble siRNA (SCR) or ISL1 siRNA (siISL1) using RNAimax reagent (Thermo Fisher Scientific) when cell

confluence was 30%–40%. After transcription for 48 hours, cells were collected and used for further experiments.

Western blotting

The tissue sample and cultured cells were lysed using RIPA buffer (Beyotime, Jiangsu, People's Republic of China) containing protease inhibitors and cocktail (Boehringer, Mannheim, Germany). The protein concentration was measured using BCA kit (Thermo Fisher Scientific). A 40 µg protein was resolved with a 10% sodium dodecyl sulfate–polyacrylamide gel (SDS-PAGE), transferred to a polyvinylidene fluoride (PVDF) membrane, and incubated with 5% skimmed milk at room temperature for 1 hour. The membranes were washed with PBST three times and then incubated with indicated antibodies at 4°C overnight. The membranes were washed with PBST three times, followed by incubation with HRP-conjugated secondary antibodies at room temperature for 1 hour. Finally, the enhanced chemiluminescence (ECL) reagent (Thermo Fisher Scientific) was used to detect the specific protein band. All the experiments were repeated at least three times. The antibodies were as follows: ISL1 antibody (1:500; Abcam, Cambridge, MA, USA), VEGF antibody (1:1,000; Abcam), and β-actin antibody (1:5,000; Sigma, Danvers, MA, USA).

CCK-8 analysis

CCK-8 analysis was performed to determine the cell proliferation. ISL1 was overexpressed or knocked down in MCF-7 cells; after a transcription of 48 hours, a total of 3×10³ MCF-7 were placed in 96-well plates. Every 24 hours, each 100 µL media was added 10 µL CCK-8 reagent (Beyotime, Jiangsu, People's Republic of China) and cells were incubated at 37°C for 1 hour. The absorbance of OD 450 nm each clearly indicated the number of surviving cells. All the experiments were repeated at least three times.

Quantitative real-time polymerase chain reaction CR (qRT-PCR)

Total RNA of tissue samples and cells were prepared using TRIzol reagent (Thermo Fisher Scientific). M-MLV Reverse Transcriptase kit (TransGen, Beijing, People's Republic of China) was used to synthesize cDNA according to the instructions. Next, SYBR (Roche Diagnostics, Mannheim, Germany) was utilized to perform qRT-PCR. The primers were as follows: ISL1 forward: 5'-TGGACATTACTCCCTCTTACAG-3', reverse: 5'-CCCGTACAACCTGATATAATCTC 775 23-3'; VEGF forward: 5'-GACATCTTCCAGGAGTACCC-3', reverse: 5'-TCTTTCTTTGGTCTGCATTAC-3'; GAPDH

forward: 5'-ATTTCCTGGTATGACAACGA-3', and reverse: 5'-TTGATGGTACATGACAAGGTG-3'. GAPDH was used as an internal control. All the experiments were repeated at least three times.

Transwell invasion analysis

The transwell invasion analysis was performed to determine the effect of ISL1 on cell invasion. In brief, each transwell invasion chambers (8- μ m pore size; Millipore Corporation, Billerica, MA, USA) was coated with 100 μ L Matrigel (BD Biosciences, San Diego, CA, USA) according to the manufacturer's protocol. 3×10^4 MDA-MB-231 cells were resuspended with serum-free DMEM medium and placed on the top of Matrigel-coated chambers; the lower chambers were filled with 600 μ L DMEM containing 10% FBS. The cells were incubated at 37°C with 5% CO₂ for 24 hours and stained with 0.5% violet crystal, and then a cotton swab was used to remove the cells that remained on the top of the filter. Finally, the number of invaded cells was counted under a light microscope. All the experiments were repeated at least three times.

In vitro tube formation analysis

In vitro tube formation analysis was performed using growth factor-reduced Matrigel (BD Biosciences), as previously described.¹⁶ Briefly, a 200 μ L growth factor-reduced Matrigel was coated in the 24-well plates and then solidified at 37°C for 30 minutes. HUVECs were cultured with CM from MCF-7 cells that were transfected with vector, ISL1 or SCR, siISL1. After incubation for 24 hours, the number of tubes formed was counted under a light microscope. All the experiments were repeated at least three times.

Statistical analysis

All experiments were repeated at least three times. All data were analyzed by SPSS 19.0 software (IBM Corp., Armonk, NY, USA) and presented as mean $\pm$ standard deviation (SD). Student's *t*-test was utilized to assess the statistical significance between two groups. The Kaplan–Meier method was used to detect the prognostic significance of ISL1. **P* < 0.05 was considered significant.

Result

ISL1 is upregulated in breast cancer, and the expression of ISL1 is corrected with tumor size and metastasis

In order to determine the role of ISL1 in breast cancer, we detected the expression of ISL1 in human breast carcinoma cells, MDA-MB-231 and MCF-7, and in human normal

breast tissue cells. MCF-10A was used as control. As shown in Figure 1A and B, both the protein and mRNA levels of ISL1 were significantly upregulated in MDA-MB-231 and MCF-7 cells (Figure 1A and B). Next, in order to determine the expression of ISL1 in breast cancer patients, we collected 108 pairs of breast cancer tissue samples and adjacent normal tissue samples. The mRNA and protein levels of ISL1 were detected by qRT-PCR and Western blotting, respectively. The results suggested that ISL1 was upregulated in the tumor tissue samples (Figure 1C and D). Moreover, we found that a high expression of ISL1 was corrected with pathological grade, tumor size, and metastasis, but there are no association between ISL expression and age, differentiation (Table 1). Moreover, we analyzed the expression of ISL1 on the bioinformatics analysis website (<http://www.cbioportal.org>), and the result indicated that ISL1 amplified in breast cancer (Figure 1E). Next, we analyzed the correlation of ISL expression and the survival curve of breast cancer patients. The results revealed that the high expression of ISL1 predicted a poor prognosis (*P* = 0.035, Figure 1F).

ISL1 facilitates proliferation of breast cancer cells

Because of the correlation between ISL1 expression and tumor size, we assumed that ISL1 might promote breast cancer cell proliferation. In order to verify our hypothesis, we performed colony formation analysis and CCK-8 analysis to analyze the effect of ISL1 on cell proliferation. ISL1 was overexpressed or knocked down in MCF-7 cells. The expression of ISL1 was detected by qRT-PCR and Western blotting, respectively (Figure 2A and B). The results of colony formation analysis suggested that ectopic expression of ISL1 strongly improved the number of colonies; however, inhibition of ISL1 obviously decreased the number of colonies (Figure 2C). In addition, CCK-8 analysis was performed every 12 hours, and the absorbance of OD 450 nm indicated the number of surviving cells. As shown in Figure 2D, overexpression of ISL1 significantly increased the proliferation rate of cells compared with vector group (Figure 2D). However, inhibition of ISL1 inhibited the proliferation rate of cells compared with SCR group (Figure 2D). To sum up, our work reveals that ISL1 plays an important role in cell proliferation.

ISL1 correlates with VEGF expression in MCF-7 cells

A previous study has found that the expression of ISL1 was correlated with VEGF expression.¹⁰ To further explore the function of ISL1 in breast cancer, we also detected the

Figure 1 ISL1 is upregulated in breast cancer, and the expression of ISL1 is corrected with tumor size and metastasis.

Notes: (A) The mRNA level of ISL1 in MDA-MB-231 and MCF-7 cells was determined by qRT-PCR; MCF-10A was used as the control group. $*p < 0.05$, MDA-MB-231 or MCF-7 vs MCF-10A. (B) The protein level of ISL1 in MDA-MB-231 and MCF-7 cells was determined by Western blotting; MCF-10A was used as the control group. (C) The protein level of ISL1 in breast cancer tissue samples was determined by Western blotting; adjacent normal tissue samples were used as the control group. $*p < 0.05$, tumor tissues vs normal tissues. (D) The mRNA level of ISL1 in breast cancer tissue samples was determined by qRT-PCR; adjacent normal tissue samples were used as the control group. (E) The expression of ISL1 in breast cancer was assessed on a website (<http://www.cbioportal.org>). MBL (sickkids 2016): Mutational profiles of metastatic breast cancer (France, 2016). (F) The survival curve in 108 breast cancer patients according to ISL1 expression was analyzed by the Kaplan-Meier method.

Abbreviations: Cum, cumulative; qRT-PCR, quantitative real-time polymerase chain reaction.

correction between ISL1 and VEGF. We overexpressed or knocked down ISL1 in MCF-7 cells and detected the protein and mRNA levels of VEGF by qRT-PCR and Western blotting, respectively. As shown in Figure 3A and B, the

mRNA level and the protein level of VEGF were regulated by ISL1. Ectopic expression of ISL1 promoted the expression of VEGF; however, inhibition of ISL1 suppressed the expression of VEGF (Figure 3A and B).

Table 1 Clinicopathologic variables in 108 breast cancer patients

Variables	Patients (n=108)	ISL1 protein expression		P-value
		Low (n=42)	High (n=66)	
Age (years)				0.982
<40	59	23	36	
≥40	49	19	30	
Tumor size (diameter)				<0.001
Small (≤3 cm)	54	30	24	
Large (≥3 cm)	54	12	42	
Pathological grade				0.007
I-II	47	25	22	
III-IV	61	17	44	
pT status				<0.001
pT1	48	30	18	
pT2-4	60	12	48	
pN status				0.018
pN0	44	23	21	
pN1-2	64	19	45	
Metastasis				0.041
Yes	57	17	40	
No	51	25	26	
Differentiation				0.059
Well/moderate	52	25	27	
Poor	56	17	39	

Downregulation of ISL1 suppresses angiogenesis in breast cancer

VEGF plays a key role in angiogenesis, and so, we detected the function of ISL1 on angiogenesis in breast cancer. HUVECs were incubated with conditional media (CM) from MCF-7 cells which were treated with vector, ISL1 or SCR, and siISL1. After incubation for 24 hours, the number of tubes was counted under a light microscope. The capillary-like areas and network structures were separately analyzed using Image J software. The relative capillary-like structure density was measured, and the number of capillary-like areas of the control group was used as 100%. As shown in Figure 4A and B, after transfection with siISL1, the number of tubes was obviously lower than that of the SCR group (Figure 4A and B). While ISL1 was overexpressed, the number of tubes was obviously higher than that of the vector group (Figure 4A and B). Taken together, our work suggests that ISL1 facilitates angiogenesis in breast cancer.

ISL1 facilitates breast cancer cells migration and invasion

Our previous work revealed that the expression of ISL1 was also correlated with metastasis. So, we detected the effect

of ISL1 on cell migration and invasion by wound healing and transwell invasion assays. MDA-MB-231 cells were transfected with vector, ISL1 or SCR, and siISL1, and the expression of ISL1 was detected by qRT-PCR and Western blotting (Figure 5A and B). The results of wound healing assay demonstrated that the siISL1 decreased inclosure of cells compared with the SCR group ($P<0.05$); however, ectopic expression of ISL1 increased the inclosure of cells, indicating a role for ISL1 in the migration of breast cancer cells (Figure 5C). Subsequently, in order to substantiate this observation, we performed a transwell invasion assay to determine the role of ISL1 on invasion of MDA-MB-231 cells. As shown in Figure 5D, ectopic expression of ISL1 increased the number of invaded cells compared with the control group, and ISL1 inhibition decreased the number of invaded cells (Figure 5D). Together, these results show that ISL1 facilitates breast cancer cells migration and invasion.

ISL1 promotes MRI Phenotyping detection

Since ISL1 facilitated angiogenesis in breast cancer, subsequently, we also performed MRI assay. The results suggested that the permeability-surface area product was obviously higher in ISL1-overexpressed group than that in the vector group; however, the permeability-surface area product was almost undetectable when ISL1 was knocked down (Figure 6).

Discussion

ISL1 has been found to play key roles in various organs and tissues.^{17,18} The expression of ISL1 is closely related to the occurrence and progression of various tumors. Moreover, ISL1 expression is high in several cancers, including pheochromocytoma,¹⁹ gastrointestinal, pancreatic and lung tumors,²⁰ and bile duct carcinoma.²¹ These studies suggested the relationship between tumorigenesis and ISL1. Recently, ISL1 has been reported to promote angiogenesis in HUVECs through regulating VEGF.¹⁰ Although a previous report indicated that ISL1 was downregulated in breast cancer,²² further details about the function of ISL1 in breast cancer remain unknown. In order to explore the function of ISL1 in breast cancer, we determined the expression of ISL1 in breast cancer cell lines and tissue sample compared with human normal breast cell lines and adjacent normal tissue samples, respectively. To our surprise, we found that ISL1 was upregulated in breast cancer, which was contrary to previous findings; the different results might be due to an antibody or some difference in the way in which the experiment was conducted.

Figure 2 ISL1 facilitates proliferation of breast cancer cells.

Notes: (A) MCF-7 cells were treated with vector, ISL1 or SCR, ISL1 siRNA (siSL1), respectively. The mRNA level of ISL1 was detected by qRT-PCR. (B) ISL1 was overexpressed or knocked down in MCF-7 cells; the protein level of ISL1 was detected by Western blotting. (C) ISL1 was overexpressed or knocked down in MCF-7 cells; the colony formation analysis was performed to detect the effect of ISL1 on cell proliferation. The graph indicated the relative number of colonies. (D) ISL1 was overexpressed or knocked down in MCF-7 cells; the CCK-8 assay was performed to assess the proliferation ability of MCF-7 affected by ISL1. * $p < 0.05$, ISL1 vs vector, siSL1 vs SCR.

Abbreviations: OD, optical density; qRT-PCR, quantitative real-time polymerase chain reaction; SCR, scramble siRNA.

Figure 3 ISL1 correlates with VEGF expression in MCF-7 cells.

Notes: (A) MCF-7 cells were treated with vector, ISL1 or SCR, ISL1 siRNA (siSL1), respectively. The mRNA level of VEGF was detected by qRT-PCR. * $p < 0.05$, ISL1 vs vector, siSL1 vs SCR. (B) ISL1 was overexpressed or knocked down in MCF-7 cells; the protein level of VEGF was detected by Western blotting.

Abbreviations: qRT-PCR, quantitative real-time polymerase chain reaction; SCR, scramble siRNA; VEGF, vascular endothelial growth factor.

Figure 4 Downregulation of ISL1 suppresses angiogenesis in breast cancer.

Notes: (A) MCF-7 cells were treated with vector, ISL1 or SCR, ISL1 siRNA (silSL1), respectively. After transfection for 48 h, the HUVECs were incubated with conditional media (CM) from MCF-7 cells. The number of tubes formed indicates the effect of ISL1 on angiogenesis. Magnification: 40 $\times$, scale bars =250 μ m. (B) The graph indicates the relative number of tubes. * p <0.05, ISL1 vs vector, silSL1 vs SCR.

Abbreviations: HUVECs, human umbilical vein endothelial cells; SCR, scramble siRNA.

Figure 5 (Continued)

Figure 5 ISL1 facilitates breast cancer cells migration and invasion.

Notes: (A) MDA-MB-231 cells were treated with vector, ISL1 or SCR, ISL1 siRNA (siSL1), respectively. The mRNA level of VEGF was detected by qRT-PCR. (B) ISL1 was overexpressed or knocked down in MDA-MB-231 cells; the protein level of VEGF was detected by Western blotting. (C) Wound healing analysis was used to detect the role of ISL1 in cell migration. ISL1 was overexpressed or knocked down in MDA-MB-231 cells. The graph indicates the relative distance of migration. Scale bars = 200 μ m; * p < 0.05, ISL1 vs vector, siSL1 vs SCR. (D) Transwell invasion analysis was used to detect the role of ISL1 in cell invasion. ISL1 was overexpressed or knocked down in MDA-MB-231 cells. The graph indicates the relative number of invaded cells. Magnification: 20 $\times$, scale bars = 200 μ m. * p < 0.05, ISL1 vs vector, siSL1 vs SCR.

Abbreviations: qRT-PCR, quantitative real-time polymerase chain reaction; SCR, scramble siRNA; VEGF, vascular endothelial growth factor.

In addition, the bioinformatics analysis also revealed that ISL1 was amplified in breast cancer. Moreover, the high expression of ISL1 was positively associated with tumor size and metastasis, suggesting that ISL1 played a key role in cell proliferation and metastasis. Previous work showed that ISL1 suppressed cell proliferation in cancer stem cells, but our work revealed that ISL1 promoted cell proliferation in breast cancer cells.

To further investigate the underlying mechanism, we cultured HUVECs with CM from MCF-7 cells that were transfected with vector, ISL1 or SCR, and siSL1 and detected the tube formation in vitro. We found that ISL1 inhibition obviously downregulated the expression of VEGF and decreased tube formation of the vascular endothelial cells. However, ectopic expression of ISL1 significantly

upregulated the expression of VEGF and increased tube formation of the vascular endothelial cells.

In summary, our work suggests that the expression of ISL1 is high in breast cancer, and its expression is associated with tumor size, metastasis, and poor prognosis. Moreover, inhibition of endogenous ISL1 expression suppresses the breast cancer cell proliferation, migration, and invasion. In addition, ISL1 regulates angiogenesis in breast cancer. The function of ISL1 in promoting angiogenesis is related to VEGF expression. Therefore, this study reveals that ISL1 regulates the process of angiogenesis accompanied by VEGF, thereby supplying nutrition for cancer cells, and promoting cell proliferation and metastasis.

Disclosure

The authors report no conflicts of interest in this work.

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Figure 6 ISL1 promotes MRI phenotyping detection.

Note: MCF-7 vector and MCF-7 ISL1 overexpression tumor-bearing mice, or MCF-7 SCR and MCF-7 shISL1 tumor-bearing mice were utilized to compare functional MRI maps of the permeability-surface area product. * p < 0.05, ISL1 vs vector, siSL1 vs SCR.

Abbreviation: SCR, scramble siRNA.

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