

Measurement of tumor necrosis factor- α , leukotriene B₄, and interleukin 8 in the exhaled breath condensate in patients with acute exacerbations of chronic obstructive pulmonary disease

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Background: Assessment of airway inflammation in the clinical course of acute exacerbations of chronic obstructive pulmonary disease (AECOPD) may advance our understanding of the pathogenesis and treatment.

Objectives: To assess airway inflammation in patients during the course of AECOPD by serial analyses of their exhaled breath condensates (EBC).

Methods: Twenty-six patients with AECOPD (22 males, mean[SD] percentage predicted forced expiratory volume in one second (FEV₁) 44.8 [14.3]), 11 with stable COPD, and 14 age and sex-matched healthy controls were studied. Patients with AECOPD were treated with systemic steroid and antibiotic for 7 days. EBC was collected from each patient with AECOPD on Day 5, 14, 30, and 60 post-hospitalization using EcoScreen (VIASYS Healthcare, USA) during tidal breathing over 10 minutes. Concentrations of tumor necrosis factor- α (TNF- α), leukotriene B₄ (LTB₄), and interleukin-8 (IL-8) were measured by enzyme-linked immunosorbent assay.

Results: The median (IQR) of TNF- α level on Day 5 was 5.08 (3.80–6.32) pg/ml, which was lower than on Day 14 (5.84 [4.91–9.14] pg/ml, $p = 0.017$), Day 30 (6.14 [3.82–7.67] pg/ml, $p = 0.045$), and Day 60 (5.60 [4.53–8.80] pg/ml, $p = 0.009$). On Day 60, subjects receiving inhaled corticosteroid (ICS) had a lower level of TNF- α than those who were not (4.82 [4.06–5.65] vs 7.66 [5.48–10.9] pg/ml, $p = 0.02$). EBC LTB₄ level did not change significantly during recovery from AECOPD whereas IL-8 was mostly undetectable.

Conclusions: EBC TNF- α level was low in patients receiving systemic steroid and antibiotic therapy for AECOPD. These findings suggest a potential role for serial EBC TNF- α for non-invasive monitoring of disease activity.

Keywords: COPD, exacerbation, exhaled breath condensate, TNF- α , LTB₄

Background

Assessment of airway inflammation in the clinical course of acute exacerbations of chronic obstructive pulmonary disease (AECOPD) may advance our understanding of the pathogenesis and the treatment effects in patients with AECOPD. Exhaled breath condensate (EBC) analysis has been developed in recent years as a novel tool to sample the lower airway lining fluid (Effros et al 2004; Hunt 2002; Montuschi and Barnes 2002; Ko et al 2007). Its noninvasive nature offers many advantages over invasive techniques such as bronchoalveolar lavage, bronchial biopsy and sputum induction, which are not so well tolerated by AECOPD patients. Because of the noninvasive nature of EBC collection, repeated measurements are feasible in the longitudinal follow-up of patients with AECOPD. Our group has previously demonstrated the repeatability of certain markers in the EBC of stable asthma and COPD patients (Ko et al 2006; Ko et al 2006; Leung et al 2005).

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Tumor necrosis factor- α (TNF- α) is a pleiotropic inflammatory cytokine produced by many different cell types. The main sources *in vivo* are stimulated monocytes, fibroblasts, and endothelial cells (Mukhopadhyay et al 2006). Leukotriene B4 (LTB4) is a potent chemoattractant of neutrophils, and it may be released by macrophages, epithelial cells and activated neutrophils (Ford-Hutchinson et al 1980). Interleukin 8 (IL-8), on the other hand, is largely produced by neutrophils and macrophages (Baggiolini et al 1989). Previous studies found significant increases in neutrophils and concentrations of TNF- α and IL-8 in spontaneous or induced sputum samples from patients with COPD when compared to smoking and non-smoking controls (Keatings et al 1996; Yamamoto et al 1997). During AECOPD, the concentrations of TNF- α and IL-8 may increase further in the induced sputum samples (Aaron et al 2001). In patients having AECOPD related to bacterial infection, increases in the concentration of neutrophil products such as myeloperoxidase and elastase, and the neutrophil chemoattractants (IL-8 and LTB4) have been observed in expectorated sputum, and their levels fell subsequently after antibiotic therapy (Crooks et al 2000). Limited data, however, are available on the changes in the EBC levels of TNF- α , LTB4, and IL-8 during AECOPD. This study aimed to assess the levels of these inflammatory mediators during the course of recovery from AECOPD by serial analyses of EBC.

Material and methods

Subject recruitment

Patients who had been admitted to the Prince of Wales Hospital with AECOPD were recruited for this study. AECOPD was defined when a patient with background COPD (National Heart, Lung and Blood Institute, World Health Organization 2006) presented with at least two of the following major symptoms (increased dyspnoea, increased sputum purulence, increased sputum volume) or one major and one minor symptom (nasal discharge/congestion, wheeze, sore throat, cough) for at least two consecutive days (Seemungal et al 1998; Patel et al 2002; Ko et al 2007). Subjects with pneumonic changes on the chest radiographs were excluded. Age and sex-matched COPD subjects who were clinically stable (without AECOPD for at least 10 weeks) and healthy, nonsmoking subjects were recruited as controls. Patients with AECOPD were managed by our standard care with administration of antibiotic (oral amoxicillin/clavulanic acid for 7 days) and systemic steroid (oral prednisolone 30mg daily for 7 days). All patients also received bronchodilators via a spacer device and if needed, oxygen therapy for respiratory failure. Current smokers (defined as cigarette smoking in the last 6 months) were excluded from

this study. Informed written consent was obtained from each subject and the study was approved by the research ethics committee of the Chinese University of Hong Kong.

Collection of exhaled breath condensate

EBC was collected using the EcoScreen (VIASYS Healthcare, Conshohocken, PA, USA) according to the manufacturer's instructions. The collection was carried out from 9–10 am. After rinsing their mouths, the recruited subjects breathed tidally through a mouthpiece that was connected through a unique one-way valve into a cooled collection tube where vapors, aerosols and moisture in the breath condensed along the walls of the tube. The design of the system prevented salivary contamination of EBC. Each subject was asked to breathe through the device, while wearing a nose-clip, for 10 minutes, and more than 1 mL of EBC could be collected from each subject. EBC was stored in 250 μ L aliquots immediately at -70°C until analysis. EBC was collected on Days 5, 14, 30, and 60 after hospitalization for subjects with AECOPD. For the stable COPD patients and normal controls, EBC collection was performed once.

Lung function measurement

Spirometry (post-bronchodilator) was performed on all subjects before each EBC collection with the Vitalograph (Model S, Buckingham, UK) spirometer according to the American Thoracic Society standards (American Thoracic Society. Standardization of spirometry – 1987). The updated predicted spirometry values for Hong Kong Chinese were adopted (Ip et al 2006).

Measurement of TNF- α , LTB4, and IL-8

The concentrations of TNF- α , LTB4, and IL-8 in EBC were measured in one batch by high-sensitivity sandwich enzyme immunoassays (TNF- α and IL-8 from BioSource International, Camarillo, CA, USA; LTB4 from Cayman Chemical Company, Ann Arbor, MI, USA) according to the manufacturers' instructions. The detection limits of TNF- α , LTB4, and IL-8 were 0.09, 4, and 0.02 pg/mL, respectively, and the intra-assay and inter-assay variability of all three markers were $<10\%$.

Intra-subject repeatability of TNF- α , LTB4, and IL-8 was assessed by collection of EBC in 6 stable COPD subjects. EBC was collected as described above at the same time (9–10 am) on 2 consecutive days.

Statistical analysis

Data were analyzed by the Statistical Package of the Social Science (SPSS) for Windows, Version 11.5 (SPSS Inc, Chicago, IL, USA). Demographic data of the subjects were presented

as means \pm standard deviation (SD). The levels of TNF- α , LTB4, and IL-8 were expressed as median and interquartile range (IQR). When the levels of these biomarkers were below the respective detection limits of the assays, the samples were assigned values at half of the lower detection limits for statistical analysis. Results were analyzed by two-tailed Students t-test or ANOVA for parametric variables and Mann-Whitney U-test or Kruskal-Wallis test for nonparametric numerical variables. The serial changes of TNF- α , LTB4, and IL-8 levels were compared at the different time points post-admission for AECOPD by Wilcoxon Signed Ranks test and Friedman test as appropriate. Correlation of lung function with the levels of EBC markers were assessed by Spearman's rank correlation test. The p values of <0.05 were considered as statistically significant. Bland and Altman's method was used to assess repeatability of TNF- α , LTB4, and IL-8 in the EBC (Bland and Altman 1986).

Results

Altogether 32 patients with AECOPD were initially recruited for this study. Two patients refused to return for EBC collection after Day 14 whereas 4 patients had another exacerbation before Day 60 and thus were excluded from the study. Finally, 26 subjects with AECOPD, 11 stable COPD patients, and 14 controls were included in the analysis. The demographics of the subjects are shown in Table 1. The AECOPD, stable COPD and control subjects were age and sex-matched. Twelve (46.1) and 5 (45.5%) subjects with AECOPD and stable COPD were on inhaled corticosteroid (ICS) whereas the median (IQR) beclomethasone-equivalent

doses were 1000 (950–2000) and 1000 (1000–1500) $\mu\text{g/day}$, respectively. None were on long term home oxygen therapy or oral steroid therapy when stable.

The detection rates of TNF- α , LTB4, and IL-8 in EBC of the subjects are shown in Table 2. The EBC levels of TNF- α and LTB4 are summarized in Table 3. The level of TNF- α in EBC was the lowest on Day 5 post-hospitalization after commencement of treatments with both systemic steroid and antibiotic. The levels of TNF- α at Day 5 was significantly lower than at Day 14 ($p = 0.017$), Day 30 ($p = 0.045$) and Day 60 ($p = 0.009$). There was however no significant difference in the levels of TNF- α between Day 14, Day 30, and Day 60 ($p = 0.89$). The level of TNF- α at Day 14 post-hospitalization for AECOPD was significantly higher than the normal controls ($p = 0.036$), but not different from the stable COPD subjects ($p = 0.26$). The EBC TNF- α level at the start of exacerbation (Day 5 post-exacerbation) was not significantly different from that during stable COPD ($p = 0.39$). There was no difference in the TNF- α level between the stable COPD and normal controls ($p = 0.41$). Figure 1 illustrates the serial levels of TNF- α during the course of AECOPD versus TNF- α levels for stable COPD and control subjects based on their single EBC collection.

The levels of LTB4 on Day 5, Day 14, Day 30, and Day 60 of AECOPD showed no statistically significant difference. In addition, there was no difference in the level of LTB4 at the different time points of the AECOPD when compared to the stable COPD patients or normal controls. IL-8 was

Table 1 Clinical characteristics of the subjects

	COPD patients with acute exacerbations (n = 26)	Stable COPD subjects (n = 11)	Normal controls (n = 14)
Age (yrs)	73.1 \pm 7.6	74.4 \pm 4.9	75.2 \pm 4.1
Sex (M/F)	22/6	8/3	9/5
Smoking history (pack years)	40.1 \pm 20.9	41.3 \pm 10.5	0
Body mass index (kg/m ²)	22.1 \pm 4.1	22.8 \pm 5.1	22.5 \pm 2.2
FEV ₁ (L)*	0.91 \pm 0.36	0.94 \pm 0.38	2.00 \pm 0.41
FVC (L)*	1.74 \pm 0.62	1.69 \pm 0.70	2.57 \pm 0.47
FEV ₁ /FVC (%)*	53.9 \pm 11.5	57.6 \pm 11.7	77.8 \pm 2.7 [#]
FEV ₁ (% predicted)*	44.8 \pm 14.3	49.4 \pm 19.6	104.1 \pm 17.0 [#]
FVC (% predicted)*	62.0 \pm 18.7	62.5 \pm 21.4	96.9 \pm 12.3 [#]
Current ICS use	12 (46.1)	5 (45.5)	0 (0)
Current theophylline use	7 (26.9)	4 (36.4)	0 (0)

Notes: Data were presented as mean \pm SD or number (%); *for the COPD patients with acute exacerbation, the lung function (post-bronchodilator) at Day 60 after admission to hospital was presented; [#]FEV₁% predicted, FVC % predicted and FEV₁/FVC ratio were higher in the normal controls when compared to COPD patients with acute exacerbations and stable COPD patients, $p < 0.001$.

Abbreviations: COPD, chronic obstructive pulmonary disease; FEV₁, forced expiratory volume in one second; FVC, forced vital capacity; ICS, inhaled corticosteroid; SD, standard deviation.

Table 2 Detection rates of TNF- α , LTB4 and IL-8 in exhaled breath condensate

	TNF- α (pg/ml)	LTB4(pg/ml)	IL-8(pg/ml)
	n (%)	n (%)	n (%)
AECOPD Day 5	26 (100)	23 (89)	1 (4)
AECOPD Day 14	26 (100)	26 (100)	4 (15)
AECOPD Day 30	26 (100)	25 (96)	3 (12)
AECOPD Day 60	26 (100)	25 (96)	2 (8)
Stable COPD	11 (100)	9 (82)	0 (0)
Normal controls	14 (100)	14 (100)	2 (14)

Abbreviations: AECOPD, acute exacerbations of chronic obstructive pulmonary disease; IL-8, interleukin-8; LTB4, leukotriene B4; TNF- α , tumor necrosis factor- α .

undetectable in most of the subjects and thus their results were not presented.

The levels of TNF- α and LTB4 did not correlate with the absolute and normal predicted values of forced expiratory volume in one second (FEV₁) at different time points of AECOPD. The levels of TNF- α and LTB4 on Day 60 post-hospitalization showed no significant difference in their levels when comparing those with FEV₁ \geq 50% against those with FEV₁ < 50% predicted normal, whereas the same observation was made among the stable state COPD subjects. On Day 60 post-hospitalization for AECOPD, subjects receiving ICS therapy had a lower level of TNF- α than those who were not on ICS (median [IQR] 4.82 [4.06–5.65] vs 7.66 [5.48–10.9] pg/ml, $p = 0.02$). The ICS was started before AECOPD and continued throughout the study. For those patients who were not on ICS before AECOPD, ICS was not introduced during the study period. The reduction of TNF- α as noted in the AECOPD patients on ICS, however, was not observed in the stable COPD patients, who had no change in their medications for 8 weeks before recruitment into this study. With reference to LTB4, there was no difference in its levels

between patients on ICS and those without ICS therapy for both the AECOPD subjects at Day 60 post-exacerbation and the stable COPD subjects.

The results of serial spirometry are shown in Table 4. There were no significant changes in the subjects' lung function in the course of AECOPD. Six stable COPD subjects (5 males) aged 66 to 74 years had EBC collected and measured for repeatability testing of the biomarkers. For TNF- α and LTB4 repeatability testing, the differences between most of the paired measurements laid within ± 2 SDs (mean differences -0.19 ± 2.01 , -0.83 ± 2.23 pg/ml for TNF- α and LTB4, respectively). Bland and Altman's plots of the repeatability tests for TNF- α and LTB4 are shown in Figure 2. IL-8 was only measurable in 1 of the 6 subjects and thus the results are not presented.

Discussion

To the best of our knowledge, this study is the first to assess inflammatory markers in the EBC serially over 60 days in patients hospitalized with AECOPD. We have shown that the TNF- α level in EBC was lower on Day 5 post-hospitalization for AECOPD when compared to the levels on Day 14, Day 30, and Day 60. The lower TNF- α level on Day 5 was likely secondary to the anti-inflammatory effect of systemic steroid and antibiotic. In contrast, there was no significant change in the serial LTB4 levels in EBC. Repeatability measurements of TNF- α and LTB4 among our stable COPD patients were satisfactory. Interestingly, the subjects receiving ICS had a lower EBC TNF- α level at Day 60 post-hospitalization for AECOPD than those not on ICS treatment. Previous studies have either measured EBC biomarkers at the start of exacerbation and compared against stable COPD and normal subjects (Gessner et al 2005), or measured the level of biomarkers serially during the first week of AECOPD (Gerritsen et al 2005; Oudijk et al 2006).

As our patients were dyspneic on admission to hospital, EBC assessment was only commenced on Day 5 when they became more stable and able to comply with EBC collection. Ideally, systemic steroids should have been withheld as it could suppress inflammation in the airway. As previous studies (Davies et al 1999; Wood-Baker et al 2005) have consistently shown that systemic steroid can reduce treatment failure and need for additional medical treatment in subjects with severe AECOPD, we felt that it was unethical to withhold the therapy, which was strongly recommended in the GOLD Guideline (National Heart, Lung and Blood Institute, World Health Organization 2006), for research purposes.

Table 3 Levels of the biomarkers

	TNF- α (pg/ml)	LTB4 (pg/ml)
AECOPD Day 5	5.08 (3.80–6.32)	6.30 (5.17–8.07)
AECOPD Day 14	5.84 (4.91–9.14)*	6.47 (5.05–8.00)
AECOPD Day 30	6.14 (3.82–7.67)*	6.26 (4.70–7.99)
AECOPD Day 60	5.60 (4.53–8.80)*	5.77 (4.82–7.37)
Stable COPD	5.39 (3.80–6.89)	5.90 (5.21–7.10)
Normal controls	4.84 (3.86–5.81)**	5.80 (5.29–7.54)

Notes: Data were presented in median (IQR); * $p < 0.05$ when compared with the level of biomarker at Day 5; ** $p = 0.036$ when comparing levels of biomarker between control subjects and Day 14 post-hospitalization for patients with AECOPD.

Abbreviations: AECOPD, acute exacerbations of chronic obstructive pulmonary disease; LTB4, leukotriene B4; TNF- α , tumor necrosis factor- α .

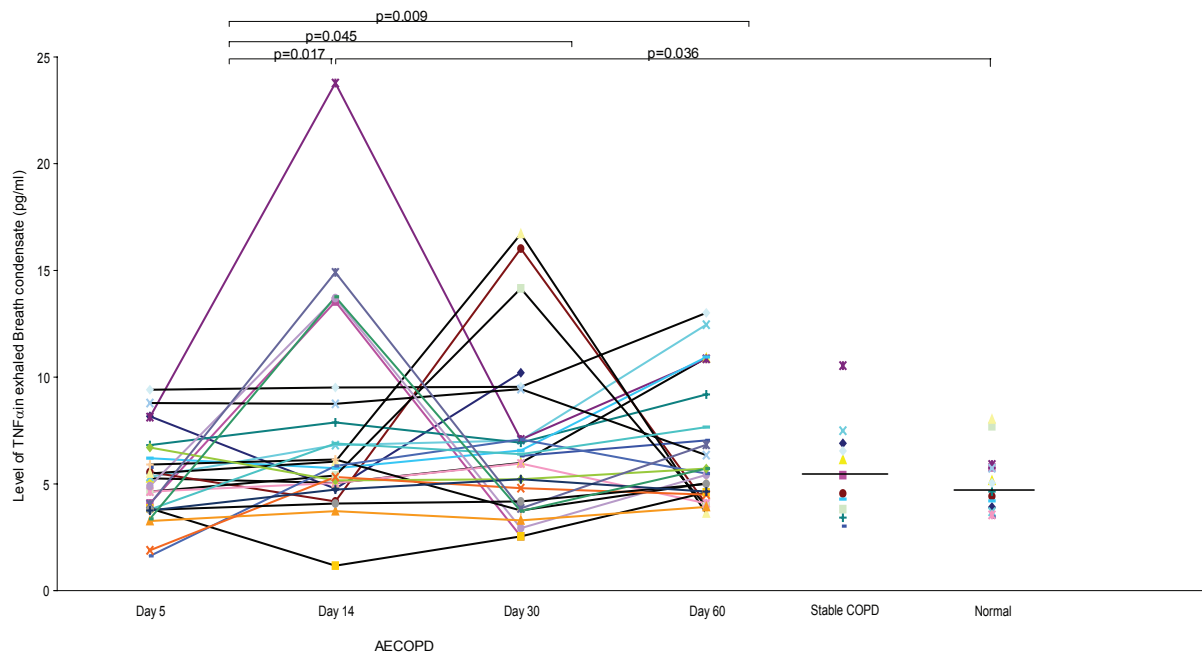


Figure 1 Levels of TNF- α in the subjects with AECOPD, stable COPD and normal controls

Abbreviations: AECOPD, acute exacerbations of chronic obstructive pulmonary disease; COPD, chronic obstructive pulmonary disease; TNF- α , tumor necrosis factor- α .

Previous studies have found that TNF- α level was raised in both EBC (Gessner et al 2005) and sputum (Aaron et al 2001) of subjects with AECOPD. However, the changes in circulating levels of TNF- α during AECOPD were less consistent. Some studies showed that its level correlated negatively with the changes in FEV₁ during the exacerbation (Pinto-Plata et al 2007), whereas others reported no change when comparing the stable baseline level with the AECOPD level (Hurst et al 2006). It is interesting to note that the level of TNF- α was similar from Day 14 onwards during the course of AECOPD in our study. It remains unclear why the TNF- α level was similar between the stable state COPD and the normal healthy controls. A previous study has suggested that markers such as LTB₄ and 8-isoprostane might take 2 months to return to normal state with antibiotic

(without systemic steroid) treatment (Biernacki et al 2003). The 1-week course of systemic steroid administered to our patients with AECOPD has possibly sped up the recovery of TNF- α .

TNF- α could be measured in the EBC in every subject in this current study, and similar observation was noted in our previous study involving asthmatic subjects (Leung et al 2005). This finding was also consistent with another study assessing TNF- α in EBC using cytometric bead array rather than ELISA (Gessner et al 2005). Our level of TNF- α was lower than that measured by cytometric bead array assay and this might be explained by the different methods of biomarker measurement (Gessner et al 2005). An experimental study with animal models showed that over-expression of TNF- α induced the pathological changes similar to emphysema and pulmonary fibrosis (Lundblad et al 2005). It is likely that TNF- α plays an important role in the pathogenesis of COPD and its level may be altered during the course of AECOPD.

We have noted that TNF- α level was lower in the EBC on Day 60 post-hospitalization for AECOPD in subjects who were on ICS therapy than those who were not. A previous study has shown that some cytokines in EBC such as IL-1 β , IL-6, IL-8, IL-12, and IL-10, but not TNF- α , were lower in stable COPD patients on ICS compared to those who were not on this therapy (Gessner et al 2005). Induced sputum study also failed to show the effect of ICS therapy on TNF- α or

Table 4 Serial spirometry results of the subjects with acute exacerbation of COPD (n = 26)

	Day 5	Day 14	Day 30	Day 60
FEV ₁ % predicted	41.5 \pm 12.9	46.1 \pm 14.2	43.6 \pm 14.2	44.5 \pm 14.6
FVC% predicted	54.8 \pm 17.8	61.5 \pm 20.5	61.7 \pm 18.2	62.1 \pm 19.0
FEV ₁ /FVC (%)	57.0 \pm 12.3	56.1 \pm 12.0	52.8 \pm 11.8	53.4 \pm 11.6
FEV ₁ (L)	0.83 \pm 0.29	0.91 \pm 0.28	0.89 \pm 0.37	0.91 \pm 0.62
FVC (L)	1.54 \pm 0.56	1.66 \pm 0.56	1.72 \pm 0.59	1.74 \pm 0.62

Note: Data were presented as mean \pm SD.

Abbreviations: FEV₁, forced expiratory volume in one second; FVC, forced vital capacity.

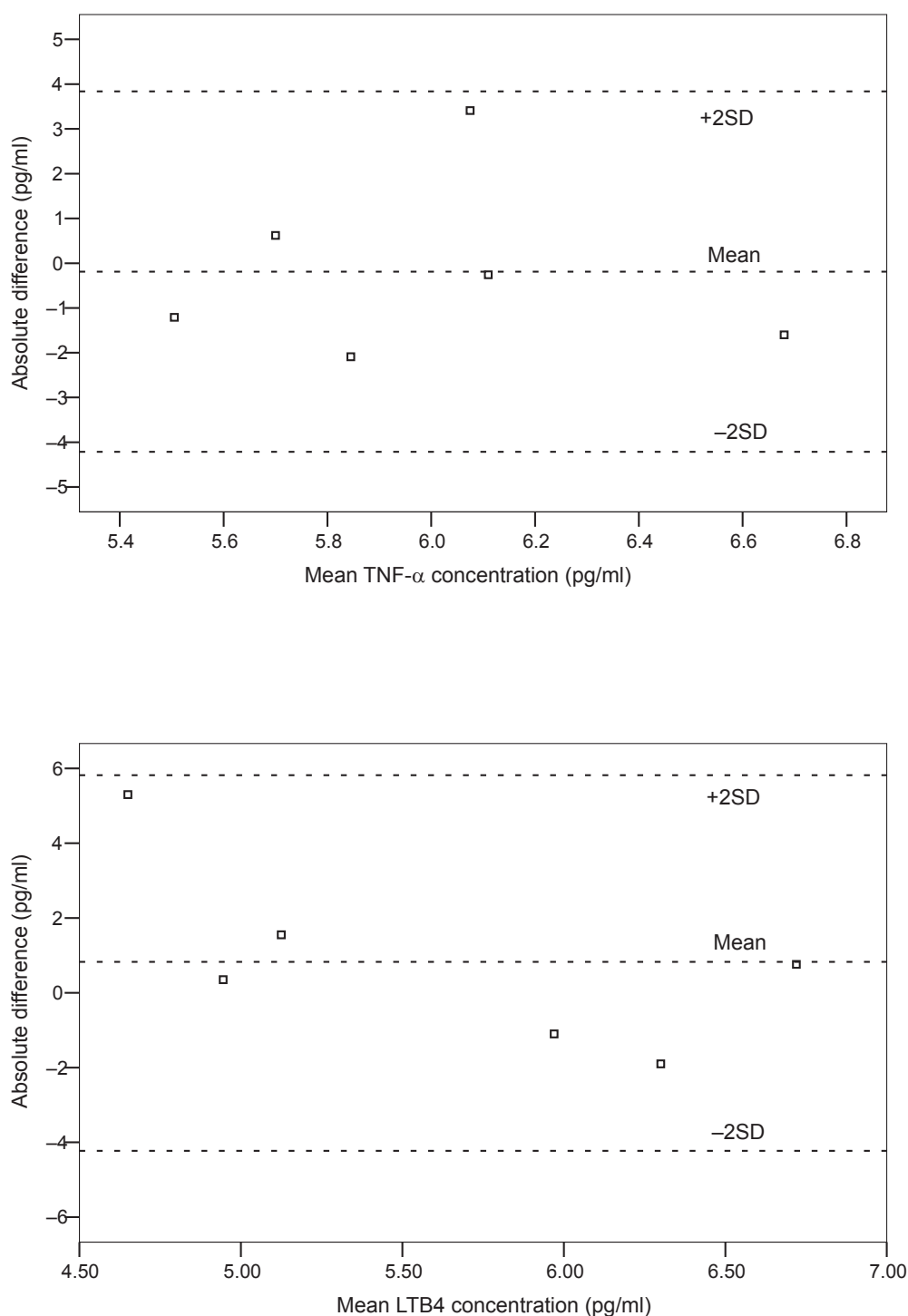


Figure 2 Bland and Altman's plot of the repeatability measurements of TNF- α and LTB4.
Abbreviations: LTB4, leukotriene B4; TNF- α , tumor necrosis factor- α .

other inflammatory indices in the sputum of COPD patients (Keatingset al 1997). Several studies have suggested that ICS could reduce the frequency of AECOPD in subjects with severe COPD (Burge et al 2000). Whether TNF- α or other biomarkers are involved in the mechanisms of AECOPD will require further exploration.

LTB4 in the COPD airway is likely mediating its effects via activation of neutrophils. Assessment of chemotactic activity of LTB4 during AECOPD using the LTB4 receptor antagonist LY293111 revealed that LTB4 contributed significantly (~30%) to neutrophil influx into the airway in COPD subjects (Crooks et al 2000). A previous study

found that LTB4 level in EBC was higher in COPD patients (for both steroid naïve and those on ICS) than among controls (Montuschi et al 2003). In our study, the changes in the level of LTB4 were not apparent during the course of AECOPD and this might be related to the treatment with both systemic steroid and antibiotic. IL-8 was not measurable in the majority of the EBC samples in this study. Previous studies on IL-8 in the airway have been inconclusive. One study found no difference in the IL-8 level in induced sputum specimen in subjects with stable or AECOPD (Bhowmik et al 2000), whereas Gessner and colleagues noted that EBC IL-8 level was higher in subjects with AECOPD admitted to the general ward or to the intensive care unit for mechanical ventilation (Gessner et al 2005). Studies on serum IL-8 level in patients with AECOPD treated with systemic steroid showed a gradual fall in its level in the first 7 days of the AECOPD (Gerritsen et al 2005). It is uncertain why we could not measure IL-8 reliably in our study. In the study by Gessner and colleagues, the IL-8 level, measured by the multiplex fluorescent bead immunoassay cytometric bead array technique, was well above the detection limit of the ELISA kit used in our current study, despite the fact that their levels of IL-8 in the stable COPD patients and controls were also quite low (Gessner et al 2005). It is possible that we might have missed the peak of IL-8 at the start of the AECOPD and thus could not detect any IL-8 after 5-days of systemic steroid.

The limitations of this study included a relatively small sample size and the ex-smoker status of the subjects was not confirmed objectively by urine cotinine assay. As smoking may affect EBC markers, we did not include active smokers in this study (Gessner et al 2005; Carpagnano et al 2003; Garey et al 2004). Ideally age- and sex-matched ex-smokers with normal lung function would serve as better controls than nonsmoking age and sex-matched subjects. In addition, we had not collected baseline EBC samples before AECOPD and thus the levels of biomarkers before therapy were unknown. Though the repeatability test of the TNF- α and LTB4 appeared satisfactory, as reflected by the differences between most of the paired measurements laying within ± 2 SDs (Figure 2), the relatively large SD values when compared to the absolute measured values of TNF- α and LTB4 would warrant cautions when interpreting the results. EBC assessment is still a relatively new tool in respiratory research with many methodological pitfalls (Effros et al 2004; Horvath et al 2005), including the variability of the measurement and the low concentrations of mediators measured (Effros et al 2004; Horvath et al 2005). There are suggestions that biomarker concentrations depend

on other factors such as the dilution of the samples (Effros et al 2004; Horvath et al 2005). Adjusting the concentrations using dilutional indicator (eg, conductivity of lyophilized EBC) in future studies may improve the accuracy of EBC measurement. As many factors were interplaying that could affect the levels of biomarkers (eg, the assay method of the biomarkers and the treatment given during the exacerbations), it is difficult to conclude whether the changes in the levels of the biomarkers were, in fact, due to “cause and effect” with respect to exacerbations and remissions.

Conclusion

In conclusion, it is feasible to obtain serial EBC in patients following AECOPD. EBC TNF- α level was lower on Day 5 post-hospitalization for AECOPD than the levels at other time points up to 60 days, and this was likely due to the effect of treatment with systemic steroid and antibiotic. The level of LTB4 did not change significantly during the course of AECOPD whereas IL-8 was mostly undetectable in the EBC. Research in EBC is a constantly evolving field and more sensitive assays and novel biomarkers may be identified in the future (Barnes et al 2006). As in the case of measuring exhaled nitric oxide level in asthma management (Smith et al 2005), prospective randomized trials are needed to evaluate if the measurement of TNF- α in EBC may guide treatment and improve outcome in patients with COPD.

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References

- Aaron SD, Angel JB, Lunau M, et al. 2001. Granulocyte inflammatory markers and airway infection during acute exacerbation of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*, 163:349–55.
- American Thoracic Society. Standardization of spirometry – 1987 update. Statement of the American Thoracic Society. *Am Rev Respir Dis*, 136:1285–98.
- Baggiolini M, Walz A, Kunkel SL, et al. 1989. Neutrophil-activating peptide-1/interleukin 8, a novel cytokine that activates neutrophils. *J Clin Invest*, 84:1045–9.
- Barnes PJ, Chowdhury B, Kharitonov SA, et al. 2006. Pulmonary biomarkers in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*, 174:6–14.
- Bhowmik A, Seemungal TA, Sapsford RJ, et al. 2000. Relation of sputum inflammatory markers to symptoms and lung function changes in COPD exacerbations. *Thorax*, 55:114–20.

- Biernacki WA, Kharitonov SA, Barnes PJ, et al. 2003. Increased leukotriene B4 and 8-isoprostane in exhaled breath condensate of patients with exacerbations of COPD. *Thorax*, 58:294–8.
- Bland JM and Altman DG. 1986. Statistical methods for assessing agreement between two methods of clinical measurement. *Lancet*, 1:307–10.
- Burge PS, Calverley PM, Jones PW, et al. 2000. Randomised, double blind, placebo controlled study of fluticasone propionate in patients with moderate to severe chronic obstructive pulmonary disease: the ISOLDE trial. *BMJ*, 320:1297–303.
- Carpagnano GE, Kharitonov SA, Foschino-Barbaro MP, et al. 2003. Increased inflammatory markers in the exhaled breath condensate of cigarette smokers. *Eur Respir J*, 21:589–93.
- Crooks SW, Bayley DL, Hill SL, et al. 2000. Bronchial inflammation in acute bacterial exacerbations of chronic bronchitis: the role of leukotriene B4. *Eur Respir J*, 15:274–80.
- Davies L, Angus RM, Calverley PM, et al. 1999. Oral corticosteroids in patients admitted to hospital with exacerbations of chronic obstructive pulmonary disease: a prospective randomised controlled trial. *Lancet*, 354:456–60.
- Effros RM, Dunning MB, 3rd, Biller J, et al. 2004. The promise and perils of exhaled breath condensates. *Am J Physiol Lung Cell Mol Physiol*, 287:L1073–80.
- Ford-Hutchinson AW, Bray MA, Doig MV, et al. 1980. Leukotriene B, a potent chemokinetic and aggregating substance released from polymorphonuclear leukocytes. *Nature*, 286:264–5.
- Garey KW, Neuhauser MM, Robbins RA, et al. 2004. Markers of inflammation in exhaled breath condensate of young healthy smokers. *Chest*, 125:22–6.
- Gerritsen WB, Asin J, Zanen P, et al. 2005. Markers of inflammation and oxidative stress in exacerbated chronic obstructive pulmonary disease patients. *Respir Med*, 99:84–90.
- Gessner C, Scheibe R, Wotzel M, et al. 2005. Exhaled breath condensate cytokine patterns in chronic obstructive pulmonary disease. *Respir Med*, 99:1229–40.
- Horvath I, Hunt J, Barnes PJ, et al. 2005. Exhaled breath condensate: methodological recommendations and unresolved questions. *Eur Respir J*, 26:523–48.
- Hunt J. 2002. Exhaled breath condensate: an evolving tool for noninvasive evaluation of lung disease. *J Allergy Clin Immunol*, 110:28–34.
- Hurst JR, Donaldson GC, Perera WR, et al. 2006. Use of plasma biomarkers at exacerbation of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*, 174:867–74.
- Ip MS, Ko FW, Lau AC, et al. 2006. Updated spirometric reference values for adult Chinese in Hong Kong and implications on clinical utilization. *Chest*, 129:384–92.
- Keatings VM, Collins PD, Scott DM, et al. 1996. Differences in interleukin-8 and tumor necrosis factor-alpha in induced sputum from patients with chronic obstructive pulmonary disease or asthma. *Am J Respir Crit Care Med*, 153:530–4.
- Keatings VM, Jatakanon A, Worsdell YM, et al. 1997. Effects of inhaled and oral glucocorticoids on inflammatory indices in asthma and COPD. *Am J Respir Crit Care Med*, 155:542–8.
- Ko FW, Ip M, Chan PK, et al. 2007. A 1-Year Prospective Study of the Infectious Etiology in Patients Hospitalized With Acute Exacerbations of COPD. *Chest*, 131:44–52.
- Ko FW, Lau CY, Leung TF, et al. 2006. Exhaled breath condensate levels of 8-isoprostane, growth related oncogene alpha and monocyte chemoattractant protein-1 in patients with chronic obstructive pulmonary disease. *Respir Med*, 100:630–8.
- Ko FW, Lau CY, Leung TF, et al. 2006. Exhaled breath condensate levels of eotaxin and macrophage-derived chemokine in stable adult asthma patients. *Clin Exp Allergy*, 36:44–51.
- Ko FW, Leung TF, Hui DS, et al. 2007. Are exhaled breath condensates useful in monitoring asthma? *Curr Allergy Asthma Rep*, 7:65–71.
- Leung TF, Wong GW, Ko FW, et al. 2005. Analysis of growth factors and inflammatory cytokines in exhaled breath condensate from asthmatic children. *Int Arch Allergy Immunol*, 137:66–72.
- Lundblad LK, Thompson-Figueroa J, Leclair T, et al. 2005. Tumor necrosis factor-alpha overexpression in lung disease: a single cause behind a complex phenotype. *Am J Respir Crit Care Med*, 171:1363–70.
- Montuschi P and Barnes PJ. 2002. Analysis of exhaled breath condensate for monitoring airway inflammation. *Trends Pharmacol Sci*, 23:232–7.
- Montuschi P, Kharitonov SA, Ciabattini G, et al. 2003. Exhaled leukotrienes and prostaglandins in COPD. *Thorax*, 58:585–8.
- Mukhopadhyay S, Hoidal JR, Mukherjee TK, et al. 2006. Role of TNFalpha in pulmonary pathophysiology. *Respir Res*, 7:125.
- National Heart, Lung and Blood Institute, World Health Organization. Global Initiative for chronic obstructive lung disease. Global strategy for the diagnosis, management and prevention of chronic obstructive pulmonary disease. Updated 2006. NHLB/WHO 2006.
- Oudijk EJ, Gerritsen WB, Nijhuis EH, et al. 2006. Expression of priming-associated cellular markers on neutrophils during an exacerbation of COPD. *Respir Med*, 100:1791–9.
- Patel IS, Seemungal TA, Wilks M, et al. 2002. Relationship between bacterial colonisation and the frequency, character, and severity of COPD exacerbations. *Thorax*, 57:759–64.
- Pinto-Plata VM, Livnat G, Girish M, et al. 2007. Systemic cytokines, clinical and physiological changes in patients hospitalized for exacerbation of COPD. *Chest*, 131:37–43.
- Seemungal TA, Donaldson GC, Paul EA, et al. 1998. Effect of exacerbation on quality of life in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*, 157:1418–22.
- Smith AD, Cowan JO, Brassett KP, et al. 2005. Use of exhaled nitric oxide measurements to guide treatment in chronic asthma. *N Engl J Med*, 352:2163–73.
- Wood-Baker RR, Gibson PG, Hannay M, et al. 2005. Systemic corticosteroids for acute exacerbations of chronic obstructive pulmonary disease. *Cochrane Database Syst Rev*, CD001288.
- Yamamoto C, Yoneda T, Yoshikawa M, et al. 1997. Airway inflammation in COPD assessed by sputum levels of interleukin-8. *Chest*, 112:505–10.