Effect of bupivacaine and adjuvant drugs for regional anesthesia on nerve tissue oximetry and nerve blood flow

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Background: Nerve blood flow has a critical role in acute and chronic pathologies in peripheral nerves. Influences of local anesthetics and adjuvants on tissue perfusion and oxygenation are deemed as relevant factors for nerve damage after peripheral regional anesthesia. The link between low tissue perfusion due to local anesthetics and resulting tissue oxygenation is unclear.

Methods: Combined tissue spectrophotometry and laser-Doppler flowmetry were used to assess nerve blood flow in 40 surgically exposed median nerves in pigs, as well as nerve tissue oximetry for 60 min. After baseline measurements, test solutions saline (S), bupivacaine (Bupi), bupivacaine with epinephrine (BupiEpi), and bupivacaine with clonidine (BupiCloni) were applied topically.

Results: Bupivacaine resulted in a significant decrease in nerve blood flow, as well as tissue oximetry values, compared with saline control. Addition of epinephrine resulted in a rapid, but nonsignificant, reduction of nerve blood flow and extensive lowering of tissue oximetry levels. The use of clonidine resulted in a reduction of nerve blood flow, comparable to bupivacaine alone (relative blood flow at T60 min compared with baseline, S: 0.86 (0.67–1.18), median (25th–75th percentile); Bupi: 0.33 (0.25–0.60); BupiCloni: 0.43 (0.38–0.63); and BupiEpi: 0.41 (0.30–0.54). The use of adjuvants did not result in any relevant impairment of tissue oximetry values (saturation values in percent at T60, S: 91.5 [84–95]; Bupi: 76 [61–86]; BupiCloni: 84.5 [76–91]; and BupiEpi: 91 [56–92]).

Conclusion: The application of bupivacaine results in lower nerve blood flow, but does not induce relevant ischemia. Despite significant reductions in nerve blood flow, the addition of clonidine or epinephrine to bupivacaine had no significant impact on nerve tissue oximetry compared with bupivacaine alone. Nerve ischemia due to local anesthetics is not enhanced by the adjuvants clonidine or epinephrine.

Keywords: peripheral regional anesthesia, complications, nerve blood flow, local anesthetics, adjuvants

Introduction
The blood supply to peripheral nerves is delivered by two integrated, but functionally independent, vascular systems.¹,² Extrinsic and intrinsic vessels form anastomoses at the level of the epineurium. Alterations in blood flow are important for peripheral nerve pathology in acute and chronic disease.³–⁶ Local anesthetics and epinephrine, used as an adjunct, are known to have a negative impact on nerve blood flow.⁴,⁷ The underlying molecular mechanisms of local anesthetic-induced reductions of nerve blood flow are still uncertain and speculative.⁹ Additionally, it is unclear whether these blood flow reductions, caused by local anesthetics, result in local hypoxic tissue conditions and act as a key factor for peripheral nerve injury.⁵,⁶ The goal of this study is to determine the effect of bupivacaine alone and in combination with the adju cents epinephrine or

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clonidine on nerve blood flow and tissue oximetry in a large animal model, simulating human conditions. Thus, data for epinephrine and clonidine, with regard to neural ischemia as a potential risk factor for nerve damage, could be investigated in our study.

Methods

General anesthesia and nerve preparation

Experiments were approved by the local authorities (Study No. 93/11; Regional Board of Animal Welfare, Giessen, Germany); study results are presented according to ARRIVE guidelines. Experiments were performed in accordance with the current German laws of animal protection. Standardized general anesthesia and euthanasia, following the experiments, were conducted as described previously. In brief, following a premedication using diazepam and ketamine, general anesthesia was induced using propofol (2–3 mg·kg⁻¹ and sufentanil). After endotracheal intubation, general anesthesia was commenced using propofol (0.2 mg·kg⁻¹·min⁻¹) and sufentanil (0.5 µg·kg⁻¹·h⁻¹). Mechanical ventilation settings were modified to obtain an end tidal CO₂ of 35–45 mmHg. Inspiratory oxygen fraction was set to 30% (to simulate clinical conditions, using low-flow oxygen supplementation in spontaneously breathing subjects). Ultrasound-guided invasive blood pressure measurement was conducted via catheter to the left femoral artery. To achieve standardized conditions and minimize motion artifacts for the measurements, pancuronium bromide (0.1 mg·kg⁻¹) was given repeatedly during the study period. Pharyngeal temperature was recorded and kept constant using warm blankets.

The anesthetized animals were placed supine, and axillary regions were dissected surgically as previously described; median nerves were exposed with preservation of the extrinsic epineural blood vessel supply.

Euthanasia was performed at the end of the experiments under general anesthesia using potassium chloride (4 mmol·kg⁻¹).

Combined tissue spectrophotometry and laser-Doppler flowmetry

Measurements of nerve blood flow (flow in arbitrary units [AU]), tissue oxygen saturation (SO₂ in %), and tissue hemoglobin level (rHb, in AU) were obtained using laser-Doppler flowmetry and tissue spectrophotometry (O2C; Lea Medizintechnik, Giessen, Germany). The system and probe (LF-1; Lea, Medizintechnik) were calibrated according to the manufacturer’s recommendation. The O2C probe was applied to the exposed nerve with a custom-made application device (Figure 1), and fixed with sutures in the surrounding tissue for constant application pressure, to acquire a test volume of ~1–2 mm deep inside the nerve, according to the manufacturer’s advice. Measurements were started 10 min before each intervention to achieve optimal measurement stability of the system. One milliliter of the randomly assigned test solution was applied directly to the median nerve next to the probe (resulting in spread under the probe due to capillary effects) immediately after acquiring data at T0.

Figure 1 O2C probe in situ, mounted on median nerve.
The O2C machine consists of two different units (Figure 2). First, blood flow (flow) is measured by using the technique of laser-Doppler flowmetry, which has been described previously.14–16 Second, the tissue spectrophotometry unit calculates tissue hemoglobin content and tissue oximetry. In brief, underlying tissue is illuminated with coherent laser light of 500–630 nm wavelength and 30 mW power through a fiber-optic cable. Probe geometry allows for the detection of blood flow at ~1–2 mm depth by analyzing backscattered light. By fitting measured spectra with spectra of known tissue saturation values (SO₂), the SO₂ of the capillary and post-capillary microvasculature is calculated by the O2C device. The tissue hemoglobin value (rHb) represents the hemoglobin concentration per tissue volume (displayed as arbitrary unit). It is dependent on microvessel density, venous filling and microvessel shunting. This parameter is helpful to rule out venous congestion (raising rHb value).

Reductions of nerve blood flow are categorized as minimal (<20% reduction of nerve blood flow [NBF] baseline), mild (20%–50% reduction of NBF), moderate (50%–80% reduction of NBF) or severe (> 80% reduction of NBF) according to structural or electrophysiological signs of tissue damage or electrophysiological conduction impairment as previously described.17–19

Study groups
Randomly, 1 mL of the following test solutions was applied topically to the nerve: bupivacaine 0.5% isobaric (Bupi) (Carbostesin, B. Braun, Melsungen, Germany); bupivacaine 0.5% with epinephrine 1:100,000 (BupiEpi) (Suprarenin, Sanofi-Aventis, Frankfurt, Germany); bupivacaine 0.5% (see previous section) with 3.75 µg/mL clonidine (BupiCloni) (Clonidin-ratiopharm, Ratiopharm, Ulm, Germany). In addition, saline was used as a negative control: saline 0.9% isotonic (NaCl, B. Braun).

Test solutions were prepared freshly before use for the experiments in a standardized fashion using single-dose vials avoiding preparations containing preservatives like sodium metabisulfite. The concentrations of epinephrine and clonidine resemble the highest concentration of epinephrine used in clinical studies.20 Only one nerve per exposed brachial plexus was exposed to only one test solution to avoid potential residual drug effects on the sample.

Outcome parameters
At the time points T0, T5, T10, T15, T20, T25, T30, T40, T50, and T60, the O2C data of blood flow (flow, in AU), tissue oximetry (SO₂, in percent), and hemoglobin (rHb, in AU) were recorded using Excel (Excel for Mac; Microsoft Corporation, Redmond, WA, USA). Parameters of systemic hemodynamics (heart rate, mean arterial pressure), as well as oxygen saturation were recorded at T0, T30, and T60.

Sample size calculation
Based on published data for nerve blood flow alterations after the application of local anesthetic, we assumed a relative change of nerve blood flow of 1.0 (no change) for saline

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**Figure 2** Schematic illustration of O2C device.

**Abbreviations:** SO₂, tissue oxygen saturation; rHb, tissue hemoglobin.
(negative control group), and compared with T0 and T60 values. Based on published data of nerve blood flow measurements, we assumed a reduction to 0.6 for bupivacaine (standard group), 0.8 for BupiCloni (intervention group), and 0.4 for BupiEpi (intervention group) at T60, compared with baseline flow at T0.9 With an assumed standard deviation of 10%, a power of 80%, and a two-sided testing, a minimum number of seven to eight nerves per group were calculated for semi-parametric testing. Due to multiple testing (two comparisons between bupivacaine and each intervention group), the sample size calculation increased to n = 9 per group. Considering a dropout rate due to accidental bleeding or technical problems of data acquisition, we planned to perform measurements in 10 nerves per group.

Exclusion criteria
Exposed nerves were not used for the experiments when epineural blood vessels were damaged during surgical dissection. Obtained measurements for each given nerve were investigated (blinded to the respective group) for signs of technical problems during the test period, or the baseline period, before starting the test period, and were excluded if:
- Continuous data acquisition was not correctly performed over the test period for a respective nerve (mostly due to inadvertent probe dislocation).
- Spectra analysis of the raw data showed incorrect values for hemoglobin extinction (tissue compression due to probe placement).

Statistical analysis
Data were analyzed using SPSS (SPSS for Windows, release 18.0). Values for SO₂ are given as percentage (%) and values for rHb are presented in arbitrary units (AU) according to the manufacturer, and are presented as absolute values at the given time points. Values for blood flow are presented as relative flow compared to baseline flow measurement, according to previous studies.5,9
Values of relative flow (AU), SO₂ (%), and rHb (AU) at T0, T30, and T60 of bupivacaine (standard group) was compared with either BupiEpi or BupiCloni (experimental groups) using Mann-Whitney U-test. The saline group served only as a negative control group in comparison with the bupivacaine group. Statistical tests for differences (two-sided) were deemed significant at a level of 0.05. A closed testing approach was used to compensate for multiple testing: Bonferroni adjustment corrected the level of significance for the two primary comparisons of groups, and the hierarchy of testing according to the primary end points and the time points was relative flow at T60, SO₂ at T60, rHb at T60, relative flow at T30, SO₂ at T30, and rHb at T30. Hodges-Lehmann (HL) estimators, with 95% confidence intervals (CI), were calculated for the interpretation of differences between microcirculation parameters (SO₂ and nerve blood flow) between bupivacaine and each respective group.

Additionally, descriptive post hoc testing was performed comparing SO₂ and nerve blood flow parameters at T0, T30, and T60 between all groups using the Kruskal–Wallis testing (two-sided testing, level of significance 0.05). In case of significance, pairwise comparisons of the saline group versus bupivacaine, BupiEpi, or BupiCloni groups were performed.

Furthermore, to exclude the potential bias of varying vital parameters between groups, vital parameters at T0, T30, and T60 were compared. In this analysis, HL estimators (95% CI) for heart rate (bpm), mean blood pressure (MAP, mmHg), and SpO₂ (SpO₂, %) were calculated between the bupivacaine group and one of the respective groups. Differences of less than 10 bpm, 5 mmHg in MAP, or 3% in SpO₂ among the compared groups were assumed to be clinically irrelevant.

Results
Number of animals and obtained measurements
Fifty-six peripheral nerves (one nerve per brachial plexus) were exposed in 28 pigs. Six nerves were excluded before randomization due to difficulties in probe mounting, or were used for surgical preparation training. According to the predefined criteria for study termination, three peripheral nerves were excluded from the study due to insufficient surgical preparation of the nerve, with suspected injury to the extrinsic nerve vasculature. Furthermore, seven nerves were excluded before data analysis, due to supposed probe dislocation and non-continuous data acquisition. Thus, 40 nerves were randomly assigned to one of the four experimental groups, and included in the final data analysis (n = 10 per group).

Vital parameters
Vital parameter data are given in Table 1. Comparison of HL estimators for mean arterial pressure, heart rate, and systemic pulse oximetry revealed no clinically relevant differences between the groups at T0, T30, and T60 min (Table 1).
Microcirculation (= flow measurements)
Analysis of flow parameters is shown in Table 2. Using the Mann–Whitney test, the bupivacaine group (standard group) showed a significant decrease of flow, compared with the saline group (control group) at T30 and T60 (both \( p < 0.0001 \)), but was not significantly different from bupivacaine with added clonidine (T30, \( p = 1.0 \); T60, \( p = 0.19 \)), or with added epinephrine (T30, \( p = 0.48 \); T60, \( p = 0.39 \)). HL estimators with 95% CI are shown in Table 2.

<table>
<thead>
<tr>
<th>Group</th>
<th>Parameter</th>
<th>T0</th>
<th>HL (95% CI)</th>
<th>T30</th>
<th>HL (95% CI)</th>
<th>T60</th>
<th>HL (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bupi (n = 10)</td>
<td>Heart rate</td>
<td>97 (81–108)</td>
<td>n/a</td>
<td>97 (83–99)</td>
<td>n/a</td>
<td>92 (84–102)</td>
<td>n/a</td>
</tr>
<tr>
<td></td>
<td>MAP</td>
<td>74 (65–79)</td>
<td>n/a</td>
<td>76 (68–86)</td>
<td>n/a</td>
<td>73 (69–83)</td>
<td>n/a</td>
</tr>
<tr>
<td></td>
<td>( \text{SpO}_2 )</td>
<td>97 (97–99)</td>
<td>n/a</td>
<td>99 (94–100)</td>
<td>n/a</td>
<td>98 (96–100)</td>
<td>n/a</td>
</tr>
<tr>
<td>Saline (n = 10)</td>
<td>Heart rate</td>
<td>96 (80–103)</td>
<td>−0.5 (−16.22)</td>
<td>89 (77–103)</td>
<td>−4.5 (−19.19)</td>
<td>101 (74–110)</td>
<td>7.5 (−10.25)</td>
</tr>
<tr>
<td></td>
<td>MAP</td>
<td>71 (59–81)</td>
<td>−2.0 (−14.12)</td>
<td>75 (68–82)</td>
<td>−2 (−12.7)</td>
<td>75 (67–86)</td>
<td>0.5 (−16.22)</td>
</tr>
<tr>
<td></td>
<td>( \text{SpO}_2 )</td>
<td>100 (96–100)</td>
<td>1.0 (−1.3)</td>
<td>100 (98–100)</td>
<td>0 (0.3)</td>
<td>99 (97–100)</td>
<td>0 (−2.2)</td>
</tr>
<tr>
<td>BupiCloni (n = 10)</td>
<td>Heart rate</td>
<td>100 (74–105)</td>
<td>−1.5 (−14.12)</td>
<td>97 (76–102)</td>
<td>−0.5 (−15.12)</td>
<td>94 (73–103)</td>
<td>0 (−15.12)</td>
</tr>
<tr>
<td></td>
<td>MAP</td>
<td>79 (71–86)</td>
<td>3.5 (−8.16)</td>
<td>80 (70–81)</td>
<td>0 (−13.9)</td>
<td>76 (63–89)</td>
<td>1.5 (−11.12)</td>
</tr>
<tr>
<td></td>
<td>( \text{SpO}_2 )</td>
<td>100 (98–100)</td>
<td>2 (0.3)</td>
<td>100 (98–100)</td>
<td>1 (0.4)</td>
<td>100 (98–100)</td>
<td>0 (−3.0)</td>
</tr>
<tr>
<td>BupiEpi (n = 10)</td>
<td>Heart rate</td>
<td>95 (84–123)</td>
<td>4.5 (−14.25)</td>
<td>100 (85–114)</td>
<td>5 (−11.23)</td>
<td>101 (88–112)</td>
<td>8 (−6.22)</td>
</tr>
<tr>
<td></td>
<td>MAP</td>
<td>73 (67–85)</td>
<td>0 (−12.14)</td>
<td>74 (67–80)</td>
<td>−3 (−14.6)</td>
<td>71 (65–78)</td>
<td>−2 (−14.9)</td>
</tr>
<tr>
<td></td>
<td>( \text{SpO}_2 )</td>
<td>100 (98–100)</td>
<td>2 (0.3)</td>
<td>100 (99–100)</td>
<td>1 (0.3)</td>
<td>100 (99–100)</td>
<td>0 (0.3)</td>
</tr>
</tbody>
</table>

Notes: Heart rate is expressed in bpm; MAP in mmHg; and \( \text{SpO}_2 \) in %. All data are given as median (25th–75th percentile), and compares each group with bupivacaine at the same time point, with differences shown as HL estimator, with 95% CI.

Abbreviations: Bupi, bupivacaine; BupiCloni, bupivacaine with clonidine; BupiEpi, bupivacaine with epinephrine; CI, confidence interval; HL, Hodges-Lehmann; MAP, mean arterial pressure; n/a, not applicable; Saline, saline control group; \( \text{SpO}_2 \), pulse oximetry; T0, time point 0 min; T30, time point 30 min; T60, time point 60 min.

Post hoc analysis (saline versus active comparator)
Descriptive post hoc testing was performed comparing \( \text{SO}_2 \) as well as nerve blood flow at the given time points between groups. Kruskal–Wallis testing revealed significant differences for nerve blood flow only for T30 (\( p < 0.0001 \)) and T60 (\( p < 0.001 \)) but not for T0 (\( p = 1.00 \)). Regarding \( \text{SO}_2 \) data, post hoc Kruskal–Wallis testing showed no significant differences between groups for T0 and T30 but a significant difference for T60 (\( p = 0.047 \)).

Thus, pairwise comparisons of saline versus each group (bupivacaine, BupiEpi, or BupiCloni) were performed when applicable. Statistical test results are shown in Table 5.

Discussion
Our results showed that the application of local anesthetics alone, such as bupivacaine, resulted in relevant nerve blood flow reductions at T30 and T60, as shown in previous studies.\(^4,5,8,9,22\) However, no significant difference for tissue oximetry between bupivacaine versus saline or versus bupivacaine in combination with clonidine or epinephrine was observed. Moreover, the addition of epinephrine or clonidine did not result in any significant alterations of nerve blood flow or tissue oximetry (\( \text{SO}_2 \)), compared with the bupivacaine standard group.
Table 2 Relative nerve blood flow

<table>
<thead>
<tr>
<th>Group</th>
<th>T5</th>
<th>T10</th>
<th>T15</th>
<th>T20</th>
<th>T25</th>
<th>T30</th>
<th>HL (95% CI)</th>
<th>T40</th>
<th>T50</th>
<th>T60</th>
<th>HL (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bupivacaine</td>
<td>0.70</td>
<td>0.61</td>
<td>0.63</td>
<td>0.57</td>
<td>0.55</td>
<td>0.50</td>
<td>n/a</td>
<td>0.36</td>
<td>0.35</td>
<td>0.33</td>
<td>n/a</td>
</tr>
<tr>
<td></td>
<td>(0.62–0.97)</td>
<td>(0.50–0.68)</td>
<td>(0.40–0.65)</td>
<td>(0.30–0.63)</td>
<td>(0.27–0.58)</td>
<td>(0.28–0.60)</td>
<td>n/a</td>
<td>(0.26–0.54)</td>
<td>(0.24–0.51)</td>
<td>(0.25–0.60)</td>
<td>n/a</td>
</tr>
<tr>
<td>Saline</td>
<td>1.02</td>
<td>1.04</td>
<td>1.09</td>
<td>1.04</td>
<td>1.20</td>
<td>1.17</td>
<td>0.738</td>
<td>1.07</td>
<td>1.02</td>
<td>0.86</td>
<td>0.527</td>
</tr>
<tr>
<td></td>
<td>(1.01–1.22)</td>
<td>(0.94–1.09)</td>
<td>(0.93–1.14)</td>
<td>(0.95–1.19)</td>
<td>(0.94–1.28)</td>
<td>(1.11–1.36)</td>
<td>(0.54–0.947)</td>
<td>(0.94–1.30)</td>
<td>(0.79–1.23)</td>
<td>(0.67–1.18)</td>
<td>(0.273–0.857)</td>
</tr>
<tr>
<td>BupiCloni</td>
<td>0.79</td>
<td>0.71</td>
<td>0.58</td>
<td>0.47</td>
<td>0.44</td>
<td>0.45</td>
<td>0.002</td>
<td>0.38</td>
<td>0.38</td>
<td>0.38</td>
<td>0.43</td>
</tr>
<tr>
<td></td>
<td>(0.68–0.97)</td>
<td>(0.59–0.81)</td>
<td>(0.50–0.67)</td>
<td>(0.40–0.63)</td>
<td>(0.32–0.73)</td>
<td>(0.35–0.57)</td>
<td>(−0.185; 0.206)</td>
<td>(0.22–0.61)</td>
<td>(0.42–0.71)</td>
<td>(0.38–0.63)</td>
<td>(−0.069; 0.312)</td>
</tr>
<tr>
<td>BupiEpi</td>
<td>0.59</td>
<td>0.45</td>
<td>0.39</td>
<td>0.41</td>
<td>0.41</td>
<td>0.38</td>
<td>−0.049</td>
<td>0.38</td>
<td>0.39</td>
<td>0.41</td>
<td>0.0897</td>
</tr>
<tr>
<td></td>
<td>(0.27–0.71)</td>
<td>(0.31–0.61)</td>
<td>(0.34–0.60)</td>
<td>(0.28–0.60)</td>
<td>(0.30–0.56)</td>
<td>(0.27–0.54)</td>
<td>(−0.274; 0.178)</td>
<td>(0.26–0.62)</td>
<td>(0.23–0.62)</td>
<td>(0.30–0.54)</td>
<td>(−0.127; 0.24)</td>
</tr>
</tbody>
</table>

Notes: All data are given as median (25th–75th percentile), and compares each group with bupivacaine at the same time point, with differences shown as HL estimator, with 95% CI. Flow, relative values of flow to baseline flow at T0.

*Significant, **not significant. Level of significance p < 0.05. For details see text.

Abbreviations: BupiCloni, bupivacaine with clonidine; BupiEpi, bupivacaine with epinephrine; CI, confidence interval; HL, Hodges-Lehmann Group; n/a, not applicable; Saline, saline control group; T5–T60, time points 5 min to 60 min.

Table 3 Tissue oximetry

<table>
<thead>
<tr>
<th>Group</th>
<th>T0</th>
<th>T5</th>
<th>T10</th>
<th>T15</th>
<th>T20</th>
<th>T25</th>
<th>T30</th>
<th>HL (95% CI)</th>
<th>T40</th>
<th>T50</th>
<th>T60</th>
<th>HL (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bupivacaine</td>
<td>92</td>
<td>92</td>
<td>90.5</td>
<td>91.5</td>
<td>92</td>
<td>90.5</td>
<td>89</td>
<td>n/a</td>
<td>84.5</td>
<td>79.5</td>
<td>76</td>
<td>n/a</td>
</tr>
<tr>
<td></td>
<td>(90–98)</td>
<td>(86–95)</td>
<td>(86–95)</td>
<td>(78–95)</td>
<td>(72–95)</td>
<td>(70–91)</td>
<td>(70–94)</td>
<td>n/a</td>
<td>(66–91)</td>
<td>(62–85)</td>
<td>(61–86)</td>
<td>n/a</td>
</tr>
<tr>
<td>Saline</td>
<td>89.5</td>
<td>90</td>
<td>89.5</td>
<td>88.5</td>
<td>88.5</td>
<td>88.5</td>
<td>88.5</td>
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<td>90.5</td>
<td>92</td>
<td>91.5</td>
<td>13.5</td>
</tr>
<tr>
<td>BupiCloni</td>
<td>89</td>
<td>88</td>
<td>89.5</td>
<td>89.5</td>
<td>93</td>
<td>91</td>
<td>90</td>
<td>3</td>
<td>90</td>
<td>86</td>
<td>84.5</td>
<td>7</td>
</tr>
<tr>
<td>BupiEpi</td>
<td>92.5</td>
<td>83.5</td>
<td>74</td>
<td>85.5</td>
<td>83</td>
<td>83.5</td>
<td>85</td>
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<td>87</td>
<td>90</td>
<td>91</td>
<td>8.5</td>
</tr>
</tbody>
</table>

Notes: All data are given as median (25th–75th percentile), and compares each group with bupivacaine at the same time point, with differences shown as HL estimator, with 95% CI. SO2 is measured in %. Statistical testing was performed for T0, T30, and T60 comparing bupivacaine with the respective group. *Significant, **not significant. Level of significance p < 0.05. For details see text.

Abbreviations: BupiCloni, bupivacaine with clonidine; BupiEpi, bupivacaine with epinephrine; CI, confidence interval; HL, Hodges-Lehmann; n/a, not applicable; Saline, saline control group; SO2, tissue oximetry; T0–T60, time points 0 min to 60 min.
Epinephrine enhances nerve block duration, mostly due to reduction of local tissue clearance and prolonged concentration of the local anesthetic in the nerve and perineural tissue. It is regularly used as an adjunct for local anesthetics to reduce systemic absorption and consecutive toxic phenomena of the injected local anesthetic. Furthermore, many clinicians assume a shortened nerve block onset time and prolonged anesthetic nerve blockade when epinephrine is used. However, some authors believe that when used as an adjunct, epinephrine might be an independent risk factor for nerve damage due to relevant ischemia. Our study showed no further statistically significant impairment of nerve blood flow (when compared with bupivacaine alone). However, two nerves of the BupiEpi group did have relevant desaturations during measurements (SO2 nadir values of 3% at T20 and 10% at T10 in another nerve). Thus, despite no significant difference between both groups of bupivacaine, with and without epinephrine, we would recommend not to use epinephrine in a patient population with preexisting peripheral nerve damage (i.e., diabetic polyneuropathy).

Clonidine is widely used as an adjunct to local anesthetic in different peripheral nerve blockades. It prolongs the duration of the blockade due to unknown mechanisms. Due to its alpha-2-adrenergic effects, vasodilatation might be assumed in peripheral tissues, without risk of nerve ischemia. However, clinical side effects, such as sedation or bradycardia, are regularly observed. In experimental models of peripheral neuropathy, clonidine acts as an anti-inflammatory drug. Thus, its clinical effects and anti-inflammatory potential are interesting factors for the regional anesthetist. Our experimental data give further evidence of safety when perineural clonidine is used in combination with local anesthetics. No further alterations of nerve microcirculation or tissue oximetry were observed, compared with bupivacaine alone.

Systemic data did not differ, in any way that was clinically relevant, between the time points T0, T30, and T60 when...
compared between each group. This is important due to the response of nerve SO$_2$ to central hypoxia and cardiovascular changes as described by Ibrahim et al.$^{29}$ Moreover, this aspect is a potential bias within previous published studies of nerve blood flow measurements in animals where changes in oxygen saturation and circulation were not, or were inadequately, controlled.

Laser-Doppler flowmetry is an established method for investigating nerve blood flow alterations.$^{4,9,30}$ Nevertheless, responses of the underlying tissues to blood flow reductions have been poorly investigated for local anesthetic applications. The use of the O$_2$C device enabled us to investigate the consequences of (microvascular) nerve blood flow alterations with regard to local post-capillary oximetry and hemoglobin content as critical parameters for tissue hypoxia and potential tissue damage.

**Limitations**

Our experimental study has several limitations, which should be kept in mind before transferring to clinical practice. First, the differences between the results in this animal study might not be easily extrapolated to humans, due to possible differences in nerve blood flow and underlying regulative mechanisms between pigs and humans. Changes in vascular tone might be influenced by general anesthesia, the surgical preparation,$^{31}$ and the mounted probe resting on the nerves. Changes in blood pressure, systemic oxygen saturation, and end tidal carbon dioxide might have an impact on nerve blood flow due to limited autoregulation of nerve blood flow.$^{30,32}$

These important bias factors are controlled for in our study by invasive blood pressure monitoring, as well as the use of controlled mechanical ventilation with fixed inspired oxygen fraction, and the use of capnometry for establishing normocapnia. Furthermore, our investigation focused only on median nerves, but potential differences between nerve vasculature must be regarded. However, Kihara et al showed no significant differences between different nerve types or fascicle size.$^{32}$ Our measurements were performed for 60 min, which is higher than in previous studies. Nevertheless, the vasoconstrictive effects of the test solutions persisted for longer time periods. Further studies should evaluate this aspect using long-term measurements and include histological or electrophysiological evaluation. Measurement of compound muscle action potential might be a helpful approach in further studies for assessing the acute and long-term effects of the investigated interventions as shown in previous studies.$^{33,34}$

“Downstream” (distal) measurements of overall tissue or nerve tissue perfusion and oxygenation by using near-infrared spectroscopy or other methods as a result of block-related vasodilation might be another interesting topic.$^{35}$

Neurotoxicity of local anesthetics is not caused solely by ischemia as a result of reduced tissue perfusion. Bupivacaine was shown to be a weak uncoupler of mitochondrial oxidative phosphorylation resulting in apoptosis in neural cells.$^{36,37}$

**Conclusion**

Based on our experimental study results, the addition of epinephrine or clonidine to bupivacaine does not result in any relevant changes of nerve blood flow, or tissue oximetry, compared with bupivacaine alone. However, bupivacaine does show a relevant depression of nerve blood flow without resulting in severe nerve tissue ischemia. Thus, impairment of nerve blood flow by bupivacaine alone is not modified in a relevant way by the combination of bupivacaine with clonidine. The combination of bupivacaine with epinephrine shows no statistically significant differences in nerve tissue perfusion or tissue oxygen saturation. Ischemia as a consequence of the use of bupivacaine alone or in combination with epinephrine or clonidine was a rare event in our experimental study. Thus, ischemia is not a potential clinical relevant risk factor of peripheral nerve damage resulting from the administration of the aforementioned drugs.

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**Disclosure**

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