Dry eye syndrome: developments and lifitegrast in perspective

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Abstract: Dry eye (DE) is a chronic ocular condition with high prevalence and morbidity. It has a complex pathophysiology and is multifactorial in nature. Chronic ocular surface inflammation has emerged as a key component of DE that is capable of perpetuating ocular surface damage and leading to symptoms of ocular pain, discomfort, and visual phenomena. It begins with stress to the ocular surface leading to the production of proinflammatory mediators that induce maturation of resident antigen-presenting cells which then migrate to the lymph nodes to activate CD4 T cells. The specific antigen(s) targeted by these pathogenic CD4+ T cells remains unknown. Two emerging theories include self-antigens by autoreactive CD4 T cells or harmless exogenous antigens in the setting of mucosal immunotolerance loss. These CD4 T cells migrate to the ocular surface causing additional inflammation and damage. Lifitegrast is the second topical anti-inflammatory agent to be approved by the US Food and Drug Administration for the treatment of DE and the first to show improvement in DE symptoms. Lifitegrast works by blocking the interaction between intercellular adhesion molecule-1 and lymphocyte functional associated antigen-1, which has been shown to be critical for the migration of antigen-presenting cells to the lymph nodes as well as CD4+ T cell activation and migration to the ocular surface. In four large multicenter, randomized controlled trials, lifitegrast has proven to be effective in controlling both the signs and symptoms of DE with minimal side effects. Further research should include comparative and combination studies with other anti-inflammatory therapies used for DE.

Keywords: lifitegrast, SAR1118, dry eye syndrome, inflammation, intercellular adhesion molecule 1, lymphocyte function-associated antigen 1

Introduction
Dry eye (DE) is a chronic ocular condition with high prevalence and morbidity.1 It leads to symptoms of ocular pain, discomfort, and visual dysfunction which can greatly impact the quality of life (QOL).2 DE is multifactorial in nature with a complex pathophysiology.3 One important facet is chronic ocular surface inflammation that can perpetuate ocular surface damage.4 Lifitegrast is the second topical anti-inflammatory agent to be approved by the US Food and Drug Administration (FDA) for the treatment of DE and the first to show improvement in DE symptoms.5 This review discusses the contribution of inflammation in DE in both animal models and humans and highlights the place of lifitegrast in the treatment of the disease.

DE disease definition, classification, and epidemiology
As per the Tear Film and Ocular Surface Dry Eye WorkShop II, DE is a multifactorial ocular surface disease causing ocular symptoms.3 It is characterized by a loss of tear film homeostasis as a result of tear film instability and hyperosmolarity, ocular surface inflammation and damage, or neurosensory pathology.3
DE is classified into two major groups based on the effect on tear film stability: aqueous deficient (decreased production of the aqueous component of the tear film) and evaporative (increased evaporation of the tear film). Aqueous deficient DE disease (ADDE) is the least common and accounts for 14% of DE. ADDE is further classified as Sjögren’s syndrome DE and non-Sjögren’s syndrome DE. Evaporative DE (EDE) is far more common and accounts for 50% of DE. The most common cause is meibomian gland dysfunction (MGD) leading to meibum deficiency. The remaining 36% of DE patients show evidence of both ADDE and EDE.

DE affects 5%–30% of the world population 50 years and older, with women representing approximately two-thirds of those affected. Many factors have been linked to DE, including hormonal alterations (eg, menopause), medication use (eg, antihistamines), and comorbidities (eg, depression). DE can also be seen after ophthalmic procedures such as LASIK, where it has been reported in up to 95% of individuals immediately after surgery and 60% of individuals 1 month after surgery. DE is also seen to a lesser degree following cataract surgery, with an estimated incidence of 9.8%.

DE has a significant impact on QOL due to chronic symptoms of ocular pain and discomfort in addition to functional vision impairment. Utility assessment is a tool for quantifying QOL and comparing QOL across different medical conditions. When applied to patients with moderate to severe DE, utility assessment scores showed a decrease in QOL for everyday and leisure activities by 60%, which was comparable to medical conditions such as severe angina, disabling hip fractures, and dialysis. Furthermore, DE can impact functional vision, interfering with everyday activities such as driving, reading, computer usage, watching television, and work performance. In addition, DE negatively impacts mental health as DE patients report higher rates of anxiety and depression. Financially, DE is estimated to cost $3.84 million per year to the health care system with an average annual cost of $783 per patient for treatment alone.

DE symptoms and signs

Currently, there is no "gold standard" test to diagnose DE. Instead, a combination of presenting signs and symptoms is used in clinical judgment. Further complicating the diagnosis of DE is the poor correlation that exists between DE symptoms and signs.

DE symptoms

The evaluation of DE symptoms can be challenging due to varying presentations and course. For example, some individuals predominantly complain of spontaneous ocular pain/discomfort characterized by different terms (eg, dryness, grittiness, burning, stinging, etc), while other patients may report evoked pain, in particular by wind and light. Yet others report visual phenomena in the form of blurry or fluctuating vision. Furthermore, some individuals describe transient symptoms, while other have a chronic disease course. Several standardized questionnaires have been developed to assess DE symptoms and their effect on QOL. These include the Dry Eye Questionnaire 5, the Ocular Surface Disease Index (OSDI), the Impact of Dry Eye on Everyday Life, and Visual Questionnaire-25. Most arrive at a DE severity score by lumping together various DE symptoms and there is no consensus on which questionnaire provides the best assessment tool.

DE signs

A careful slit-lamp evaluation with a few additional tests can reveal many signs of DE. These signs include decreased tear production measured by Schirmer tear test, tear film instability measured by tear-film breakup time, and corneal or conjunctival epithelial damage measured by the degree of punctate epithelial erosions, best seen after administration of fluorescein, rose bengal, or lissamine green dyes. Signs of MGD such as plugging of the meibomian glands, eyelid telangiectasias, and abnormal meibum quality may also be present.

Point-of-care tests can be used to evaluate subclinical markers of DE, namely, ocular surface inflammation and tear osmolarity. Ocular surface inflammation is assessed by measuring the levels of matrix metalloproteinase (MMP)-9, a protease produced by the epithelial cells in response to inflammatory stress (Inflammadry; Quantel, San Diego, CA, USA). Tear film osmolarity is measured with a portable osmometer (TearLAB, San Diego, CA, USA).

Lacrimal functional unit and DE pathophysiology

The lacrimal functional unit (LFU) is composed of the eyelids, lacrimal glands (main and accessory), meibomian glands, cornea, conjunctiva, and their accompanying sensory, autonomic, and motor innervation. The LFU has three major functions as follows:

- to maintain a pure optical surface for light refraction,
- to maintain ocular surface homeostasis, and
- to upregulate and downregulate immune response as needed.

The LFU accomplishes these goals by regulation of neuronal and hormonal pathways that ultimately control tear
flow and quality. In a commonly presented schematic, the tear film is statically illustrated as having three layers—an outer lipid layer, a middle aqueous layer, and an inner mucinous layer. The reality, however, is likely more complex as dynamic interactions have been demonstrated between lipids (phospholipids, fatty acids, and cholesterol) in meibum and proteins (lipocalin) in the aqueous layer.\(^{39,40}\)

Despite the complexity, it is generally agreed that lipids are produced by meibomian glands located in the tarsal plate.\(^{41}\) They drain a clear oil called meibum that helps reduce the rate of evaporation of the aqueous layer.\(^{42}\)

The aqueous layer is the largest component of the tear film and is produced by the lacrimal glands (main and accessory) with smaller contributions from the conjunctival epithelium.\(^{43}\) It is mostly composed of salts that maintain an adequate osmolar gradient and proteins with a wide array of functions. These include growth factors (eg, epidermal growth factor, hepatocyte growth factor, platelet-derived growth factor)\(^{44}\) and defense proteins (eg, lactoferrin, lysozyme, immunoglobulin A)\(^{45}\) that are important for ocular surface repair and immunologic defenses. Anti-inflammatory proteins are also found, including transforming growth factor (TGF)-\(\beta\), which suppresses antigen-presenting cells (APCs), and tissue inhibitor metalloproteinases (TIMP-1 and TIMP-2), which suppress corneal neovascularization and limit inflammatory cell trafficking into the ocular surface.\(^{36-48}\) Anti-inflammatory omega-3-derived lipids (lipoxins, resolvins, and neuprotectin) are released into the tear film by resident regulatory polymorphonuclear neutrophils, corneal epithelial cells, and the lacrimal gland.\(^{49-54}\)

The mucin layer is produced by conjunctival goblet cells (GCs) which secrete a large gel-forming mucin (MUC5AC) with contributions by the lacrimal glands producing a smaller and soluble mucin (MUC7).\(^{55}\) The mucin layer interacts with the glyocalyx of ocular surface epithelial cells, forming a dense barrier particularly against pathogens, thereby playing an important role in mucosal immunotolerance.\(^{55,56}\) In addition, the mucin layer serves to trap debris and sloughed off epithelial cells as well as facilitates sliding of the eyelid against the corneal and conjunctival epithelium during blinking.\(^{57,58}\)

A unifying mechanism in DE is the disruption of one or more components of the LFU, leading to tear film instability and impairment of one or more of its three main functions. The loss of a pure optical surface can lead to aberrant light refraction which can create various visual phenomena.\(^{54,57}\) Tear film instability can give rise to a hyperosmolar tear film either by rapid evaporation of the aqueous component or by normal evaporation of a reduced aqueous component.\(^{59}\) This creates a hostile environment for ocular surface epithelial cells which respond by undergoing apoptosis and releasing proinflammatory mediators as they are injured.\(^{60,61}\) Inflammation is amplified by the loss of anti-inflammatory mechanisms of the LFU.\(^{38}\) Persistent inflammation further damages the ocular surface and can impact sensory nerves, thus dampening reflex tear secretion and causing further tear film instability.\(^{1}\) A feedback cycle can ensue with chronic inflammation leading to more ocular surface damage and, thus, more inflammation.

### Inflammation in animal models of DE

Ocular surface inflammation begins with a rapid, but nonspecific innate immune response to stress. In mice, hyperosmolar stress on the ocular surface triggered the release of inflammatory mediators such as interleukins (IL)-1\(\beta\), tumor necrosis factor (TNF)-\(\alpha\), and MMP-9, which initiate the mitogen-activated protein kinase (MAPK) signaling pathways c-jun n-terminal kinases, extracellular-regulated kinases, and p38.\(^{62}\) In a similar manner, desiccating stress (DS), created by scopolamine plus low humidity, leads to the induction of a wide variety of inflammatory mediators on the ocular surface, including IL-1\(\alpha\), IL-1\(\beta\), CC chemokine ligand (CCL)2, CCL3, CCL5, C-X-C motif chemokine ligand (CXCL)10, TNF-\(\alpha\), interferon (IFN)-\(\gamma\), IL-2, IL-6, IL-10, and MMP-9. Again, this inflammatory milieu was found to activate the MAPK signaling pathways.\(^{63,64}\) Interestingly, aging mice were found to spontaneously develop ocular surface inflammation, with elevated IFN-\(\gamma\) and IL-7\(\alpha\) on the ocular surface.\(^{65}\)

This innate immune response gives way to a slower, but more specific adaptive immune response requiring a complex interaction between APCs, namely, macrophages and dendritic cells (DCs), and CD4\(^+\) T cells. In a study by Schaumburg et al,\(^{66}\) acute cytokine production from the initial innate immune response phase of DS-induced DE mice was associated with an increased number of CD11c\(^+\) DCs and increased expression of DC maturation markers (major histocompatibility complex II, CD83, CD86, C-C motif receptor 7) which preceded activation of CD4\(^+\) T cells.

Once activated by APCs, CD4\(^+\) T cells infiltrate and damage the ocular surface with corneal irregularity, corneal barrier disruption, and decreased conjunctival GCs noted.\(^{67}\) The specific antigen(s) targeted by these pathogenic CD4\(^+\) T cells remains unknown.

One possibility is that T cell response is aimed at self-antigens of the cornea and conjunctiva. In fact, adoptive transfer of CD4\(^+\) T cells from DS-induced DE mice to nude mice was enough to reproduce disease with CD4\(^+\) T cells.
localizing to the lacrimal gland, cornea, and conjunctiva in the nude mice. Adhesive transfer of CD4+ T cells from aging mice with signs of DE was similarly effective in replicating disease. The important interplay between the APCs and T cells became evident as APC deletion prior to DS induction mitigated the ability of T cells to recreate the disease in nude mice recipients.

T regulatory cells (Tregs) also play an important role as inflammatory modulators in DE mice. Adoptive transfer of pathogenic CD4+ T cells from DS-induced DE mice to nude mice is effectively attenuated by reconstitution of Treg cells in nude recipient mice. In DE mice, Treg dysfunction contributes to pathogenesis as Tregs from DS-induced DE mice were less capable of suppressing proliferation of T helper (Th)17 cells, leading to higher expression of IL-17 and higher number of Th17 cells in regional lymph nodes.

Another possibility is that T cell response is aimed at harmless exogenous antigens as part of a derailed immunologic response in the setting of dysfunctional mucosal immunotolerance. Guzman et al studied the role of mucosal tolerance in DS-induced DE mice. Ocular exposure to ovalbumin (OVA) antigen in wild-type (WT) mice led to immunotolerance demonstrated by reduced in vitro antigen-specific T cell proliferation and in vivo delayed-type hypersensitivity (DTH) response to OVA immunization. Immunotolerance was also retained early on in DS-induced DE mice when they were exposed to ocular OVA on day 1 of DS. However, when OVA instillation was applied on day 4 of DS, DS-induced DE mice exhibited loss of immunotolerance with elevated in vitro antigen-specific T cell proliferation and in vivo DTH response to OVA immunization, suggesting time-dependent deterioration of ocular mucosal tolerance. Inhibition of NF-κB, a key regulator of mucosal tolerance, restored mucosal tolerance and decreased corneal staining and inflammatory markers (IL-1β and IL-6) in DS-induced DE mice. Similarly, time-dependent mucosal tolerance loss and mitigation of corneal damage were demonstrated in a DE mouse model created by resection of extraorbital lacrimal glands.

Loss of GCs is often noted in DE models and may contribute to mucosal immunotolerance loss, given their critical role in modulating antigen distribution and exposure to adjacent APCs. Barbosa et al studied the role of GCs in immune tolerance using SAM pointed domain ETS transcription factor knockout (Spdef<sup>−/−</sup>) mice, a DE mouse model that lacks GCs and exhibits signs of DE. In WT mice, topically applied antigen OVA was effectively delivered to the stroma through GC-associated passages for uptake by adjacent APCs (CD11b<sup>+</sup> F4/80<sup>+</sup> macrophages), while Spdef<sup>−/−</sup> mice retained OVA within the epithelium. APCs isolated from conjunctival draining cervical lymph nodes of Spdef<sup>−/−</sup> mice showed stronger induction of antigen-specific lymphoproliferation, greater IFN-γ production, and lesser Treg proliferation, compared to WT. These findings were consistent with the loss of immune tolerance observed in Spdef<sup>−/−</sup> mice compared to WT, as assessed by cutaneous DTH to OVA following immunization with complete Freund’s adjuvant mixed with OVA.

In addition to regulating antigen exposure to DCs, GCs play a role in modulation of DCs phenotype. Studies by Contreras-Ruiz and Masli revealed that GCs TSP-1-dependent expression of TGF-β, particularly the TGF-β<sub>2</sub> isoform, plays a role in modulation of DC phenotype. When DCs were co-cultured with WT globet cells from mice, they were found to have reduced major histocompatibility complex II and costimulatory molecules (CD80 and CD86) expression, compared to cultures of DCs alone. This effect was dependent on TSP-1 expression by GCs.

Irrespective of whether T cell activation is triggered by self-antigens or exogenous antigens, T cell migration to the ocular surface is a key event in DE and is driven by a variety of inflammatory cytokines and chemokines as well as other proteins. MRL/lpr mice are homozygous for the recessive <i>lpr</i> (lymphoproliferative) gene and used as Sjögren’s syndrome DE models. These mice have high expression of intercellular adhesion molecule (ICAM)-1, an adhesion molecule important for homing and activation of infiltrating lymphocytes, in the conjunctival epithelium and vascular endothelium along with infiltrating lymphocytes within the lacrimal gland tissue. In fact, ICAM-1 expression is positively correlated with disease progression and severity.

The importance of these inflammatory mediators is further highlighted by mouse studies showing improvement of DE with the use of anti-inflammatory therapy targeting IL-1, IL-17, and C-C motif receptor 2. Furthermore, T cell infiltration decreased with the use of monoclonal antibodies against ICAM-1 and its receptor lymphocyte functional associated antigen-1 (LFA-1).

**Inflammation in humans with DE**

Hyperosmolarity is a known trigger of inflammation. Cell culture experiments demonstrated increased proinflammatory mediators (eg, IL-1, IL-6, TNF-α, and MMP-9) after human corneal epithelium was subjected to hyperosmolar stress. Similar results were noted in limbal epithelial cells with elevated levels of IL-1β, IL-8, and TNF-α found through
the c-jun n-terminal kinase and extracellular-regulated kinase MAPK signaling pathway.\textsuperscript{61}

Similar to mice, patients with DE (defined in a number of different ways) have increased tear levels of proinflammatory cytokines, chemokines, and chemokine receptors including IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IL-13, IL-17, IL-21, IFN-\(\gamma\), TNF-\(\alpha\), CXCL9, CXCL10, CXCL11, CXCR3, IL-1Ra, IP-10/CXCL10, fractalkine/CX3CL1, CCL-5, and vascular endothelial growth factor.\textsuperscript{83–90} In fact, levels of IL-6, IL-17, IL-21, TNF-\(\alpha\), CXCL-9, CXCL-10, and CXCL-11 were found to be positively correlated with DE signs and symptoms (eg, Schirmer test, tear-film breakup time, punctate epithelial erosions, GC density, OSDI).\textsuperscript{98,90–92}

Patients with DE have other features in common with mouse models, including T cell infiltration of the ocular surface and elevated ICAM expression. Stern et al\textsuperscript{93} studied patients with moderate to severe ADDE and found CD4\(^+\) T cell infiltration in the conjunctiva, along with increased markers of immune activation (HLA-DR and HLA-DQ). ADDE patients were also found to have increased ICAM-1 expression in the conjunctival epithelium and lacrimal gland venule endothelium, near CD4\(^+\) T cells.\textsuperscript{77–95}

Inflammation in DE can also affect the corneal nerves, disrupting normal tearing reflex and blink rate.\textsuperscript{96} Many studies using in vivo confocal microscope, a minimally invasive tool for imaging ocular surface, have shown decreased sub-basal nerve density of the cornea\textsuperscript{97–103} and morphologic changes of the sub-basal nerves (increased tortuosity, nerve sprouting, increased bead-like formation, and decreased reflectivity) in DE patients.\textsuperscript{97–99,101,104,105} Additionally, DE patients generally show higher DC density and morphologic changes including larger DC size and additional dendrites at the sub-basal nerve plexus.\textsuperscript{98,100,104,106,107} A few studies have attempted to correlate in vivo confocal microscope parameters to DE signs and symptoms. Decreased sub-basal nerve density is correlated to decreased corneal sensitivity,\textsuperscript{99,102} increased DC density at the sub-basal nerve plexus,\textsuperscript{98} and increased DE symptoms and signs.\textsuperscript{97,98,108} On the other hand, studies correlating DC density to DE signs and symptoms have yielded conflicting results.\textsuperscript{100,107,109}

\section*{Treatment modalities: targeting inflammation}

\subsection*{Corticosteroids}

\textbf{Mechanism of action}

Topical corticosteroids decrease inflammation primarily by binding to glucocorticoid receptors and regulating the expression of anti-inflammatory and proinflammatory genes.\textsuperscript{110} Specifically, corticosteroids suppress NF-\(\kappa\)B, a key transcription factor in inflammation, leading to suppression of proinflammatory mediators and promoting lymphocyte apoptosis. There is a wide range of proinflammatory mediators that are suppressed by corticosteroids, including ICAM-1, MMPs, prostaglandins, cytokines, chemokines, and phospholipase A2. Non-genomic effects of corticosteroids also aid in suppressing leukocyte infiltration into areas of inflammation.\textsuperscript{111}

\textbf{Clinical data in DE}

Topical corticosteroids have proven effective in the treatment of DE in several studies.\textsuperscript{112–116} One of these studies included a multicenter, randomized, double-masked, placebo-controlled clinical trial with 64 moderate to severe ADDE patients receiving either 0.5\% loteprednol or placebo four times a day for 4 weeks.\textsuperscript{115} This study evaluated symptoms on a visual analog scale and signs by corneal staining, conjunctival injection, and lid margin injection. At 2 weeks, treatment with 0.5\% loteprednol improved the signs of DE, including lid margin injection and conjunctival injection. Furthermore, in a subset analysis of patients with moderate clinical inflammatory component, treatment with 0.5\% loteprednol improved corneal staining, conjunctival injection, and DE symptoms (eye redness).

\textbf{Side effects}

The use of corticosteroids is limited due to a wide range of side effects including glaucoma, ocular infection, corneal thinning, and formation of cataracts.\textsuperscript{117} Given the chronic nature of DE, topical corticosteroid use tends to be limited to short-term treatment of DE exacerbations.

\subsection*{Cyclosporine}

\textbf{Mechanism of action}

Cyclosporine is FDA approved for the treatment of DE signs. It suppresses inflammation by binding to cyclophilins and inhibiting calcineurin, a calcium-dependent phosphatase, thereby preventing nuclear factor of activated T-cells, cytoplasmic 1 (NFATc1) dephosphorylation which reduces IL-2 levels suppressing T cell activation.\textsuperscript{118}

\textbf{Formulation}

Cyclosporine requires suspension and emulsion forms due to its poor solubility in water. Restasis is formulated with 0.05\% cyclosporine in an emulsion of glycerin (2.2\%), castor oil (1.25\%), polysorbate 80 (1.00\%), carbomer copolymer type A (0.05\%), purified water (to 100\%), and sodium hydroxide for pH adjustment.\textsuperscript{119}
Clinical data in DE
Studies with topical cyclosporine treatment in patients with moderate to severe DE generally showed significant improvement of DE signs and variable improvement of DE symptoms. Large multicenter, randomized, double-blind, controlled studies with moderate to severe DE patients have shown improvement in DE signs (corneal staining, tear production) over emulsion placebo. Symptomatic improvement in these studies was more variable, with the most consistent improvement found in reports of “dryness” symptoms. Similarly, smaller prospective randomized studies of moderate to severe DE patients treated with cyclosporine or combinations of cyclosporine (cyclosporine plus artificial tears [AT] or cyclosporine plus vitamin A) consistently showed improvement in DE signs (corneal staining, conjunctival staining, GC density, tear production, tear film stability) and, in some cases, DE symptoms (OSDI, blurry vision, burning, and photophobia).

Side effects
Cyclosporine use is limited by its side effects of mild to moderate burning and irritation of the eye. Tolerance is improved with the use of concomitant topical corticosteroids.

Lifitegrast
Mechanism of action
Lifitegrast (formerly SAR1118) is the only FDA-approved treatment for both signs and symptoms of DE. Lifitegrast suppresses inflammation by mimicking ICAM-1, thus blocking the interaction between ICAM-1 and LFA-1, a cell surface protein belonging to the β2 family of integrins and found on leukocytes (Figure 1). LFA-1/ICAM-1 interaction has been implicated in various aspects of lymphocyte activation and migration. It promotes migration of DC to lymph nodes where they can activate naïve T cells. It also plays a direct role in T cell activation by maintaining initial contact between naïve T cells and APCs in lymph nodes and enhancing T cell sensitivity to antigens by stabilization of the immunologic synapse between naïve T cells and APCs in the lymph nodes. Beyond T cell activation, LFA-1/ICAM-1 interaction also plays a role in T cell migration by allowing firm adhesion to the endothelium with subsequent transmigration into inflamed tissue. By blocking the interaction between LFA-1 and ICAM-1, lifitegrast is able to disrupt various key steps in T cell–mediated inflammation.

Figure 1  Mechanism of action of lifitegrast.
Notes: Lifitegrast blocks ICAM-1 and LFA-1 interaction, which is critical in migration of DCs to lymph nodes, naïve T cell activation by DCs, and T cell transmigration into the ocular surface.
Abbreviations: APCs, antigen-presenting cells; DC, dendritic cell; ICAM-1, intercellular adhesion molecule; LFA-1, lymphocyte functional associated antigen-1.
Lifitegrast is prepared in PBS with a pH, tonicity, and osmolality range consistent with other currently approved topical ophthalmic solutions. It is preservative-free and comes in single-unit dose ampules.

Animal studies with lifitegrast in non-DE conditions
A few animal studies have shown the anti-inflammatory effect of lifitegrast in various inflammatory-based ocular diseases. A study by Sun et al tested the efficacy of lifitegrast in inhibiting inflammation in mice with corneal inflammation induced by antibiotic-killed Pseudomonas aeruginosa and Staphylococcus aureus in the presence of contact lenses. Lifitegrast reduced neutrophilic infiltration into the cornea, reduced clinical signs of corneal inflammation, and prevented P. aeruginosa- and S. aureus- induced inflammation compared to controls. In rat streptozotocin model of diabetic retinopathy, lifitegrast was able to significantly reduce the number of adherent leukocytes and level of myeloperoxidase, a leukocyte-derived proinflammatory protein, to levels comparable to those of normal retina. Lifitegrast also significantly reduced blood–retinal barrier breakdown, compared to vehicle-control.

Animal studies with lifitegrast in DE
A study of 10 idiopathic keratoconjunctivitis sicca dogs receiving 1% lifitegrast three times a day for 12 weeks showed a significant increase in tear production from baseline for 11 of 18 study eyes, which corresponded with a decreased inflammatory cell infiltrate of the conjunctiva from baseline.

Human studies with lifitegrast
In vitro studies demonstrate lifitegrast’s ability to inhibit human T cell binding to ICAM-1 and inhibit the release of proinflammatory cytokines including IFN-γ, TNF-α, macrophage inflammatory protein-1α, IL-1α, IL-1β, IL-2, IL-4, and IL-6 from activated lymphocytes. Additionally, imaging studies reveal inhibition of immunologic synapse formation in the presence of lifitegrast.

The clinical efficacy of lifitegrast in the treatment of signs and symptoms of DE has been demonstrated across four prospective, multicenter, randomized, double-masked, vehicle-controlled trials (Table 1). The first of these trials was a phase II clinical trial of 230 patients with moderate to severe DE receiving either 0.1%, 1%, or 5% lifitegrast vs vehicle twice a day for 84 days. Key eligibility criteria included adult DE patients with corneal staining score ≥2, un-anesthetized Schirmer >1 and <10, no active lid margin disease, and worsening of signs (inferior corneal staining score [ICSS] increase ≥1 point) and symptoms (ocular discomfort score [ODS] increase ≥1 point) in response to controlled adverse environment (CAE), an environment with specified humidity, temperature, airflow, and lighting designed to worsen DE. The use of topical cyclosporine and AT was prohibited 6 weeks and 3 days from visit 1, respectively. Three visits occurred during the treatment period at days 14, 42, and 84 of treatment during which patients were exposed to additional 90-minute sessions of CAE. Lifitegrast (1% and 5%) showed significant improvement in ICSS (primary sign endpoint) at 84 days from baseline, compared to vehicle. This study also showed improvement in tear production with 5% lifitegrast at 14 days from baseline, compared to vehicle. In terms of symptoms, all lifitegrast groups showed significant improvement in the OSDI at 14 days and 1% and 5% lifitegrast showed significant improvement in the vision-related subscale score of OSDI at 14 and 84 days from baseline, compared to vehicle. Baseline ODS showed trends toward improvement, but was not significantly different from vehicle.

This was followed by the first Phase III trial, OPUS-1, consisting of 588 patients with moderate to severe DE receiving 5% lifitegrast vs vehicle twice a day for 84 days. Unlike the previously described Phase II study, CAE was only used during screening visits and not during the in-treatment visits. Once again, lifitegrast significantly improved ICSS (primary end point) at 84 days from baseline, compared to vehicle. Additional improvements in DE signs over vehicle included superior corneal staining, total corneal staining, nasal conjunctival staining, and conjunctival staining, with some of these changes starting as early as at 14 days of treatment. Lifitegrast significantly improved DE symptoms over vehicle, including decreased ODS and visual analog eye dryness score (VAS-EDS); however, this study failed to show significant improvement in vision-related subscale score of OSDI (primary symptom end point) over vehicle.

The next Phase III trial was OPUS-2 consisting of 718 patients with moderate to severe DE receiving 5% lifitegrast vs vehicle twice a day for 84 days. Study methods were similar to the previous two studies; however, no CAE was used in this study and AT use within 30 days and VAS-EDS ≥40 were added to the inclusion criteria based on post hoc analysis from OPUS-1, suggesting that the drug effect is increased in patients with recent AT use and VAS-EDS ≥40. In this study, primary symptom end point was met as lifitegrast showed significant improvement from baseline in VAS-EDS.
### Table 1: Summary of studies using LIF for the treatment of dry eye

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<th>Reference</th>
<th>Evidence level</th>
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<th>Population</th>
<th>Dose/treatment length</th>
<th>Efficacy LIF vs control</th>
<th>LIF vs baseline*</th>
<th>Side effects</th>
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<td>(84 days with 1% and 5% LIF)</td>
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<td>Symptoms: OSDI (14 days)</td>
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<td>(84 days with 1% LIF) VR-OSDI (14 and 84 days with 1% and 5% LIF),</td>
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<td>VAS (84 days, burning/stinging only)</td>
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<td>VR-OSDI, ODSI, VAS</td>
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<td>Sheppard et al.</td>
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<td>• Conjunctival redness in any eye</td>
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<td>• CSS = 2 in any eye</td>
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<td>vehicle-</td>
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<td>• Schl ≥1 and &lt; 10 in any eye</td>
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<td>• BCVA ≥0.7 logMAR in both eyes</td>
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<td>study</td>
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<td>• CAE-induced increase in ODS ≥1 point in designated eye</td>
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<td>• Vs vehicle twice daily for 84 days. No CAE</td>
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<td>Signs: ICSS (84 days), SCSS</td>
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<td>(14 and 84 days)</td>
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<td>VR-OSDI, ODSI, VAS</td>
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<td>Tauber et al.</td>
<td>Level 1A</td>
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<td>• AT use within 30 days</td>
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<td></td>
<td>• Site irritation</td>
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<td></td>
<td>• BCVA ≥0.7 logMAR in both eyes</td>
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<td>• Mild to</td>
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<td></td>
<td>double-blind,</td>
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<td>• CSS ≥2 in ≥1 eye region, CRS ≥1 in ≥1 eye</td>
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<td>• Transient</td>
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<td></td>
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<td>• VAS-EDS ≥40</td>
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<td>LIF better:</td>
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<td>• Vs vehicle twice daily for 84 days. No CAE</td>
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<td>Signs: ICSS, nasal conjSS, TCSS</td>
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<td>No difference: ICSS, TCSS, nasal conjSS</td>
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<td>• Dysgeusia</td>
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<td>Holland et al.</td>
<td>Level 1B</td>
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<td>• Instillation</td>
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<td>• AT use within 30 days</td>
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<td>• Site irritation</td>
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<td>• BCVA ≥0.7 logMAR in both eyes</td>
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<td>double-blind,</td>
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<td>• CSS ≥2 in ≥1 eye region, CRS ≥1 in ≥1 eye</td>
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<td>• Transient</td>
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<td></td>
<td>vehicle-</td>
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<td>• VAS-EDS ≥40</td>
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<td>• Schl ≥1 and ≤10 in designated eye</td>
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<td>study</td>
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<td>• ICSS ≥0.5 in designated eye</td>
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<td></td>
<td>• Vs vehicle twice daily for 84 days. No CAE</td>
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<td>Signs: ICSS (84 days, ad hoc analysis)</td>
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<td>Symptoms: VAS-EDS (14, 42, and 84 days), VAS (itching, foreign body sensation, eye discomfort only at 42 days)</td>
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<td>• Site reaction</td>
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<td></td>
<td>No difference: ODS, VAS (burning, photophobia, pain)</td>
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<td>• Dysgeusia</td>
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**Notes:** Each study has been categorized from Level 1 through 4 as follows: Level 1A- a randomized, double-masked design; Level 1B- a randomized, double-masked design, with weak patient masking due to significant variations in treatment vs placebo side effect profiles; Level 2A- a randomized, single-masked design; Level 2B- a randomized, single-masked design, with weak masking due to significant variations in treatment vs placebo side effect profiles; Level 3- randomized, non-masked; Level 4- nonrandomized, non-masked. Primary end points are shown in bold, while secondary end points are shown in italics. *Signs and symptoms are listed where the LIF group had improved parameters compared to baseline but not compared to control.

**Abbreviations:** AT, artificial tears; BCVA, best-corrected visual acuity; CAE, controlled adverse environment; conjSS, conjunctival staining score; CRS, conjunctival redness score; CSS, corneal staining score; ICSS, inferior corneal staining score; LIF, lifitegrast; logMAR, log of minimum angle of resolution; ODS, ocular discomfort score; OSDI, Ocular Surface Disease Index; SCSS, superior corneal staining score; Schl, Schirmer’s without anesthesia; TBUT, tear breakup time; TCSS, total corneal staining score; VAS, visual analog scale; VAS-EDS, visual analog scale eye dryness score; VR-OSDI, vision-related subscale score of OSDI.
as early as 14 days and continuing to 84 days, over vehicle. Additionally, lifitegrast was superior in decreasing ODS at 84 days from baseline, compared to vehicle. Primary sign end point of ICSS showed improvement from baseline, but did not reach significant difference compared to vehicle. The authors hypothesize that the lack of ICSS improvement was related to patients’ recent AT use which hindered detection of drug effect, or the enrollment of more severe DE patients with more advanced ocular surface damage.

The final phase III trial was OPUS-3 consisting of 711 patients with moderate to severe DE receiving 5% lifitegrast vs vehicle twice a day for 84 days. Study methods were similar to those of OPUS-2. This study also met its primary symptom end point of VAS-EDS improvement from baseline compared to vehicle starting at 14 days and continuing to 84 days. Other symptoms significantly improved with the use of lifitegrast over vehicle, including visual analog scale for itching, foreign body sensation, and eye discomfort at 42 days of treatment.

**Safety studies with lifitegrast**

The safety profile of lifitegrast has been explored in several dose-escalation tolerance studies (Table 2). Dose-escalation tolerance studies using healthy dogs with a maximum

### Table 2 Summary of safety studies using LIF for the treatment of dry eye

<table>
<thead>
<tr>
<th>Reference</th>
<th>Evidence level</th>
<th>n</th>
<th>Population</th>
<th>Dose/treatment length</th>
<th>TEAEs</th>
</tr>
</thead>
</table>
| Semba et al 131     | Level IA       | 28 | Americans 18–50 years of age with the following:  
• Nonsmoker status  
• BCVA ≥20/40  
• BMI of 19.9–29.9  
• No clinically significant acute or chronic medical condition  
• No history of ocular inflammation | LIF 0.1%, 0.3%, 1%, 5% vs placebo.  
Three dosing periods (QD × 10 days, TiD × 10 days) separated by 3 days treatment | Ocular: Eye irritation (n=5 with LIF), ocular hyperemia (n=3 with LIF)  
Non-ocular: Headache (n=4 with LIF), erythema (n=2 with LIF), and musculoskeletal pain (n=2 with LIF)  
No difference: IOP, vital signs, ECG, slit-lamp exam, BCVA, TBUT, Schl, chronic suppression of lymphocyte subsets (CD3, CD4, CD8 counts), hematologic and chemistry panel  
Overall TEAEs severity: Mild |
| Paskowitz 132       | Level IA       | 13 | Americans ≥18 years old undergoing pars plana vitrectomy for various indications including epiretinal membrane, vitreomacular traction, vitreous hemorrhage, dislocated IOL, and intraocular inflammation | LIF 0.1%, 1%, 5% BID for 1 week | Ocular: Transient stinging (31%) with LIF 5%  
Non-ocular: Dysgeusia (23%) with LIF 5%  
No difference: No LIF-related delays in postoperative healing |
| Donnenfeld et al 133 | Level IA       | 332| Americans ≥18 years old with the following:  
• Self-reported history of dry eye  
• BCVA ≥0.7 logMAR  
• CSS ≥2 in ≥1 region  
• VAS ≥40 in either eye dryness or eye discomfort  
• Use or desire to use AT substitute in the past 6 months  
• Schl ≥1 and ≤10 | LIF 5% vs vehicle BID for 1 year | Ocular: Instillation site irritation (15% LIF vs 4.5% placebo), instillation site reaction (13.2% LIF vs 1.8% placebo). Drop comfort improved at each visit. Minimal visual acuity reduction (11.4% LIF vs 6.3% placebo), not considered LIF related. Dry eye (1.8% LIF vs 5.4% placebo), not considered LIF related  
Non-ocular: Dysgeusia (16.4% LIF vs 1.8% placebo)  
No difference: IOP, cataracts, chronic suppression of lymphocyte subsets (CD3, CD4, CD8 counts), hematologic panel, renal panel, and liver panel  
Overall TEAEs severity: Mild to moderate  
Concomitant AT use: Higher rates of ocular TEAEs in AT + LIF vs LIF (67.2% vs 45%) and AT + placebo vs placebo (44.2% vs 32.7%). Higher rates of non-ocular TEAEs in AT + LIF vs LIF (60.9% vs 42.7%) and AT + placebo vs placebo (44.2% vs 32.7%). Lower discontinuation due to TEAEs in AT + LIF vs LIF (3.1% vs 5.4%) and AT + placebo vs placebo (0% vs 4.4%) |

**Notes:** Each study has been categorized from Level I through 4 as follows: Level IA- a randomized, double-masked design; Level IB- a randomized, double-masked design, with weak patient masking due to significant variations in treatment vs placebo side effect profiles; Level 2A- randomized, single-masked design; Level 2B- a randomized, single-masked design, with weak masking due to significant variations in treatment vs placebo side effect profiles; Level 3- randomized, non-masked; Level 4- non-randomized, non-masked.

**Abbreviations:** AT, artificial tears; BCVA, best-corrected visual acuity; BID, twice a day; BMI, body mass index; CSS, corneal staining score; ECG, electrocardiogram; IOL, intraocular lens; IOP, intraocular pressure; LIF, lifitegrast; logMAR, log of minimum angle of resolution; QD, daily; Schl, Schirmer’s without anesthesia; TBUT, tear breakup time; TEAE, treatment-emergent adverse event; TiD, three times a day; VAS, visual analog scale.
dosage of 10% lifitegrast administered three times per day for 1 month did not demonstrate any adverse effect on the ocular surface.

Similarly, dose-escalation tolerance assessment with a maximum dosage of 3% lifitegrast administered three times per day for 13 weeks did not show any adverse effect on the ocular surface, but was associated with a transient period of blinking and squinting upon drop administration, which appeared to improve after the first few days of instillation.

Short-term dose-escalation tolerance studies of healthy individuals and patients undergoing pars plana vitrectomies have tested lifitegrast in dosages ranging from 0.1% to 5% at frequencies as high as three times per day. In these studies, adverse events from lifitegrast were mild to moderate in severity and consisted of transient ocular irritation and ocular hyperemia. Non-ocular adverse events included dysgeusia, headaches, erythema, and musculoskeletal pain. No effects were seen in intraocular pressure, visual acuity, ocular surface exams, healing time, and hematologic and chemistry panels. SONATA was a multicenter, randomized, double-blind, vehicle-controlled trial that was conducted to assess the long-term safety profile of lifitegrast 5% vs vehicle twice daily for 1 year in 331 DE patients. Eligibility criteria included Schirmer tear test result ≥1 and ≤10 mm in 5 minutes, no active lid margin disease, corneal staining score ≥2.0, EDS ≥40, and use and/or desire to use AT in the past 6 months. Patients receiving lifitegrast had a higher percentage of treatment-emergent adverse effects (TEAEs; 53.6% lifitegrast vs 34.2% vehicle), most of which were mostly transient and mild to moderate in severity. TEAEs included instillation site irritation (15.0% lifitegrast vs 4.5% vehicle), instillation site reaction (13.2% lifitegrast vs 1.8% vehicle), reduction in visual acuity (11.4% lifitegrast vs 6.3% vehicle), DE (1.8% lifitegrast vs 5.4% vehicle), and dysgeusia (16.4% lifitegrast vs 1.8% vehicle). Drop comfort improved at each visit. Dysgeusia was likely the result of tear drainage into the oropharynx, while decreased visual acuity may have been due to transient alterations in the tear film leading to altered refractive properties. Despite the decreased visual acuity, changes in best-corrected visual acuity from baseline to 1 year were minimal in both groups. No serious ocular adverse events occurred in this study. All non-ocular TEAEs, except for dysgeusia, were considered to be unrelated to lifitegrast. There was no evidence of immunosuppression as per CD3, CD4 and CD8 serum levels and no alterations in hematologic, renal and liver panels. Concomitant AT use appeared to increase the rates of ocular TEAEs in both groups, but was associated with lower discontinuation rates due to TEAEs in both groups.

Conclusion and remaining questions
Evidence suggests that ocular surface inflammation is an important component of DE. Stress to the ocular surface stimulates production of proinflammatory mediators inducing maturation of resident APCs that migrate to the lymph nodes to activate autoreactive CD4\(^+\) T cells which migrate to the ocular surface causing more inflammation and damage. Lifitegrast blocks the interaction between ICAM-1 and LFA-1, which has been shown to be critical for APC migration to the lymph nodes as well as CD4\(^+\) T cell activation and migration to the ocular surface. In four large multicenter, randomized controlled trials, lifitegrast has proven to be effective at controlling both the signs and symptoms of DE with minimal side effects.

Despite its success, many questions remain. It is known that not all individuals with DE symptoms have detectable levels of inflammation as measured by ocular surface MMP-9 levels. DE patients with ADDE are more likely to have inflammation, especially in the setting of systemic inflammation such as Sjögren’s syndrome and graft vs host disease. Could these patients also be more likely to respond to anti-inflammatory therapy such as lifitegrast?

Another question is whether anti-inflammatory therapies for DE could work better in combination. While studies of topical cyclosporine and methylprednisolone combinations showed faster symptomatic relief, at this time, no combination studies have been done with lifitegrast. Additionally, no comparative studies have been done with lifitegrast to determine if one anti-inflammatory agent could be better than the others. These are all important avenues of future investigation.

Of note, we focused our review on the inflammatory component of DE, given our focus on lifitegrast as a new therapeutic modality in DE. However, it is known that not all patients with DE symptoms have ocular surface inflammation (as measured by Inflam madry). Other contributors to DE include MGD and resultant EDE. The role of inflammation in the initiation and propagation of MGD is less well clarified. Furthermore, many other exposures contribute to DE, which do not clearly fit into the autoreactive and/or loss of mucosal tolerance story. These include environmental exposures such as air pollution and low humidity, dietary patterns such as a high consumption of free fatty acids, and psychosocial considerations such as depression and chronic widespread pain, to name a few. The role of inflammation,
along with the other contributors, needs to be considered when evaluating a patient with DE.

Acknowledgments
This study was supported by the Department of Veterans Affairs, Veterans Health Administration, Office of Research and Development, Clinical Sciences Research EPID-006-15S (Dr Galor), R01EY026174 (Dr Galor), NIH Center Core Grant P30EY014801, and Research to Prevent Blindness Unrestricted Grant.

Disclosure
The authors report no conflicts of interest in this work.

References


Lifitegrast for treatment of dry eye syndrome