In vitro binding and receptor-mediated activity of terlipressin at vasopressin receptors $V_1$ and $V_2$

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Abstract: Terlipressin, a synthetic, systemic vasoconstrictor with selective activity at vasopressin-1 ($V_1$) receptors, is a pro-drug for the endogenous/natural porcine hormone [Lys8]-vasopressin (LVP). We investigated binding and receptor-mediated cellular activities of terlipressin, LVP, and endogenous human hormone [Arg8]-vasopressin (AVP) at $V_1$ and vasopressin-2 ($V_2$) receptors. Cell membrane homogenates of Chinese hamster ovary cells expressing human $V_1$ and $V_2$ receptors were used in competitive binding assays to measure receptor-binding activity. These cells were used in functional assays to measure receptor-mediated cellular activity of terlipressin, LVP, and AVP. Binding was measured by [3H]AVP counts, and the activity was measured by fluorometric detection of intracellular calcium mobilization ($V_1$) and cyclic adenosine monophosphate ($V_2$). Binding potency at $V_1$ and $V_2$ was AVP > LVP > terlipressin. LVP and terlipressin had approximately sixfold higher affinity for $V_1$ than for $V_2$. Cellular activity potency was also AVP > LVP > terlipressin. Terlipressin was a partial agonist at $V_1$ and a full agonist at $V_2$; LVP was a full agonist at both $V_1$ and $V_2$. The in vivo response to terlipressin is likely due to the partial $V_1$ agonist activity of terlipressin and full $V_2$ agonist activity of its metabolite, LVP. These results provide supportive evidence for previous findings and further establish terlipressin pharmacology for vasopressin receptors.

Keywords: hormones, in vitro techniques, pharmacology, hepatorenal syndrome

Introduction

Terlipressin ($N^8$-tryglycyl-8-lysine-vasopressin), a pro-drug for the endogenous/natural porcine hormone [Lys8]-vasopressin (LVP),1,2 is a synthetic vasopressin analog derived from the natural/endogenous human hormone [Arg8]-vasopressin (AVP). Terlipressin differs from AVP in the substitution of lysine for arginine at the eighth position of endogenous AVP and the addition of three glycy1 residues at the amino terminus (Figure 1). Large randomized clinical studies in patients with hepatorenal syndrome type 1 have shown that intravenous administration of terlipressin improves renal function in this setting.3,4 Terlipressin exerts its effects via its partial vasopressin-1 ($V_1$) agonist activity and the full $V_2$ agonist activity of its metabolite, LVP.5 Findings from experimental studies conducted in rat and human tissues provide convincing evidence of the rapid, acute vasoconstrictor function of terlipressin following intravenous bolus administration of clinically relevant doses, as well as the longer term vasoconstrictive effects achieved through terlipressin conversion to LVP.6

$V_1$ receptors (previously known as $V_{1a}$ receptors) are located on vascular smooth muscle in the kidney, myometrium, bladder, liver, spleen, platelets, and fat tissue; $V_2$ receptors mediate vasoconstriction, hepatic glycogenolysis, and platelet aggregation.7,8
Vasopressin-2 (V₂) receptors are located on the basolateral membrane of the distal tubule and collecting ducts in the kidney; V₂ receptors mediate the antidiuretic effects of vasopressin. Cleavage of the N-triglycyl residue on terlipressin occurs in vivo, resulting in a prolonged duration of action for terlipressin compared with LVP. The purpose of this study was to investigate the binding activities and receptor-mediated cellular activities of terlipressin, LVP, and AVP at vasopressin receptor subtypes V₁ and V₂ in vitro. Notably, vasopressin-3 receptors (previously known as V₁b receptors) were not investigated in this study because they are not relevant to the hepatorenal syndrome type 1 disease state.

Materials and methods

Cell lines

Chinese hamster ovary (CHO) cells expressing human V₁ receptors (line CHO-S; study numbers 20404, 21104, and 10009187; Cerep, Celle l’Evescault, France) and human V₂ receptors (line CHO-K₁; study numbers 20404, 21104, and 10009187; Cerep) were used for the competitive binding and functional assays (Table 1). The expression of vasopressin receptors was approximately fivefold higher in the V₂ cell line; thus, different amounts of cell membrane proteins or cells were used in the assays (Table 2).

Competitive binding assays

Cell membrane homogenates were prepared from cells of the CHO-S and CHO-K₁ cell lines (study numbers 20404, 21104, and 10009187; Cerep). [3H]AVP at a concentration of 0.3 nM was added to each cell membrane homogenate (V₁ assay: 40-µg protein; V₂ assay: 16-µg protein), which was then incubated (V₁ assay: 60 minutes; V₂ assay: 120 minutes) at 22°C with various concentrations of terlipressin, LVP, and AVP in assay buffer containing Tris–HCl 50 mM (pH 7.4), MgCl₂ 5 mM, and 0.1% bovine serum albumin (Table 2). The following terlipressin concentrations were tested (molar): 1.0 × 10⁻³, 3.0 × 10⁻⁴, 1.0 × 10⁻⁴, 3.0 × 10⁻⁵, 1.0 × 10⁻⁵, 3.0 × 10⁻⁶, 1.0 × 10⁻⁶, 3.0 × 10⁻⁷, 1.0 × 10⁻⁷, and 1.0 × 10⁻⁸ M. The following LVP and AVP concentrations were tested (molar): 1.0 × 10⁻⁶, 1.0 × 10⁻⁷, 3.0 × 10⁻⁸, 1.0 × 10⁻⁸, 3.0 × 10⁻⁹, 1.0 × 10⁻⁹, 3.0 × 10⁻¹⁰, 1.0 × 10⁻¹⁰, 3.0 × 10⁻¹¹, 1.0 × 10⁻¹¹, and 1.0 × 10⁻¹² M. Nonspecific binding was determined in the presence of AVP 1 µM. All assays were performed in triplicate.

After incubation, samples were filtered rapidly under vacuum through glass fiber filters (GF/B; Packard Biosciences Company, Meriden, CT, USA [now PerkinElmer, Inc., Waltham, MA, USA]) presoaked with 0.3% polyethylenimine and rinsed several times with ice-cold Tris–HCl 50 mM buffer in a 96-sample cell harvester (UniFilter; PerkinElmer, Inc.). The filters were air dried, and the radioactivity retained in the filters was counted in a scintillation counter (TopCount; PerkinElmer, Inc.) in a scintillation cocktail (MicroScint 0; PerkinElmer, Inc.). The standard reference agonist for V₁ was [d(CH₂)₅₁,Tyr(Me)₂]-AVP, and the standard reference agonist for V₂ was AVP. These reference compounds were tested at

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**Table 1** Cells and reference compounds

<table>
<thead>
<tr>
<th>Assays</th>
<th>Cells</th>
<th>Receptor density (Bₘₐₓ, fmol/mg protein)</th>
<th>Reference compound</th>
</tr>
</thead>
<tbody>
<tr>
<td>Binding</td>
<td>Human V₁ (agonist radioligand)⁺</td>
<td>Human recombinant (CHO-S cells)</td>
<td>880</td>
</tr>
<tr>
<td></td>
<td>Human V₂ (agonist radioligand)⁺</td>
<td>Human recombinant (CHO-K₁ cells)</td>
<td>4000</td>
</tr>
<tr>
<td>Functional</td>
<td>Human V₁ (agonist effect)⁹</td>
<td>Human recombinant (CHO-S cells)</td>
<td>880</td>
</tr>
<tr>
<td></td>
<td>Human V₂ (agonist effect)¹⁰</td>
<td>Human recombinant (CHO-K₁ cells)</td>
<td>4000</td>
</tr>
</tbody>
</table>

**Abbreviations:** AVP, [Arg₈]-vasopressin; LVP, [Lys₈]-vasopressin; V₁, vasopressin-1; V₂, vasopressin-2.

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**Figure 1** Molecular formulas for terlipressin, lysine-vasopressin, and arginine-vasopressin.

Abbreviations: AVP, [Arg₈]-vasopressin; LVP, [Lys₈]-vasopressin.
several concentrations to obtain competition curves from which the concentration achieving half-maximal inhibition concentration (IC_{50}) was calculated.

**Receptor-mediated cellular activity assays**

Cell cultures were established from the same CHO cell lines used in the binding assays described earlier (Table 1). \(^9,10\) Cells expressing human V1 receptors were suspended in Dulbecco’s Modified Eagle Medium (Thermo Fisher Scientific, Waltham, MA, USA), complemented with 0.1% fetal calf serum, and then distributed in microplates at a density of 45,000 cells per well (Table 2). The fluorescent probe (Fluo4 NW; Thermo Fisher Scientific) was mixed with probenecid in Hank’s balanced salt solution buffer (Thermo Fisher Scientific) and added into each well after equilibration with the cells for 60 minutes at 37°C, and then for 15 minutes at 22°C. After 5 minutes of equilibration, the plates were positioned in a microplate reader (CellLux; PerkinElmer, Inc.) for the addition of various concentrations of terlipressin, LVP, and AVP and for subsequent measurements of activity. The following terlipressin concentrations were tested (molar): 1.0 \(\times\) 10^{-3}, 1.0 \(\times\) 10^{-4}, 1.0 \(\times\) 10^{-5}, 3.0 \(\times\) 10^{-6}, 1.0 \(\times\) 10^{-7}, 1.0 \(\times\) 10^{-8}, 3.0 \(\times\) 10^{-9}, 1.0 \(\times\) 10^{-9}, 1.0 \(\times\) 10^{-10}, and 1.0 \(\times\) 10^{-11} M. The following LVP and AVP concentrations were tested (molar): 1.0 \(\times\) 10^{-3}, 1.0 \(\times\) 10^{-4}, 1.0 \(\times\) 10^{-5}, 1.0 \(\times\) 10^{-6}, 1.0 \(\times\) 10^{-7}, 1.0 \(\times\) 10^{-8}, 3.0 \(\times\) 10^{-9}, 1.0 \(\times\) 10^{-9}, 1.0 \(\times\) 10^{-10}, and 1.0 \(\times\) 10^{-11} M. The LVP and AVP concentrations were tested (molar): 1.0 \(\times\) 10^{-3}, 1.0 \(\times\) 10^{-4}, 1.0 \(\times\) 10^{-5}, 1.0 \(\times\) 10^{-6}, 1.0 \(\times\) 10^{-7}, 1.0 \(\times\) 10^{-8}, 3.0 \(\times\) 10^{-9}, 1.0 \(\times\) 10^{-9}, 1.0 \(\times\) 10^{-10}, 3.0 \(\times\) 10^{-10}, 1.0 \(\times\) 10^{-11}, 3.0 \(\times\) 10^{-12}, 1.0 \(\times\) 10^{-12}, 1.0 \(\times\) 10^{-13}, and 1.0 \(\times\) 10^{-14} M. All assays were performed in triplicate.

The reference compounds for V_1 and V_2 were tested at several concentrations to obtain concentration–response curves from which the concentration producing half-maximal effective concentration (EC_{50}) or IC_{50} was calculated.

**Data analysis**

For the binding assays, specific ligand binding to receptors was defined as the difference between the total binding and

### Table 2 Experimental conditions

<table>
<thead>
<tr>
<th>Binding assays</th>
<th>Cell membrane homogenates</th>
<th>Ligand</th>
<th>Concentration</th>
<th>Nonspecific</th>
<th>Incubation</th>
<th>Method of detection</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human V_1 (agonist receptor)</td>
<td>40 (\mu)g protein</td>
<td>[^{[3]H}AVP]</td>
<td>0.3 nM</td>
<td>AVP (1 (\mu)M)</td>
<td>60 minutes at room temperature</td>
<td>Scintillation counting</td>
</tr>
<tr>
<td>Human V_2 (agonist receptor)</td>
<td>16 (\mu)g protein</td>
<td>[^{[3]H}AVP]</td>
<td>0.3 nM</td>
<td>AVP (1 (\mu)M)</td>
<td>120 minutes at room temperature</td>
<td>Scintillation counting</td>
</tr>
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</table>

<table>
<thead>
<tr>
<th>Functional assays</th>
<th>Cell density/well</th>
<th>Stimulus</th>
<th>Measured component</th>
<th>Incubation</th>
<th>Method of detection</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human V_1 (agonist effect)</td>
<td>45,000</td>
<td>None (1 (\mu)M AVP for control)</td>
<td>Intracellular (calcium)</td>
<td>60 minutes at room temperature</td>
<td>Fluorometry</td>
</tr>
<tr>
<td>Human V_2 (agonist effect)</td>
<td>3000</td>
<td>None (1 (\mu)M AVP for control)</td>
<td>cAMP</td>
<td>30 minutes at room temperature</td>
<td>HTRF</td>
</tr>
</tbody>
</table>

**Abbreviations:** AVP, [Arg^8]-vasopressin; cAMP, cyclic adenosine monophosphate; HTRF, homogeneous time-resolved fluorescence; V_1, vasopressin-1; V_2, vasopressin-2.
the nonspecific binding determined in the presence of an excess of unlabeled ligand. Results were expressed as a percentage of control-specific binding (measured specific binding/control-specific binding × 100) and as percent inhibition of control radioligand-specific binding (100 – [measured specific binding/control-specific binding] × 100) obtained in the presence of terlipressin, LVP, or AVP. Selectivity of terlipressin and LVP for V₁ and V₂ receptors was calculated as compared with AVP.

Values for the IC₅₀ and the Hill coefficient (nH) were determined by nonlinear regression analysis of the competition curves generated with mean replicate values using Hill equation curve fitting (Y = D + [A – D]/[1 + (C/C₅₀)ⁿH]), where Y = specific binding, D = minimum specific binding, A = maximum specific binding, C = test article concentration, C₅₀ = IC₅₀, and nH = slope factor). Inhibition constants (Ki) were calculated from the Cheng Prusoff equation (Ki = IC₅₀/[L/KD]), where L = concentration of radioligand in the assay and KD = affinity of the radioligand for the receptor).

For the functional assays, results were expressed as a percentage of control agonist response (measured response/control response × 100) in the presence of the test article. The EC₅₀ values were determined by nonlinear regression analysis of the functional assay curves generated with mean replicate values using the Hill equation curve fitting (Y = D + [A – D]/[1 + (C/C₅₀)ⁿH]), where Y = response, D = right asymptote of the curve, A = left asymptote of the curve, C = test article concentration, C₅₀ = EC₅₀, and nH = slope factor). The results for V₁ were expressed as a percentage of the control response to AVP 1 µM, and the results for V₂ were expressed as percent inhibition of the control response to AVP 0.03 nM.

Results

Binding affinity

Results of the binding affinity assays are summarized in Table 3 and illustrated in Figure 2. The potency of binding affinity of the tested compounds to both V₁ and V₂ receptors was AVP > LVP > terlipressin. For V₁ receptors, the binding affinity of LVP was about twofold less than that of AVP, and the binding affinity of terlipressin was about 600-fold less than that of LVP. For V₂ receptors, the binding affinity of LVP

Table 3 Binding affinity assay results

<table>
<thead>
<tr>
<th>Compound tested</th>
<th>IC₅₀ (moles)</th>
<th>IC₅₀ Selectivity for V₁</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Human V₁</td>
<td>Human V₂</td>
</tr>
<tr>
<td>Terlipressin</td>
<td>1.7 × 10⁻⁴</td>
<td>9.6 × 10⁻⁴</td>
</tr>
<tr>
<td>LVP</td>
<td>2.9 × 10⁻⁶</td>
<td>1.8 × 10⁻⁸</td>
</tr>
<tr>
<td>AVP</td>
<td>1.3 × 10⁻⁹</td>
<td>8.0 × 10⁻¹⁰</td>
</tr>
</tbody>
</table>

Abbreviations: AVP, [Arg₈]-vasopressin; IC₅₀, half-maximal inhibition concentration; Kᵱ, inhibition constant; LVP, [Lys₈]-vasopressin; V₁, vasopressin-1; V₂, vasopressin-2.

Figure 2 Binding affinity as a function of concentration for the tested compounds.

Abbreviations: AVP, [Arg₈]-vasopressin; LVP, [Lys₈]-vasopressin; V₁, vasopressin-1; V₂, vasopressin-2.
was about 12-fold less than that of AVP, and the binding affinity of terlipressin was about 700-fold less than that of LVP.

AVP bound V₁ and V₂ receptors with similar high affinities (8.0 × 10⁻¹⁰ and 8.5 × 10⁻¹⁰, respectively). LVP also bound V₁ and V₂ receptors with high affinity, but affinity for V₁ was about sixfold higher than that for V₂ (1.8 × 10⁻⁹ and 1.0 × 10⁻⁸, respectively). Terlipressin bound V₁ and V₂ receptors with low affinity, but affinity for V₁ was about sixfold higher than that for V₂ (1.1 × 10⁻⁶ and 6.9 × 10⁻⁶, respectively).

### Cellular activity

Results of the vasopressin-mediated functional activity assays are summarized in Table 4 and illustrated in Figure 3. The order of potency for cell activation was the same as for the binding assay: AVP>LVP>>terlipressin. LVP was a full agonist at both V₁ and V₂ receptors, producing maximal cell activation about equal to that of AVP. Terlipressin produced maximal cell activation about equal to that of AVP at V₂ receptors, and thus was a full agonist at V₂ receptors, but it produced only 41% of the maximal effect observed with AVP at V₁ receptors, indicating that it is a partial agonist at V₁ receptors.

### Discussion

Terlipressin is a pro-drug for the endogenous/natural porcine hormone LVP. In the current study, the results of radioligand competitive binding assays in cell membrane homogenates of CHO cells expressing human V₁ and V₂ receptors demonstrated that terlipressin binds to V₁ and V₂ receptors with much lower affinities than LVP or AVP. Terlipressin was selective for V₁ receptors at a V₁:V₂ ratio of 6:1. Results of the functional assays indicated that terlipressin behaves as a partial agonist at V₁ receptors and a full agonist at V₂ receptors.

Our results support findings from previously reported studies and provide additional evidence regarding the pharmacologic properties of terlipressin. Colson et al. using vascular smooth muscle cells isolated from rat aorta and human uterine artery, found that the relative binding affinity (Kᵢ [nM]) of LVP, AVP, and terlipressin for human and rat V₁ receptors was AVP>LVP>>terlipressin. The mean binding

<table>
<thead>
<tr>
<th>Compound tested</th>
<th>EC₅₀ (moles)</th>
<th>Human V₁</th>
<th>Human V₂</th>
<th>Intrinsic activity (maximal activity of test compound/maximal activity of AVP)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Terlipressin</td>
<td>5.0 × 10⁻⁸</td>
<td>2.8 × 10⁻⁴</td>
<td>41</td>
<td>≈100</td>
</tr>
<tr>
<td>LVP</td>
<td>2.9 × 10⁻¹⁰</td>
<td>2.4 × 10⁻¹¹</td>
<td>≈100</td>
<td>≈100</td>
</tr>
<tr>
<td>AVP</td>
<td>1.8 × 10⁻¹⁰</td>
<td>4.4 × 10⁻¹²</td>
<td>100</td>
<td>100</td>
</tr>
</tbody>
</table>

Abbreviations: AVP, [Arg⁸]-vasopressin; EC₅₀, half-maximal effective concentration; Kᵢ, inhibition constant; LVP, [Lys⁸]-vasopressin; V₁, vasopressin-1; V₂, vasopressin-2.
affinity of terlipressin for human V₁ receptors (1229 nM) was about 120-fold less than the binding affinity of LVP (10 nM); about a 700-fold difference in terlipressin (852 nM) versus LVP binding (1.2 nM) was observed at rat V₁ receptors.¹¹

Mean maximal cell activation with terlipressin, as measured by intracellular calcium mobilization, was 48% at human V₁ receptors and 66% at rat V₁ receptors versus 100% with LVP.¹¹

The selectivity of terlipressin for V₁ receptors was 1.9-fold higher than that for V₂ receptors compared with the 6.3-fold higher selectivity observed in our study. This difference may be a reflection of the different cell lines utilized for competitive binding assays (rat and human vascular smooth muscle cells versus CHO cells, respectively) in the two studies. Bernadich et al¹² found that the mean potency of terlipressin (852 nM) versus CHO cells, respectively) in the two studies.

Results from the current study provide additional, supportive evidence for previous findings and further establish the binding affinity, potency, and selectivity of terlipressin at vasopressin receptors.

**Conclusion**

These data suggest that the in vivo pharmacologic response to the administration of terlipressin primarily reflects LVP-mediated activation of V₁ receptors. However, terlipressin demonstrated agonist activity at both V₁ and V₂ receptors. Results from the current study provide additional, supportive evidence for previous findings and further establish the binding affinity, potency, and selectivity of terlipressin at vasopressin receptors.

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**Disclosure**

Khurram Jamil and Krishna R Devarakonda are employees of Mallinckrodt Pharmaceuticals. Stephen Chris Pappas is an employee of Orphan Therapeutics, LLC. The authors report no other conflicts of interest in this work.

**References**
