Protein phosphatase 2Cδ/Wip1 regulates phospho-p90RSK2 activity in lesional psoriatic skin

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Objectives: P90 ribosomal S6 kinase (RSK) 1 and 2 are serine/threonine protein kinases believed to mediate proliferation and apoptosis via the extracellular signal-regulated kinases (ERK1/2) signaling pathway. Macrophage migration inhibitory factor (MIF) and epidermal growth factor (EGF) are activators of this pathway and are elevated in the serum of patients with psoriasis compared with healthy controls. Studies on COS-7 cell cultures have shown that protein phosphatase 2Cδ (PP2Cδ) decreases the activity of RSK2 following EGF stimulation. We therefore hypothesize that PP2Cδ regulates RSK2 activity in psoriasis.

Methods: In paired biopsies from nonlesional (NL) and lesional (L) skins, we analyzed the level of RSK1, 2 phosphorylation and the expression of PP2Cδ isoforms, integrin-linked kinase-associated serine/threonine phosphatase (ILKAP) and wild-type p53-induced phosphatase 1 (Wip1) by Western blotting, immunofluorescence and coimmunoprecipitation with monoclonal antibody for RSK2. The induction of Wip1 by MIF or EGF was studied in cultured normal human keratinocytes.

Results: The protein level of RSK1, 2 phosphorylated at T573/T577 was significantly increased in L compared with NL psoriatic skin, while phosphorylation at S380/S386 was reduced in L compared with NL psoriatic skin when assayed by Western blotting and immunofluorescence microscopy. ILKAP expression was significantly higher in L than in NL skin, whereas Wip1 was expressed in similar amounts but showed increased coimmunoprecipitation with RSK2 in L compared with NL psoriatic skin. In cultured normal human keratinocytes stimulated with MIF, Wip1 phosphorylation and Wip1 expression were increased after 24 hours, but not when costimulated with dimethyl fumarate (DMF). The increased coimmunoprecipitation of Wip1 with RSK2 was significantly induced by EGF or MIF activation at 24 hours and could be significantly inhibited by DMF or the ERK1/2 inhibitor PD98059.

Conclusion: The complex formation of Wip1 with RSK2 indicates a direct interaction reducing P-RSK2 (S386) activation in L skin and indicates that Wip1 has a role in the pathogenesis of psoriasis.

Keywords: P90 RSK1, 2, EGF, MIF, PP2Cδ/ILKAP, PP2Cδ/Wip1

Introduction
It has been suggested that extracellular signal-regulated kinase (ERK1/2) and mitogen- and stress-activated kinase (MSK1/2) are involved in the pathogenesis of psoriasis. These kinases are activated by macrophage migration inhibitory factor (MIF) and epidermal growth factor (EGF); both are overexpressed in serum from psoriasis patients.
The p90 ribosomal S6 kinases, RSK1–3, are a family of serine/threonine kinases activated by phosphorylation through ERK1/2 signaling. The phosphorylation sites in RSK1–3 are highly conserved amino acid sequences.\textsuperscript{6–8} Phospho-RSK1 was induced by MIF and EGF in cultured human keratinocytes,\textsuperscript{9} but the expression of phospho-RSK1, 2 in psoriatic skin has not been tested.

RSK1, 2, and 3 isoforms are present in most tissues; they activate genes involved in proliferation and inactivate proapoptotic proteins like the BCL2-associated agonist of cell death.\textsuperscript{10,11} In resting cells, ERK1/2 is bound to the C-terminal site of RSK kinases, and upon stimulation, RSK1, 2 become phosphorylated at T573/T577. This is followed by autophosphorylation of homologous sites at S380/S386 in the middle region of RSK1, 2. The middle region cooperates with phosphoinositide-dependent protein kinase to dock and subsequently phosphorylate the N-terminal kinase domain of RSK1, 2 at S221/S227, respectively, which is needed for full RSK activation.\textsuperscript{12–15}

Any inhibition of these steps will prohibit the effect of RSK on substrates and block the feedback reaction from the N-terminal kinase domain, controlling binding to P-ERK1/2.

Dimethyl fumarate (DMF) is a well-known oral drug for the treatment of psoriasis.\textsuperscript{16} DMF inhibits the induced phosphorylation of MSK1/2 and nuclear factor-κB (NF-κB)/p65 in cultured human keratinocytes, which is needed for cytokine transcription and is believed to be important in the pathogenesis of psoriasis.\textsuperscript{5,17,18} The structurally related MSK1/2 and RSK1, 2 are both activated by the ERK1/2 signaling pathway. In keratinocytes, preincubation with DMF inhibited the MIF- or EGF-induced activation of P-MSK1 and P-MSK1 at multiple sites, similar to the ERK1/2 inhibitor PD98059.\textsuperscript{9} These specific effects led to the inhibition of keratinocyte proliferation mediated by the induction of p-p53 (S15).

Members of the protein phosphatase family (PP2C) play a role in reversing protein kinase activation. PP2C is a highly conserved family of serine/threonine phosphatases.\textsuperscript{19–22} PP2Cα, PP2Cβ, and PP2Cδ/wild-type p53-induced phosphatase 1 (Wip1) isoforms inhibit the activation of Jun N-terminal kinase (JNK) and p38 mitogen-activated protein kinase (MAPK) kinases and apoptosis. In contrast, the human PP2Cδ/integrin-linked kinase-associated serine/threonine phosphatase (ILKAP; 46 kDa) isoform inhibits proliferation and oncogenic transformation by suppressing integrin-linked kinase 1 (ILK1) activity, causing the activation of p38 MAPK and JNK/c-Jun kinases and apoptosis.\textsuperscript{21–23}

The mammalian PP2Cδ/Wip1 isoform is induced by ionizing radiation in a p53-dependent manner.\textsuperscript{5,22} Wip1 (67 kDa) is considered an oncogene as it inactivates the tumor suppressor protein p53.\textsuperscript{26}

In COS-7 cells transfected with empty vector or human influenza hemagglutinin-tagged RSK2 or myc-PP2Cδ/ILKAP plasmids, RSK2 activity was regulated by complex formation with PP2Cδ. The overexpression of PP2Cδ/ILKAP decreased RSK2 kinase activity by 50% by dephosphorylating p-RSK2 at S386. This complex formation could, in turn, be induced by phorbol 12-myristate 13-acetate through ERK1/2-mediated phosphorylation of PP2Cδ/ILKAP.\textsuperscript{20}

Thus, the interaction of RSK2 and PP2Cδ/ILKAP appears to play an important role in the modulation of the ERK1/2 signaling cascade, at least in vitro, and could potentially play a role in psoriasis. To investigate this further, we therefore determined the protein expression of total and phosphorylated RSK1 and RSK2 and the expression of the PP2Cδ isoforms ILKAP and Wip1 in lesional (L) and nonlesional (NL) psoriatic skin. In addition, we used coimmunoprecipitation to investigate the complex formation between RSK2 and Wip1 and ILKAP.

The Regional Ethical Committee of Region Midtjylland, Denmark, approved the experiments with psoriatic patients (M20090102). A signed consent was obtained from each patient who donated skin samples, according to the Declaration of Helsinki principles.

Materials and methods

Cell cultures

Normal adult human keratinocytes were obtained by trypsinization of skin samples, as described before.\textsuperscript{27} Second-passage keratinocytes were trypsinized and seeded in 10 cm petri dishes, $4.5 \times 10^{6}$ cells/plate in keratinocyte basal medium (Gibco 17005; Thermo Fisher Scientific, Waltham, MA, USA), with only bovine pituitary growth hormone added from the supplement\textsuperscript{9} Gibco 37000-015, 50 μg/mL, gentamicin (Gibco 15710, 5 μg/mL), and 2% fetal calf serum (FCS; Gibco 160000 44). After 24 hours, the cells were either left alone or preincubated with DMF (140 μM) or 50 μM of PD98059 for 1 hour and stimulated with recombinant MIF (100 ng/mL; R&D Systems, Oxon, UK) or h-EGF (2 ng/mL; PeproTech, London, UK) for 24 hours. Alternatively, the cells were stimulated with MIF for 0 and 30 minutes and 2, 12, 24, 48, and 96 hours alone or after preincubation with...
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140 µM DMF (47967-25-G-F; Sigma-Aldrich, Brøndby, Denmark).

DMF was dissolved as described before. The ERK1/2 inhibitor PD98059 and MSK1/RSK1 inhibitor R0-318220 from Calbiochem (La Jolla, CA, USA) were dissolved in dimethyl sulfoxide (DMSO; 1% vol/vol). No-drug controls contained an equal volume of DMSO.

Human breast cancer cell line MCF-7 from the European Collection of Cell Cultures (Wiltshire, UK) was cultured in Dulbecco’s Modified Eagle Medium (DMEM; Gibco 61965-026) with 10% FCS, gentamicin (Gibco 15710), penicillin, streptomycin (Gibco 15140-148), and 2.5% Hepes until 60% confluence. Cells were trypsinized and seeded in 10 cm petri dishes, 4.0×10⁶ cells/plate in DMEM, supplemented with pituitary growth hormone (50 µg/mL), 2% FCS, and 2.5% Hepes.

Tissue material from donors

Keratome and punch skin biopsies were obtained from volunteers with moderate-to-severe plaque psoriasis. The biopsies were collected during two different time periods. In the first group, 5 patients’ paired biopsies were analyzed by Western blotting (Figures 1A and B and 2A and B) and immunohistochemistry (results similar to those seen in Figure 3A and B). Biopsies from the second group of 6 patients were analyzed by immunohistochemistry (Figure 3A and B), Western blotting (Figure 4A and C) and immunoprecipitation (Figure 5A–C).

The third group of 7 normal healthy donors who had donated keratome biopsies were analyzed by Western blotting and immunoprecipitation (Figure 4B and C). Each participant with psoriasis was untreated for at least 1 month prior to the time of biopsy.

Immunohistochemistry

Punch biopsies from NL and L plaque-type psoriatic skin were obtained as described before. Then, 4-µm sections were heated in 10 mM sodium citrate buffer for 10 minutes in a microwave oven (750 W). The sections were further permeabilized in Tris-buffered saline (TBS) with 1% Triton X-100 for 10 minutes and washed with TBS with 0.3% Triton X-100. Nonspecific binding sites were blocked with Image-iT FX signal enhancer (136933; Thermo Fisher Scientific, Waltham, MA, USA). Tissue sections were incubated with the primary antibody dissolved in TBS med 0.3% Triton X-100 (T-9284; Sigma-Aldrich) with 5% of goat nonimmune serum (DAKO X0907) at 4°C overnight. Antibodies were rabbit anti-P-RSK1 (T573) or P-RSK1 (S380; Cell Signalling Technology, Beverly, MA, USA), isotype control (sc-2027) normal rabbit IgG, and goat anti-rabbit Alexa Flour 594 (A-11037) or Alexa Flour 488 (A 31628; Invitrogen, Molecular Probes®). The samples were mounted in ProLong.
anti-fade reagent with 4',6-diamidine-2'-phenylindole (DAPI; P36935; Invitrogen, Roskilde, Denmark). Samples were viewed by using an epifluorescence microscope (Leica Microsystems, Wetzlar, Germany).

In other experiments, tissue sections were incubated with rabbit anti-ILKAP (ab70019; Abcam, Cambridge, UK) at 4°C overnight, developed with goat anti-rabbit Alexa Flour 488 (A-31628; Invitrogen) for 1 hour, and mounted with DAPI.

Immunoprecipitation
Immunoprecipitation was performed using protein isolated from keratome biopsies or cultured human keratinocytes, extracted in 1× lysis buffer (#9803; Cell Signalling Technology) with added 22 µL/mL protease inhibitor cocktail (ethylenediaminetetraacetic acid-free complete; Hoffman-La Roche Ltd., Basel, Switzerland), 10 µL/mL of 100 nM phenylmethylsulfonyl fluoride (PMSF)/mL, and nuclear lysis benzonase (1 µL/100 µL). An amount of protein (500 µg) in 500 µL of lysis buffer was added to 2 µg of mouse monoclonal RSK2 antibody (Santa Cruz, CA, USA) and incubated overnight at 4°C on a rotating table. Then, 40 µL of the suspended volume of Protein A/G Plus-Agarose (Santa Cruz, CA, USA) was added at 4°C for 4 hours. The pellets were collected by centrifugation at 2500 rpm for 5 minutes, washed 4 times with sterile PBS, and resuspended in 40 µL of sample buffer containing 10 mM dithiothreitol and 2.5% sodium dodecyl sulfate (SDS). Samples were boiled for 2–3 minutes and centrifuged to pellet the agarose beads. Then, they (20 µL) were separated on gels.

Western blotting
Cell extracts from keratome biopsies or cell cultures were prepared in 1× lysis buffer supplemented with 22 µL/mL of protease inhibitor and 10 µL/mL of 100 mM PMSF/mL buffer. Equal loads of protein (50 µg) were added to 3× SDS lysis buffer and separated on SDS-polyacrylamide gel electrophoresis (PAGE) 8%–16% Tris–Glycine Gels (Invitrogen) or on NuPAGE 4%–12% Bis-Tris Gel, (Novex; Life Technology; Thermo Fisher Scientific). Proteins were blotted and incubated as described before. Equal load was controlled by anti-β-actin (A-1978; Sigma-Aldrich). Densitometric analysis of the band intensity was performed on a scanner (Epson PERFECTION V75 PRO) and quantitated by Jelly Quant (Mads Rasmussen 2004, Aarhus, Denmark).
The following antibodies were used: anti-P-RSK1 (T573) and (S380), anti-RSK1, 2, 3, and HRP anti-rabbit Ab #7074 (Cell Signalling Technology); rabbit anti-RSK1 and monoclonal anti-RSK2 (E-1), HRP anti-mouse (sc-2060), and HRP anti-rabbit (sc-2054); immunoprecipitation: monoclonal anti-RSK2 (E-1); sc-9986 and rabbit anti-Wip1 (H-300; sc-20712) epitope C-terminus AA 306-605 (Santa Cruz Biotech, Santa Cruz, CA, USA); rabbit anti-PP2Cδ/ILKAP (ab70001; Abcam); HRP-conjugated goat anti-rabbit #7074, HRP-conjugated horse anti-mouse #7076 (Cell Signalling Technology); and rabbit anti-PP2Cδ/Wip1:PPM1D Center AA 182-212 (Cat. No. AP13875c) that does not recognize P-Wip1 (Abgent Europe, London, UK).

**Statistical methods and calculation of the experimental data**

Antibody binding analyzed by Western blotting and densitometry analysis of immune reactive band intensity were carried out as described previously. Graphs were made with Sigma Plot Version 11. STATA was used to test for normal distribution and for performing Student’s t-tests. A probability of $p<0.05$ was regarded as statistically significant.

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**Figure 3 (A)** Nuclear staining of keratinocytes in biopsies from NL and L skin by P-RSK1 (T573) antibodies, NL (a, b) and plaque-type L (e, f) psoriatic skin, isotype control Ab (i, j) and Alexa Flour 594 (red) (bars 50 µm; resp. 20 µm). (B) Cytosolic staining of keratinocytes in NL (a, b) and L (e, f) skin by P-RSK1 (S380) antibodies, and Alexa Flour 488 (green), nuclear staining DAPI (blue) (c, d, g, h, i, k) (bars 100 µm a, c, e, g, i, l; resp. 20 µm b, d, f, h, j, k).

**Abbreviations:** DAPI, 4',6-diamidine-2'-phenylindole; L, lesional; NL, nonlesional; non-sp, nonspecific; RSK, p90 ribosomal S6 kinase; sp, specific.
Western blotting using anti-P-RSK1 (T573) or anti-P-RSK1 (S380) antibodies recognize a conserved area expressed in all of RSK1, 2, and 3; therefore, an anti-RSK1, 2, or 3 antibody was used.

The level of P-RSK1 (T573) was significantly higher ($p^*=0.006$) in paired samples of L skin than in NL psoriatic skin (Figure 1A). In contrast, the P-RSK1 (S380) immunoactive bands normalized to β-actin showed that P-RSK1 (S380); n=5) was significantly lower ($p^*=0.049$) in L skin than in NL skin (Figure 1B). In order to localize P-RSK1 (T573) by immunostaining, sections of paraffin-embedded skin from NL and L skin were incubated with P-RSK1 (T573) antibodies and anti-rabbit Alexa Flour 594 (red color) and DAPI (blue color). This showed nuclear staining of keratinocytes in the epidermis (Figure 3A). The number of stained nuclei (manual counting) in (20×20) µm area/patients was higher in L skin (L, 10.2±1.2) than in NL skin (NL, 7.8±1.2). The number of cytosolic-stained cells in the epidermis with anti-P-RSK1 (S380) and anti-rabbit Alexa Flour 488 (green color) in (100×150) µm area/patients was lower in L skin (L, 20±3) than in NL skin (NL, 5±1; Figure 3B). One of four patients is shown. Isotype control showed that immunostaining with anti-P-RSK1 (S380) Ab was nonspecific in the dermis and specific in the epidermis.

### Expression of ILKAP is higher in L than in NL psoriatic skin

To investigate the presence of ILKAP, paired biopsies from NL and L skin were analyzed by Western blotting using anti-ILKAP antibodies and anti-β-actin for normalization. Data from 5 patients were represented as mean values ± SD. These values show that the level of ILKAP (46 kDa) expression was significantly higher ($p^*=0.008$) in L skin than in NL skin (Figure 2A).

In order to localize ILKAP by immunostaining, paired biopsies from NL and L skin were incubated with anti-ILKAP, anti-rabbit Alexa Flour 488 (green), and DAPI. ILKAP staining in the nuclei in L skin was 53 dots/equal area compared with NL skin with 32 dots/equal area (Figure 2B).

### Wip1 was expressed at a similar level in NL and L psoriatic skin

Whole cell protein extracts (50 µg/lane) from paired biopsies were prepared as before and analyzed by Western blotting.
using anti-Wip1 antibodies. The blots were reincubated with a monoclonal anti-RSK2 or with polyclonal anti-RSK1 and anti-β-actin. This showed that the level of Wip1 (67 kDa) was not significantly different in NL compared with L skin when quantitated to β-actin (Figure 4A). Wip1 was expressed as a double band in psoriatic skin. Because RSK1, 2 but not RSK3 are involved in the regulation of proliferation, we had to confirm the amount of RSK1 and RSK2.10 The levels of RSK1 and RSK2 were not equal in NL and L skin, as seen before.

The level of Wip1 in healthy normal skin biopsies was tested. Because MCF-7 breast cancer cells have previously been shown to express constitutive high levels of Wip1,24,26 we used these cells as a positive control and tested for association with RSK2 (Figure 4B).

MCF-7 cells were preincubated with vehicle or DMF (140 µM) for 1 hour and stimulated with tumor necrosis factor-α (TNF-α; 10 ng/mL) for 24 hours. Whole cell protein extracts were immunoprecipitated with anti-RSK2 Ab and analyzed by Western blotting using anti-Wip1 and anti-RSK2 Ab. Wip1 was expressed as a single band in normal, healthy skin, at the molecule weight of Wip1 when compared with MCF-7 cells expressing P-Wip1/Wip1 (antibody recognizing both forms) in complex with RSK2. Preincubation with DMF strongly reduced the complex formation of P-Wip1/Wip1 with RSK2 (Figure 4B). We tested the Wip1 and RSK2 complex formation in psoriatic and normal skin biopsies. As before, proteins were immunoprecipitated with anti-RSK2 antibody and analyzed by Western blotting (Figure 4C). The analysis showed that there was a complex formation of Wip1 with RSK2 in psoriatic skin, but not in normal skin samples.

RSK2 forms more complexes with both Wip1 and ILKAP in L psoriatic skin than in NL skin

Complex formation between PP2Cδ/ILKAP and RSK2 was previously identified in immunoprecipitates from COS-7 cells.20 We therefore investigated the complex formation between RSK2 and Wip1 or ILKAP in psoriatic skin. Whole cell protein extracts (see “Materials and methods” section) from NL and L skin (n=4) were immunoprecipitated with monoclonal antibodies for RSK2 and analyzed with antibodies for Wip1, ILKAP, and RSK2 by Western blotting. The expression of Wip1 and ILKAP was normalized against the expression of RSK2. The ratio of Wip1 to RSK2 indicated a significant association of Wip1 in complex with RSK2.
in L skin (grey graphs) compared with NL skin (black graphs; \( p^* = 0.0168 \)). The ratio of ILKAP to RSK2 indicated that there was also significant complex formation between ILKAP and RSK2 in L skin (\( p^* = 0.033 \)) compared with NL skin (Figure 5A).

In order to confirm the association between Wip1 or ILKAP and RSK2, proteins from NL and L skin from two new patients were immunoprecipitated with anti-Wip1 Ab (H-300: sc-20712)28 and tested with anti-Wip1 Ab (AP12876c; which does not recognize P-Wip1) and anti-RSK2 Ab. This confirmed that the amount of Wip1 associated with RSK2 was higher in L skin than in NL skin (Figure 5B). Immunoprecipitation with anti-ILKAP Ab and testing with anti-ILKAP and anti-RSK2 Ab confirmed that the amount of ILKAP associated with RSK2 was higher in L than in NL skin (Figure 5C).

### Activation and new synthesis of Wip1 are regulated by MIF and DMF in normal human keratinocytes

The induction of P-Wip1/Wip1 was studied in a time-dependent manner. Keratinocytes were preincubated with either vehicle or DMF (140 µM) for 1 hour and then stimulated with MIF (100 ng/mL) for 30 minutes or 2 or 24 hours (Figure 6A). Alternatively, keratinocytes were stimulated with MIF (100 ng/mL) for 12, 24, 48, and 96 hours (Figure 6B). Whole cell extracts were analyzed with anti-Wip1 Ab (H-300; sc20712)28 and Western blotting. MIF stimulation induced P-Wip1 and Wip1 at 24 hours, which was inhibited by DMF (Figure 6A). MIF stimulation induced a second new synthesis of Wip1 at 48 hours, which was inhibited by DMF (Figure 6B). Because MIF stimulation has been shown to inhibit the expression of p-P53 (S15) from 24 to 96 hours, we tested the effect of DMF on the new synthesis of Wip1. Preincubation with DMF reduced the new synthesis of Wip1 (Figure 6B) and simultaneously activated the expression of p-P53 (S15), as seen before.9

### Complex formation of Wip1 and RSK2 is induced by EGF and MIF in normal human keratinocytes

In order to investigate the role of the ERK signaling pathway in the association of Wip1 with RSK2, keratinocytes were preincubated with DMF (140 µM) or PD98059 (50 µM) for 1 hour and then stimulated with EGF (2 ng/mL) or MIF (100 ng/mL) for 24 hours. Proteins (400 µg/sample) were immunoprecipitated with anti-RSK2 Ab and analyzed by Western blotting by anti-Wip1 and anti-RSK2 antibodies. Stimulation with EGF or MIF induced the immunoprecipitated complexes of P-Wip1/Wip1 with RSK2 at 24 hours. Figure 7A shows a representative blot. Densitometry of 6 different experiments showed that the

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**Figure 6** (A) MIF stimulation activated P-Wip1 at 24 hours and the new synthesis of Wip1 at 48 hours, which were inhibited by costimulation with DMF. Keratinocytes were preincubated with vehicle or DMF (140 µg/mL) and stimulated with MIF (100 ng/mL) at 30 minutes and 2 and 24 hours. (B) Keratinocytes were preincubated with vehicle or DMF (140 µM) and stimulated with MIF (100 ng/mL) for 12, 24, 48, and 96 hours. Whole cell extracts were analyzed by Western blotting with anti-Wip1 and reprobed with anti-p-P53 (S15), anti-p53, and anti-RSK2 antibodies.

**Abbreviations:** DMF, dimethyl fumarate; MIF, macrophage migration inhibitory factor; RSK, p90 ribosomal S6 kinase; Wip1, wild-type p53-induced phosphatase 1.
Figure 7 (A) Complex formation of Wip1 with RSK2 in human keratinocytes was induced by EGF and MIF. Keratinocytes were either left alone or preincubated with DMF (140 µM) or PD98059 (50 µM) for 1 hour and stimulated with EGF (2 ng/mL) or MIF (100 ng/mL) for 24 hours. Whole cell extracts were immunoprecipitated with anti-RSK2 antibody and analyzed by WB with anti-Wip1 (H-300) epitope C-terminus and anti-RSK2; 1 representative gel is shown. (B) Densitometry of 6 different experiments from pooled data shows the ratio of P-Wip1/Wip1 (both) associated with RSK2. EGF-stimulated versus (co) untreated cells *p=0.003; MIF stimulated versus (co) untreated cells ¤p=0.037; EGF-stimulated versus (DMF and EGF) **p=0.008; MIF-stimulated versus (DMF and MIF) p=0.059; EGF-stimulated versus (PD98059 and EGF) ***p=0.001; MIF-stimulated versus (PD98059 and MIF) *p=0.0045. (C) Phosphorylation status of RSK2 at 24 hours. Keratinocytes were preincubated with vehicle, DMF, PD98059, or Ro-318220 and stimulated as before and immunoprecipitated with anti-RSK2 Ab. The protein complexes were analyzed with anti-PP2Cδ/Wip1 (epitope Center AA 182-212; antibody does not recognize P-Wip1). The activation of P-RSK2 was tested by anti-RSK2 antibody.

**Abbreviations:** DMF, dimethyl fumarate; EGF, epidermal growth factor; MIF, macrophage migration inhibitory factor; PP2Cδ, protein phosphatase 2Cδ; RSK, p90 ribosomal S6 kinase; Wip1, wild-type p53-induced phosphatase 1.

PD98059 significantly inhibited the ratio of EGF- or MIF-induced association of total Wip1 with RSK2. Preincubation with DMF significantly reduced the EGF-induced association of total Wip1 with RSK2, while the ratio of P-Wip1/Wip1 (both) complexes with RSK2 was significantly increased by EGF as well as by MIF compared with controls (co; anti-Wip1 (H-300) sc-20712, epitope C-terminus). Preincubation with the ERK1/2 inhibitor PD98059 significantly inhibited the ratio of EGF- or MIF-induced association of total Wip1 with RSK2. Preincubation with DMF significantly reduced the EGF-induced association of total Wip1 with RSK2, while the
inhibitory effect of DMF on the MIF-induced association of total Wip1 with RSK2 was only borderline significant (Figure 7B). The slow-migrating band P-Wip1 (Figure 7A) was not inhibited by DMF, similar to Ro318220, which did not inhibit P-PP2Cδ/ILKAP.

To identify the phosphorylation status of RSK2 at 24 hours, keratinocytes were stimulated as before, immunoprecipitated with anti-RSK2 antibodies, and analyzed with anti-PP2Cδ/Wip1 (epitope PPMID Center AA 182-212 [Cat. No. AP13875c]) and anti-RSK2 Ab. This showed that EGF or MIF with EGF activated P-RSK2 expression at 24 hours in complex with PP2Cδ/Wip1, and this was inhibited by DMF, PD98059, and Ro-318220 (Figure 7C). Anti-PP2Cδ/Wip1 (epitope Center AA 182-212) Ab did not recognize the slow-migrating band of P-Wip1.

Discussion

In this study, we made 3 novel observations that provide information on how RSK2 activity is regulated by PP2Cδ/Wip1 in L skin of patients with psoriasis. First, the level of P-RSK1, 2 (T573)/(T577) was significantly higher in L than in NL psoriatic skin, while the opposite was true for P-RSK1, 2 S380/S386 (Figure 1A and B). Second, the level of ILKAP was significantly higher in L than in NL skin (Figure 2A and B). Third, although the level of Wip1 was similar in L and NL psoriatic skin (Figure 4A), we observed a pronounced complex formation between Wip1 and RSK2 in L skin compared with NL skin (Figure 5A and B). There was no complex formation in normal skin (Figure 4C). ILKAP also formed significant complexes with RSK2 in L skin compared with NL skin (Figure 5A and B). Based on these findings, we propose that Wip1 and ILKAP regulate the activity of RSK2 and the phosphorylation of RSK2 (S386) in L psoriatic skin. Because phosphorylation sites of RSK1-3 are highly conserved amino acid sequences, antibodies for P-RSK1, 2 S380/S386 (Figure 1A and B) detect also P-RSK2 (T577) or P-RSK2 (S386) (Figure 1A and B). Because phosphorylation is considered to stabilize protein expression, we normalized P-RSK1, 2 to β-actin and not to RSK1, 2, and 3.

Previous research shows that nonactive RSK1-3 is localized in cytoplasm, while active RSKs are translocated to the nucleus. Ultraviolet type B stimulation induced the nuclear expression of P-RSK2 (T577) in normal skin and skin cancer cells. In psoriatic skin, the number of keratinocytes expressing nuclear P-RSK1, 2 T573/T577 was higher in L than in NL epidermis (Figure 3A). The number of cells expressing cytoplasmic P-RSK1, 2 S380/S386 in the epidermis was reduced (Figure 3B), pointing to dephosphorylation by Wip1 in psoriatic epidermis.

Two different gene loci are described for the PP2Cδ/Wip1 and ILKAP isoforms. ILKAP (46 kDa) has been shown to regulate ILK1 activity physiologically. Nuclear localization of ILK in keratinocytes was associated with increased DNA synthesis. In accordance with this, we found more anti-ILKAP staining in the nuclei of keratinocytes in L than in NL epidermis (Figure 2B). Two anti-ILKAP positive dots/nucleus indicate cell division.

Phosphorylation at Ser and Thr residues in peptides was important for binding ILKAP or Wip1 to RSK2. Affinity was higher for pSer386 peptides than for pThr577, which indicates that ILKAP in partnership with Wip1 controls P-RSK2 activity.

Unlike ILKAP, Wip1 is an oncogene overexpressed in breast and ovarian carcinoma, and Wip1 enhances growth by dephosphorylating tumor suppressor proteins (p53, AIM, INK4A, and ARF). Wip1 mediates negative feedback of p38 MAPK/p53 signaling by dephosphorylating p-p53 (S15) in response to ultraviolet radiation. The p53 gene and protein have been previously reported to be reduced in L psoriatic skin compared with NL normal skin. This is in accordance with our findings that Wip1 is active in L psoriatic skin, forms complexes with p-RSK2, deregulates P-RSK2 (S386) activity, and dephosphorylates p-p53 (S15).

In cultured keratinocytes, MIF stimulation increased the new synthesis of Wip1 and inhibited P-p53 (S15), but costimulation with DMF inhibited Wip1 and induced p-p53 (S15) in accordance with the function of Wip1 and our earlier results (Figure 6B).

Deleting Wip1 phosphatase activity inhibited tumor growth. In MCF-7 cells, immunoprecipitation with RSK2 confirmed the high level of association of RSK2 with P-Wip1/Wip1. Earlier, TNF-α induced the transcription of Wip1 by activating NF-κB in MCF-7 cells. In the promoter region of Wip1, potential binding sites were found for transcription factors NF-κB, c-Jun, and CREB/ATF.

MSK1 activation was shown to induce NF-κB gene transcription, which is in accordance with the new synthesis of Wip1. Along that line, preincubating cells with DMF resulted in the inhibition of P-MSK1/2, P-RSK1-2, and NF-κB activation, reducing the MIF-induced transcription of Wip1 and the association of Wip1 with RSK2 in keratinocytes and in MCF-7 cells, according to our hypothesis (Figure 8).

IL-20 is produced by suprapapillary keratinocytes in L psoriatic skin, and IL-20 polymorphism has been suggested to determine susceptibility to plaque-type psoriasis.
gene expression of IL-20 is induced through the activation of MSK1 and NF-κB in keratinocytes, and DMF inhibited IL-20 mRNA expression in vitro. Therefore, treatment with DMF will also reduce the level of IL-20 in the psoriatic skin and inhibit psoriasis-like morphological changes and inflammation.

**Conclusion**

Our results indicate that the constitutive, high levels of EGF and MIF in psoriatic patients\(^\text{3,4,39}\) result in the activation of the ERK signaling pathway, phosphorylation of MSK1/2 and RSK1/2 kinases, and activation and new synthesis of Wip1. The complex formation with Wip1 results in disruption of the full-length activation of RSK2 at S386, significantly interfering with the regulation of proliferation and apoptosis in keratinocytes in L psoriatic skin.

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**Disclosure**

The authors report no conflicts of interest in this work.

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