Clinical significance of miR-34a expression in thyroid diseases – an 18F-FDG PET-CT study

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Purpose: To evaluate the possible roles of miR-34a expression in thyroid lesions, to unravel the correlation between fluorodeoxyglucose (FDG) uptake and miR-34a expression and moreover, to discover the underlying mechanisms by which miR-34a regulates FDG avidity.

Methods: We retrospectively reviewed 75 patients with pathology-confirmed thyroid diseases who underwent 18F-FDG positron emission tomography/computed tomography (PET/CT) within 3 months before undergoing thyroid surgery and miR-34a analysis from June 2012 to July 2017. 18F-FDG uptake of thyroid lesions was also analyzed semiquantitatively using maximum standardized uptake value (SUVmax). The association between miR-34a expression and clinicopathological variables (age, sex, TNM stage, histopathology, lesion numbers, location and 18F-FDG avidity) was investigated. When there were multiple lesions in thyroid bed, only the one with the highest 18F-FDG uptake was analyzed. Next, we inhibited the miR-34a expression in TPC-1 cells and detected the expression of glucose transporter 1 (GLUT1) mRNA and protein.

Results: In the patients cohort, miR-34a was upregulated in those with malignant thyroid diseases compared with benign lesions. The expression of miR-34a was associated with tumor stages, histopathological types and SUVmax. There was an inverse relationship between miR-34a expression and SUVmax in patients with thyroid diseases (Spearman correlation coefficient = –0.553, P < 0.0001). With an SUVmax of 4.3 as the threshold, sensitivity and specificity of the prediction of miR-34a expression (low or high) were 70% and 94.3%, respectively. The area under the receiver operating characteristic curve was 0.843 (95% confidence interval: 0.749, 0.936; P = 0.001). Inhibiting miR-34a in TPC-1 cells significantly increased GLUT1 mRNA and protein expression.

Conclusion: miR-34a expression was upregulated in thyroid lesions, negatively correlated with SUVmax and can be predicted by FDG SUVmax. In addition, miR-34a may regulate FDG avidity via targeting GLUT1.

Keywords: miR-34a, thyroid cancer, PET/CT

Introduction

Thyroid lesions are one of the most common endocrine-related diseases worldwide. The types of thyroid tumors include benign adenoma and malignant papillary thyroid carcinoma (PTC), follicular thyroid carcinoma (FTC), medullary thyroid carcinoma (MTC) and anaplastic thyroid carcinoma (ATC). PTC, which is relatively less harmful compared to other cancers, is the most common type of thyroid carcinoma, contributing to 80% of all thyroid malignancies, with an overall 10-year survival rate of more than 90%.1 However, a significant increase in annual incidence is observed for papillary
cancers (from 6.7% to 10.3%) around the world, without statistically significant decrease in mortality. Using ultrasonography (US) and US-guided fine-needle aspiration (FNA) biopsies, impalpable mini-sized thyroid disease could be detected easily.8 Thyroid carcinoma is considered a curable disease; however, local recurrence and/or distant metastasis may occur in some patients, which indicates that accurate diagnosis and appropriate risk evaluation are still necessary. US-guided FNA biopsy has been thought to be the gold standard for thyroid nodule diagnosis, with correct final diagnoses in 70%–80% of cases, leaving 20%–30% of cases ambiguous.7 New biomarkers which may change clinical outcomes, including operative strategy and prognosis, remain to be discovered.

MicroRNAs (miRNAs), which were firstly identified in Caenorhabditis elegans, are noncoding, endogenous small RNA molecules containing 18–23 nucleotides and regulate gene expression by binding to the 3′-UTR of specific “target” mRNA.8 By regulating the expression of specific target mRNAs at the posttranscriptional level, miRNAs act as oncogenes as well as tumor suppressors. Recent studies have shown that miR-34a is involved in numerous biological processes, especially in cancers (Table 1).

miR-34 is a key regulator of tumor development, and miR-34a, miR-34b, and miR-34c are the major members of the miR-34 precursor family. miR-34a has been reported to be upregulated in PTC tissues and cell lines.25,26 However, no research has thoroughly elucidated the relationship between miR-34a and clinical characteristics so far.

Glucose metabolism can be molecularly imaged in vivo with 18F-fluorodeoxyglucose positron emission tomography (18F-FDG PET), and the uptake of FDG has been correlated with tumor proliferation in thyroid carcinomas. FDG PET scans are gaining an important role in thyroid cancer for

### Table 1 Overview of miR-34a roles in cancers

<table>
<thead>
<tr>
<th>Cancer type</th>
<th>miR-34a significance</th>
<th>Authors</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Breast carcinoma</td>
<td>Met oncogene is unaffected by miR-34a in nonmetastatic and metastatic breast carcinomas, while in bone metastasis miR-34a correlates with Met, a key player of the bone-metastatic process, expression</td>
<td>Maroni et al</td>
<td>9</td>
</tr>
<tr>
<td>Head and neck cancer</td>
<td>Forced miR-34a expression induced cell cycle arrest and senescence; genes, including FUT1, AXL and MAP2K1 were identified as the novel targets of miR-34a by qPCR and luciferase assay</td>
<td>Wang et al</td>
<td>10</td>
</tr>
<tr>
<td>Gastric cancer</td>
<td>Silencing miR-34a activates IGF2BP3, which ranked number 1 of the upregulated genes and associated with poor disease-specific survival</td>
<td>Zhou et al</td>
<td>11</td>
</tr>
<tr>
<td>Osteosarcoma</td>
<td>miR-34a targets DLL1, leading to the ATF2/ATF3/ATF4 signaling pathway, significantly altering and negatively regulating the multi-chemoresistance of osteosarcoma</td>
<td>Pu et al</td>
<td>12</td>
</tr>
<tr>
<td>Ovarian</td>
<td>Levels of miR-34a are downregulated in ovarian cancer patients. Introduction of miR-34a in ovarian cancer 94 cells resulted in reduced proliferation, motility and invasion</td>
<td>Yan et al</td>
<td>13</td>
</tr>
<tr>
<td>Myeloma</td>
<td>Therapeutic potential of synthetic miR-34a against human multiple myeloma was shown partially by repressing BCL-2 and CDK6</td>
<td>Di Martino et al</td>
<td>14</td>
</tr>
<tr>
<td>Hepatocellular cancer</td>
<td>miR-34a, by mimicking ST3GAL5, significantly changes HCC malignant behaviors and oncogenicity in vitro and in vivo assays, acting as tumor suppressor</td>
<td>Cai et al</td>
<td>15</td>
</tr>
<tr>
<td>Pancreatic cancer</td>
<td>miR-34a is downregulated in the majority of pancreatic cancers and inhibited the growth of MiaPaCa-2 subcutaneous xenografts</td>
<td>Pramanik et al</td>
<td>17</td>
</tr>
<tr>
<td>Melanoma</td>
<td>Overexpression of miR-34a decreased the invasiveness of melanoma cells, partially involved in P53</td>
<td>Yamazaki et al</td>
<td>18</td>
</tr>
<tr>
<td>Colorectal cancer</td>
<td>miR-34a mediates OXA resistance of CRC by inhibiting macroautophagy via TGF-β/Smad4 pathway</td>
<td>Sun et al</td>
<td>19</td>
</tr>
<tr>
<td>Lung cancer</td>
<td>Hypermethylation of miR-34a promoter was detected in 68.7% lung adenocarcinoma</td>
<td>Daugaard et al</td>
<td>20</td>
</tr>
<tr>
<td>Leukemia</td>
<td>miR-34a promoted cell apoptosis and inhibited autophagy by targeting HMGB1 in acute myeloid leukemia</td>
<td>Liu et al</td>
<td>21</td>
</tr>
<tr>
<td>Renal cell cancer</td>
<td>miR-34a was upregulated in 198 RCC patients and correlated positively with HNF4A</td>
<td>Fritz et al</td>
<td>22</td>
</tr>
<tr>
<td>Cervical cancer</td>
<td>Forced expression of miR-34a inhibited proliferation, migration and invasion by reducing HMGB1 mRNA and protein levels</td>
<td>Chandrasekaran et al</td>
<td>23</td>
</tr>
<tr>
<td>Prostate cancer</td>
<td>Involved in chemoresistance and therapeutic efficacy of PTX in PTC-resistant prostate cancer</td>
<td>Wen et al</td>
<td>24</td>
</tr>
</tbody>
</table>

**Abbreviations:** HCC, hepatocellular carcinoma; OXA, oxaliplatin; CRC, colorectal cancer; RCC, renal cell carcinoma; PTX, paclitaxel; PTC, papillary thyroid carcinoma; qPCR, quantitative polymerase chain reaction.
initial staging, radiation therapy planning and restaging after therapy.27–29 Furthermore, miR-34a was found to be significantly upregulated in type 2 diabetes,30 indicating that miR-34a possibly regulates glycolysis, glucose metabolism and FDG avidity. Researches have revealed that glucose transporter 1 (GLUT1) plays crucial roles in transporting $^{18}$F-FDG and FDG avidity. Researchers have revealed that glucose transporter 1 (GLUT1) plays crucial roles in transporting $^{18}$F-FDG into the cell by acting as a key rate-limiting enzyme.31,32 Thus, quantification of glucose metabolism with FDG PET may present a noninvasive method to predict miR-34a expression. We retrospectively examined a cohort of patients with thyroid diseases who had underwent miR-34a expression testing and pretreatment FDG PET, and explored the clinical significance of miR-34a expression as well as the relationship between miR-34a expression and FDG avidity. The study flow chart is shown in Figure 1.

Materials and methods

Patients, human tissues and cell culture

This study was approved by the Institutional Review Board of the Third Affiliated Hospital of Kunming Medical University. All patients whose tissue samples were used were informed about the study, and a written informed consent was obtained from them. Due to the retrospective design of this study, written informed patient consent for reviewing their medical records was waived by our institutional review board, but confidentiality of patient data was still confirmed. The records of patients with newly diagnosed thyroid diseases who underwent FDG PET/CT prior to thyroid surgery at our institution from June 2012 to July 2017 were reviewed. We identified a total of 81 patients with thyroid lesions who underwent FDG PET/CT within 1 month prior to operation. Of these patients, 6 with initial other primary cancers (4 had in lung and the other 2 in breast) were excluded. Ultimately, 75 patients were enrolled in this retrospective study, and the corresponding formalin-fixed paraffin-embedded (FFPE) surgical specimens were obtained from the Department of Pathology. Human TPC-1 cell line was purchased from Shanghai Cancer Institute (Shanghai, People’s Republic of China). TPC-1 cells were maintained in Dulbecco’s Modified Eagle’s Medium supplemented with 10% fetal bovine serum (Thermo Fisher Scientific, Waltham, MA, USA) at 37°C with 5% CO$_2$ in a humidified incubator.

Clinicopathological data analysis

Medical records were reviewed to determine sex and age at diagnosis. Pathology reports were reviewed for the following data: tumor size, multifocality, histopathology and location. TNM stage was assigned based on the AJCC Cancer Staging Manual 8th edition.33 None of the patients had a history of neck radiation. Thyroid cancer was diagnosed pathologically after surgery. Patients were divided into 2 groups according to miR-34a expression (0.36 is the average relative expression of miR-34a among all the patients). Basic information of the patients is shown in Table 2.

Interpretation and analysis of FDG PET/CT scan

$^{18}$F-FDG PET/CT scans were obtained with an advanced integrated PET/CT scanner (Biograph 16; Siemens). All patients fasted for at least 6 h before FDG was intravenously injected in the resting state. A standard dose of 0.12 mCi/kg$^{-1}$ $^{18}$F-FDG tracer was used for all patients. In the period between injection of $^{18}$F-FDG tracer and image acquisition, the patients were instructed to remain seated or recumbent and silent in order to minimize muscular $^{18}$F-FDG uptake. Patients were kept warm 30–60 min prior to tracer injection and throughout the uptake period in order to minimize $^{18}$F-FDG accumulation in brown fat. Blood glucose was measured in all diabetic

Figure 1 Study flowchart showing the inclusion and exclusion criteria and the entire study design. Abbreviations: FDG, fluorodeoxyglucose; GLUT1, glucose transporter 1; PET/CT, positron emission tomography/computed tomography; qRT-PCR, quantitative reverse transcription-polymerase chain reaction; SUVmax, maximum standardized uptake value.
surrounding thyroid tissue was categorized as non-FDG-avid while tumor with no discernible FDG uptake higher than pathological reports was categorized as FDG-avid tumor, the thyroid that corresponds to the location recorded in categorized: tumor with a focal discrete FDG uptake in CT images. Based on FDG uptake in thyroid, tumors were more than 10 years experience assessed the FDG PET/CT images. Two nuclear medicine physicians with acquired using a conventional 3-dimensional protocol with rotation time per rotation, 1.5 pitch and a section thickness of 120 kV X-ray voltage, 50 mA tube current, a 0.75 s tube The CT scan was performed using a standardized protocol field of view was from the skull base to the mid-thigh level. A Siemens scanner, with the patients positioned with both arms down. The CT scan comprised dual-slice CT. The scan field of view was from the skull base to the mid-thigh level. The CT scan was performed using a standardized protocol of 120 kV X-ray voltage, 50 mA tube current, a 0.75 s tube rotation time per rotation, 1.5 pitch and a section thickness of 5 mm. Immediately after the CT scan, PET images were acquired using a conventional 3-dimensional protocol with 2.5 min per frame. Two nuclear medicine physicians with more than 10 years experience assessed the FDG PET/CT images. Based on FDG uptake in thyroid, tumors were categorized: tumor with a focal discrete FDG uptake in the thyroid that corresponds to the location recorded in pathological reports was categorized as FDG-avid tumor, while tumor with no discernible FDG uptake higher than surrounding thyroid tissue was categorized as non-FDG-avid tumor. In the case of multifocal tumors, the largest tumor was selected for interpretation. After the acquisition, maximum standardized uptake value (SUVmax) was assessed on the Siemens syngo MultiModality WorkPlace system by a single nuclear medicine physician. The SUVmax was defined as the decay-corrected radioactivity per unit volume divided by the injected radioactivity per body weight of the patient. SUVmax was determined by manually placing a cylindrical region of interest (ROI) over the basin of the primary tumor site. This was done on trans-axial images by an experienced nuclear medicine physician. ROIs were drawn to cover the whole tumor. In cases of multiple malignant nodules, an ROI was drawn on the largest one.

### miR-34a inhibitor transfection

To assess the effect of miR-34a inhibitor on GLUT1 expression, we employed the miR-34a inhibitor (5′-ACAAC-CAGCUAAGACACUGCC A-3′) chemically synthesized by GenePharma (Shanghai, People’s Republic of China) using Lipofectamine 2000 (Thermo Fisher Scientific) according to the manufacturer’s protocol. The nonsense single-stranded RNA (5′-CAGUACUUUGUGUAGUA CAA-3′), chemically synthesized by GenePharma, was transfected into the negative control (NC) group.

### RNA extraction, reverse transcription and RT-qPCR

Twenty-micrometer sections were cut from FFPE surgical specimen of each patient. Macrodissection of specimens was performed to obtain at least 75% of tumor cells in samples before RNA extraction. Total RNA was isolated from tissue samples using RecoverAll™ total nucleic acid isolation kit for FFPE specimen (AM1975; Thermo Fisher Scientific) according to the manufacturer’s instructions. Concentration and purity of total RNA was assessed by monitoring A260/230 ratio and A260/280 ratio using NanodropND-1000 (Thermo Fisher Scientific). A260/280 ratios of 1.8–2.1 indicated acceptable RNA purity. The integrity of RNA was then analyzed using 1% agarose gel electrophoresis, and the gel was then stained using ethidium bromide. A total of 1 µg RNA was used to produce cDNA using a TaqMan miRNA reverse transcription kit according to the manufacturer’s instructions (Thermo Fisher Scientific). Real-time PCR was carried out using SYBR green Real-time PCR Master Mix on an ABI 7500 Real-time PCR instrument (Applied Biosystems). The conditions of real-time PCR were as follows: 95°C for 30 s, 95°C for 5 s, 55°C for 10 s to anneal and 72°C for 15 s to elongate followed by 35 cycles. U6 was used as internal control. Relative gene expression levels were calculated

### Table 2 miR-34a expression in thyroid lesions and clinical characteristics

<table>
<thead>
<tr>
<th>Clinical characteristics</th>
<th>Patients (n)</th>
<th>miR34a expression (mean ± SD)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>≤45</td>
<td>41</td>
<td>0.3495 ± 0.02</td>
<td>0.2729</td>
</tr>
<tr>
<td>&gt;45</td>
<td>34</td>
<td>0.3759 ± 0.01</td>
<td></td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>43</td>
<td>0.3551 ± 0.02</td>
<td>0.5398</td>
</tr>
<tr>
<td>Female</td>
<td>32</td>
<td>0.3700 ± 0.02</td>
<td></td>
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<tr>
<td>Tumor stage</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>I/II</td>
<td>47</td>
<td>0.4111 ± 0.01</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>III/IV</td>
<td>18</td>
<td>0.2811 ± 0.02</td>
<td></td>
</tr>
<tr>
<td>Histopathology</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>PTC</td>
<td>41</td>
<td>0.4251 ± 0.08</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>FTC</td>
<td>12</td>
<td>0.3217 ± 0.04</td>
<td></td>
</tr>
<tr>
<td>FTC</td>
<td>8</td>
<td>0.3050 ± 0.05</td>
<td></td>
</tr>
<tr>
<td>FTC</td>
<td>4</td>
<td>0.2650 ± 0.05</td>
<td></td>
</tr>
<tr>
<td>Goiter</td>
<td>10</td>
<td>0.1980 ± 0.03</td>
<td></td>
</tr>
<tr>
<td>Lesion numbers</td>
<td></td>
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<tr>
<td>Single</td>
<td>51</td>
<td>0.3629 ± 0.02</td>
<td>0.4865</td>
</tr>
<tr>
<td>Multiple</td>
<td>24</td>
<td>0.3442 ± 0.02</td>
<td></td>
</tr>
<tr>
<td>Location</td>
<td></td>
<td></td>
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<tr>
<td>Left</td>
<td>38</td>
<td>0.3597 ± 0.11</td>
<td>0.8570</td>
</tr>
<tr>
<td>Right</td>
<td>26</td>
<td>0.3600 ± 0.11</td>
<td></td>
</tr>
<tr>
<td>Both</td>
<td>11</td>
<td>0.3400 ± 0.12</td>
<td></td>
</tr>
<tr>
<td>SUVmax</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Low (≤3.63)</td>
<td>27</td>
<td>0.4326 ± 0.02</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>High (&gt;3.63)</td>
<td>48</td>
<td>0.3098 ± 0.01</td>
<td></td>
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</table>

**Abbreviations:** SD, standard deviation; PTC, papillary thyroid carcinoma; FTC, follicular thyroid carcinoma; MTC, medullary thyroid carcinoma; ATC, anaplastic thyroid carcinoma; SUVmax, maximum standardized uptake value.
using the 2-ΔΔCt method. The sequences of primers used for miR-34a expression analysis were as follows: forward, 5’-GTCTCTAGACTAAAGGGTTGCATGGTGT-3’; reverse, 5’-GTCCGCCGCGCCAGAGGCAATATA-CATTCTCCCGCAG-3’.

Western blot analysis
Samples of 20 μg total protein were separated on 10% SDS-PAGE, and were transferred onto a polyvinylidene difluoride (PVDF) membrane (Bio-Rad, Hercules, CA, USA). The PVDF membrane was incubated with phosphate-buffered saline, containing 5% milk, overnight at 4°C. Subsequently, the PVDF membrane was incubated with monoclonal mouse anti-human GLUT1 and β-actin primary antibodies (1:2000 and 1:2500; Abcam) at room temperature for 1 h. This was followed by incubation with rabbit anti-mouse IgG-HRP (1:2000; Santa Cruz Biotechnology) at room temperature for 1 h. An enhanced chemiluminescence kit (Pierce Biotechnology) was then used to detect chemiluminescence. The relative protein expression was analyzed by Image-Pro Plus software (version 6.0; Media Cybernetics, Inc., Rockville, MD, USA), represented as the density ratio versus β-actin.

Statistical analysis
All statistical analyses were performed using IBM SPSS version 22 by 2 bio-statisticians. Univariate analysis of SUVmax was performed using univariate analysis. Patients’ age, tumor and lymph node stage and, miR-34a expression were included as independent variables into the model. Spearman rank correlation was used to determine the association between miR-34a expression and SUVmax. The receiver operating characteristic (ROC) curve was used to assess the best threshold of SUVmax which was used to predict miR-34a expression. The best threshold of SUVmax was based on the highest Youden index. The miR-34a expression and SUVmax of thyroid lesions were evaluated for calculating the area under the ROC curve and Youden index for determining the cutoff value for SUVmax. The Youden index, which is a comprehensive measurement for the performance of a diagnostic test, was generated considering every possible cutoff point. The value that generates the highest Youden index for a particular ratio is considered as the best cutoff for that ratio, as it provides highest discrimination between pathology and no pathology. The data are represented as mean ± standard deviation. A P-value of 0.05 or less was considered statistically significant.

Results

Patients and tumor characteristics
A total of 75 patients were enrolled in the present study, including 32 males (42.7%) and 43 females (57.3%), with a mean age of 38 ± 21.8 (range 26–66) years at the time of surgery. All the patients underwent a total thyroidectomy with or without neck dissection. Three patients underwent level II–VI lymph node dissection, 3 underwent level II and level III lymph node dissection, 5 underwent level II lymph node dissection, 3 underwent level IV and central lymph node dissection and 49 underwent central neck dissection. In addition, 12 patients underwent subtotal thyroidectomy without lymph node dissection. Among the 75 cases, 41 (54.6%) were diagnosed as PTC, 12 (16%) as FTC, 8 (10.6%) as MTC, 4 (5.3%) as ATC and 10 (13.3%) as simple goiter (Table 2). We also calculated the average miR-34a expression and considered 0.36 as the cutoff value. Expression lower than 0.36 was labeled as miR-34a-L and higher than 0.36 labeled as miR-34a-H. Among the 75 patients, miR-34a-L was identified in 41 cases (54.6%) and miR-34a-H in 34 cases (45.3%).

Association between miR-34a expression and clinicopathological features of thyroid carcinoma
As shown in Table 2, the miR-34a expression between I/II and III/IV changed significantly (0.41 ± 0.01 and 0.28 ± 0.02, respectively; P < 0.001). The mean expression of miR-34a in benign thyroid lesions (goiter) was 0.19 ± 0.01, while it was 0.38 ± 0.01 in thyroid carcinomas (PTC, FTC, MTC and ATC) (P < 0.001). In the malignant thyroid lesions, the miR-34a expression varied significantly among PTC, FTC, MTC and ATC with the mean value of 0.43 ± 0.08, 0.32 ± 0.04, 0.31 ± 0.05 and 0.27 ± 0.05, respectively (P < 0.001). As shown in Table 2 and Figure 2, the tumor stage, histopathology types and SUVmax demonstrated significant correlation with miR-34a expression (Table 2 and Figure 2).

FDG PET characteristics
All patients had SUVmax data for their primary thyroid lesions, and in patients with low (less than 3.63) and high SUVmax (equal to or more than 3.63), the mean expression of miR-34a was 0.43 ± 0.02 and 0.31 ± 0.01, respectively (P = 0.0476). There was a trend toward a lower normalized miR-34a expression in the primary tumor of patients with higher SUVmax than in patients with lower SUVmax (P < 0.001) (Table 2 and Figure 2D).
ROC curves
The correlation between SUVmax and miR-34a expression was analyzed. An obvious negative correlation was observed with a Spearman correlation coefficient of −0.5528 (P < 0.001) (Figure 3A). ROC curve was generated using the logistic regression model for the normalized SUVmax of the primary tumor (Figure 4B); the area under the curve was 0.754. With an SUVmax of 4.3 as the threshold, sensitivity and specificity of the prediction of miR-34a expression were 70% and 94.3%, respectively. The area under the ROC curve was 0.843 (95% confidence interval [CI]: 0.749, 0.936; P = 0.001) (Figure 3B).

Inhibition of miR-34a expression upregulated GLUT1 expression in vitro
TPC-1 cells transfected with 45 nM miR-34a inhibitor or NC were cultured in full media. There was an approximately 55% reduction of the miR-34a level in TPC-1 cells treated with the inhibitor (Figure 4A). As depicted in Figure 5B and C, both the mRNA and protein of GLUT1 were increased in TPC-1 cells treated with miR-34a inhibitor for 12 h. As expected, TPC-1 cells transfected with miR-34a inhibitor exhibited a significant upregulation of GLUT1 mRNA and protein (2.3-fold and 1.8-fold, respectively) compared with NC and control group (P < 0.001) (Figure 4B and C). Collectively, these results demonstrated that miR-34a could regulate the GLUT1 expression in TPC-1 cells.

Discussion
In the present study, we demonstrate that miR-34a is upregulated in malignant thyroid diseases and its expression is inversely associated with tumor stage and histopathology types. SUVmax can be used to predict miR-34a expression, and an inverse correlation between SUVmax and miR-34a expression between I/II and III/IV (relative to U6)miR-34a expression among different pathology types (relative to U6) miR-34a expression between low and high FDG avidity (relative to U6) miR-34a expression between benign and malignant tissues (relative to U6)I/II***III/IV

**Abbreviations:** FDG, fluorodeoxyglucose; FTC, follicular thyroid carcinoma; MTC, medullary thyroid carcinoma; ATC, anaplastic thyroid carcinoma; SUVmax, maximum standardized uptake value; PTC, papillary thyroid carcinoma.
Figure 3 Graphs show the relationship between miR-34a expression and SUVmax in patients with thyroid diseases. (A) There was an inverse relationship between miR-34a expression and SUVmax in patients with thyroid diseases (Spearman correlation coefficient = –0.533, P < 0.0001). (B) ROC curve analysis of SUVmax to predict miR-34a expression. The miR-34a expression was divided into 2 different groups, named as miR-34a-L (whose miR-34a expression is lower than or equal to 0.36) and miR-34a-H (miR-34a expression is higher than 0.36). With an SUVmax of 4.3 as the threshold, sensitivity and specificity of the prediction of miR-34a expression (low or high) were 70% and 94.3%, respectively. The area under the ROC curve was 0.843 (95% confidence interval: 0.749, 0.936; P = 0.001).

Abbreviations: SUVmax, maximum standardized uptake value; ROC, receiver operating characteristic.

Figure 4 Inhibiting miR-34a expression increased GLUT1 expression. Graphs show the influence of miR-34a inhibition on glucose transporters in cultured TPC-1 cells. miR-34a may decrease 18F-FDG uptake via targeting GLUT1 expression. Data are mean ± standard deviation. (A) miR-34a inhibitor successfully downregulated the miR-34a expression (P < 0.001), while there is no significant statistical difference between TPC-1 and NC group (P > 0.05). (B and C) Influence of miR-34a inhibition on expression of GLUT1 mRNA and protein in TPC-1 cells. miR-34a inhibition significantly increased expression of GLUT1 mRNA and protein (both P < 0.001). ***P < 0.001.

Abbreviations: GLUT1, glucose transporter 1; FDG, fluorodeoxyglucose; NC, negative control.
expression was identified. With an SUVmax of 4.3 as the threshold, sensitivity and specificity of the prediction of miR-34a expression were 70% and 94.3%, respectively. The area under the ROC curve was 0.843 (95% CI: 0.749, 0.936; \( P = 0.001 \)). We further inhibited the miR-34a expression in TPC-1 cells and both elevated GLUT1 mRNA and protein were detected, indicating that miR-34a possibly reduced the FDG avidity partially via targeting GLUT1 expression in thyroid cancers.

The incidence of thyroid disease has been increasing in the past decades throughout the world.34–36 Microscopic nodules have been reported in approximately half of individuals, 3.5% have occult papillary carcinoma and 15% have palpable goiters.37 Currently, the gold standard for thyroid nodule diagnosis is US-guided FNA biopsy, achieving correct final diagnoses in 70%–80% of cases; FNA biopsy is a reasonable approach to thyroid nodules for it has decreased costs substantially by facilitating selection of patients who need to undergo surgical excision.38 However, the remaining 20%–30% of cases are considered indeterminate for malignancy.39 Therefore, additional methods which can improve the sensitivity and specificity of diagnosis are highly desirable. Overcoming the challenges of accurate assessments of the risk for individual patients is important to establish appropriate treatment plans and optimize outcomes.

Numerous researches have shown that miRNAs play important roles in multiple biological and metabolic processes, including cell differentiation, proliferation, survival and malignancy.5 miR-34 is a key regulator of tumor suppression. miR-34a, miR-34b and miR-34c are major members of the miR-34 precursor family. miR-34a is produced by its own transcript located in the second exon of chromosome 1, whereas miR-34b and miR-34c are produced by a common primary transcript derived from the second exon of chromosome 11.40 Previous studies have shown that miR-34a was downregulated in a considerable number of cancers via repressing an assembly of genes promoting cell proliferation (Table 1). However, in contrast to its frequent decreased expression in numerous cancers, miR-34a has been found to be increased in thyroid carcinomas and cell lines.41–43 In the present study, we confirmed that miR-34a is upregulated in malignant thyroid carcinoma, including PTC, FTC, MTC and ATC, compared with the benign tissues (Figure 2, \( P < 0.001 \)). Importantly, although statistical difference is not significant, miR-34a expression was associated with the histopathology types and expressed lower in ATC, which is the most malignant among thyroid cancers, indicating that miR-34a may also play a tumor suppressor role in thyroid carcinoma. We not only confirmed the changes in miR-34a mRNA expression in thyroid cancers which was previously identified but also give insight into the different roles of miR-34a in thyroid cancer.42 Ma et al revealed that overexpressing miR-34a promoted PTC cell proliferation and colony formation and inhibited apoptosis, whereas knockdown of miR-34a showed the opposite effects; furthermore, miR-34a overexpression led to activation of PI3K/Akt/Bad signaling pathway in PTC cells, and depletion of Akt reversed the pro-growth and antiapoptotic effects of miR-34a; thus, they concluded that miR-34a promotes proliferation and suppresses apoptosis in PTC cells via PI3K/Akt/Bad pathway, functioning as an oncogene in PTC.25 However, we may not completely agree with their conclusions. First, they carried...
out the study in TPC-1 cells, one of the human PTC cell lines. Although there were many common properties of the cell lines, each line had a unique profile of parameters, which most likely reflects the individuality of the tumors of origin and individual genotypes and capacity for a range of phenotypic expression of the cells. Therefore, their conclusions may not represent the miR-34a function among other thyroid cell lines. Second, they carried out the experiment in TPC-1 cell line without analyzing the clinical significance of miR-34a in clinical samples, so possibly their study cannot reflect the virtual roles of miR-34a in clinical samples. In the present study, we analyzed the correlation between clinical characteristics and miR-34a expression based on clinical samples and confirmed that the tumor stage, histopathology types and SUVmax indeed influence the miR-34a expression. Moreover, our results demonstrated that tumors with high-level malignancy expressed lower miR-34a; elevated miR-34a in thyroid carcinoma, including PTC, FTC, MTC and ATC, may be an outcome of cells’ reaction trying to protect themselves. Cong et al analyzed miRNA using 499 PTC samples and 58 normal thyroid tissues obtained from The Cancer Genome Atlas (TCGA) database and identified that miR-34a was upregulated in PTC tissues, which is consistent with our research. Moreover, they revealed that miR-34a expression was higher in later tumor stage (2.69 in III/IV stage vs. 2.61 in I/II stage, $P = 0.004$), which seems different from our study. This disparity may be caused on the one hand by the different patient cohorts (PTC only in Cong et al’s study, while PTC, FTC, MTC and ATC in the current study); on the other hand, Cong’s research included different patients of different regions, ages, races and so on, so possibly heterogeneity within the group is inevitable. Fritz et al confirmed that miR-34a function as a tumor suppressor in renal cell carcinoma (RCC) whereas they found miR-34a to be upregulated in a larger cohort of RCC patients, indicating that tumor suppressor does not have to be downregulated. Although Dalgard et al confirmed tumor suppressor activity for miR-34a, they also observed differential levels of miR-34a expression in different retinal blastoma cells and clinical sample tissues. Moreover, differential miR-34a expression is also observed in breast, melanoma, pancreatic and prostate cancer cell lines. These phenomena can be explained by gain or loss of 1p36 at the miR-34a loci, CpG methylation silencing or genomic alteration at the p53-binding site in the miR-34a gene. In the present study, no significant correlations were identified between miR-34a expression and other clinicopathological features of thyroid diseases, including age, gender, lesion numbers and the position of lesions.

$^{18}$F-FDG uptake is based on enhanced aerobic glycolysis in cancer cells, known as Warburg effect. Several investigators have shown that FDG positive avidity is likely to be associated with a larger tumor size, the presence of extrathyroid extension and lymph node metastasis in patients with PTC; however, others have declared that there is no correlation between the SUVmax in primary PTC and the presence of multifocality, extrathyroid extension of primary PTC or lymph node metastasis. As miR-34a could serve as a therapeutic target, using miR-34a mimics may lead to restoration of the level of miR-34a and many target proteins, and thus to an improvement of the disease. miR-based treatment is currently becoming a reality as there are many biotech companies focusing on the use of miRNAs. Using SUVmax to predict the miRNA expression is safe and noninvasive. In the current study, we demonstrated that there is an obvious correlation between SUVmax and miR-34a expression, and with a cutoff value of 4.3, SUVmax can effectively predict the miRNA expression (Figure 3).

Previous studies have shown that there was significant correlation between the expression of GLUT1 and SUVmax in $^{18}$F-FDG PET/CT, indicating that GLUT1 plays an important role in increasing FDG uptake. To explore the potential mechanisms by which miR-34a regulates FDG avidity, we carried out other experiments in TPC-1 cells. After inhibiting the miR-34a, we detected the GLUT1 expression and found increased GLUT1 mRNA and protein (Figure 4), indicating that miR-34a possibly inhibits FDG avidity by targeting GLUT1 in thyroid cancers (Figure 5).

Our study has several unique strengths. First, this single-center study on thyroid lesions limits participant heterogeneity and variations in tumor subtypes. Second, our finding that the miR-34a expression was associated with clinicopathological parameters suggests the utility of preoperative miR-34a analysis in risk stratification and surgical management. Lastly, we analyzed the relationship between miR-34a expression and $^{18}$F-FDG PET/CT features in both malignant thyroid carcinoma and benign lesions and found out that the SUVmax can ideally be used to predict the miR-34a expression. Some limitations in our study need to be declared. First, the sample size (75 patients) was relatively small and this is partially due to the strict inclusion criteria (primary diagnosis, untreated and complete pathology results). Second, the current study was conducted at one hospital, which means the present results are relatively limited. Third, this was a retrospective study, and there was unavoidable selection bias. Further large prospective studies that include more hospitals and clinical samples are urgently needed to consolidate our
results. Finally, the interaction between miR-34a and GLUT1 is indirect and more substantial evidence should be dug out. In addition, more specific experiments illustrating the interaction between miR-34a and GLUT1 (luciferase reporter essay, for example) should be carried out.

**Conclusion**

Collectively, we found that in patients with malignant thyroid lesions, miR-34a expression is upregulated compared with the benign tumors or goiters and associated with tumor stage and histopathology types; moreover, an inverse correlation between 18F-FDG PET/CT avidity was confirmed and miR-34a may regulate FDG uptake partially via targeting GLUT1, which needs to be validated in our future study.

**Acknowledgments**

This study was supported by the Initiation Foundation for Doctors of Yunnan Tumor Hospital (BSKY201706).

**Disclosure**

The authors report no conflict of interest in this work.

**References**


