

Light-triggered methylcellulose gold nanoparticle hydrogels for leptin release to inhibit fat stores in adipocytes

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Abstract: Leptin is released in response to increased triglyceride storage in adipocytes and impacts body weight, but has drawbacks such as poor therapeutic effect and side effects when delivered systemically. Leptin also modifies adipocyte sensitivity to insulin to inhibit lipid accumulation. Here, light-triggered degradation of hydrogels was used to improve accuracy and effectiveness for sustained and controllable release. In our approach, leptin was entrapped within methylcellulose (MC)-based hydrogels, with incorporation of gold nanoparticles (NP). The incorporation of gold NP into MC hydrogels led to a tunable light irradiation response that dictated the hydrogel release rate of leptin. This manuscript demonstrates feasibility in designing tunable thermosensitive hydrogels for loading multimodality therapeutic agents to enhance the bioactivity of leptin for obesity therapy.

Keywords: leptin, adipocytes differentiation, methylcellulose, gold nanoparticle, degradable hydrogel

Introduction

Leptin is produced primarily by adipose tissue, and circulating concentrations correlate with the amount of body fat stores.¹ Although in vitro studies have tested the ability of leptin to directly influence adipocyte metabolism, further evidence indicates leptin inhibits insulin responsiveness of adipose tissue.² Furthermore, leptin modifies adipocyte sensitivity to insulin to inhibit lipid accumulation. In addition, glucose uptake by adipose tissue showed no response to leptin when leptin was administered intravenously³ or directly⁴ to adipocytes. Prolonged exposure to leptin did, however, cause a substantial dose-dependent inhibition of insulin-stimulated glucose uptake by the adipocytes. Leptin is a major factor in the modulation of body weight, but suffers from poor therapeutic effect and side effects after systemic administration.⁵

Hydrogels have shown broad utility for tissue regeneration and drug delivery.⁶⁻⁹ Furthermore, swelling of pH-sensitive hydrogels in environments with lower pH – consistent with injured tissues (~pH 6.0)⁶ or tumor microenvironment (~pH 6.7)⁷ – has been promoted by inclusion of amino acids in the polymeric structure. Specifically, protonation of amine groups in response to pH change increased absorption of water, thus promoting degradation of the hydrogel and enhanced localized release.^{6,7} Among the hydrogels reported on in the literature, a methylcellulose (MC), composed of hydrophilic moieties that are non-immunogenic and non-biofouling of proteins, has shown interesting utility due to its thermogelling properties.¹⁰ The inverse MC gel remains injectable, allowing for easy storage, facile timing of injection, and no further manipulation after in vivo injection.

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Concurrently, the photothermal effect of metallic nanoparticles following light excitation has been employed in drug release.^{11,12} Dense, metallic nanoparticles are capable of attenuating incident light via scattering and absorption, leading to localized increase in temperature. In the current work, we designed a drug-loaded injectable MC hydrogel including gold nanoparticles (NPs) to impart light irradiation sensitivity to stimulate thermally triggered drug release. When illuminated, gold NP absorbed the light, resulting in a localized thermal effect and a consequent increase in the release of loaded materials (Figure 1A). Due to the physical effects of biological tissues attenuating light, we further tested long wavelengths of 808 and 980 nm, able to penetrate to depths consistent with subcutaneous adipocytes. MC–gold NP hydrogels were loaded with leptin to regulate fat storage in adipocytes, controlled with light irradiation. Cytotoxicity, photothermal effect, and leptin released from the hydrogels were measured. The release of leptin from hydrogels at different conditions toward adipocytes was used to verify light irradiation-mediated release with in-vitro studies.

Materials and methods

Materials

We purchased MC (Mw=14,000 g mol⁻¹), phosphate-buffered saline (PBS, pH 7.4), Oil Red O dye, dexamethasone, insulin, isobutylmethylisobutylxanthine (IBMX), leptin from mouse (>98%), and 4',6-diamidino-2-phenylindole dihydrochloride (DAPI) from Sigma-Aldrich Co. (St Louis, MO). CellTiter 96[®] AQ_{ueous} One Solution Cell Proliferation Assay kit for the 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) assay was purchased from Promega (Madison, WI). Bare gold NPs (A11-20, size: 20 nm) were purchased from Nanopartz[™] Inc (Loveland, Colorado). For transmission electron microscopy (TEM, JEOL JEM-1400) analysis, a drop of gold NP solution was allowed to air-dry onto a Formvar-carbon-coated 200-mesh copper grid. TEM images were imaged on a JEOL-1400 transmission electron microscope operating at an accelerating voltage of 100 kV. The UV–vis absorption spectra of the gold NP solutions that were prepared under deionized water were measured using a UV–vis spectrophotometer (Tecan's Sunrise; Tecan Trading AG, Männedorf, Switzerland).

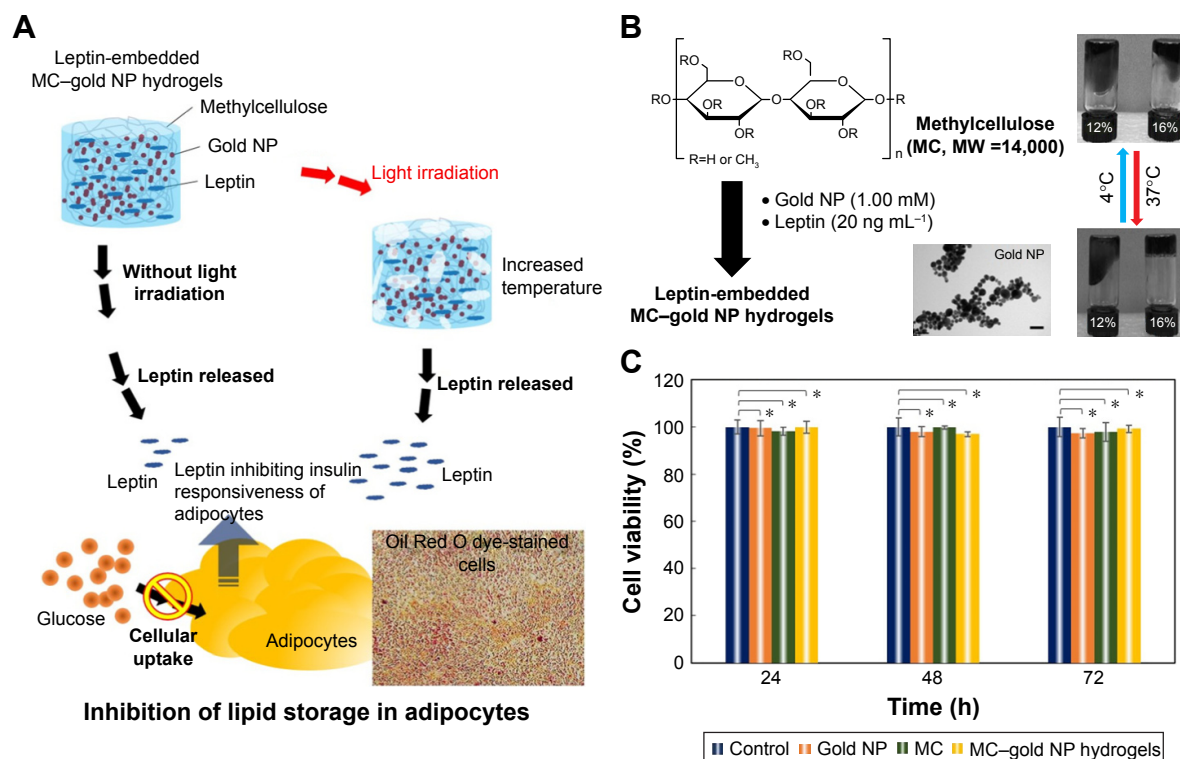


Figure 1 Light-sensitive MC–gold NP hydrogels.

Notes: (A) A conceptual representation of a light-triggered MC–gold NP hydrogel for leptin release to inhibit fat stores in adipocytes. Light irradiation increases porosity and promotes efflux of payload material in a controlled, tunable manner. (B) Schematic illustration of the synthesis of leptin-embedded MC–gold NP hydrogels. Leptin was embedded into MC–gold NP hydrogels and exposed to 37°C for 5 min to facilitate the formation of MC–gold NP hydrogels. The TEM image is of gold NPs. Bar = 50 nm. Photographs (right panel) of MC–gold NP hydrogels with 12% or 16% (v/v) MC. A complete hydrogel was achieved at 16% (v/v). (C) Viability of 3T3-L1 cells after exposure to various treatments. Cell viability is given as the percentage of viable cells remaining after treatment for 24, 48, or 72 h when compared with unexposed cells. Cell numbers were determined by the standard MTS assay (**P*>0.15; based on a two-tailed t-test, assuming unequal variances). The bars represent the mean ± standard deviation (n=6).

Abbreviations: MC, methylcellulose; MTS, 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium; NP, nanoparticle; TEM, transmission electron microscopy.

Cell culture and differentiation

The Mouse embryonal fibroblast 3T3-L1 (CL-173, ATCC) cell lines were maintained in Dulbecco's Modified Eagle's Medium (DMEM) with 4,500 mg L⁻¹ glucose, 10% fetal bovine serum (FBS), 100 U mL⁻¹ penicillin, and 100 µg mL⁻¹ streptomycin.

For differentiation protocol of 3T3-L1 cells to adipocytes,¹³ 3T3-L1 cells were cultured in Basal Medium I (BMI; DMEM supplemented with 4,500 mg L⁻¹ glucose, 10% FBS, 100 U mL⁻¹ penicillin, and 100 µg mL⁻¹ streptomycin) at designated Day 1. At 3 days post-confluence (Day 3), cells were induced to differentiate with Differentiation Medium I (DMI; DMEM supplemented with 1 µM dexamethasone, 0.5 mM isobutylmethylxanthine, and 10 µg mL⁻¹ insulin). After 2 days (Day 5), the medium were replaced with Differentiation Medium II (DMII; DMEM supplemented with 10% FBS and 10 µg mL⁻¹ insulin). At Day 7, the cells were maintained in the BMI and, subsequently, the cells were re-fed every 24 h with BMI. All cells were cultured in a 37°C incubator with 5% CO₂.

Preparation of hydrogels incorporating metal nanoparticles

The MC was dissolved in PBS to a final concentration of 12% or 16% (v/v) at 4°C. Subsequently, the gold NP solution was added into the MC solution, with stirring, resulting in final concentrations of 0.125, 0.25, 0.5, 1.00, or 2.00 mM. The mixed solutions were placed in cylindrical containers and exposed to 37°C, from 4°C onward, for 5 min to facilitate the formation of MC–gold NP hydrogels (6.0 mm diameter, 2.0 mm height), shown schematically in Figure 1B.

Toxicity of laser irradiation or hydrogels by MTS assay

For the MTS assay, 2×10⁵ 3T3-L1 cells were seeded in each of the wells of a 24-well plate and fed with culture medium overnight. A porous polyester Transwell™ insert with a pore size of 0.4 µm was placed in each well of the 24-well plate. Then, tested hydrogels were placed into the Transwell™ insert. The 3T3-L1 cells were then exposed to test hydrogels or laser wavelength of 520 nm (1.0 mW), 808 nm (1.5 mW), or 980 nm (1.5 mW) for different periods of irradiation for 24, 48, or 72 h incubation. The CellTiter 96® AQueous One solution cell proliferation assay system (Promega, Madison, WI, USA) was used to evaluate cell proliferation and observation of the optical density (OD) of formazan at 490 nm, quantified cell viability. The reagent contained a tetrazolium compound 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium and inner salt (MTS), and the reduction of MTS achieved by untreated cells was set at

100%, and that of test cells was expressed as a percentage of untreated cells.^{14–16} Data are shown as the mean ± the standard deviation for six independent experiments.

Temperature measurement of hydrogels

The concentration of 16% (v/v) MC hydrogel was chosen for subsequent measurements. Observation of MC–gold NP hydrogel at various gold NP concentrations (0.125, 0.25, 0.5, 1.00, or 2.00 mM) and temperature triggered by different laser wavelengths of 520 nm (1.0 mW), 808 nm (1.5 mW), or 980 nm (1.5 mW) for irradiation of various time periods was measured with a thermometer. The tip of the laser fiber was mounted above the hydrogel. Data are shown as the mean ± the standard deviation for three independent experiments.

Leptin release from hydrogels

For preparation of leptin-embedded MC–gold NP hydrogels, the aqueous leptin solution (20 ng mL⁻¹) was added into the mixture (1 mL) as described earlier. To assess the leptin released from these hydrogels, the leptin-embedded MC–gold NP hydrogels were placed in the Transwell™ insert and incubated in complete DMEM (0.5 mL) at 37°C for irradiation of laser wavelength (980 nm: 10 min) every day for 5 days. Leptin released from these hydrogels was determined by the Mouse Leptin ELISA Kit (Sigma-Aldrich Co., USA). Data are shown as the mean ± the standard deviation for four independent experiments.

In vitro studies and analysis

We seeded 2×10⁵ 3T3-L1 cells in each well of a 24-well plate, and followed the protocol as described in the subsection on differentiation of adipocytes. On Day 5 in the process of adipocyte differentiation, a porous polyester Transwell™ insert (Corning, NY) with a pore size of 0.4 µm was placed in each well. The leptin-embedded MC–gold NP hydrogels (6.0 mm diameter, 2.0 mm height) were placed in the Transwell™ insert and incubated in complete DMEM (0.5 mL) at 37°C for laser irradiation (520 nm: 4 min; 808 nm: 2 min; or 980 nm: 10 min) every day for 5 days. The tip of the laser fiber was mounted above the hydrogel. Untreated adipocytes were used as the control.

To identify adipocyte accumulation, fat droplets in 3T3-L1 cells were stained with Oil Red O dye.^{17,18} Before the Oil Red O dye staining, treated cells were washed with PBS, fixed with 4% paraformaldehyde, and washed twice with PBS. Cells were then treated with isopropanol (60%) for 1 min and then stained in the Oil Red O dye solution (0.18% [w/v] Oil Red O dye in 60% isopropanol) for 30–40 mins.

The treated cells were washed with isopropanol (60%) and twice with PBS. The Oil Red O dye-stained cells were observed in PBS under a microscope. Quantification of Oil Red O dye was performed by extracting the dye using 100% isopropanol and OD was then measured at a wavelength of 520 nm.

Total RNA was extracted from homogenized cell lysates using the Total RNA Isolation Kit (GenedireX, CA, USA), and first-strand cDNA was synthesized with the GScript Enzyme Mix (GeneDireX, CA, USA) according to the manufacturer's instructions. The peroxisome proliferator-activated receptor γ (PPAR- γ) cDNA was measured for quantitative real-time reverse-transcription polymerase chain reaction (qRT-PCR) analysis using the following sequences of primers of PPAR- γ : sense-CGCTGATGCACTGCCTATGA; antisense-AGAGGTCCACAGAGCTGATTCC. β -actin: sense-TGTCCACCTTCCAGCAGATGT; antisense-AGCTCAGTAACAGTCCGCCTAGA.

Results and discussion

Toxicity and temperature of MC–gold NP hydrogels

For preparation of injectable leptin-embedded MC–gold nanoparticles (NP) hydrogels (Figure 1B), an effective loading-matrix should exhibit minimal toxicity. Cytotoxicity of MC hydrogels, gold NP, or leptin-embedded MC–gold NP hydrogels to 3T3-L1 cells was evaluated by MTS assay after culturing for 24, 48, or 72 h. Cells with no treatment were used as a control group. No cytotoxicity was observed for any of MC hydrogels, gold NP (Conc: 1.00 mM), or leptin-embedded MC–gold NP hydrogels incubated with mouse embryonal fibroblast 3T3-L1 cells ($P>0.15$) (Figure 1C).

The photo-thermal effect of metallic nanoparticles upon light-excitation can trigger release of pre-loaded drugs from gels and carriers.^{11,12} In our experiment, the commercial gold NP yield the highest absorbance peak around 520 nm (Figure S1). Due to the major concern of light bio-penetration, we also applied two wavelengths of 808 and 980 nm into the photo-thermal treatment to be more consistent with treatment at a depth comprising subcutaneous tissues. To determine the effect of incorporated gold NP concentration on the temperature of MC–gold NP hydrogels, hydrogels were immersed and incubated at pH 7.4. The tip of the laser fiber was mounted above the hydrogel. The results showed that the temperature of the MC–gold NP hydrogels increased from $33^{\circ}\text{C}\pm 0.3^{\circ}\text{C}$ to $48^{\circ}\text{C}\pm 0.6^{\circ}\text{C}$ with the incorporation of gold NP at 1.00 mM for light irradiation using 980 nm at 0 and 10 min, respectively (Figure 2).

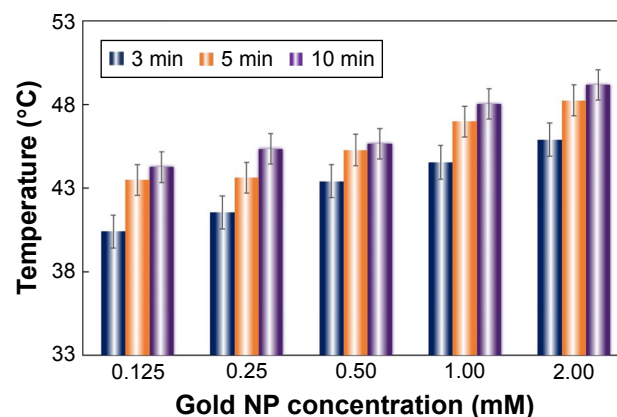


Figure 2 Temperature of MC–gold NP hydrogels under light irradiation.

Notes: Temperature achieved by irradiation with 980 nm (1.5 mW) for 3, 5, or 10 min for varying NP content. The tip of the laser fiber was mounted above the hydrogel. The bars represent the mean \pm standard deviation ($n=3$).

Abbreviations: MC, methylcellulose; NP, nanoparticle.

The incorporation of gold NP had a positive correlation on the temperature with considerable increases achieved even for the lowest concentration of 0.125 mM. At 2.00 mM gold NP concentration, some aggregation was observed. The temperature of MC–gold NP hydrogels under light irradiation using 520 nm or 808 nm also presented similar increase (Figure S2). In addition, when laser wavelength of 980 nm for light irradiation was increased to 10 min, cell viability was still maintained at $\sim 90\%$ ($P>0.15$) (Figure S3A). In contrast, laser wavelength of 808 nm for light irradiation of 2 min, resulted in cell viability $>75\%$ compared with laser wavelength of 520 nm ($P<0.05$), indicating that laser wavelength of 808 nm exhibited a significantly toxicity toward the cells. (Figure S3B). We subsequently chose a gold NP concentration of 1.00 mM and laser wavelength of 980 nm for light irradiation of 10 min in subsequent in vitro studies.

Differentiation of 3T3-L1 cells to adipocytes

In this culture protocol of adipocyte differentiation, mature adipocytes normally differentiated after Day 7. Staining for lipids, or fat, is commonly achieved with Oil Red O dye.¹⁹ Consequently, the quantification of lipid storage in cells and its correlation with Oil Red O dye-stained cells was determined by measuring OD of Oil Red O dye extracted from treated cells (Figure 3A). The OD value of Oil Red O dye increased gradually during the incubation period for the Basal Medium I (BMI, Day 0–Day 3), Differentiation Medium I (DMI, Day 3–Day 5), and Differentiation Medium II (DMII, Day 5–Day 7). The highest OD value was achieved at Day 9, and subsequently OD value decreased slowly after this day. Consistent with OD measurements,

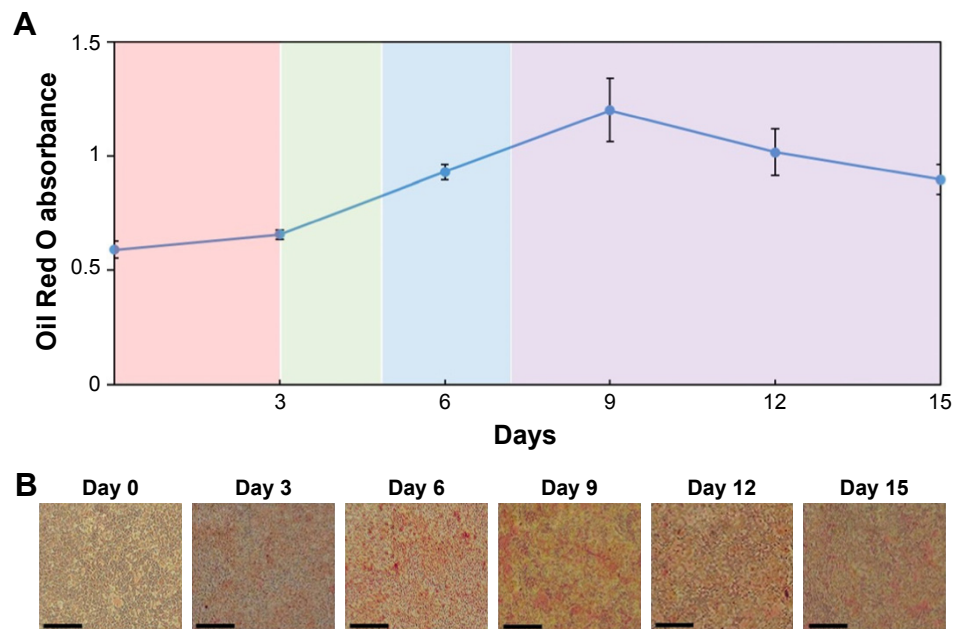


Figure 3 Differentiation of 3T3-L1 cells to adipocytes.

Notes: (A) Quantification of lipid storage in cells was measured by the optical density of Oil Red O dye, which was extracted from treated cells. The bars represent the mean \pm standard deviation ($n=3$). (B) Photographs of Oil Red O dye-stained cells at different periods. Bars =200 μ m.

we observed significant Oil Red O dye-stained adipocytes at Day 6, Day 9, and Day 12 with microscopy as shown in Figure 3B. Overall, we chose the mature adipocytes at Day 5–Day 9 for the measurement of effect of leptin on fat stores in subsequent *in vitro* studies.

Leptin released from leptin-embedded MC–gold NP hydrogels

To evaluate leptin concentration post-treatment, we used Mouse Leptin ELISA Kit to detect leptin released from these leptin-embedded MC–gold NP hydrogels. The concentration of leptin released from hydrogels increased gradually during the incubation period for the leptin-embedded MC–gold NP hydrogels incubated with light irradiation (980 nm; 10 min) every day (Figure 4A). Leptin release was significantly stimulated with light irradiation compared with treatment without light irradiation ($P<0.05$). In addition, control samples of leptin indicated a similar bioactivity with or without exposure to light irradiation. Taken together, the results imply that leptin released post-treatment with light irradiation did not affect bioactivity.

To investigate the effect of leptin released from these hydrogels on lipid storage in adipocytes, hydrogels were placed into the Transwell™ insert. The 3T3-L1 cells after post-differentiation (Day 5) were then exposed to these hydrogels under different conditions. On Day 7 or 9, leptin-embedded MC–gold NP hydrogels subjected to light irradiation

(980 nm) every day for 10 min and leptin with or without light irradiation showed similar inhibition of lipid storage ($P<0.05$; Figure 4B). Furthermore, the leptin-embedded MC–gold NP hydrogels treated with light irradiation displayed an ~50% inhibition of lipid storage compared with control on day 7 (222%, $P<0.05$) or on day 9 (198%, $P<0.05$). In Figure 4C, we observed micrographs of adipocytes when the leptin-embedded MC–gold NP hydrogels under light irradiation were utilized and coincides with the relative lipid storage shown in Figure 4B. In addition, leptin-embedded MC–gold NP hydrogels under light irradiation using 520 or 808 nm showed consistent results when using 980 nm (Figure S4). Overall, these results confirmed the considerable effect of leptin released from leptin-embedded MC–gold NP hydrogels on decreasing the amount of fat stores in adipocytes after light irradiation (Figure 4B).

In vitro assay

In addition to its importance in adipogenesis, PPAR- γ plays an important role in regulating fat storage.²⁰ Overall, leptin modulates mRNA levels of PPAR- γ in adipocytes (Figure 5A).²¹ Data analyzed by gel electrophoresis for PPAR- γ cDNA are consistent with the previous finding that the leptin released from MC–gold NP hydrogels under light irradiation (Lanes G–I) and that leptin under various conditions (Lanes B, D, E, and F) resulted in lower PPAR- γ cDNA (Figure 5B). Conversely, negative control (Lane A) and

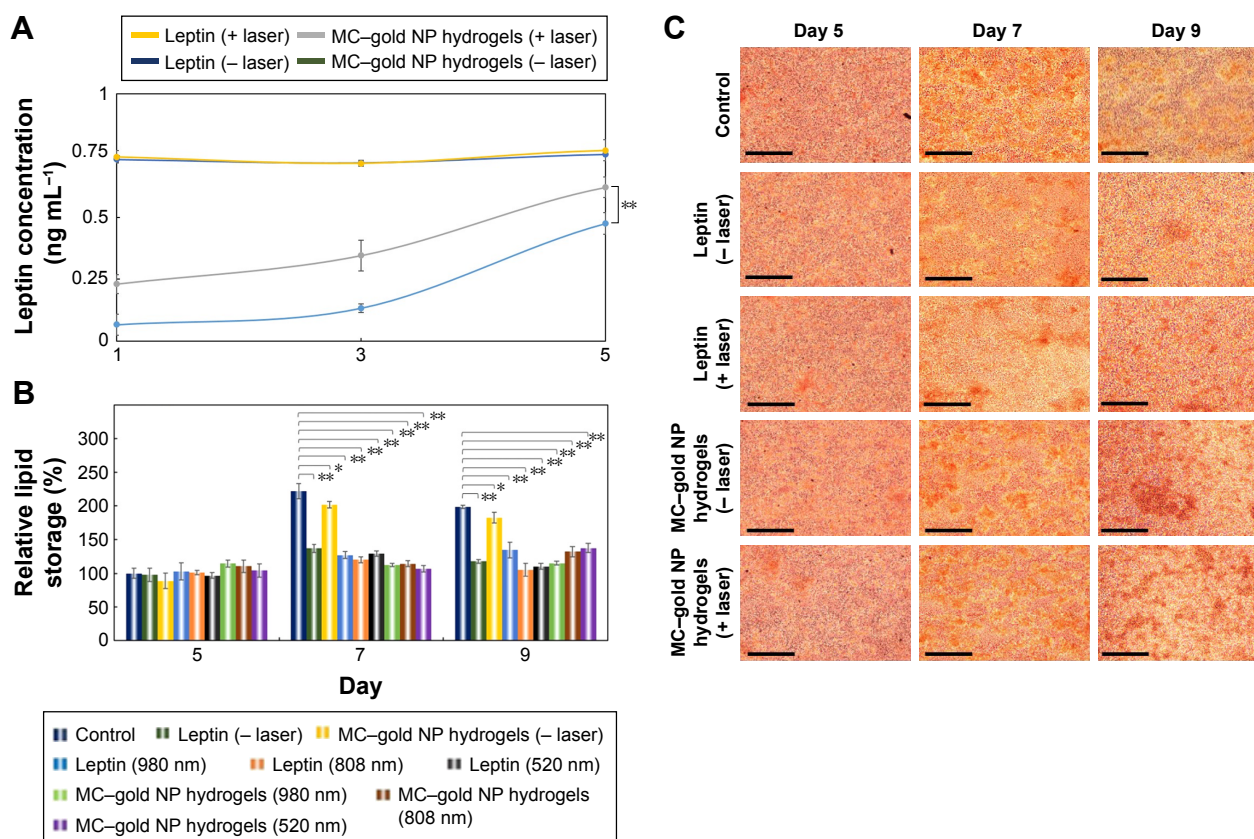


Figure 4 Light-triggered release of leptin from leptin-embedded MC-gold NP hydrogels.

Notes: (A) Measurement of the leptin release. Leptin-embedded MC-gold NP hydrogels were placed in Transwell™ inserts (Corning, New York, NY, USA) and incubated in complete DMEM at 37°C for irradiation (980 nm; 10 min) every day for 5 days. Leptin release was determined by Mouse Leptin ELISA Kit (Sigma-Aldrich Co., St Louis, MO, USA; ** $P < 0.05$; based on a two-tailed t -test, assuming unequal variances). The bars represent the mean \pm standard deviation ($n=4$). Lines are included to guide the eye. (B) Quantification of lipid storage in cells was measured by the optical density (OD) of Oil Red O dye post-incubation under various conditions of light irradiation (* $P > 0.15$; ** $P < 0.05$; based on a two-tailed t -test, assuming unequal variances). The bars represent the mean \pm standard deviation ($n=3$). (C) Photographs of Oil Red O dye-stained cells at different periods post-treatment. Bars = 200 μ m.

Abbreviations: -, without; +, with; MC, methylcellulose; NP, nanoparticle.

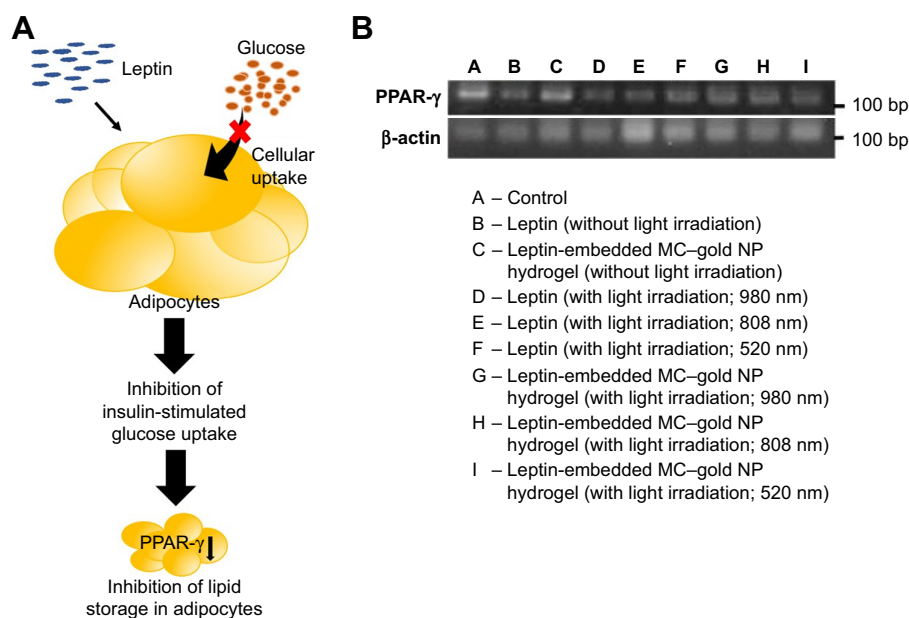


Figure 5 Effect of leptin on the mRNA levels of PPAR- γ in adipocytes.

Notes: (A) A conceptual representation of the leptin-modulated mRNA levels of PPAR- γ in adipocytes. (B) Gel electrophoresis. PPAR- γ cDNA was measured for quantitative real-time reverse transcription polymerase chain reaction analysis. β -actin cDNA was used as an internal standard.

Abbreviations: MC, methylcellulose; NP, nanoparticle.

MC–gold NP hydrogels without light irradiation (Lane C) achieved significantly greater PPAR- γ cDNA levels than the other groups. These data imply that successful passive delivery of leptin released from these hydrogels to adipocytes results in localized and highly efficient inhibition of fat storages.

Conclusion

MC–gold NP hydrogels with potential for efficient and localized release triggered by light irradiation were developed. The real-time temperature-dependent degradation rate of the MC–gold NP hydrogels could be controlled by the proportion of gold NP included. Leptin was embedded into MC–gold NP hydrogels and then effectively released with light irradiation. The released leptin could accumulate in cells efficiently. Such light-triggered leptin release from hydrogels has broad potential for drug delivery.

Acknowledgment

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Disclosure

The authors report no conflicts of interest in this work.

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Supplementary materials

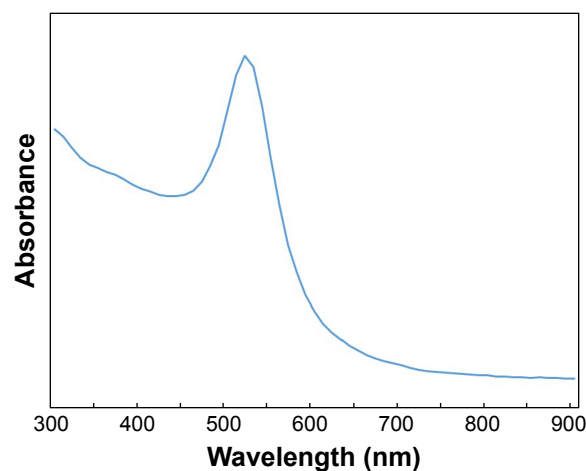


Figure S1 Characterization of gold nanoparticles (NPs).

Note: UV-vis absorption spectra of gold NPs, revealing absorption peak at approximately 520 nm.

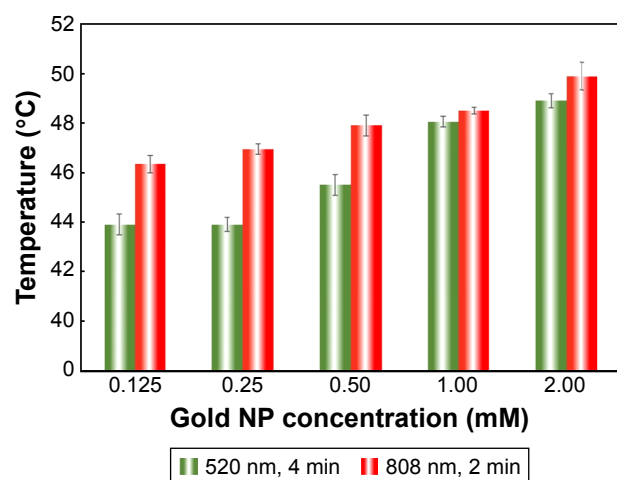


Figure S2 Temperature of MC-gold NP hydrogels under light irradiation.

Notes: Temperature was measured for MC-gold NP hydrogels at various gold NP concentrations after irradiation (520 nm: 4 min, 808 nm: 2 min). The tip of the laser fiber was mounted above the hydrogel. The bars represent the mean \pm standard deviation ($n=3$).

Abbreviations: MC, methylcellulose; NP, nanoparticle.

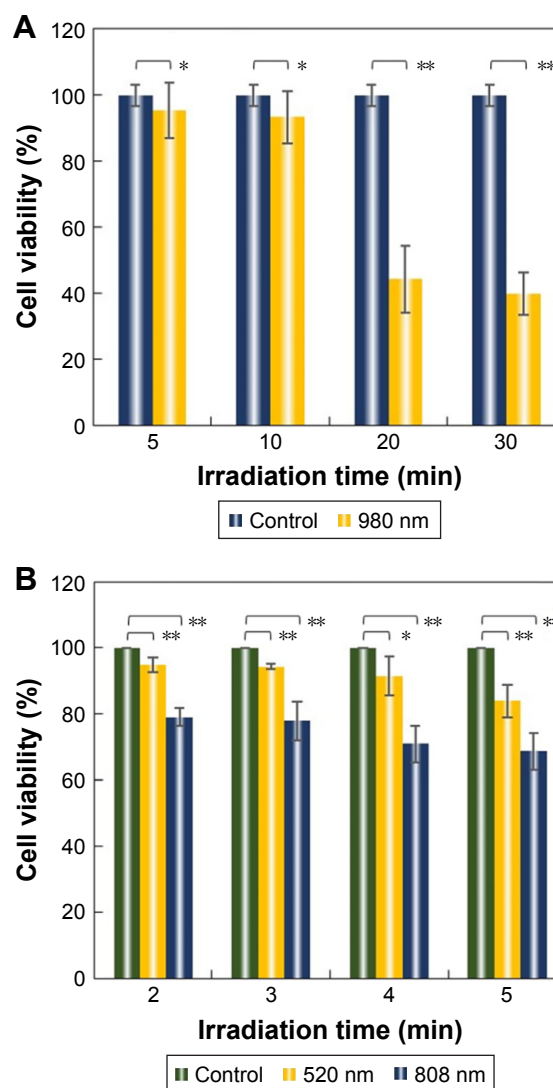


Figure S3 Cell viability after light irradiation.

Notes: Viability of 3T3-L1 cells after exposure to laser wavelengths of 980 nm (A) and 520 and 808 nm (B). Cell viability is given as the percentage of viable cells remaining after various treatments compared against the unexposed cells. Cell numbers were determined by the standard MTS assay (* $P>0.15$; ** $P<0.05$; based on a two-tailed t-test, assuming unequal variances). The bars represent the mean \pm standard deviation ($n=6$).

Abbreviation: MTS, 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulphophenyl)-2H-tetrazolium.

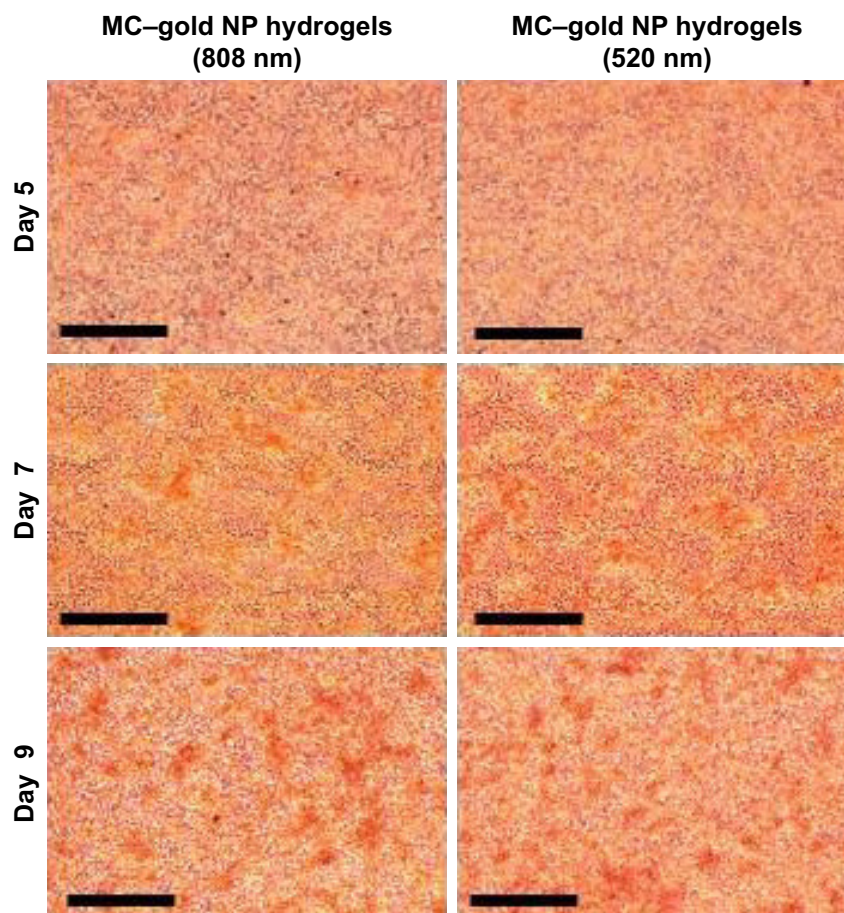


Figure S4 The lipid storage in adipocytes post-treatment.

Notes: Photographs of Oil Red O dye-stained cells at different periods post-treatment. Bars =200 μ m.

Abbreviations: MC, methylcellulose; NP, nanoparticle.

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