Investigating a rare methicillin-resistant Staphylococcus aureus strain: first description of genome sequencing and molecular characterization of CCI5-MRSA

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Correspondence: Abiola C Senok Mohammed Bin Rashid University of Medicine and Health Sciences, Building 14, Dubai Healthcare City, PO Box 505055, Dubai, United Arab Emirates Tel +971 4 383 8717 Email abiola.senok@mbru.ac.ae **Purpose:** Methicillin resistant *Staphylococcus aureus* CC15 strains (CC15-MRSA) have only been sporadically described in literature. This study was carried out to describe the genetic make-up for this rare MRSA strain.

Methods: Four CC15-MRSA isolates collected in Riyadh, Saudi Arabia, between 2013 and 2014 were studied. Two isolates were from clinical infection and 2 from retail meat products. Whole genome sequencing was carried out using Illumina HiSeq2500 genome analyzer.

Results: All the CC15-MRSA isolates had the multilocus sequence typing profile ST1535, 13–13-1–1-81-11-13, which is a single locus variant of ST15. Of the 6 contigs related to the SCC element, one comprised a recombinase gene ccrAA, ccrC-PM1, fusC and a helicase, another one included mvaS, dru, mecA and 1 had yobV and Q4LAG7. The SCC element had 5 transposase genes, namely 3 identical paralogs of tnpIS431 and 2 identical paralogs of tnpIS256. Two identical copies of a tnpIS256-based insertion element flank the aacA-aphD gene. Two copies of this insertion element were present with 1 located in the SCC element and another inserted into the sasC gene. A short 3 kb region, which lacks any bacteriophage structural genes and site-specific DNA integrase, was inserted into the hlb gene. The hsdM and the 5'-part of the hsdS gene are replaced by a copy of the hsdM/hsdS paralogs from $vSa\beta$ giving rise to a new chimeric paralog ofhsdS in vSaa.

Conclusion: CC15-MRSA shows a novel SCC*mecV/SCCfus* composite element. Its variant of *hsdM/hsdS* probably facilitated uptake of foreign mobile genetic elements that promoted emergence of CC15-MRSA. Close surveillance is needed to monitor spread and emergence of further CC15 MRSA strains.

Keywords: whole genome sequencing, MRSA, MLST, clonal complex, SCCmec, Saudi Arabia

Introduction

In recent years, the landscape of the molecular epidemiology of methicillin resistant *Staphylococcus aureus* (MRSA) has been characterized by the emergence and dissemination of new strains. Clonal complex 15 (CC15) is ubiquitous and widely described in the literature, but these isolates are mostly methicillin susceptible *S. aureus* (MSSA). CC15-MSSA was recently identified as a predominant nasal colonizer in a report from Saudi Arabia. Previously, methicillin resistant CC15 strains (CC15-MRSA) have only been sporadically described in literature. In a large scale genotyping study of MRSA isolates, no CC15-MRSA was identified. Two isolates of CC15-MRSA associated with nasal colonization have been reported in Iran and Saudi Arabia. While whole genome sequencing data are available for CC15-MSSA, there are, to the best of our knowledge, no publications on the genomic data for the rare CC15-MRSA. Recently, we reported the first

identification of CC15-MRSA from clinical infections and retail meat products in the Middle East.^{6,7} In light of the emergence of CC15-MRSA in our setting and to provide much-needed insight into the genetic make-up of this rare MRSA clone, we have carried out whole genome sequencing of these isolates.

Materials and methods

The human isolates were identified as part of a larger MRSA study for which ethical approval was obtained from the Institutional Review Board, King Khalid University Hospital, Riyadh, Kingdom of Saudi Arabia. Patient consent was waived as the study involved use of archived isolates from specimens submitted for routine diagnostic tests and without use of patient identifiers. Four CC15-MRSA isolates collected in Riyadh, Saudi Arabia between 2013 and 2014 were studied. Two isolates (RUH-2 and RUH-71) were from patients with sepsis and wound infection, respectively, while the other 2 (RUH-98 and RUH-99) were from retail camel meat. S. aureus identification and confirmation of methicillin resistance was performed as previously described.^{6,7} Genomic DNA was extracted using Qiagen DNA isolation kit (Qiagen, Hilden, Germany) in accordance with manufacturer's instructions. Whole genome sequencing was carried out using the Illumina HiSeq2500 genome analyzer.

Sequencing reads were assembled de-novo with SPAdes and the final assembly was done with SPAdes version 3.10.1 (http://bioinf.spbau.ru/spades).8 Contigs shorter than 500 nt were dropped. Reads were mapped to the SPAdes contigs but also to the reference sequence (ST15-MSSA strain ST20130938, GenBank: CP012972.19) with the Burrow-Wheeler aligner "bwa" using the local aligning algorithm "mem" ("bwa" version 0.7.12-r1039, https://github.com/ <u>lh3/bwa</u>). 10 We also used "bwa-mem" to map the whole SPAdes contigs on the reference sequence (ST15-MSSA strain ST20130938, GenBank: CP012972.1). Read mappings and coverage were visually inspected with "tablet" ("tablet" version 1.14.10.21, https://ics.hutton.ac.uk/tablet/).11 We manually scaffolded and annotated the contigs from isolate RUH-2, which cover the genomic islands of $vSa\beta$ and vSaa, a 3 kb element inserted into the hlb gene and the SCC element. We used the GenomeDiagram module from Biopython to draw sketches from the manually annotated sequences. 12 The reads and the SPAdes contigs were submitted to NCBI sequence database. The manually scaffolded and annotated regions were submitted to Genbank as short sequences.

Results

For each isolate, a de-novo assembly of the genomic sequence was carried out. The assemblies comprised 73 and 71 contigs

for the human isolates RUH-2 and RUH-71, respectively. Isolates RUH-98 and RUH-99 from camel meat had 72 and 66 contigs, respectively. The overall G/C content for the chromosomal contigs was 33%. All the CC15-MRSA isolates had the MLST profile 13–13-1–1-81–11-13. All of the 4 isolates sequenced carried a 30-kb plasmid harboring additional antibiotic resistance genes, namely *cadD*, *cadX*, *blaI*, *blaR*, *blaZ*, *lnuA*, *aadD*. In addition, isolate (RUH-71, from human wound infection) harbored another putative plasmidic contig encoding *tetK*. A comparison of the genomic features of the 4 CC15-MRSA isolates reported in this study (RUH-2, RUH71, RUH-98, RUH-99), with CC15-MSSA sequences in the NCBI GenBank (VCU006, MPROS1797, 08–02119, ST20130938, ST20130940, ST20130941) is given in Table S1.

The 6 contigs related to the SCC element (Figure 1) were identical in all isolates. One contig comprised a recombinase gene "ccrAA", ccrC-PM1, fusC and a helicase; another contig included mvaS, dru, mecA and 1 contig had yobV and Q4LAG7 (putative protein associated with SCCmec V/VT, GenBank AM990992.1: 52112 to 54100) (Figure 1). The SCC element presumably comprises 5 transposase genes, namely 3 identical paralogs of tnpIS431 (size 675 nt) and 2 identical paralogs of tnpIS256 (size 1173 nt) (Figure 1). Two identical copies of a tnpIS256-based insertion element flank the bifunctional kanamycin resistance determinant aacA-aphD. Two copies of this insertion element were present in the genome with 1 copy located in the SCC element and another copy inserted into the sasC gene encoding a surface protein.

The CC15-MRSA isolates had a short 3 kb region inserted into the *hlb* gene (Figure 2). The 3 kb insertion element lacks any bacteriophage structural genes and site-specific DNA integrase. The insertion element comprised 5 genes, including *scn* (staphylococcal complement inhibitor) and *chp* (chemotaxis inhibitor), but *sak* (staphylokinase) was absent (Figure 2).

The CC15-MRSA isolates showed a variant of hsdM/hsdS at the major pathogenicity island vSaa compared with the reference CC15-MSSA genome (Figure 3). The hsdM and the 5'-part of the hsdS gene were replaced by a copy of the hsdM/hsdS paralogs from vSaβ. This gives rise to a new chimeric paralog of hsdS in vSaa (Figure 3). The chimeric hsdS has an intact reading frame. We can see this recombination in all of the 4 CC15-MRSA isolates. Furthermore, a Sau3AI restriction system is present in all of the CC15 isolates analyzed, while the type IV restriction system SauUSI is absent (Table S1).

Discussion

All the CC15-MRSA isolates had the MLST profile 13–13-1–1-81–11-13, which is a single locus variant of ST15. This

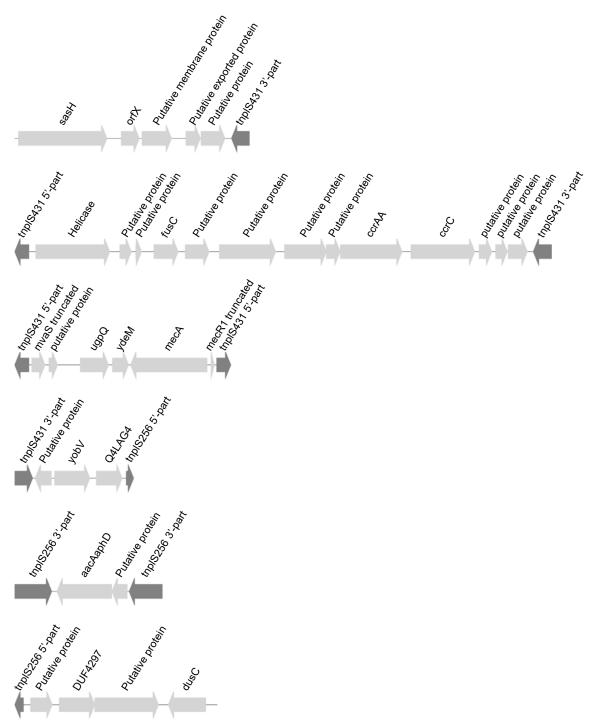


Figure 1 CC15-MRSA SCC element.

Notes: Six contigs related to the SCC element (MF185204 to MF185209). The first contig comprises the flanking orfX gene, and the last contig the flanking dusC gene. In between, four contigs are shown carrying genes typically found in SCC elements. Contig 2 has the fusidic acid resistance gene fusC and the SCC recombinase genes ccrAA and ccrC. Contig 3 comprises the mecA gene cluster with mecR1 truncated by tnplS431. Contig 5 constitutes a true insertion element, where the bifunctional kanamycin resistance determinant aacAaphD is flanked by two copies of tnplS256.

 $\textbf{Abbreviation:} \ \mathsf{MRSA}, \ \mathsf{methicillin-resistant} \ \mathsf{Staphylococcus} \ \mathit{aureus}.$

MLST profile has been assigned to ST1535 (https://pubmlst.org/bigsdb?db=pubmlst-saureus-isolates&page=profiles) and comprises pta-81 instead of pta-12 in canonical ST15. This pta-81 differs from pta-12 by only 1 single-nucleotide

polymorphism, which was present in all our isolates. Three of the 4 isolates assigned to ST1535 in the PUBMLST database are MSSA (https://pubmlst.org/bigsdb?db=pubmlst_sau-reus_isolates&page=profiles). The fourth is MRSA isolate

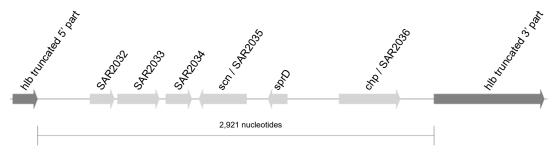


Figure 2 The hlb-3kb-insert in CC15-MRSA.

Notes: The hemolysin beta gene (hlb) is interrupted by a 3 kb insertion element in CC15-MRSA genomes.

Abbreviation: MRSA, methicillin-resistant Staphylococcus aureus.

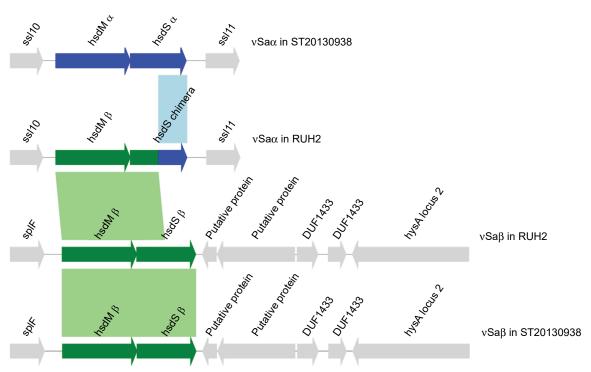


Figure 3 hsdM/hsdS recombination in CC15-MRSA.

Notes: The Figure shows the contents of genomic islands vSaa and $vSa\beta$ in isolate RUH-2 (ST1535/CC15, MF185202, MF185203) and in ST20130938 (ST15/CC15, Genbank accession CP012972.1). The reference genome CP012972.1 comprises two distinct paralog of hsdM/hsdS in genomic islands alpha and beta. The mapping of the sequencing reads from isolate RUH-2 onto the reference sequence CP012972.1 reveals, that hsdM-alpha and the 5'-end of hsdS- alpha are missing in RUH-2, while the coverage of hsdM-beta and the 5'-end of hsdS- beta is doubled with respect to other chromosomal genes, indicating that this stretch of DNA is duplicated in RUH-2. We extracted the duplicated region of $vSa\beta$ from the SPAdes contigs and were able to link it to contigs mapping to vSaa.

Abbreviation: MRSA, methicillin-resistant Staphylococcus aureus.

MPROS1797, which has a similar SCC element as the CC15 MRSA in this study (https://www.ncbi.nlm.nih.gov/biosample/SAMEA2664415; Table S1). Due to the presence of repeats, the SCC element could not be scaffolded into a single contiguous sequence. The overall constellation of the SCC element as shown in Figure 1 was interpreted as a novel SCC*mecV/SCCfus* composite element. A very similar element has also been found by microarray hybridization in CC97-MRSA from Saudi Arabia. Furthermore, reports from Saudi Arabia have described MRSA isolates from other lineages that also harbored SCC*fus* in addition to SCC*mec* IV

or V elements.^{6,13} Insertion elements flanked by 2 antiparallel copies of a transposase are common in bacteria, and often found in association with antibiotic resistance genes. The *sasC* gene, which is interrupted by insertion of another copy of the tnpIS256-based insertion element, has been linked with biofilm production in *S. aureus*.¹⁴

In *S. aureus*, an insert in the *hlb* gene is typically a prophage comprising several structural genes encoding the capsule, head and tail of the phage alongside an integrase at the terminus. It also frequently carries virulence associated genes like *sea*, *sep* (N315), *see*, *chp*, *sak* and *scn* in

various combinations. 15 The Riyadh CC15-MRSA isolates (as well as other published CC15 genomes) had a short 3 kb region inserted into the *hlb* gene (Figure 2). The absence of bacteriophage structural genes and site-specific DNA integrase suggests that it is no longer mobile but blocking the *hlb* insertion site. This insertion element seems to be a remnant of a bacteriophage as it is homologous to the terminus of the hlb converting phage in MRSA252 (Genbank accession BX571856.1, genes SAR2032 to SAR2036). In MRSA252, the *hlb* converting phage has a size of about 44 kb. However, we did not find any resemblance between the putative SCC elements in our CC15-MRSA isolates and the SCC element in MRSA252. MRSA252 has a full mec gene cluster comprising mecA/mecR1/mecI/mecR2, while the mec cluster is truncated in *mecR1* in the CC15-MRSA isolates. MRSA252 has cassette recombinase ccrA-2/ccrB-2, while we find ccrC in the CC15-MRSA isolates. The sak gene is associated with tissue invasion. 16,17 Its absence could be the reason why most CC15 isolates are associated with carriage rather than invasive infection.

Two distinct copies of the hsdM and hsdS genes are present in most genomes of S. aureus. These genes encode components of a type I restriction-modification (R-M) system (hsdM encodes a DNA methylase, hsdS encodes the specificity determinate). One pair of hsdM/hsdS is typically located in the genomic island vSaa between the superantigen-like genes ssl10 and ssl11. A second pair of hsdM/hsdS genes resides in genomic island $vSa\beta$. The CC15-MRSA isolates showed a different variant of hsdM/ hsdS at the major pathogenicity island vSaa compared with the reference CC15-MSSA genome. This has arisen presumably due to intrachromosomal recombination, resulting in a repertoire of hsdM/hsdS restriction enzymes that deviate from the CC15 parent. This might have played a role in acquiring an SCC element as bacteria use the type I R-M system to control uptake of foreign DNA. In the type I R-M system, the hsdM/hsdS gene products are required in this process.¹⁸ Usually, the composition of the genomic island vSaa and $vSa\beta$ is highly conserved within clonal complexes. It has been shown that the type I R-M system has facilitated the evolution of distinct S. aureus lineages and controls the horizontal transfer of mobile genetic elements.¹⁸ The Sau3A is a type II system that digests DNA at GATC sites and that was first described by Seeber et al. 19 Interestingly, Sau3AI is present in all CC15 isolates, while SauUSI is absent (Table S1). This is rather unusual as Sau3AI is uncommon in S. aureus. Indeed, most S. aureus isolates harbor the SauUSI (type IV) at the DNA locus for *Sau3AI*.^{19,20} Therefore, based on our findings, we suggest that changes in the *hsdM/hsdS* system and the type II R-M locus facilitated uptake of foreign mobile genetic elements, that is, of SCC*mec/SCCfus* by the ancestral CC15-MSSA promoting emergence of CC15-MRSA.

The limitation of our work is that the gaps between the contigs, which are presumably caused by repeated sequence elements, could not be resolved since the average fragment size of the Illumina library was only about 250 nt. Also, we were unable to determine the spa type reliably from our assembly since spa is a highly repetitive locus of a variable number of imperfect repeats. This genomic arrangement typically provokes artifacts in read assembly.

Accession numbers

The raw read sequences have been deposited in the Sequence Read Archive database (Bioproject PRJNA386092) with accession numbers: SAMN06925301, SAMN06925302, SAMN06925303, SAMN06925304.

De-novo assembled contigs have been deposited at DDBJ/ENA/GenBank under the accession NHZU00000,000, NHZV000000000, NHZW000000000, NHZX000000000. The version described in this paper is version NHZU01000000, NHZV01000000, NHZV01000000, NHZX010000000.

The manually scaffolded sequences for *hlb*_3kb_insert, *vSaa*, *vSaβ* and the 6 SCC element contigs have been submitted to the GenBank under the following accession numbers: MF185201, MF185202, MF185203, MF185204, MF185205, MF185206, MF185207, MF185208, MF185209

Conclusion

We provide the molecular characterization of a MRSA strain from a common lineage that until recently gave rise only to very few MRSA. The findings indicate that CC15-MRSA has a novel SCC*mecV*/SCC*fus* composite element. Changes in the *hsdM/hsdS* system and the type II R-M locus probably played role in the emergence of this rare MRSA strain. Close surveillance is needed, especially with regard to spread among humans and livestock in the Middle East and emergence of further CC15 MRSA strains.

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Disclosure

The authors report no conflicts of interest in this work.

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References

- Monecke S, Coombs G, Shore AC, et al. A field guide to pandemic, epidemic and sporadic clones of methicillin-resistant *Staphylococcus aureus*. PLoS One. 2011;6(4):e17936.
- Sarkar A, Raji A, Garaween G, et al. Antimicrobial resistance and virulence markers in methicillin sensitive *Staphylococcus aureus* isolates associated with nasal colonization. *Microb Pathog.* 2016;93:8–12.
- Abou Shady HM, Bakr AE, Hashad ME, Alzohairy MA. Staphylococcus aureus nasal carriage among outpatients attending primary health care centers: a comparative study of two cities in Saudi Arabia and Egypt. Braz J Infect Dis. 2015;19(1):68–76.
- Campanile F, Bongiorno D, Borbone S, Stefani S. Hospital-associated methicillin-resistant *Staphylococcus aureus* (HA-MRSA) in Italy. *Ann Clin Microbiol Antimicrob*. 2009;8:22.
- Japoni-Nejad A, Rezazadeh M, Kazemian H, Fardmousavi N, van Belkum A, Ghaznavi-Rad E. Molecular characterization of the first community-acquired methicillin-resistant *Staphylococcus aureus* strains from Central Iran. *Int J Infect Dis.* 2013;17(11):e949–e954.
- Senok A, Ehricht R, Monecke S, Al-Saedan R, Somily A. Molecular characterization of methicillin-resistant *Staphylococcus aureus* in nosocomial infections in a tertiary-care facility: emergence of new clonal complexes in Saudi Arabia. *New Microbes New Infect*. 2016;14: 13–18.
- Raji MA, Garaween G, Ehricht R, Monecke S, Shibl AM, Senok A. Genetic Characterization of *Staphylococcus aureus* Isolated from Retail Meat in Riyadh, Saudi Arabia. *Front Microbiol.* 2016;7:911.
- Bankevich A, Nurk S, Antipov D, et al. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol*. 2012;19(5):455–477.
- Trouillet-Assant S, Lelievre L, Martins-Simoes P, et al. Adaptive processes of *Staphylococcus aureus* isolates during the progression from acute to chronic bone and joint infections in patients. *Cell Microbiol*. 2016;18(10):1405–1414.

- Li H. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. 2013; arXiv:1303.3997v2. Available from: https:// arxiv.org/abs/1303.3997. Accessed August 24, 2017.
- Milne I, Stephen G, Bayer M, et al. Using Tablet for visual exploration of second-generation sequencing data. *Brief Bioinform*. 2013;14(2):193–202.
- Pritchard L, White JA, Birch PR, Toth IK. GenomeDiagram: a python package for the visualization of large-scale genomic data. *Bioinformatics*. 2006;22(5):616–617.
- Monecke S, Skakni L, Hasan R, et al. Characterization of MRSA strains isolated from patients in a hospital in Riyadh, Kingdom of Saudi Arabia. BMC Microbiol. 2012;12:146.
- Speziale P, Pietrocola G, Foster TJ, Geoghegan JA. Protein-based biofilm matrices in Staphylococci. Front Cell Infect Microbiol. 2014;4:171.
- van Wamel WJ, Rooijakkers SH, Ruyken M, van Kessel KP, van Strijp JA. The innate immune modulators staphylococcal complement inhibitor and chemotaxis inhibitory protein of *Staphylococcus aureus* are located on beta-hemolysin-converting bacteriophages. *J Bacteriol*. 2006;188(4):1310–1315.
- Monecke S, Luedicke C, Slickers P, Ehricht R. Molecular epidemiology of Staphylococcus aureus in asymptomatic carriers. Eur J Clin Microbiol Infect Dis. 2009;28(9):1159–1165.
- Wieckowska-Szakiel M, Sadowska B, Rozalska B. Staphylokinase production by clinical *Staphylococcus aureus* strains. *Pol J Microbiol*. 2007;56(2):97–102.
- Waldron DE, Lindsay JA. Sau1: a novel lineage-specific type I restriction-modification system that blocks horizontal gene transfer into *Staphylococcus aureus* and between *S. aureus* isolates of different lineages. *J Bacteriol*. 2006;188(15):5578–5585.
- Seeber S, Kessler C, Gotz F. Cloning, expression and characterization of the Sau3AI restriction and modification genes in *Staphylococcus carnosus* TM300. *Gene.* 1990;94(1):37–43.
- Xu SY, Corvaglia AR, Chan SH, Zheng Y, Linder P. A type IV modification-dependent restriction enzyme SauUSI from Staphylococcus aureus subsp. aureus USA300. Nucleic Acids Res. 2011;39(13):5597–5610.

(Continued)

Supplementary material

Table S1 Comparison of CC15-MRSA and CC15-MSSA

						MPROSI /9/	08-02,119	512,01,30,938	512,01,30,940	ST2,01,30,938 ST2,01,30,940 ST2,01,30,941
Biosample accession	SAMN06925302	SAMN06925301	SAMN06925303	SAMN06925304	SAMN00138234	SAMEA2664415	SAMEA2664415 SAMN04939716 SAMN04166246 SAMN04166494 SAMN04166543	SAMN04166246	SAMN04166494	SAMN04166543
Collection date	07-Nov-2013	04-Apr-2014	26-Oct-2014	26-Oct-2014						
Collection place	Saudi Arabia:	Saudi Arabia:	Saudi Arabia:	Saudi Arabia:						
	Riyadh	Riyadh	Riyadh	Riyadh						
Host	Homo sapiens	Homo sapiens	Camelus	Camelus						
			dromedarius	dromedarius						
Host disease	Sepsis	Wound infection								
Isolation source			Retail meat,	Retail meat,						
			neighborhood	neighborhood						
			meat shop	meat shop						
MLST	1535	1535	1535	1535	15	1535	582	15	15	15
Clonal complex	15	15	15	15	15	15	CC15/CC20	15	15	15
							chimera			
SCC element	SCCmecV /	SCCmecV /	SCCmecV /	SCCmecV /	None	SCCmecV /	None	None		
	SCCfus	SCCfus	SCCfus	SCCfus		SCCfus				
Paired end sequencing	2×5 I	2×51	2×51	2×51		2×100				
Average insert size	260	270	290	310		260				
Fragments sequenced	10461449	9646853	8433245	7667462		2852668				
Total number of bases	1067067798	983979006	066061098	782081124		570533600				
Estimated coverage	360	330	290	260		061				
WGS accession	SAMN06925302		SAMN06925303	SAMN06925304	AGTZ00000000.1		CP0156451	CP012972.1	CP012979.1	CP012978.1
	NHZU000000000	00000000XZHN	NHZW000000000	NHZX000000000						
Number of contigs	73 contigs	71 contigs	72 contigs	66 contigs						
capsular genotype	8	8	8	8	8	8	8	8	8	8
(assembly)										
agr type (assembly)	=	=	=	=	=	=	=	=	=	=
RIDOM spa type	t328, uneven	t328, uneven	t328, uneven	t328, uneven	t393	t084	t084	t385	t084	t084
(assembly)	coverage	coverage	coverage	coverage						
RIDOM spa profile	07-23-12-34-	07-23-12-34-	07-23-12-34-34	07-23-12-34-	07:23:12:34:12:12: 07:23:12:34:3	07:23:12:34:3	07:23:12:34:3	07-23-12-34-	07:23:12:34:3	07:23:12:34:3
	34-12-12-23-	34-12-12-23-	12-12-23-02-12-	34-12-12-23-	12:23:02:12:23	4:12:12:23:02:	4:12:12:23:02:	34-12-12-23-	4:12:12:23:02:	4:12:12:23:02:
	02-12-23-02-	02-12-23-02-	23-02-12-23	02-12-23-02-		12:23	12:23	12–23	12:23	12:23
	12–23	12–23		12–23						
RIDOM spa repeat	4	<u>+</u>	4	<u>4</u>	=	=	=	01	=	=
count										

Table SI Comparison of CCI5-MRSA and CCI5-MSSA

Isolate name	RUH-2	RUH-71	RUH-98	RUH-99	VCU006	MPROSI797	08-02,119	ST2,01,30,938	ST2,01,30,938 ST2,01,30,940 ST2,01,30,94	ST2,01,30,941
cna (assembly)	Missing	Missing	Missing							
sarT/sarU (assembly)	Present	Present	Present							
sasC	Truncated	Truncated	Truncated	Truncated	Intact	Truncated	Intact	Intact	Intact	Intact
tetK	Missing	Present	Missing	Missing	Missing	Missing	Missing	Missing	Missing	Missing
blaZ	Present	Present	Missing							
fusC	Present	Present	Present	Present	Missing	Present	Missing	Missing	Missing	Missing
mecA	Present	Present	Present	Present	Missing	Present	Missing	Missing	Missing	Missing
hlb	Truncated	Truncated	Truncated							
scn	Present	Present	Present							
chp	Present	Present	Present							
3 kb hlb insert	Present	Present	Present							
sau3Al	Present	Present	Present							
sauUSI	Missing	Missing	Missing							

Notes: Comparison genome properties of CC15-MRSA from this study with those of CC15-MSSA/MRSA from the NCBI GenBank.

Abbreviations: MRSA, methicillin-resistant Stophylococcus aureus; MSSA, methicillin susceptible S. aureus; MLST, multilocus sequence typing: WGS, whole genome shotgun.

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