Biochemical assessments of thyroid profile, serum 25-hydroxycholecalciferol and cluster of differentiation 5 expression levels among children with autism

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Tarek Desoky¹
Mohammed H Hassan²
Hanan M Fayed³
Hala M Sakhr⁴
¹Department of Neuropsychiatry, ²Department of Medical Biochemistry and Molecular Biology, ³Department of Clinical and Chemical Pathology, ⁴Department of Pediatrics, Qena Faculty of Medicine, Qena University Hospitals, South Valley University, Qena, Egypt

Background: The exact pathogenesis of autism is still unknown. Both thyroid hormones and 25(OH)D are important for brain development, in addition to CD5; all have immunomodulatory actions by which their dysregulation may have a potential role in autism pathogenesis.

Objectives: The objectives of this study were to assess the thyroid profile, serum 25(OH)D levels and CD5 expression levels among autistic patients and to find out the correlations between the measured biomarkers with each other on one side and with the disease severity on the other side.

Patients and methods: This cross-sectional case–control study has been conducted on 60 children with autism and 40 controls, recruited from Qena Governorate, Upper Egypt. Childhood Autism Rating Scale (CARS) score was used to assess the included patients. Biochemical assays of thyroid function in the form of free triiodothyronine (FT3), free tetraiodothyronine (FT4), thyroid-stimulating hormone (TSH) and 25(OH)D were done using commercially available enzyme-linked immunosorbent assay (ELISA) kits, while CD5 expression levels were measured using flow cytometry (FCM) analysis for all the included patients and controls.

Results: The overall measurement results show significant higher mean serum TSH levels, mean CD5 expression levels with significant lower mean serum 25(OH)D levels among autistic children when compared with the control group (p<0.05 for all). Significant negative correlations between CD5 with FT3, FT4 and 25(OH)D were observed. CARS score showed significant negative correlations with both FT3 and 25(OH)D, while it was positively correlated with CD5 in a significant manner (p<0.05 for all).

Conclusion: Elevated CD5 expression and decreased 25(OH)D stores could play a potential role in the pathogenesis of autism via their immune-modulator actions. High TSH serum levels among autistic children, although within the physiological range, reflect the presence of thyroid dysfunction among such children, which needs further assessment.

Keywords: thyroid hormones, 25(OH)D, CD5, autism, flow cytometry, ELISA

Introduction
Autism is a complicated syndrome that affects neurological development characterized by pervasive deficits in the social interaction, verbal and nonverbal communication impairment and stereotyped patterns of interests and activities.¹² The severity of autism can be assessed using Childhood Autism Rating Scale (CARS).³

The exact pathogenesis of autism is still unknown; however, genetic, neurologic, environmental and immunological factors may be involved.⁴
Vitamin D plays an important role in brain homeostasis and has immunomodulatory, neuroprotective and anti-inflammatory functions. Previous studies reported an association between vitamin D deficiency and neuropsychiatric diseases involving autism. Only 25(OH)D; calcidiol is currently used for the assessment of vitamin D saturation worldwide and reflects the total body stores of vitamin D. Serum 1,25-(OH)2D3 cannot be used as an indicator for vitamin D3 status as its level may decrease only in severe vitamin D3 deficiency. Both vitamin D3 and thyroid hormones belong to group-I hormones using steroid hormone receptors. Thyroid hormones play an essential role in early brain development and significantly affect the whole body activities and functions, and their decrease to a certain degree may cause multiorgan failure. Vitamin D deficiency may associate with autoimmune thyroid dysfunction. The CD5 is a transmembrane protein present on the surface of T cells and on many thymocytes and a subset of B cells surfaces. High CD5 levels are reported in many autoimmune diseases, providing evidence of CD5 role in the pathogenesis of autoimmunity.

Based on literature, the present novel study aimed to assess and investigate the status and the correlations of 25(OH)D3, thyroid hormones and CD5 in children with autism, all of which have immunomodulatory actions. Their disturbances may play a role in the occurrence of autoimmunity, supporting the autoimmune theory of autism. In addition, both 25(OH)D3 and thyroid hormones are important for brain development, and their disturbances may have a role in various neurodevelopmental diseases including autism.

Patients and methods
Study design
This cross-sectional case–control study has been conducted on 60 newly diagnosed autistic children (55 males and five females); their mean age was 7.03±2.34 years. They were selected from the outpatients’ psychiatric clinics of the Neuropsychiatric and Pediatric Departments of Qena University Hospital(s), South Valley University, Upper Egypt, after approval from the ethics committee of the Qena University. This was in addition to 40 apparently healthy, age-matched volunteers selected as controls (20 males and 20 females). The recruitment of patients and controls in this study was according to the guidelines laid down in the Declaration of Helsinki, and every child’s parents were informed about the aim of the study and written consent was given. The total duration of the study was one year, from May 1, 2016 to April 30, 2017.

Patients’ evaluation
All included patients were subjected to detailed history and thorough neuropsychiatric examination. Individual interviews with every included child and his/her parents for clinical assessment of autistic symptoms severity using CARS was conducted. CARS assessed behavior guided by 14 parameters by which autism affected them in addition to one parameter of general autism impression. A score from 1 to 4 for each parameter was assigned as follows: 1= no signs of autism (normal behavior for age) and 4= severe symptoms of autism (severe affection of behavior). The CARS maximum score of 60 was divided as follows: no autism (15–29.5), mild to moderate autism (30.5–37) and severe autism (37.5–60).

Laboratory workup
A total of 10 mL of peripheral blood venous samples were drawn from an antecubital vein, which were divided into two vacutainer tubes. A total of 2 mL was collected on an EDTA tube for flow cytometry (FCM) analysis of the CD5 expression levels using FACSCalibur FCM (BD Biosciences, San Jose, CA, USA). The remaining 8 mL of venous blood was evacuated into serum separator gel tubes, where the samples were allowed to clot for 30 minutes at 37°C before centrifugation for 15 minutes at 3,000 rpm. Separated sera were aliquoted into 1 mL cryotubes and stored at −20°C till the time of biochemical analysis. All samples were measured in a single assay to avoid repeated freeze–thaw cycles. Commercially available sandwich enzyme-linked immunosorbent assay (ELISA) kits were used for thyroid-stimulating hormone (TSH) and 25(OH)D biochemical analyses, and solid-phase competitive ELISA assay kits were used for assay of free triiodothyronine (FT3) and free tetraiodothyronine (FT4) using ELISA Multiskan EX Microplate Photometer (Stat Fax 2100; Thermo Scientific, Palm city, FL, USA). FT3, FT4 and TSH ELISA assay kits were supplied by Calbiotech Inc. (Spring Valley, CA, USA). The catalog numbers were F3106T for FT3, F4107T for FT4 and TS227T for TSH, while that for 25(OH)D was supplied by Chongqing Biosope Co., Ltd (Chongqing, People’s Republic of China). with the catalog number: BYEK1472. Reference values according to the used ELISA kits for the assessment of thyroid profile were 0.7–6.4 µIU/mL for TSH, 1.2–4.4 pg/mL for FT3 and 0.8–2 ng/dL for FT4, while for 25(OH)D reference values, deficiency was considered if its serum level was <20 ng/mL, insufficiency was considered if its serum level was ≥20 and <30 ng/mL and sufficiency was considered if its serum level was ≥30 ng/mL.
Biomarkers in autism

Figure 1 Flow cytometer dot plot analysis of CD5 expression level subsets.
Notes: (A) Forward scatter and side scatter of peripheral blood, R1 (gated lymphocytes). (B) CD5^+ lymphocytic expression.

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Notes: (A) Forward scatter and side scatter of peripheral blood, R1 (gated lymphocytes). (B) CD5^+ lymphocytic expression.

FCM analysis of CD5 expression
CD5 surface marker expression levels were measured using polyclonal anti-CD5 antibody conjugated to fluorescein isothiocyanate (FITC; Chongqing Biospes Co., Ltd, catalog number: YPA1579). Briefly, as manufacturer’s instructions, 100 µL of whole blood was stained with 10 µL of FITC-conjugated, lineage-specific anti-CD5 marker. After 15 min of incubation with the antibodies at room temperature in the dark, RBCs were lysed using FACS lysing solution and washed with phosphate-buffered saline (PBS). After proper setting, calibration and compensation, an isotype-matched control was used with each sample. The labeled lymphocytes were acquired via the properties of a forward scatter and side scatter dot plot histogram, which was used to define the lymphocyte population (R1) on FCM (Figure 1), and analysis was done using FACSCalibur FCM with CellQuest software version 5.2 (BD Biosciences).

Statistical analysis
The normality of data and homogeneity variances prior to further statistical analysis were tested using the Anderson–Darling test. Categorical variables were described by number and percent (n, %), whereas continuous variables were described by mean and standard error of mean (mean, SEM) or mean and standard deviation (mean, SD). Chi-square test and Fisher’s exact test were used to compare categorical variables, while Student’s t-test and analysis of variance (ANOVA) were used to compare continuous quantitative variables of parametric data. Spearman’s rank correlation coefficient was used to explore the relationship between quantitative variables. A two-tailed test was considered significant when \( p < 0.05 \). All analyses were performed using the IBM SPSS version 20.0 software (IBM Corporation, Armonk, NY, USA).

Table 1 Demographic data of the studied groups

<table>
<thead>
<tr>
<th>Variables</th>
<th>Patients group</th>
<th>Control group</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Patients (total), (N=60)</td>
<td>Patients with mild to moderate autism (n=30)</td>
<td>Patients with severe autism (n=30)</td>
</tr>
<tr>
<td>Age, mean ± SD</td>
<td>7.03±2.34</td>
<td>6.8±2.08</td>
<td>7.27±2.6</td>
</tr>
<tr>
<td>Sex, n (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Males</td>
<td>55 (92.6)</td>
<td>28 (93.3)</td>
<td>27 (90.0)</td>
</tr>
<tr>
<td>Females</td>
<td>5 (8.3)</td>
<td>2 (6.7)</td>
<td>3 (10.0)</td>
</tr>
<tr>
<td>CARS score, mean ± SD</td>
<td>–</td>
<td>32.67±1.99</td>
<td>46.8±4.23</td>
</tr>
</tbody>
</table>

Note: *Statistically significant difference (p<0.05).
Abbreviations: SD, standard deviation; CARS, Childhood Autism Rating Scale.
Table 2 Mean ± SEM of the serum levels of FT3, FT4, TSH, 25(OH)D and CD5 in children with autism compared with the control group

<table>
<thead>
<tr>
<th>Variables</th>
<th>Patients</th>
<th>Control</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>FT3 (pg/dL)</td>
<td>3.53±0.54</td>
<td>3.46±0.36</td>
<td>0.064</td>
</tr>
<tr>
<td>FT4 (ng/dL)</td>
<td>1.45±0.13</td>
<td>1.38±0.09</td>
<td>0.580</td>
</tr>
<tr>
<td>TSH (µIU/mL)</td>
<td>2.25±1.05</td>
<td>1.45±0.96</td>
<td>&lt;0.05*</td>
</tr>
<tr>
<td>25(OH)D (ng/mL)</td>
<td>18.63±10.8</td>
<td>45.9±8.85</td>
<td>&lt;0.001*</td>
</tr>
<tr>
<td>CD5 (%)</td>
<td>7±1.55</td>
<td>2.85±0.95</td>
<td>&lt;0.01*</td>
</tr>
</tbody>
</table>

Note: *Statistically significant difference (p<0.05).

Abbreviations: SEM, standard error of mean; FT3, free triiodothyronine; FT4, free tetraiodothyronine; TSH, thyroid-stimulating hormone.

Discussion

Autism spectrum disorder (ASD) is a severe disorder that affects the neurological development, characterized by disturbances in the social interaction and communication, in addition to stereotyped repetitive behaviors. Its prevalence differs by the region, but a higher prevalence of up to 1:88 child has been reported. Because the etiology of autism is a highly contemporary field of research in neuroscience and psychiatry and can be considered as a challenging issue as there is no direct mechanism that can simply explain the pathogenesis of autism, the present study investigated a group of biomarkers with different degrees of autism severity and searched the possible resulting correlations that may provide new insights on autism diagnosis and therapy.

There is increasing knowledge that normal thyroid function is required for normal neurological development and thyroid dysfunction associated with many neurodevelopmental disorders including ASD. The findings of the present study revealed significantly higher TSH serum levels with nonsignificant differences in the serum levels of FT3 and FT4 among children with autism when compared with the controls with significantly higher serum levels among children with severe degree of autism versus those having mild to moderate autism, suggesting the presence of subclinical hypothyroidism in autistic children, although the serum levels of TSH still fall within the normal range, which is wide. In addition, there was a significant negative correlation between FT3 and CARS score. These findings were in agreement with the study of Khan, who reported a high rate of hypothyroidism in children with autism. Nir et al reported larger diurnal variation in TSH serum levels in comparison with controls explaining the possible thyroid dysfunction in ASD in relation to hypothalamic–pituitary axis. Although the

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![Figure 2](https://www.dovepress.com) Comparisons between the mean levels of FT3, FT4, TSH, CD5 (A) and 25(OH)D (B) in patients with autism versus the control group.

Abbreviations: FT3, free triiodothyronine; FT4, free tetraiodothyronine; TSH, thyroid-stimulating hormone.
Table 3  Mean ± SEM of the serum levels of FT3, FT4, TSH, 25(OH)D and CD5 among children with autism according to CARS degree versus the control group

<table>
<thead>
<tr>
<th>Variables</th>
<th>Children with mild to moderate degree of autism</th>
<th>Children with severe degree of autism</th>
<th>Control group (n=40)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>FT3 (pg/dL)</td>
<td>3.67±0.53</td>
<td>3.39±0.52</td>
<td>3.46±0.36</td>
<td>0.060</td>
</tr>
<tr>
<td>FT4 (ng/dL)</td>
<td>1.47±0.13</td>
<td>1.42±0.12</td>
<td>1.38±0.09</td>
<td>0.080</td>
</tr>
<tr>
<td>TSH (µIU/mL)</td>
<td>2.11±1.03</td>
<td>2.38±1.08</td>
<td>1.45±0.96</td>
<td>&lt;0.05*</td>
</tr>
<tr>
<td>25(OH)D3 (ng/mL)</td>
<td>24.08±10.62</td>
<td>13.19±7.94</td>
<td>45.9±8.85</td>
<td>&lt;0.001*</td>
</tr>
<tr>
<td>CD5 (%)</td>
<td>5.8±0.76</td>
<td>8.2±1.16</td>
<td>2.85±0.95</td>
<td>&lt;0.001*</td>
</tr>
</tbody>
</table>

Note: *Statistically significant difference (p<0.05).

Abbreviations: SEM, standard error of mean; FT3, free triiodothyronine; FT4, free tetraiodothyronine; TSH, thyroid-stimulating hormone; CARS, Childhood Autism Rating Scale.

The findings of the present study revealed significant lower serum levels of 25(OH)D among autistic children, especially those having severe degree when compared with the control group with higher frequency of 25(OH)D deficiency in severe autism than mild to moderate autism in which higher frequency of 25(OH)D insufficiency was noticed. In addition, there was significant negative correlation between 25(OH)D and CARS score reflecting an inverse association with disease severity. These findings were in agreement with several studies, explaining that the immunoregulatory function of 25(OH)D has been lost by the presence of lower vitamin D stores, contributing to the occurrence and severity of autism through the raised autoimmunity, and this was confirmed in one of the previously mentioned studies by the improvement in autism symptoms following vitamin D supplementation to such patients. Although MacKawy et al, reported that serum levels of 25(OH)D was positively correlated with T3, while, negatively correlated with TSH among hypothyroid patients, in a statistically significant manner, the present study didn’t show any significant correlations between serum levels of 25(OH)D with FT3 , FT4 or TSH, which could be explained by the mild degree of thyroid dysfunction noticed among the included autistic children.

Table 4  Classification of the included children with autism according to the status of 25(OH)D

<table>
<thead>
<tr>
<th>25(OH)D status</th>
<th>Children with mild to moderate autism (n=30)</th>
<th>Children with severe autism (n=30)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sufficient (&gt;30 ng/mL), n (%)</td>
<td>6 (13.6)</td>
<td>4 (9.1)</td>
<td>&lt;0.05*</td>
</tr>
<tr>
<td>Insufficient (20–30 ng/mL), n (%)</td>
<td>6 (13.6)</td>
<td>0 (0.0)</td>
<td></td>
</tr>
<tr>
<td>Deficient (&lt;20 ng/mL), n (%)</td>
<td>18 (40.9)</td>
<td>26 (59.1)</td>
<td></td>
</tr>
</tbody>
</table>

Note: *Statistically significant difference (p<0.05).

Table 5  Correlations of the serum levels of FT3, FT4, TSH, 25(OH)D and CD5 among children with autism

<table>
<thead>
<tr>
<th>Variables</th>
<th>FT3 (pg/dL)</th>
<th>FT4 (ng/dL)</th>
<th>TSH (µIU/mL)</th>
<th>25(OH)D (ng/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>25(OH)D</td>
<td>r</td>
<td>-0.080</td>
<td>-0.200</td>
<td>0.109</td>
</tr>
<tr>
<td>(ng/mL)</td>
<td>p-value</td>
<td>0.544</td>
<td>0.126</td>
<td>0.407</td>
</tr>
<tr>
<td>CD5 (%)</td>
<td>r</td>
<td>-0.371</td>
<td>-0.297</td>
<td>0.322</td>
</tr>
<tr>
<td>p-value</td>
<td>&lt;0.01*</td>
<td>&lt;0.05*</td>
<td>&lt;0.05*</td>
<td>&lt;0.05*</td>
</tr>
</tbody>
</table>

Note: *Statistically significant difference (p<0.05).

Abbreviations: FT3, free triiodothyronine; FT4, free tetraiodothyronine; TSH, thyroid-stimulating hormone.
Table 6 Correlations of the serum levels of FT3, FT4, TSH, 25(OH)D and CD5 among autistic children with CARS score

<table>
<thead>
<tr>
<th>Variables</th>
<th>CARS score</th>
<th>r</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>FT3 (pg/dL)</td>
<td>−0.329</td>
<td>&lt;0.05*</td>
<td></td>
</tr>
<tr>
<td>FT4 (ng/dL)</td>
<td>−0.195</td>
<td>0.135</td>
<td></td>
</tr>
<tr>
<td>TSH (µIU/mL)</td>
<td>0.183</td>
<td>0.162</td>
<td></td>
</tr>
<tr>
<td>25(OH)D (ng/mL)</td>
<td>−0.582</td>
<td>&lt;0.001*</td>
<td></td>
</tr>
<tr>
<td>CD5 (%)</td>
<td>0.808</td>
<td>&lt;0.001*</td>
<td></td>
</tr>
</tbody>
</table>

Note: *Statistically significant difference (p<0.05).

Abbreviations: FT3, free triiodothyronine; FT4, free tetraiodothyronine; TSH, thyroid-stimulating hormone; CARS, Childhood Autism Rating Scale.

Our study showed significantly higher expression levels of CD5 among autistic patients, especially those with severe degree, in comparison with those in the control group with significant positive correlation with CARS score and hence severe degree of autism. Only one study could be traced in the literature investigating CD5 plasma levels and their associations with CARS score in autism, which was in agreement with our findings, strengthening the immune dysregulation hypothesis and the raised autoimmunity in autism pathogenesis.27 Our study differs from theirs in the method of CD5 assay, in which Halepoto et al27 used ELISA in the assessment of CD5 expression levels, while we used FCM. Explanation for such findings needs further molecular research.

To the best of our knowledge, the present study is the first original research investigating the correlations between thyroid hormones, 25(OH)D and CD5 in autism, and we report the following correlations that may be promising in understanding the pathophysiology of autism. There were significant negative correlations of CD5 with FT3, FT4 and 25(OH)D, which indicate that the higher the CD5, the lower the FT3, FT4 and 25(OH)D and the higher the CARS score and hence the increased degree of autism severity. These findings may open new roads regarding the therapeutic approach in autism and future studies regarding the combined use of vitamin D supplementations with immunomodulatory drugs specifically act on CD5 should be researched as a new guideline in autism therapy.

Conclusion
The findings of the present study support the autoimmunity theory in autism pathogenesis as well as all the studied biomarkers (thyroid hormones, 25(OH)D and CD5) have direct or indirect immunoregulatory role in the human body and are disturbed in autistic patients.

Study limitations
The relatively small study sample and the cross sectional nature of the study.

Disclosure
The authors report no conflicts of interest in this work.

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