Predictors of high on-aspirin platelet reactivity in elderly patients with coronary artery disease

Objectives: Previous studies have illustrated the link between high on-aspirin platelet reactivity (HAPR) with increasing thrombotic risks. The aim of our study was to investigate relative risk factors of HAPR in elderly patients with coronary artery disease.

Methods: Elderly, hospitalized coronary artery disease patients on regular aspirin treatment were enrolled from January 2014 to September 2016. Medical records of each patient were collected, including demographic information, cardiovascular risk factors, concomitant drugs and routine biological parameters. Arachidonic acid (AA, 0.5 mg/mL) and adenosine diphosphate (ADP, 5 µmol/L) induced platelet aggregation were measured via light transmission assay (LTA) to evaluate antplatelet responses, referred as LTA–AA and LTA–ADP.

Results: A total of 275 elderly patients were included, with mean age of 77.2±8.1 years, and males accounted for 81.8%. HAPR was defined as LTA–AA in the upper quartile of the enrolled population. HAPR patients tended to have lower renal function (P=0.052). Higher serum uric acid (SUA) level, as well as lower platelet count, hemoglobin and hematocrit were observed in HAPR patients, with a higher proportion of diuretics use (P<0.05). Multivariate analysis revealed that SUA (OR: 1.004, 95% CI: 1.000–1.007, P=0.048), platelet count (OR: 0.994, 95% CI: 0.989–1.000, P=0.045), hematocrit (OR: 0.921, 95% CI: 0.864–0.981, P=0.011) and concomitant P2Y12 receptor inhibitors use (OR: 1.965, 95% CI: 1.075–3.592, P=0.028) were correlated with HAPR. Spearman’s correlation analysis demonstrated an inverse association of LTA–AA with hematocrit (r=0.234, P<0.001), hemoglobin (r=0.209, P<0.001) and estimated glomerular filtration rate (r=−0.132, P=0.031).

Conclusion: SUA, platelet count, hematocrit and P2Y12 receptor inhibitors use were independently correlated with HAPR. These parameters might provide novel therapeutic targets for optimizing antplatelet therapy.

Keywords: aspirin, platelet reactivity, elderly, risk factors,coronary artery disease

Introduction

Thrombosis, considered as pathological hemostasis, is one of the severe complications for coronary artery disease (CAD), which poses a great threat to public health. Platelet activation plays crucial roles in the process of thrombosis. For one thing, during the early stage of thrombosis, vascular injury initiates the recruitment of platelets to the damaged sites, then the activated platelets adhere to the vessel wall and release platelet agonists, which promote additional platelet activation and aggregation. For another, platelets exert procoagulant effects and release mediators that support the recruitment of leukocytes.\(^1,2\) In addition, recent studies revealed that platelet-derived miRNAs, miRNAs and exosomes are also involved in thrombus formation.\(^3\)

Aspirin, exerting inhibitory effects on platelet activation by acetyling the serine 529 residue of cyclooxygenase (COX), reduces the risk of myocardial infarction
and stroke, and is thus widely used in the prevention and treatment of cardiovascular diseases.\(^4,5\) However, individual variations in the platelet response to aspirin limits its use in some patients, including patients who experience ischemic events despite regular aspirin therapy.\(^6\)–\(^8\)

High on-aspirin platelet reactivity (HAPR) is defined as insufficient inhibition of platelet activation in patients on regular aspirin treatment. As a consequence, aspirin fails to prevent thrombotic complications in a significant proportion of patients, in the range of 7%–20% of adults.\(^9,10\) Several mechanisms of HAPR have been put forward, such as the decreased biological availability, drug–drug interactions, inadequate doses or poor compliance, reduced absorption due to advanced age or concurrent proton pump inhibitors (PPIs), non-platelet thromboxane A2 (TXA2) synthesis, accelerated platelet turnover, polymorphism of COX/glycoprotein IIb/IIIa (GPIIb/IIIa) receptors/collagen receptors and others.\(^7,11,12\)

The link between HAPR with increasing risk of ischemic events has been demonstrated in previous studies.\(^13,14\) A number of platelet function tests (PFTs) have been developed to distinguish HAPR patients, intending to reduce thrombotic risks as well as to guide individual treatment.\(^15,16\) Among which light transmission assay (LTA) was regarded as the traditional “gold standard” of PFTs. Using different platelet agonists, such as arachidonic acid (AA), adenosine diphosphate (ADP), collagen and epinephrine, platelet inhibition due to aspirin or thienopyridines could be distinguished. However, controversies still existed in optimal cut-off for aspirin response, which should be capable of predicting both thrombotic events and bleeding complications.\(^17\)

Older age was associated with an increasing incidence of high platelet reactivity (HPR) in patients receiving antiplatelet therapy, which might partially elucidate the relatively higher thrombosis risks and poorer prognosis in this particular population.\(^18,19\) In this study, we aimed to investigate potential risk factors for HAPR in elderly CAD patients (aged >60 years), thus identifying “at-risk” patients and promoting the optimization of antiplatelet therapy.

### Methods and materials

#### Study design

Elderly patients on regular aspirin treatment (100 mg/d), hospitalized in the Department of Geriatrics of Peking University First Hospital were enrolled from January 2014 to September 2016. Inclusion criteria were as follows:

1. Age: >60 years.

2. Presence of at least one of the following: stable angina pectoris; acute coronary syndrome (ACS); previous percutaneous coronary intervention (PCI); coronary artery bypass graft; confirmed coronary atherosclerotic plaques via computed tomography angiography or coronary angiography.

3. Platelet count in the range between 75×10\(^9\)/L and 600×10\(^9\)/L.

Patients were excluded when there existed contraindications for aspirin, concomitant with non-steroidal anti-inflammatory drugs, GPIIb/IIIa receptor inhibitors, Vitamin K antagonists or novel anticoagulants; patients with severe renal or liver dysfunction, peptic ulcer or history of gastrointestinal hemorrhage were excluded.

This study complied with the Declaration of Helsinki, and the protocol was approved by the Ethical Review Committee of Peking University First Hospital. Written informed consent was obtained from all the enrolled participants.

#### Light transmission assay

All patients were on aspirin 100 mg/day treatment for at least 1 week before evaluating platelet function by LTA. LTA using AA, 0.5 mg/mL induced platelet aggregation (LTA–AA) was measured in order to evaluate aspirin responses as described.\(^20,21\) Peripheral blood samples were drawn in a sodium citrate tube (1:9), and all measurements were conducted within 2 hours. Platelet-rich plasma (PRP) and platelet-poor plasma (PPP), used as reference, were prepared through centrifugation at separate speeds (200×g 10 min for PRP; 2,000×g 10 min for PPP). AA-induced platelet aggregation was performed using an LBY-NJ4 platelet aggregometer (PRECIL, Beijing, China). The percentage of platelet aggregation was defined as the maximal light transmittance after AA addition. Besides this, ADP, 5 μmol/L induced platelet aggregation (LTA–ADP) was also measured to exclude patients with poor response to thienopyridines.\(^22,23\) Platelet aggregation results were further normalized based on platelet counts of each blood sample to mitigate bias.

#### Hospitalization records collection

For each patient, detailed medical records during the hospitalization were obtained from the electronic system, including age, gender, cardiovascular disease status, cardiovascular risk factors, combined drugs, co-morbidities, routine biological parameters such as routine blood test, glucose, coagulation function, renal or liver function parameters, uric acid, lipids and others (Table 1).
Table 1  Clinical features in patients with different platelet reactivity status

<table>
<thead>
<tr>
<th>Clinical features</th>
<th>LAPR (n=69)</th>
<th>MAPR (n=138)</th>
<th>HAPR (n=68)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>76.0±8.6</td>
<td>77.2±8.3</td>
<td>78.4±7.2</td>
<td>0.266</td>
</tr>
<tr>
<td>Female gender, n (%)</td>
<td>17 (24.6)</td>
<td>24 (17.4)</td>
<td>9 (13.2)</td>
<td>0.217</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>24.9±3.7</td>
<td>24.3±3.2</td>
<td>24.5±3.0</td>
<td>0.596</td>
</tr>
<tr>
<td>Previous PCI, n (%)</td>
<td>38 (55.1)</td>
<td>80 (58.0)</td>
<td>38 (55.9)</td>
<td>0.912</td>
</tr>
<tr>
<td>Hypertension, n (%)</td>
<td>51 (73.9)</td>
<td>102 (73.9)</td>
<td>53 (38.4)</td>
<td>0.798</td>
</tr>
<tr>
<td>Diabetes, n (%)</td>
<td>28 (40.6)</td>
<td>59 (42.8)</td>
<td>33 (48.5)</td>
<td>0.617</td>
</tr>
<tr>
<td>Ischemic cerebrovascular disease, n (%)</td>
<td>14 (20.3)</td>
<td>42 (30.4)</td>
<td>24 (35.3)</td>
<td>0.137</td>
</tr>
<tr>
<td>Hyperlipidemia, n (%)</td>
<td>57 (82.6)</td>
<td>115 (83.3)</td>
<td>59 (86.8)</td>
<td>0.767</td>
</tr>
<tr>
<td>Peripheral artery disease, n (%)</td>
<td>57 (82.6)</td>
<td>111 (80.4)</td>
<td>54 (79.4)</td>
<td>0.887</td>
</tr>
<tr>
<td>Current smoking, n (%)</td>
<td>13 (18.8)</td>
<td>16 (11.6)</td>
<td>9 (13.2)</td>
<td>0.358</td>
</tr>
<tr>
<td>LVEF (%)</td>
<td>60.0 (32.0–70.0)</td>
<td>61.0 (40.0–80.0)</td>
<td>61.0 (43.0–77.0)</td>
<td>0.861</td>
</tr>
<tr>
<td>eGFR (mL/min)</td>
<td>66.2±16.9</td>
<td>65.0±15.2</td>
<td>59.8±18.2</td>
<td>0.052</td>
</tr>
<tr>
<td>25-(OH)-D (pg/mL)</td>
<td>40.5±7.0</td>
<td>40.6±16.2</td>
<td>43.4±16.6</td>
<td>0.410</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>6.0 (5.4–9.0)</td>
<td>6.0 (5.2–9.8)</td>
<td>6.1 (5.2–8.4)</td>
<td>0.478</td>
</tr>
<tr>
<td>Hcy (µmol/L)</td>
<td>14.7±5.7</td>
<td>14.2±6.1</td>
<td>13.2±4.3</td>
<td>0.185</td>
</tr>
<tr>
<td>hsCRP (mg/L)</td>
<td>1.9 (0.0–16.1)</td>
<td>2.8 (0.0–31.5)</td>
<td>2.5 (0.0–19.7)</td>
<td>0.951</td>
</tr>
<tr>
<td>TG (mmol/L)</td>
<td>1.2 (0.4–3.5)</td>
<td>1.4 (0.4–5.6)</td>
<td>1.2 (0.4–4.8)</td>
<td>0.253</td>
</tr>
<tr>
<td>TCHO (mmol/L)</td>
<td>3.4 (2.4–5.1)</td>
<td>3.4 (1.7–6.4)</td>
<td>3.4 (2.0–5.9)</td>
<td>0.735</td>
</tr>
<tr>
<td>HDL-C (mmol/L)</td>
<td>1.0 (0.7–1.7)</td>
<td>1.0 (0.4–4.8)</td>
<td>1.0 (0.5–2.1)</td>
<td>0.243</td>
</tr>
<tr>
<td>LDL-C (mmol/L)</td>
<td>1.9 (0.3–3.0)</td>
<td>1.8 (0.9–5.2)</td>
<td>1.9 (1.1–4.3)</td>
<td>0.418</td>
</tr>
<tr>
<td>SUA (µmol/L)</td>
<td>344±69.5</td>
<td>337±85.2</td>
<td>368±89.7</td>
<td>0.010</td>
</tr>
<tr>
<td>PLT x10^9/L</td>
<td>202.9±93.6</td>
<td>186.6±53.2</td>
<td>172±51.2</td>
<td>0.025</td>
</tr>
<tr>
<td>MPV (fl)</td>
<td>8.5 (6.9–13.8)</td>
<td>8.4 (6.2–12.4)</td>
<td>8.6 (7.0–11.3)</td>
<td>0.651</td>
</tr>
<tr>
<td>Hb (g/L)</td>
<td>134.6±16.6</td>
<td>132±16.1</td>
<td>126±17.4</td>
<td>0.033</td>
</tr>
<tr>
<td>Hct (%)</td>
<td>39±4.6</td>
<td>38±4.6</td>
<td>36±4.5</td>
<td>0.005</td>
</tr>
<tr>
<td>PT (s)</td>
<td>10±1.5</td>
<td>11±2.9</td>
<td>11.5±3.0</td>
<td>0.981</td>
</tr>
<tr>
<td>APTT (s)</td>
<td>32.7±5.6</td>
<td>31.9±4.9</td>
<td>32.3±4.7</td>
<td>0.804</td>
</tr>
<tr>
<td>FIB-c (g/L)</td>
<td>2.9 (1.8–5.5)</td>
<td>2.8 (1.7–4.4)</td>
<td>2.9 (1.3–6.1)</td>
<td>0.552</td>
</tr>
<tr>
<td>FDP (mg/L)</td>
<td>1.3 (0.0–7.4)</td>
<td>1.5 (0.0–31.4)</td>
<td>1.8 (0.0–11.3)</td>
<td>0.112</td>
</tr>
<tr>
<td>Thienopyridines, n (%)</td>
<td>29 (42.0)</td>
<td>69 (50.0)</td>
<td>41 (60.3)</td>
<td>0.099</td>
</tr>
<tr>
<td>Statins, n (%)</td>
<td>64 (92.7)</td>
<td>130 (94.2)</td>
<td>65 (95.6)</td>
<td>0.778</td>
</tr>
<tr>
<td>ACEI/ARB, n (%)</td>
<td>29 (18.8)</td>
<td>60 (20.3)</td>
<td>35 (20.6)</td>
<td>0.442</td>
</tr>
<tr>
<td>β-blockers, n (%)</td>
<td>50 (72.4)</td>
<td>94 (68.1)</td>
<td>49 (72.1)</td>
<td>0.753</td>
</tr>
<tr>
<td>CCB, n (%)</td>
<td>27 (39.1)</td>
<td>53 (38.4)</td>
<td>29 (42.6)</td>
<td>0.839</td>
</tr>
<tr>
<td>Diuretics, n (%)</td>
<td>8 (11.6)</td>
<td>18 (13.0)</td>
<td>17 (25.0)</td>
<td>0.048</td>
</tr>
<tr>
<td>Nitrates, n (%)</td>
<td>25 (36.2)</td>
<td>55 (39.9)</td>
<td>26 (38.2)</td>
<td>0.878</td>
</tr>
<tr>
<td>Hypoglycemics, n (%)</td>
<td>19 (27.5)</td>
<td>44 (31.9)</td>
<td>23 (33.8)</td>
<td>0.710</td>
</tr>
<tr>
<td>PPI, n (%)</td>
<td>21 (30.4)</td>
<td>43 (31.2)</td>
<td>24 (35.3)</td>
<td>0.796</td>
</tr>
</tbody>
</table>

Notes: Values are mean ± SD or median (range) unless stated otherwise. *P<0.05, **P<0.01.
Abbreviations: ACEI, angiotensin-converting enzyme inhibitor; APTT, activated prothromboplastin time; ARB, angiotensin receptor antagonist; BMI, body mass index; CCB, calcium channel blockers; eGFR, estimated glomerular filtration rate; FDP, fibrin degradation product; Fb-c, Fibrinogen c; HAPR, high on-aspirin platelet reactivity; Hb, hemoglobin; HbA1c, Hemoglobin A1c; Hct, hematocrit; Hcy, homocysteine; hDl-C, high density lipoprotein cholesterol; hsCRP, hypersensitive C reactive protein; LAPR, low on-aspirin platelet reactivity; LDL-C, low density lipoprotein cholesterol; LVEF, left ventricular ejection fraction; MAPR, moderate on-aspirin platelet reactivity; MPV, mean platelet volume; PCI, percutaneous coronary intervention; PLT, platelet count; PPI, proton pump inhibitors; PT, prothrombin time; SUA, serum uric acid; TCHO, total cholesterol; TG, triglycerides.

Statistical analysis
Continuous variables were expressed as mean ± SD or median (range), while categorical variables were described as frequency and percentage. Conformity to normal distribution was evaluated for continuous variables using both Kolmogorov–Smirnov and Shapiro–Wilks tests. One-way analysis of variance test or nonparametric Kruskal–Wallis test was used to make comparisons for continuous variables, while chi-square or Fisher exact test was applied for categorical variables. Chi-square and linear trend test was performed to make comparison for age composition in patients with different platelet reactivity status.

Receiver operating characteristic (ROC) curve and multivariate logistic regression analysis was performed to investigate risk factors for HAPR, and Spearman’s correlation test was used to identify the factors correlated with LTA–AA. A 2-tailed P-value <0.05 was considered as statistically significant for all analysis executed. Statistical analysis was carried out using SPSS version 14.0 software (SPSS Inc., Chicago, IL, USA).

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Results

Enrollment
A total of 289 elderly CAD patients on regular aspirin treatment were enrolled, but 14 patients with high on-clopidogrel platelet reactivity (5 μmol/L ADP-induced platelet aggregation more than 50%) were excluded to avoid bias, resulting in a total of 275 elderly CAD patients with the mean age of 77.2±8.1 years. Males accounted for 81.8% (225 of 275), and 156 of the 275 CAD patients (56.7%) had received PCI previously. The enrollment flowchart is shown in Figure S1.

Distribution of platelet aggregation
Light transmission assay of AA-induced platelet aggregation (LTA–AA) was measured to evaluate aspirin response, with the quartiles 9.54%, 11.63% and 13.93% of residual platelet aggregation respectively. HAPR was defined as LTA–AA in the upper quartile, that is, more than 13.93% residual platelet aggregation (n=68). Meanwhile, low on-aspirin platelet reactivity (LAPR) was defined as LTA–AA in the lower quartile of the enrolled population (≤9.54%, n=69). The remaining 138 patients in the middle 2 quartiles were assigned into moderate on-aspirin platelet reactivity (MAPR) group.

As shown in Figure 1A, age composition varied among LAPR, MAPR and HAPR patients, and the proportion of senile patients (aged ≥80 years) increased gradually from LAPR group to HAPR group (P for linear trend=0.046). Distribution of LTA–AA and LTA–ADP in patients receiving dual antiplatelet therapy (DAPT) is shown in Figure 1B.

Clinical features of HAPR
Patients were divided into 3 groups, HAPR, MAPR and LAPR, according to quartiles of LTA–AA. Comparisons were made in terms of age, gender composition, cardiovascular risk factors, co-morbidities, routine biological parameters and concomitant drugs.

As shown in Table 1, HAPR patients had a tendency toward reduced renal function (P=0.052). Higher serum uric acid (SUA), as well as lower platelet count, hematocrit and hemoglobin were observed in HAPR patients (P<0.05). In terms of combined drugs, diuretics use was more frequently prescribed in HAPR patients (P<0.05).

The cut-off for hyperuricemia in the elderly was defined as the SUA concentration >7 mg/dL (416.5 mmol/L).24 Patients who met that criteria (n=47) for hyperuricemia were more common in HAPR patients, when compared with non-HAPR patients (14.5%, 12.3%, 29.4% for LAPR, MAPR and HAPR, P=0.023).

Multivariate regression analysis
Predictive value for HAPR of each of the variables listed in Table 1 with a P-value <0.10 were evaluated using ROC curve, including SUA, platelet count, estimated glomerular filtration rate (eGFR), hemoglobin and hematocrit. Areas under the ROC curve are exhibited in Table S1. Furthermore, a combination of these variables had an improved predictive value when compared with each variable alone (area under the curve [AUC]: 0.661, 95% CI: 0.581–0.740, P<0.001).

To investigate related factors for HAPR, multivariate regression analysis was performed. Previously reported

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**Figure 1** Light transmission assay.

**Notes:** (A) Patients were divided into 3 groups according to the quartile of LTA–AA, and age composition varied among LAPR, MAPR and HAPR patients. The percentage of senile patients (aged more than or equal to 80 years) increased gradually, from LAPR group to HAPR group (P for linear trend=0.046). (B) The distribution of LTA–AA and LTA–ADP in patients receiving dual antiplatelet therapy. The vertical dotted line indicates the cut-off of HAPR (13.93%), while the horizontal line marked the cut-off for poor clopidogrel response (50%). HAPR (LTA–AA >13.93%, n=68); LAPR (LTA–AA ≤9.54%, n=69); MAPR (9.54% < LTA–AA ≤13.93%, n=138).

**Abbreviations:** ADP, adenosine diphosphate; HAPR, high on-aspirin platelet reactivity; LTA–AA, light transmission assay-arachidonic acid; LAPR, low on-aspirin platelet reactivity; MAPR, moderate on-aspirin platelet reactivity.
Predictors for HAPR in the elderly

Variables such as age, gender, type 2 diabetes mellitus, current smoking and serum lipids were considered. Variables listed in Table 1 with a *P*-value <0.10 were also selected, including eGFR, SUA, platelet count, hemoglobin, hematocrit, concomitant P2Y12 receptor inhibitors and diuretics use. Table 2 lists the variables included in the equation at the last step, and all *P*-values have been adjusted for age and gender. It was revealed that SUA (OR: 1.004, 95% CI: 1.000–1.007, *P*=0.048), platelet count (OR: 0.994, 95% CI: 0.989–1.000, *P*=0.045), hematocrit (OR: 0.921, 95% CI: 0.864–0.981, *P*=0.011) and concomitant P2Y12 receptor inhibitors use (OR: 1.965, 95% CI: 1.075–3.592, *P*=0.028) were independently correlated with HAPR.

Spearman’s correlation analysis was performed to evaluate the correlation between LTA–AA and these variables with a *P*-value <0.10. As shown in Figure 2, hemoglobin (*r*=-0.209, *P*<0.001), hematocrit (*r*=-0.234, *P*<0.001) and eGFR (*r*=-0.132, *P*=0.031) were negatively associated with LTA–AA.

**Discussion**

Advanced age is associated with higher risk of thrombotic complications, with increased mortality and worse prognosis, which might be partly due to the increasing occurrence of HPR in this population. In our study, the percentage of senile patients gradually increased from LAPR to HAPR group. The relatively higher proportion of senile patients in HAPR group might be due to declined absorption of aspirin, drug–drug interactions and age-related hemodynamic changes in the elderly. As the link between HAPR with

**Table 2 Risk factors associated with HAPR**

<table>
<thead>
<tr>
<th>Variables</th>
<th>B</th>
<th>SE</th>
<th>Wald</th>
<th><em>P</em>-value</th>
<th>OR</th>
<th>95% Lower</th>
<th>95% Upper</th>
</tr>
</thead>
<tbody>
<tr>
<td>SUA</td>
<td>0.00</td>
<td>0.002</td>
<td>3.920</td>
<td>0.048*</td>
<td>1.004</td>
<td>1.000</td>
<td>1.007</td>
</tr>
<tr>
<td>PLT</td>
<td>-0.01</td>
<td>0.003</td>
<td>4.011</td>
<td>0.045*</td>
<td>0.994</td>
<td>0.989</td>
<td>1.000</td>
</tr>
<tr>
<td>Hct</td>
<td>-0.08</td>
<td>0.032</td>
<td>6.515</td>
<td>0.011*</td>
<td>0.911</td>
<td>0.864</td>
<td>0.981</td>
</tr>
<tr>
<td>Diuretics</td>
<td>0.64</td>
<td>0.379</td>
<td>2.867</td>
<td>0.090</td>
<td>1.900</td>
<td>0.904</td>
<td>3.994</td>
</tr>
<tr>
<td>P2Y12 receptor inhibitors</td>
<td>0.68</td>
<td>0.308</td>
<td>4.820</td>
<td>0.028*</td>
<td>1.965</td>
<td>1.075</td>
<td>3.592</td>
</tr>
<tr>
<td>Constant</td>
<td>1.24</td>
<td>1.478</td>
<td>0.703</td>
<td>0.402</td>
<td>3.453</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Note:* *P*<0.05.

**Abbreviations:** HAPR, high on-aspirin platelet reactivity; Hct, hematocrit; OR, odds ratio; PLT, platelet count; SE, standard error; SUA, serum uric acid.

**Figure 2** Spearman’s correlation analysis.

Notes: (A) Hematocrit (*r*=-0.234, *P*<0.001) was negatively associated with LTA–AA. (B) Hemoglobin (*r*=-0.209, *P*<0.001) was inversely correlated with LTA–AA. (C) eGFR (*r*=-0.132, *P*=0.031) was inversely correlated with LTA–AA.

**Abbreviations:** eGFR, estimated glomerular filtration rate; Hb, hemoglobin; Hct, hematocrit; LTA–AA, light transmission assay-arachidonic acid.
cardiovascular events has been substantially illustrated, it is crucial to reveal predictors of HAPR and uncover potential mechanisms, in order to identify at-risk patients and to optimize antiplatelet therapy, especially in the elderly.

In our study, HAPR patients tended to have reduced renal function, and Spearman’s correlation analysis confirmed an inverse association between LTA–AA and eGFR. Similarly, Blann et al observed a weak relationship between lower eGFR and HAPR, regardless of the method for platelet reactivity evaluation. What is more, eGFR even showed an inverse correlation with platelet expression of CD62P. Aksu et al observed higher incidence of HAPR in patients with severe chronic kidney disease (CKD), but not in mild or moderate CKD. HAPR might contribute to the increasing risk of cardiovascular complications in patients with declined renal function. However, Tello–Montoliu et al failed to track the link between antiplatelet response with renal dysfunction in non-diabetic patients with CAD. Our study excluded patients with severe CKD, thus minimizing the impact.

In our study, higher SUA level, as well as higher incidence of hyperuricemia was observed in HAPR patients. Previous studies reported the increasing risk of recurrent gout attacks in patients on low-dose aspirin. However, the link between SUA with aspirin response in the elderly has not been reported before. In the Nardin et al study, SUA level increased in patients receiving aspirin in combination with ticagrelor, but not in those with clopidogrel. However, their work failed to confirm effects of SUA on residual platelet reactivity. Similarly, Barbieri et al did not observe an association between SUA and HAPR. However, a weak correlation between HAPR with SUA was shown in our enrolled patients, and higher SUA level might potentially indicate the relatively poor response to aspirin. A possible explanation was that elevation of uric acid promotes the formation of a pro-inflammatory and prothrombotic state, with an increase in non-platelet TXA2 synthesis, thus leading to insufficient platelet inhibition.

Elderly HAPR patients had lower platelet count, when compared with non-HAPR patients. Likewise, platelet count was weakly correlated with HAPR in our study. It was conventionally thought that platelet counts might influence the results of platelet aggregation studies. However, recent studies revealed that LTA is not sensitive to platelet count changes, at least within a broad range of 150–600 x 10^9/L. In ADAPT–DES (Assessment of Dual AntiPlatelet Therapy with Drug-Eluting Stents) study, lower platelet count was associated with higher P2Y12 reaction units (PRU), but after multivariate adjustment, higher platelet count was independently associated with increased thrombosis risks.

It was also reported that aspirin might promote platelet apoptosis via several mechanisms, including the activation of p38 map kinase (MAPK), protein kinase C/B (PKC/PKB) and caspase-3, inhibition of proteasome function, triggering phosphatidylserine externalization and so forth. An inverse association of LTA–AA with hemoglobin and hematocrit was observed in our study. Previous studies reported the negative association between hematocrit or hemoglobin with VerifyNow assays or multiple electronic assay results, but not with LTA. In Voisin et al study, while PRU significantly declined, hematocrit and hemoglobin increased, but the percentage of platelet inhibition did not vary. In Pendyala et al study, lower baseline hematocrit was linked with HPR evaluated by the VerifyNow assay. Kim et al confirmed an inverse correlation between VerifyNow assay results with hematocrit (r=-0.409, P=0.001), and after adjustment, anemia was an independent risk factor for HPR. A possible explanation is that hematocrit changes might influence calcium concentration in citrated blood used for PFTs. Thus, Kakoulos et al developed a formula for PRU adjustment according to hematocrit.

Concomitant drugs might influence antiplatelet effects as well. Our study revealed the proportion of diuretic use was higher in HAPR patients when compared with non-HAPR patients. However, it is inadequate to establish a causal relationship, and the potential mechanisms warrant further study. Further, concomitant P2Y12 receptor inhibitor use was correlated with HAPR, possibly due to the tendency of more P2Y12 receptor inhibitor use in patients with poor aspirin response to obtain sufficient platelet inhibition. Furthermore, previous studies reported possible effects of PPIs, calcium channel blockers (CCBs) or statins on antiplatelet responses, while controversies existed. In Liu et al, esomeprazole or rabeprazole use did not affect antiplatelet response after initiation of DAPT for 30 days. ADAPT–DES study demonstrated the interaction between combined PPIs use with clopidogrel (OR: 1.38, 95% CI: 1.25–1.52, P<0.001). Tsukahara et al reported that CCB use was associated with higher incidence of HPR (OR: 1.93, 95% CI: 1.18–3.18, P<0.05). In Gremmel et al, co-administration with CCB was an independent risk factor of HPR, based on both LTA assay and VerifyNow assay. In terms of statins effects, it was reported that switching to a non-CYP3A4 metabolized statin might minimize drug–drug interactions and improve antiplatelet responses. However, Malmstrom et al revealed that lipid-lowering treatments did not exert additional inhibitory effects on platelets in CAD patients with impaired glucose tolerance.

The present studies have some advantages. First, the mean age of our enrolled patients were 77.2 years, higher
than most previous studies, which focused on HAPR. As older age has been recognized as a crucial risk factor for HAPR, investigating predictors of HAPR in this population might promote the optimization of antiplatelet therapy and uncover potential mechanisms. Further, ADP-induced platelet aggregation was used to exclude patients with poor response to thienopyridines, thus minimizing interference factors. Furthermore, all LTA assays in this study were performed by skilled technicians in our clinical laboratory, and results were normalized according to platelet counts.

Nevertheless, there are a few limitations. Mainly, this is a single-center study and other PFTs such as VerifyNow aspirin, multiple electrode platelet aggregometry or PFA-100 were not performed simultaneously.

In conclusion, elderly HAPR patients tended to have reduced renal function, higher SUA level as well as lower platelet count, hemoglobin and hematocrit. Platelet count, SUA, hematocrit and concomitant P2Y12 receptor inhibitors use were independently correlated with HAPR. These parameters might provide novel therapeutic targets for optimizing antiplatelet therapy.

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Disclosure

The authors report no conflicts of interest in this work.

References


Supplementary materials

**Table S1** Predictive value for HAPR (ROC curve)

<table>
<thead>
<tr>
<th>Variables</th>
<th>AUC</th>
<th>P-value</th>
<th>95% Lower</th>
<th>95% Upper</th>
</tr>
</thead>
<tbody>
<tr>
<td>SUA</td>
<td>0.625</td>
<td>0.002**</td>
<td>0.545</td>
<td>0.706</td>
</tr>
<tr>
<td>PLT</td>
<td>0.580</td>
<td>0.052</td>
<td>0.499</td>
<td>0.662</td>
</tr>
<tr>
<td>eGFR</td>
<td>0.593</td>
<td>0.024*</td>
<td>0.511</td>
<td>0.676</td>
</tr>
<tr>
<td>Hb</td>
<td>0.578</td>
<td>0.060</td>
<td>0.500</td>
<td>0.656</td>
</tr>
<tr>
<td>Hct</td>
<td>0.595</td>
<td>0.021*</td>
<td>0.518</td>
<td>0.673</td>
</tr>
<tr>
<td>Combination (SUA + PLT + eGFR + Hb + Hct)</td>
<td>0.661</td>
<td>&lt;0.001***</td>
<td>0.581</td>
<td>0.740</td>
</tr>
</tbody>
</table>

Notes: *P<0.05, **P<0.01, ***P<0.001.

Abbreviations: AUC, area under the curve; eGFR, estimated glomerular filtration rate; HAPR, high on-aspirin platelet reactivity; Hb, hemoglobin; Hct, hematocrit; PLT, platelet count; ROC, receiver operator characteristic; SUA, serum uric acid.

**Figure S1** Flowchart of the study.

Abbreviations: CAD, coronary artery disease; HAPR, high on-aspirin platelet reactivity; LAPR, low on-aspirin platelet reactivity; LTA–AA, light transmission assay-arachidonic acid; MAPR, moderate on-aspirin platelet reactivity.