Scrub typhus: risks, diagnostic issues, and management challenges

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Abstract: Scrub typhus is an acute febrile illness in the “tsutsugamushi triangle”, transmitted by chiggers that can be treated effectively if detected early. Laboratory testing, including molecular and serological assays, is needed for confirming the diagnosis, especially in the absence of the pathognomonic eschar. In this review, factors that play a role in disease occurrence and clinical clues for diagnosis, in addition to risk factors contributing to disease severity, including mortality, are discussed in detail. Moreover, issues related to diagnostic assays, treatment, and mixed infections are also enumerated and described.

Keywords: Orientia tsutsugamushi, disease severity, mortality predictors, diagnosis, coinfections, treatment

Introduction
Scrub typhus is a vector-borne zoonosis endemic in South Asia, Southeast Asia, East Asia, the Pacific Islands, and Northern Australia (the “tsutsugamushi triangle”), with reports of similar infections from Africa, the Middle East, and South America. This infection is caused by Orientia tsutsugamushi, which is transmitted to humans by the bite of infected chiggers (larvae) of trombiculid mites. The name “tsutsugamushi disease” was given by Hashimoto in 1810. The tsutsugamushi triangle is home to more than half the world’s population, with 2 billion at risk and 1 million cases of scrub typhus occurring per year. Clinical manifestations range from asymptomatic to severe disease. The mortality rate varies and can be as high 50%, such that the mortality among 1 million infections in a single year is likely enormous. This is because the organism responsible affects the vascular endothelium and mononuclear macrophages. Therefore, all organs, including the lungs, liver, kidneys, and central nervous system, can be affected. Misdiagnosis and underdiagnosis is also known to occur due to lack of availability of diagnostic tests and the aspecific nature of symptoms, especially when the characteristic eschar is not present.

Scrub typhus is not transmitted directly from person to person; it is only transmitted by the bites of vectors. The vector responsible is the chigger of the trombiculid mite belonging to the genus Leptotrombidium, but recently newer vector genera have been discovered that are capable of transmitting this agent. Tilak et al reported that Schoengastia ligula (northeast India) transmitted O. tsutsugamushi in tea-garden workers, while Lee et al discovered this agent could be transmitted by Euschoengastia koreaensis in South Korea. Knowledge of the vector, including species, distribution, density, and habitats, is important to understand the epidemiology of scrub typhus in...
a given area or region. An increase in vector density contributes to increased transmission, due to more humans being bitten by infected chiggers.

*O. tsutsugamushi* serotype distribution varies from region to region in the tsutsugamushi triangle, and strain types are identified by sequencing the 56 kDa gene. In South Korea, the Boryong serotype is predominantly encountered, and the Karp and Gilliam serotypes are common in Taiwan, and the Gilliam serotype is prevalent in China. In India, based on a 56 kDa analysis, strains similar to Kato and Karp are common, whereas in Japan Kato, Karp, Gilliam, Kawasaki, and Kuroki types are observed. Scrub typhus without the eschar is a febrile illness without any evidence of localization, and is hence termed “acute undifferentiated fever.” This illness is thus clinically indistinguishable from malaria, dengue fever, other rickettsioses, leptospirosis, and enteric fever, which are common causes of acute undifferentiated fever in the Asia-Pacific region. In this review, factors describing populations at risk, severity predictors, clinical clues, diagnostic assays, coinfections observed, and drugs available for treatment of scrub typhus are described.

**Risk factors for acquiring scrub typhus**

The abundance of the chigger of the trombiculid mite, which is the vector for scrub typhus, determines the chance of acquiring scrub typhus, which in turn determines the prevalence of scrub typhus in a given region. There is always a spurt of cases during certain seasons in the endemic areas described, and this varies from country to country and is dependent on the climate and environment. Additionally, within a country, certain regions show increased prevalence.

Chiggers are abundant in locales with high relative humidity (60%–85%), low temperature (20°C–30°C), low incidence of sunlight, and a dense substrate-vegetative canopy. As such, they are found in great numbers in forest clearings, riverbanks, and grassy regions. Humans acquire scrub typhus on exposure to infected larvae (chiggers) of trombiculid mites. The density of chiggers of *Leptotrombidium pallidum* and *L. scutellare* is very high from September to November in South Korea, with a consequent rise in scrub typhus in humans. This provides evidence that an increase in chigger density of the vector species is responsible for the high seasonal prevalence in endemic areas. This variation in chigger activity gives rise to the seasonality of scrub typhus.

Peak prevalence in South Korea occurs in autumn (September–November), in Japan increased cases are observed in autumn and winter, in north China (Shandong) in September–November, and in south China in June–September. In south India, scrub-typhus cases occur mostly in the cooler months (August–January), while in Southeast Asia scrub-typhus cases are highest in July–November. Incidence of scrub typhus can vary from country to country and also region in large countries like India and China. It has been reported that each 1°C and 1% change in temperature can cause an increase in incidence, as evidenced by a 15% rise in scrub-typhus cases in Guangzhou, China.

In addition to temperature, secondary vegetation and rainfall increase the incidence of scrub typhus. Occupational risk is higher in farmers (aged 50–69 years), females, and those working in vegetable fields, harvesting in autumn, and rural highlands. In a study by Kweon et al, outdoor activities like resting on a grass field without a mat, working in short sleeves and bare hands, and defecating and/or urinating outdoors while squatting increased risk for scrub typhus. In a case-control study from South Korea, individuals engaged in fruit farming, gathering chestnuts, and taking breaks in areas adjacent to agricultural operations had an increased risk of contracting scrub typhus compared to controls. The authors opined that providing a health-education program would lower the risk in these individuals and similar groups. Land use is another determinant, as scrub-typhus incidence increases when forest lands are converted to fields, palm oil, and rubber plantations, and also when urbanization occurs.

**Clinical clues favoring a diagnosis of scrub typhus**

**Presence of eschars**

The presence of eschars is considered pathognomonic. It has been reported that eschar incidence varies from 7% to 97% in endemic areas and is painless. Eschars are often found in covered areas of the body, such as the groin, axilla, chest, and lower back, including the buttocks. Recently, there have been reports of multiple eschars and atypical eschars, which were punched-out ulcers with slough. This could also be due to the original eschar scab being removed by scratching or falling away, especially during bathing. This seems plausible, because the eschar appears at the chigger-bite site a few days before the onset of symptoms. Therefore, a thorough examination becomes necessary and improves eschar detection, leading in turn to improvement in the diagnosis of scrub typhus in a clinical setting. This has been observed at our center: eschar incidence improved from...
<10% in 200342 to 55% in 2013.46 Eschars of scrub typhus appear a few days after at chigger-bite sites, before the disease manifests. The eschar is painless and consists of a black scab, with an erythematous halo and minimal edema.44,47 Detection of eschars is dependent not only on the clinical experience of the examining physician but is also influenced by skin color (eschars are more easily seen on the fair-skinned than the dark-complexioned) and also on the thoroughness of the physical examination.48 The differentials for a scrub-typhus eschar include insect bites (including spider bites) and post-traumatic scabs,49 which can all be ruled out with a little patience and perseverance. The eschar of anthrax, though painless, is surrounded by extensive or marked edema that is gelatinous and stretches the skin, and is often preceded by a pruritic papule.50,51 Table 1 gives a summary of clinical and laboratory findings favoring or not favoring a diagnosis of scrub typhus.23,25,28,52–58

Use of clinical prediction rules
As given in Table 1, a few clinical clues other than eschars are available for suspecting or diagnosing scrub typhus in the clinic or at the bedside. It is to be noted that even at a referral hospital in endemic countries, the diagnosis of scrub typhus is usually based on the clinical findings.54 As definitive diagnosis requires laboratory testing, clinical prediction rules have been tried. Chen et al observed a 100% negative predictive value for clinical criteria combining the presence of eschars, atypical lymphocytes in peripheral blood smear, and contact history.59 A clinical rule formulated for scrub typhus by Jung et al uses five predictors with a maximum score of 8 points. The scoring criteria include age $\geq 65$ years (2 points), recent history of fieldwork/outdoor activities (1 point), onset of illness during an outbreak period of scrub typhus (2 points), myalgia (1 point), and eschars (2 points). A score below 3 rules out scrub typhus, while 100% sensitivity was observed for a score $\geq 3$. The authors felt that this could be used for selecting patients for empirical therapy in resource-poor situations or for performing specific laboratory tests.23 Similarly, Siriwongpan et al devised and validated a clinical risk scoring system using a set of 526 patients with scrub typhus based on the World Health Organization (WHO) case definition.60–62

<table>
<thead>
<tr>
<th>Supporting a diagnosis of scrub typhus</th>
<th>Against a diagnosis of scrub typhus (usually diagnosed)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Eschar</strong></td>
<td>Bone pain (dengue)</td>
</tr>
<tr>
<td>Regional lymphadenopathy</td>
<td>Bleeding manifestations (dengue)</td>
</tr>
<tr>
<td>Total fever $\geq 8$ days</td>
<td>Loose stools (enteric fever)</td>
</tr>
<tr>
<td>CRP $&gt;32$ mg/L</td>
<td>White blood-cell counts $&lt;5,000$/mm$^3$ (dengue)</td>
</tr>
<tr>
<td>ALT/AST $&gt;1$</td>
<td>Platelets $&lt;50,000$/mm$^3$ (dengue)</td>
</tr>
<tr>
<td>Defervescence within 48–72 hours of specific therapy</td>
<td>Bilirubin $&gt;2$ mg/dL (malaria, hepatitis A)</td>
</tr>
<tr>
<td></td>
<td>AST $&gt;500$ U/L (dengue)</td>
</tr>
<tr>
<td></td>
<td>ALT $&lt;100$ U/L (malaria)</td>
</tr>
<tr>
<td></td>
<td>ALT $&gt;500$ U/L (hepatitis A)</td>
</tr>
</tbody>
</table>

**Risk factors determining severity and outcome in scrub typhus**
Severity of disease based on genotype
The severity of scrub typhus varies considerably, which might correlate with the virulence of the particular *O. tsutsugamushi* strain responsible for the infection. There is evidence that frequency of eschars and rash in scrub typhus is dependent on the infecting genotype. South Korean individuals with the Boryong genotype have significantly higher incidence (97%) of eschars and skin rash compared to 74% with the Karp genotype.53 Karp genotypes (summer scrub typhus, isolated in Guangdong, Fujian, Hainan provinces in southern China) were found to be more virulent and caused more severe disease than Kawasaki genotypes (autumn–winter scrub typhus) isolated in Shandong and northern Jiangsu provinces in northern China.48 Eschar presence was not significant in severe and nonsevere scrub typhus.64

Clinical and laboratory parameters predicting severity in scrub typhus
Patients with possible scrub typhus with low body temperature, rapid pulse rate, presence of crepitation, low percentage of lymphocytes, low serum albumin, elevated aspartate aminotransferase, elevated serum creatinine, and positive urine albumin should be monitored closely for severity progression.60 Tables 2 and 3 provide a summary of the significant features predicting severity and outcome of scrub typhus. Parameters that show very significant $P$-values (0.01) by multivariate logistic regression analysis have been included in the tables. In a meta-analysis of 89 studies (19,644 patients with scrub typhus), fatal outcome was reported in 2,488 patients, with an overall mortality of 12.7%. Though increasing age
was associated with fatality, presence or absence of eschars did not affect the outcome.65

**Laboratory diagnosis of scrub typhus**

Scrub typhus can mimic other acute febrile illnesses common in the tropics, especially when pathognomonic eschars are absent.10 Therefore, laboratory tests become mandatory for confirmation of the diagnosis.38,75,76 Methods available include direct methods like isolation of the pathogen in cell cultures (HeLa, L929, Vero, and BHK21) and detection of scrub typhus-specific DNA like 56 kDa, 47 kDa, 16S ribosomal RNA, and GroEL gene targets by polymerase chain reaction (PCR). Indirect methods include detection of antibodies to *O. tsutsugamushi* by immunofluorescence assay (IFA), enzyme-linked immunosorbent assay (ELISA),77 and rapid diagnostic assays.78,79 Tables 4 and 5 give performance characteristics of the available assays for laboratory confirmation of scrub typhus.

Real-time PCR assays like the 47 kDa, 56 kDa, and GroEL type-specific genes are used (eg, 56 kDa and 47 kDa genes for *O. tsutsugamushi*) than if genus-specific genes are used for confirmation of the diagnosis.38,75,76 Methods available include direct methods like isolation of the pathogen in cell cultures (HeLa, L929, Vero, and BHK21) and detection of scrub typhus-specific DNA like 56 kDa, 47 kDa, 16S ribosomal RNA, and GroEL gene targets by polymerase chain reaction (PCR). Indirect methods include detection of antibodies to *O. tsutsugamushi* by immunofluorescence assay (IFA), enzyme-linked immunosorbent assay (ELISA),77 and rapid diagnostic assays.78,79 Tables 4 and 5 give performance characteristics of the available assays for laboratory confirmation of scrub typhus.

Real-time PCR assays like the 47 kDa, 56 kDa, and GroEL were increasingly used, and these detect 10–50 copies/μL of *O. tsutsugamushi*. Real-time PCR specificity is higher if type-specific genes are used (eg, 56 kDa and 47 kDa genes for *O. tsutsugamushi*) than if genus-specific genes are used (17 kDa genes for *Rickettsia* spp.), which again are stronger than nonspecific conserved “housekeeping” genes like *HSPD1* (GroEL) and 16S ribosomal RNA.27 The drawback...
Table 4 Performance of nonmolecular diagnostic tests used for detection of scrub typhus

<table>
<thead>
<tr>
<th>Type of assay</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cell culture</td>
<td>5–56</td>
<td>100</td>
</tr>
<tr>
<td>Antigen detection</td>
<td>65–100</td>
<td>100</td>
</tr>
<tr>
<td>IgM IFA</td>
<td>70–100</td>
<td>84–100</td>
</tr>
<tr>
<td>IgM + IgG IFA</td>
<td>78–97</td>
<td>98–100</td>
</tr>
<tr>
<td>IgM ELISA</td>
<td>70–100</td>
<td>87–100</td>
</tr>
<tr>
<td>IgG ELISA</td>
<td>58–96</td>
<td>92–98</td>
</tr>
<tr>
<td>IgM ICT</td>
<td>47–99</td>
<td>95–100</td>
</tr>
<tr>
<td>IgM + IgG ICT</td>
<td>61–100</td>
<td>74–100</td>
</tr>
</tbody>
</table>

**Abbreviations:** IFA, immunofluorescence assay; ELISA, enzyme-linked immunosorbent assay; ICT, immunochromatographic test.

Table 5 Summary of performance characteristics of molecular assays for diagnosis of scrub typhus

<table>
<thead>
<tr>
<th>Target</th>
<th>Assay</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>56 kDa</td>
<td>Conventional PCR</td>
<td>0–96</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Nested PCR</td>
<td>16–100</td>
<td>88–100</td>
</tr>
<tr>
<td></td>
<td>qPCR</td>
<td>65–73</td>
<td>100</td>
</tr>
<tr>
<td>47 kDa</td>
<td>Conventional PCR</td>
<td>3–7</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Nested PCR</td>
<td>81–85</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>qPCR</td>
<td>63–81</td>
<td>90–100</td>
</tr>
<tr>
<td>16S rRNA</td>
<td>LAMP</td>
<td>52</td>
<td>94</td>
</tr>
<tr>
<td></td>
<td></td>
<td>45–87</td>
<td>100</td>
</tr>
<tr>
<td>GroEL</td>
<td>Conventional PCR</td>
<td>66</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Nested PCR</td>
<td>90.4</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>qPCR</td>
<td>56.4</td>
<td>96.2</td>
</tr>
<tr>
<td></td>
<td>LAMP</td>
<td>87.5</td>
<td>100</td>
</tr>
</tbody>
</table>

**Note:** An evaluation done using 24 eschar samples from scrub typhus–confirmed cases showed sensitivity and specificity of 83.3% and 100%, respectively (Prakash, unpublished data, 2012).

**Abbreviations:** qPCR, quantitative polymerase chain reaction; LAMP, loop-mediated isothermal amplification; rRNA, ribosomal RNA.
used in resource-poor situations as point-of-care diagnostic tests, whereas performance of a PCR would be very useful in centers with facilities for same.49

**Coinfections and scrub typhus**

In endemic areas, coinfections have been described, and these include infections with other pathogens causing similar illness. Table 6 enumerates the grading of coinfections according to Phommasone et al,27 and Tables 7 and 8 describe such infections.

A few scenarios are described for our understanding. The first is by Sonthayanon et al, who found that among the 82 serological coinfections observed, molecular assays were positive for leptospirosis in 43 (52%), scrub typhus in nine (11%), and both in five (6%), whereas 25 (30%) were negative for both leptospirosis and scrub typhus. Possible explanations for the difference observed between serologic and molecular results include low sensitivity of the molecular assay, failure to test a sample obtained during the window of bacteremia in leptospirosis, serologic cross-reactivity, and acute infection caused by one pathogen in the background of a recent but inactive infection caused by the second pathogen.128

Second, in the presence of eschars, testing for *Leptospira* serology is unwarranted, according to Lee and Liu, as four of the seven cases who were *Leptospira* serology-positive had eschars.26 As treatment with doxycycline or azithromycin is very effective against *Leptospira* and *Orientia*, serological cross-reaction or coinfection does not matter, as treatment with either will be beneficial to the patient with an acute febrile illness when both serologies are positive.129

Third, dual and triple infections occur, as reported by Ahmad et al, who described malaria, dengue, and scrub typhus in five cases, 21 were dengue cross-reactive, malaria smears were positive in 14, and nine individuals had IgM antibodies to scrub typhus and dengue. Further clarification regarding which was a cross-reaction could have been determined if information regarding presence or absence of

<table>
<thead>
<tr>
<th>Evidence grade</th>
<th>Coinfecting pathogen (test positive)</th>
<th>Positive</th>
<th>Diagnostic test positive for scrub typhus</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grade I</td>
<td><em>Rickettsia typhi</em> (PCR)132</td>
<td>3</td>
<td>PCR</td>
</tr>
<tr>
<td></td>
<td><em>Plasmodium falciparum</em> (smear)133</td>
<td>2</td>
<td>PCR</td>
</tr>
<tr>
<td></td>
<td><em>Leptospira</em> (culture, PCR, MAT)133</td>
<td>5</td>
<td>Culture and/or PCR</td>
</tr>
<tr>
<td></td>
<td>Dengue (NS1 antigen, PCR)133</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td></td>
<td><em>Rickettsia</em> spp. (PCR)133</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td></td>
<td><em>Mycoplasma pneumoniae</em> (PCR)134</td>
<td>1</td>
<td>PCR</td>
</tr>
<tr>
<td></td>
<td><em>Leptospira</em> (PCR)134</td>
<td>5</td>
<td>PCR</td>
</tr>
<tr>
<td>Grade I and II</td>
<td><em>Leptospira</em> (culture, PCR, MAT)135</td>
<td>4</td>
<td>Culture, PCR, and IFA</td>
</tr>
<tr>
<td>Grade II</td>
<td><em>Leptospira</em> (culture, MAT, IFA)136</td>
<td>62</td>
<td>IFA</td>
</tr>
<tr>
<td></td>
<td><em>Leptospira</em> (MAT)136</td>
<td>7</td>
<td>IFA</td>
</tr>
<tr>
<td></td>
<td><em>Leptospira</em> (culture, MAT)137</td>
<td>11</td>
<td>IFA</td>
</tr>
<tr>
<td></td>
<td><em>Coxiella burnetii</em> (IFA)137</td>
<td>5</td>
<td>IFA</td>
</tr>
<tr>
<td></td>
<td>Dengue (NS1-antigen ELISA)138</td>
<td>1</td>
<td>IFA</td>
</tr>
<tr>
<td></td>
<td><em>JEV</em> (IgM ELISA)139</td>
<td>26</td>
<td>Culture and/or PCR</td>
</tr>
<tr>
<td></td>
<td><em>Leptospira</em> (MAT)139</td>
<td>1</td>
<td>IFA</td>
</tr>
<tr>
<td></td>
<td><em>Leptospira</em> (MAT)140</td>
<td>1</td>
<td>IFA</td>
</tr>
<tr>
<td></td>
<td>Severe fever with thrombocytopenia syndrome (PCR)141</td>
<td>3</td>
<td>IFA</td>
</tr>
<tr>
<td>Grade III</td>
<td><em>Leptospira</em> (MAT)142</td>
<td>9</td>
<td>Dot-blot assay</td>
</tr>
<tr>
<td></td>
<td>Dengue (NS1 antigen)143</td>
<td>6</td>
<td>IgM ELISA</td>
</tr>
<tr>
<td></td>
<td><em>Leptospira</em> (IgM ELISA)143</td>
<td>8</td>
<td>IgM ELISA</td>
</tr>
<tr>
<td></td>
<td>Dengue (IgM ELISA)143</td>
<td>21</td>
<td>IgM ELISA</td>
</tr>
<tr>
<td></td>
<td>Malaria (smear)144</td>
<td>14</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Spotted fever, group rickettsia (IFA)144</td>
<td>3</td>
<td>IFA</td>
</tr>
<tr>
<td></td>
<td><em>R. typhi</em> (IFA)144</td>
<td>1</td>
<td></td>
</tr>
</tbody>
</table>

**Table 6 Grading of coinfections**

<table>
<thead>
<tr>
<th>Grade</th>
<th>Tests</th>
<th>Specificity</th>
<th>Sensitivity</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Culture, NAATs, and antigen detection</td>
<td>Best</td>
<td>Poor</td>
</tr>
<tr>
<td>II</td>
<td>Seroconversion</td>
<td>Good</td>
<td>Good</td>
</tr>
<tr>
<td>III</td>
<td>Single serological value above cutoff</td>
<td>Poor</td>
<td>Very good</td>
</tr>
</tbody>
</table>

Abbreviation: NAATs, nucleic acid-amplification tests.

**Table 7 Coinfections (dual) demonstrated in scrub-typhus patients**

**Abbreviations:** PCR, polymerase chain reaction; MAT, microscopic agglutination test; IFA, immunofluorescence assay; ELISA, enzyme-linked immunosorbent assay; JEV, Japanese encephalitis virus.
eschars was available or determination of NS1-antigen positivity, as was done by Basheer et al.  

# Management challenges

Atypical clinical features and absence of eschars may result in delayed diagnosis, complications, or death. 

Scrub typhus responds promptly to effective treatment, with patients becoming afebrile within 24–48 hours, so much so that when enteric fever, septicemia, and malaria are ruled out, empirical treatment with doxycycline (even when given late in the disease) is clinically useful. Therefore, empirical therapy with doxycycline is to be encouraged in regions or locales where scrub typhus is endemic or reemerging. This will lead to a reduction in complications, with a corresponding decrease in morbidity and mortality. Treatment for scrub typhus has been reviewed extensively by Peter et al and Rajapakse et al. Doxycycline is useful as an empirical treatment, because of its high cost-effectiveness and wide spectum of activity, and is considered safe in children <8 years of age. There is grade B evidence for lack of dental staining in children given short-course doxycycline, as may be given in scrub typhus. Moreover, doxycycline reaches good concentrations in cerebrospinal fluid, as does minocycline, though use of the latter is limited by dose-related vestibular side effects. Fluoroquinolones are not good drugs for treatment, nor are penicillins, clarithromycin, and cephalosporins. O. tsutsugamushi with reduced susceptibility has been observed for doxycycline and chloramphenicol in Chiang Rai, northern Thailand. This may not have been true resistance, but due to delayed treatment or tolerance.

Data on scrub typhus in pregnancy are scanty. Among the 82 cases reviewed from the literature till 2014 by McGready et al, 2.5% were associated with maternal mortality. Miscarriage occurred in 17%, and poor neonatal outcomes (stillbirth, preterm labor, and low birth weight) were documented in 42%. Macrolide antibiotics, such as azithromycin, are safe in pregnancy, but doxycycline, which is cheaper, can also be used if the former is not available. The aim of therapy is to save both mother and child, and the benefits of therapy with doxycycline outweigh the risks. Cross et al opined that doxycycline treatment should be used in children and pregnant women for treating scrub typhus, as the infection-associated risks are too large and thus overwhelmingly against avoiding therapy with this agent. Recently, Jang et al reported that intravenous azithromycin was efficacious in the treatment of severe scrub typhus.

The major concern is that misdiagnosis occurs when the characteristic eschars are absent. This is of importance, as treatment with an appropriate antibiotic (doxycycline, tetracycline, or chloramphenicol) renders patients afebrile within 48 hours, such that pyrexia persisting beyond 72 hours rules out scrub typhus. Such patients have jaundice (icteric sclera and/or total bilirubin >1.5 mg/dL), no headache, and relative bradycardia (<110/min).

# Conclusion

Scrub typhus is an important cause of febrile illness in the Asia-Pacific region. The main management challenge is institution of specific therapy in a timely and an effective manner, as stated elsewhere in this review. For this, rapid and accurate diagnosis becomes necessary, especially in the absence of eschars. In resource-poor endemic settings, clinical prediction rules have been defined and found useful. In addition, a battery of tests is needed for increasing diagnostic yield and sorting out the issue of coinfections. Finally, appropriate treatment should be initiated, keeping in mind the risk and benefits afforded by such treatment.

# Disclosure

The author reports no conflicts of interest in this work.

# Table 8 Details of infections with two other pathogens in scrub-typhus patients

<table>
<thead>
<tr>
<th>First pathogen (test)</th>
<th>Second pathogen (test)</th>
<th>Third pathogen (test)</th>
<th>Number (evidence)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Orientia tsutsugamushi (PCR)</td>
<td>Rickettsia typhi (PCR)</td>
<td>Mycobacterium tuberculosis (culture)</td>
<td>1 (Grade I)</td>
</tr>
<tr>
<td>O. tsutsugamushi (PCR)</td>
<td>R. typhi (PCR)</td>
<td>Salmonella enterica, group D (culture)</td>
<td>1 (Grade I)</td>
</tr>
<tr>
<td>O. tsutsugamushi (PCR)</td>
<td>Leptospirosis (PCR)</td>
<td>JEV (IgM-capture ELISA)</td>
<td>2 (Grade II)</td>
</tr>
<tr>
<td>Plasmodium vivax (RDT)</td>
<td>O. tsutsugamushi (PCR)</td>
<td>Dengue (NS1 antigen)</td>
<td>1 (Grade I)</td>
</tr>
<tr>
<td>Malaria (smear)</td>
<td>Dengue (IgM ELISA)</td>
<td>O. tsutsugamushi (IgM ELISA)</td>
<td>5 (Grade III)</td>
</tr>
<tr>
<td>O. tsutsugamushi (IFA)</td>
<td>R. typhi (IFA)</td>
<td>Spotted fever, group rickettsia (IFA)</td>
<td>6 (Grade III)</td>
</tr>
</tbody>
</table>

**Abbreviations:** PCR, polymerase chain reaction; JEV, Japanese encephalitis virus ELISA, enzyme-linked immunosorbent assay; RDT, rapid diagnostic test; IFA, immunofluorescence assay.

# References


