Photothermal therapy of melanoma tumor using multiwalled carbon nanotubes

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Abstract: Photothermal therapy (PTT) is a therapeutic method in which photon energy is transformed into heat rapidly via different operations to extirpate cancer. Nanoparticles, such as carbon nanotubes (CNTs) have exceptional optical absorbance in visible and near infrared spectra. Therefore, they could be a good converter to induce hyperthermia in PTT technique. In our study, for improving the dispersibility of multiwalled CNTs in water, the CNTs were oxidized (O-CNTs) and then polyethylene glycol (PEG) was used for wrapping the surface of nanotubes. The formation of a thin layer of PEG around the nanotubes was confirmed through Fourier transform infrared, thermogravimetric analysis, and field emission scanning electron microscopy techniques. Results of thermogravimetric analysis showed that the amount of PEG component in the O-CNT-PEG was approximately 80% (w/w). Cell cytotoxicity study showed that O-CNT was less cytotoxic than pristine multiwalled nanotubes, and O-CNT-PEG had the lowest toxicity against HeLa and HepG2 cell lines. The effect of O-CNT-PEG in reduction of melanoma tumor size after PTT was evaluated. Cancerous mice were exposed to a continuous-wave near infrared laser diode (λ=808 nm, P=2 W and I=8 W/cm²) for 10 minutes once in the period of the treatment. The average size of tumor in mice receiving O-CNT-PEG decreased sharply in comparison with those that received laser therapy alone. Results of animal studies indicate that O-CNT-PEG is a powerful candidate for eradicating solid tumors in PTT technique.

Keywords: Photo thermal therapy, CNTs, hyperthermia, PEG, melanoma

Introduction

Cancer still remains one of the world’s most disastrous diseases. Conventional cancer treatments like surgical intervention, radiation, and chemotherapeutic drugs also kill healthy cells and cause toxicity to the body.1 A promising area that is making revolutionary strides in methods of cancer treatment is cancer nanotechnology, which includes the integration of fields like engineering, material sciences, optics, pharmaceutics, chemistry, and physics with cancer biology.2

Hyperthermia is a simple method of cancer therapy in which irradiation of near infrared (NIR) light increases the local temperature of tumor and disturbs the cancer cells. Internalized nanoparticles (NPs) to the tumor sites could be stimulated by laser irradiation, to produce localized heat in the range of 40°C–45°C so as to destroy cancer cells.2-5 NPs such as single-walled carbon nanotubes,6 multiwalled carbon nanotubes (MWNTs),7,8 graphene,9,10 iron oxide NPs,11 gold nanorods,12 and gold nanoshells13,14 are used to transform NIR radiation to vibrational energy. Therefore, heat generation based on laser irradiation could elevate malignant tissues’ temperature and destroy tumors.15 Inducing high temperature for a sufficient time causes physical damage like protein denaturation and membrane lysis and could increase oxidative stress, which can lead to cell death.2

The use of CNTs in photothermal therapy (PTT) has been of much interest due to their unique properties as photothermal agents. CNTs absorb light in the visible and near infrared (NIR) regions of the electromagnetic spectrum, and thus may be converted into heat by exposure to light.7,16-19

The aim of this study was to produce an effective therapeutic agent that can be used to treat melanoma tumors in photothermal therapy (PTT) using multiwalled carbon nanotubes.
stress, finally causing coagulative necrosis or apoptosis.\textsuperscript{5,15} Utilizing the NIR light in the range 700–1,100 nm to induce hyperthermia is really efficient because biological systems mainly lack chromophores to absorb NIR light. NIR wavelength, compared to other range of wavelengths, is more transmissive through the body and is poorly attenuated by biological systems.\textsuperscript{16,17} By localization and excitation of potent NPs in the tumor region, lesions could be cured without direct access to the tumor site and selectively destroyed.\textsuperscript{18} This method promises to ablate recurrent, unconventional, and inaccessible tumors. The wide electromagnetic absorbance spectrum of carbon nanotubes (CNTs) creates exceptional properties compared to other plasmonically heated nanomaterials (like gold nanoshells and nanorods) that is dependent on the size and shape of CNTs.\textsuperscript{19} CNTs promise an extraordinary combination of attributes in improving the next generation of photothermal agents because of their strong ability to transform NIR radiation into heat.\textsuperscript{20} NIR exposure of CNTs excites electrons to the excited state and releases vibrational energy through heat, which could be used to induce cell death.\textsuperscript{21,22} Studies show that CNTs can achieve thermal destruction using tenfold-lower doses in solution and using threefold-lower laser power than that required for gold nanorods.\textsuperscript{23} and these also indicate that MWNTs are more potent than bulk single-walled nanotubes in transferring the NIR light into heat.\textsuperscript{2} Studies by Murphy et al\textsuperscript{24} reported that the inflammation and fibrosis induced by CNTs could be reduced by shortening the length of CNTs. One of the conventional methods to shorten and exfoliate the CNT bundles is oxidation. Oxidation also enhances the wettability, surface reactions, functionality, and cell penetration of CNTs.\textsuperscript{25–27} To improve the poor dispersibility of CNTs into aqueous media, different hydrophilic polymers can be used. Functionalization of CNTs with polyethylene glycol (PEG) moieties (PEGylation) has been utilized to enhance the solubility, biocompatibility, and cell membrane penetration of the moieties.\textsuperscript{28,29}

In this study, at first we solubilized oxidized-MWNTs with PEG to enhance their hydrophilicity and biocompatibility and then evaluated their efficacy in treatment of melanoma via hyperthermia treatment in vivo.

Materials and methods
Materials
Purified MWNTs (number of walls: 3–15, outer diameter: 5–20 nm, length: 1–10 μm) were purchased from Plasmachem (Berlin, Germany). PEG1000 was purchased from Sigma-Aldrich (St Louis, MO, USA). All solvent were of analytical grade.

Preparation of CNTs-PEG
For wrapping of CNTs with PEG, the CNTs should be oxidized first. For oxidation of CNTs, 1 g of MWNTs was sonicated with 20 mL nitric acid and sulfuric acid solution (1:3 v/v) for 30 minutes. Then, the solution was refluxed for 21 hours at 100°C. The solution was diluted with 1 L of deionized water, then filtered, and finally washed till the pH adjusted to 6. The filtrate was dried in an oven at a temperature of 40°C for 24 hours.\textsuperscript{30–32} The oxidized-CNTs (O-CNTs) were ready for following steps.

50 mg of O-CNTs was added to 50 mL of deionized water containing 500 mg of PEG1000.\textsuperscript{33} The mixture was sonicated for 3 minutes and stirred overnight for wrapping hydrophilic polymer around the O-CNTs. The resulting solution was centrifuged at 4,000 rpm for 10 minutes to remove any aggregates, and the supernatant containing O-CNT-PEG was collected. Excess PEG was removed by dialysis.

Characterization of O-CNT-PEG
IR measurements were performed by (Perkin Elmer Spectrum One, Fourier transform infrared [FT-IR]; Shelton, CT, USA). Thermogravimetric analysis (TGA) was carried out (BAHR-Thermoanalyse Gmbh, TGA; Hüllhorst, Germany) under dynamic atmosphere of an inert gas (N\textsubscript{2}) at 30 mL/min (room temperature). Field emission scanning electron microscopic (FESEM) analyses were performed (TESCAN MIRA 3-XMU, FESEM; Brno, Czech Republic). UV–Vis absorption of O-CNT-PEG was analyzed (PG instruments Ltd., T80+ UV–Vis spectrophotometer, Lutterworth, UK) to determine the best photoabsorption wavelength for excitation.

Cytotoxicity assay for O-CNT-PEG
To verify the cytotoxicity of modified CNTs, the cytotoxicity of MWNT, O-CNT, and O-CNT-PEG was assessed by the standard MTT assay. Two human cell lines, human cervical cancer cells (HeLa) and human hepatocellular carcinoma cell line (HepG2), were purchased from the National Cell Bank of Pasteur Institute (Tehran, Iran). HeLa and HepG2 were cultured in Roswell Park Memorial Institute medium (RPMI)-1640 medium supplemented with 10% fetal bovine serum and 1% penicillin–streptomycin at 37°C in a humidified incubator with 5% CO\textsubscript{2}. Cells in the exponential growth phase were seeded in 96-well plates at a density of 1x10\textsuperscript{4} viable cells/well. After overnight incubation, cells were exposed to different concentrations of MWNT, O-CNT, and O-CNT-PEG. After 24 hours, 20 μL of MTT (5 mg/mL) and 100 μL of medium were introduced. The plates were incubated for 3–4 hours. The formazan crystals in each well
were dissolved in 100 μL of dimethyl sulfoxide. After complete solubilization of the dye, plates were read at 570 nm against 690 nm on an ELISA reader. The percentage of cell viability was calculated. The cell viability was estimated as the reduction of values from a dimethyl sulfoxide control, and the values were the mean of three different experiments.

Tumor induction
The metastatic murine melanoma cell line B16/F10 (NCBI C540 was purchased from the National Cell Bank of Pasteur Institute of Iran, Tehran, Iran) was cultured in RPMI 1640 medium, under 5% CO₂ at 37°C. Then it was supplemented with 10% fetal bovine serum, 100 IU/mL of penicillin, and 100 μg/mL streptomycin.

For tumor induction, female C57BL/6J mice that were inbred, weighing 20–30 g, and aged 6–8 weeks were selected. Murine melanoma cells at a count of 10⁶–10⁷ were suspended in 200 μL culture medium and injected hypodermically. This examination was done in the Center of Experimental and Comparative Medicine, Shiraz University of Medical Sciences, Shiraz, Iran and was cleared by the Ethical Committee of Shiraz University of Medical Sciences generously. Mice selection, methods of care, and sacrificing were the same and adhered to the guideline of Animal Care Committee of Iran Veterinary Organization. Experiments were done under aseptic situation, and the protocol of anesthesia, postoperative care, and surgical methods were the same for all mice.

In vivo photothermal therapy (PTT) of tumors
The therapeutic effect of ablation with O-CNT-PEG and laser irradiation was evaluated by monitoring the size of tumor inoculated in mice. Two weeks after melanoma cell line injection, the tumors had grown adequately (about 1 cm³) to start hyperthermia therapy. The mice were randomized into 3 groups (n=5) and anesthetized with a combination of ketamine 10% (100 mg/kg) and xylazine 2% (10 mg/kg). After sedation, the hair on tumor was shaved and the skin was cleansed. Tumor sizes were measured using a caliper and evaluated with an ultrasonography machine (Ultrasonix SonixOP; Burnaby, BC, Canada) before the treatment and 3 days after the treatment. The tumor size was estimated with the following Equation:

\[
\text{Tumor volume} = \left(\frac{L}{W}\right)^2 \times 2 \ (\text{mm}^3)
\]

where L and W represent length and width of the tumor, respectively.

The mice received treatment as follows:
Group I (O-CNT-PEG): O-CNT-PEG (1 mg/mL) was injected into the tumor at a dose of 200 μL/cm³ (tumor volume).
Group II (Laser therapy): laser therapy was done without any pretreatment with NPs.
Group III (Control): did not receive any treatment.

The tumor area in groups I and II were irradiated using an 808 nm continuous-wave NIR laser diode 808-2W with the intensity of 8 W/cm² and spot size of 0.25 cm² for 10 minutes. After treatment, all mice were sacrificed, the tumor size was measured, and the mass was excised for histological studies.

Statistical analysis
Data are expressed as mean ± SD. Significant differences of the values were statistically tested using paired-sample t-test in each group. Multiple comparisons at multiple time points were tested using analysis of variance with repeated measures. The statistical analyses were performed by SPSS® statistical software, version 20.0 (SPSS Inc., Chicago, IL, USA) for Windows®. A P-value of <0.05 was regarded as significant.

Results
Preparation and characterization of O-CNT-PEG
TGA measurements and FT-IR spectroscopy were employed to determine wrapping the CNTs with PEG chains. In the FT-IR spectrum of O-CNT, the peaks observed at 3,409.36, 2,870, and 1,750 cm⁻¹ were correlated to O-H (carboxylic acid), C-H, and C=O bonds, respectively (Figure 1). In the O-CNT-PEG, PEG-related peaks covered the peaks of O-CNT. Increase in the intensity and width of O-H peak (3,409.69 cm⁻¹), C-H peak (2,872.13 cm⁻¹), and C-O peak (1,108 cm⁻¹) were due to the presence of PEG. Indeed, the peaks at 1,250, 1,298 and 1,351 cm⁻¹ were due to C-H scissoring and bending in PEG (Figure 1).

TGA measurements provided further evidence regarding the content of polymer on the surfaces of CNTs. Figure 2 shows the TGA thermograms of O-CNT and O-CNT-PEG. Wrapped PEG started to thermally degrade at 330°C, and at 450°C decomposed completely. The plateau in the thermogram of O-CNT-PEG trace after 450°C was attributed to the carbon nanotube. According to the weight loss region, the amount of O-CNT component in the O-CNT-PEG was approximately 20% (w/w).

Morphological changes of functionalized CNTs were assessed by FESEM (Figure 3). It was observed that a
A continuous layer of PEG had been formed on the surface of O-CNTs.

These results indicated that PEG chains were successfully wrapped onto the CNT surfaces. Therefore, the solution of O-CNT-PEG was stable in water without aggregation over long periods of storage (several months).

UV-Vis absorption spectrum of O-CNT-PEG is shown in Figure 4. According to this spectrum curve, the maximum absorption wavelength of O-CNT-PEG was in the range of 670–940 nm. Penetration of light in skin and cells in the NIR spectrum is more convenient than visible spectrum. Therefore, the wavelength of 808 nm was chosen for photoexcitation.

Cell cytotoxicity of O-CNT-PEG

The cytotoxicity profile of MWNT, O-CNT, and O-CNT-PEG against the cell lines was assessed to confirm the modification of CNTs with PEG. For this purpose, the cytotoxicity of MWNT, O-CNT, and O-CNT-PEG was evaluated using the MTT assay. HeLa and HepG2 cells were exposed to that NPs at different concentrations for 24 hours and MTT assay was carried out. As shown in Figure 5A and B, at concentrations of up to 1,000 ng/mL, the O-CNT-PEG NPs showed relatively low toxicity in both cell lines. Presence of PEG around the CNTs increased the cell viability significantly. Oxidation and then coating of MWNTs with PEG increased the cell viability in HepG2 and HeLa cell lines.

PTT of tumors

The in vivo effects of O-CNT-PEG with laser irradiation on the subcutaneously implanted murine melanoma were evaluated by monitoring the tumor size.

Tumor sizes were recorded before and 3 days after the irradiation. Data were analyzed, and significant difference was seen between groups I and II. The shrinking of tumor size in O-CNT-PEG group is clearly evident in Figure 6. As can be seen from this figure, the tumor size before and after the treatment in each mouse was recorded. In the control group, it is clear that the tumor growth was faster where the initial size was greater, but in the O-CNT-PEG group the tumor size was decreased in all cases. By using O-CNT-PEG...
NPs in laser therapy, the large tumor (975 mm$^3$) shrank to a small size (125 mm$^3$), while in the control group the mouse with a tumor of 1,080 mm$^3$ died before day 3 of the study. In the laser therapy group, the tumors grew at a slower rate, and in two cases the size of the tumors decreased. This indicates that the average size of tumor before and 3 days after the treatment was increased in the control group (from 406 mm$^3$ to 745.31 mm$^3$, respectively), but these values decreased in the O-CNT-PEG group (from 566.4 mm$^3$ to 174.69 mm$^3$, respectively). The $P$-value between all groups was significantly different after treatment.

The slope of tumor size against time in the control, laser therapy, and O-CNT-PEG groups was about $+113.10$ mm$^3$/d, $+14.47$ mm$^3$/d, and $-130.57$ mm$^3$/d, respectively (+ indicates increasing and – indicates decreasing of tumor size). The slope of tumor size reduction in O-CNT-PEG group is remarkable. Additionally, ultrasound images were taken to define the depth of tumors in different groups. The stages of tumor treatment in O-CNT-PEG group is clear in Figure 7.

Histopathologic evaluation was performed for professional scrutiny. Gross evaluation of tumors indicated severe shrinkage of tumor size in O-CNT-PEG group in comparison with control and laser therapy groups. Microscopic evaluation showed the presence of nodular subtype malignant melanoma in all cases. Necrosis was found to be the most important discriminator between the cases, and its percentage was higher in cases allocated in O-CNT-PEG group compared to control and laser therapy groups. In the O-CNT-PEG group, a direct association was seen with the site of necrosis and deposition of NPs, which is shown in Figure 8. Mitosis was higher in the control cases while compared with the other cases. There is no evidence of regressive fibrosis, lymphocytic infiltration, vascular invasion, neurotropism, ulceration, and microsatellites in cases. The results are detailed in Table 1.

**Discussion**

CNTs can absorb NIR light and convert its energy into heat effectively. In this study, we used these NPs to evaluate their ability for tumor size reduction by PTT technique. At first, for improving the dispersibility of MWNTs in water, the hydrophilic polymer PEG was used to wrap the surface of the nanotubes. Modified MWNTs were characterized, and the formation of a thin layer of PEG around the nanotubes was established. Results of TGA showed that the amount of PEG component in the O-CNT-PEG was approximately 80%
Cell cytotoxicity study showed that O-CNT was less cytotoxic than pristine MWNTs, and O-CNT-PEG had the lowest toxicity against HeLa and HepG2 cell lines. The cell viability after exposure to different NPs in HeLa cell line at all concentrations was significantly different \((P<0.05)\). But in the HepG2 cell line, the cytotoxicity of O-CNT-PEG and O-CNT in some concentrations (500 ng/mL and 1,000 ng/mL) was not significantly different \((P>0.05)\). The sensitivity of different cell lines to the same materials is different, and the concentration and duration of exposure time to the materials are also important factors for toxicity evaluation.

Melanoma was induced in mice by injection of B16/F10 cell line. For evaluation the effect of ablation with O-CNT-PEG and laser irradiation, the tumor area was irradiated by an 808 nm CW NIR laser diode after the injection of O-CNT-PEG into the tumor. The average size of tumor in mice receiving O-CNT-PEG decreased sharply in comparison with the laser therapy alone.

Pristine, as-produced CNTs have hydrophobic surface and tend to bundle up. So, these NPs are insoluble in most solvents and biological medium.\(^4\) Besides, presence of residual metal catalysts and also the insolubility of pristine CNTs cause they show more toxic effects than functionalized form of them.\(^4\) Surfactants, nucleic acids, peptides, and polymers are used for functionalization of CNTs.\(^4\) Functionalization of CNTs was done through two main approaches: noncovalent and covalent functionalization. In noncovalent functionalization, the electronic structure of the nanotubes was preserved. Indeed, functionalization through this method is often simple and reproducible. So, many researchers have recommended this approach for modification the surface of CNTs.\(^4\) Our strategy in this

![Figure 5](https://www.dovepress.com/)

**Figure 5** MTT assay of MWNT, O-CNT, and O-CNT-PEG against HepG2 (A) and HeLa (B) cell lines.

**Abbreviations:** Conc, concentration; MWNT, multiwalled carbon nanotube; O-CNT, oxidized carbon nanotube; Peg, polyethylene glycol.

![Figure 6](https://www.dovepress.com/)

**Figure 6** The size of tumors before and 3 days after the treatment with PTT in different groups.

**Abbreviations:** O-CNT, oxidized carbon nanotube; Peg, polyethylene glycol; PTT, photothermal therapy.
Photothermal therapy of melanoma tumor

A study involved the generation of dispersed MWNTs by using PEG wrapping. After oxidation of MWNTs, carboxylic acid and hydroxyl groups formed at the side and at the end of nanotubes. These functional groups can interact with PEG through hydrogenic bonds, and so a thin layer of polymer was formed around the surface of MWNTs. PEG is a hydrophilic polymer which can improve water dispersibility of MWNTs. In addition, coating NPs with this biocompatible and biodegradable polymer could enhance the localization of NPs in tumor region by the enhanced permeation and retention effect. The results of cytotoxicity evaluation of MWNTs showed that oxidation and then functionalization of MWNTs with PEG reduced the cytotoxicity of the NTs against two cell lines (Figure 5). Oxidation removes residual metal catalysts, and wrapping PEG around the side of nanotubes improves the water dispersibility.

CNTs can be used as plasmonic NPs that can absorb NIR light and convert its energy to heat. This heat can destroy the tumor cells, which are more sensitive to elevated temperature (hyperthermia) than normal cells. Hyperthermia also increases the vascular permeability of tumor compared with normal vasculature, which can facilitate the delivery of anticancer drugs into tumors. Therefore, localization of plasmonic NPs in the tumor region can be an efficient strategy for treatment of solid tumors. In this in vivo ablation experiment, O-CNT-PEG was injected into the melanoma tumor, and then the solid tumor region was irradiated. The rate of...
Table 1 Results of histopathologic evaluation

<table>
<thead>
<tr>
<th>Groups</th>
<th>Necrosis (%)</th>
<th>Breslow’s thickness</th>
<th>Tumor stage after treatment according AJCC 2010</th>
</tr>
</thead>
<tbody>
<tr>
<td>O-CNT-PEG</td>
<td>More than 90%</td>
<td>About 3 mm</td>
<td>IIA</td>
</tr>
<tr>
<td>Laser therapy</td>
<td>About 25%</td>
<td>&gt;4 mm</td>
<td>IIB</td>
</tr>
<tr>
<td>Control</td>
<td>Less than 5%</td>
<td>&gt;4 mm</td>
<td>IIB</td>
</tr>
</tbody>
</table>

Abbreviations: O-CNT, oxidized carbon nanotube; PEG, polyethylene glycol.

References