Does intraperitoneal injection of propofol prior to detorsion improve testes weight and histopathological findings in a rat model?

Objectives: To determine the long-term preventive effects of intraperitoneal propofol on testicular ischemia–reperfusion injury in a rat model.

Materials and methods: Forty adult male albino Wistar rats were divided randomly into the following four groups according to the planned treatment (n=10 per group): group I, control; group II, sham-operated; group III, torsion/detorsion (T/D); and group IV, T/D plus propofol. Testicular ischemia was achieved by twisting the left testis 720° clockwise (ie, applying torsion) for 1 h. In the T/D plus propofol group (group IV), 50 mg/kg propofol was administered intraperitoneally 30 minutes before detorsion. Ipsilateral orchiectomy was performed under general anesthesia to determine the mean testicular weight and to enable histopathological examination of the testes using Johnsen’s mean testicular biopsy score 30 days after the surgical procedure in all groups.

Results: The testicular weights in groups I, II, III, and IV were 1.65±0.32, 1.59±0.33, 1.11±0.56, and 1.08±0.50 g (mean±SD), respectively. Testicular weight was significantly lower in the T/D groups (III and IV) than in both the control and sham-operated groups (I and II), but there was no improvement in testicular weight as a result of propofol administration. Similarly, Johnsen’s mean testicular biopsy score was lower in groups III and IV than in groups I and II, but no positive effect was conferred by the administration of propofol in group IV.

Conclusion: The use of propofol in the treatment of testicular ischemia–reperfusion injury caused by testis torsion has no significant long-term therapeutic potential.

Keywords: testis, reperfusion, injury, propofol, rats

Introduction

Testicular torsion is a surgical emergency that mainly affects newborns, children, and adolescent boys. The prevalence of torsion in the USA is reportedly 8.6 per 100,000 males aged 10–19 years per year. The direct precipitating events are unknown, but may include undescended testis, cold temperature, and trauma or rapid growth at puberty, with some evidence for a familial predisposition. Previous studies have demonstrated a clear correlation between the duration of torsion and subsequent testicular function and suggest that at least partial testicular atrophy is not rare in patients who undergo detorsion as little as 4 hours after the onset of pain. Therefore, surgical detorsion should be performed as early as possible.

While the mechanism underlying testicular tissue damage during torsion is not completely understood, current studies have demonstrated that the primary pathophysiological event is ischemia followed by reperfusion injury, which results in the production of abun-
Like possibly due to its phenolic hydroxyl group, which acts as an applied anesthetic agent that exhibits potent antioxidant activity. Several antioxidants or anti-inflammatory drugs such as allopurinol, caffeic acid phenyl ester, dehydroepiandrosterone, erydosteine, ibuprofen, morphine, N-acetylcyesteine, propofol, resveratrol, and selenium have been investigated with a view to preventing ischemia–reperfusion injury in the testis. Propofol (2, 6-diisopropylphenol) is a widely applied anesthetic agent that exhibits potent antioxidant activity against lipid peroxidation both in vitro and in vivo, possibly due to its phenolic hydroxyl group, which acts like α-tocopherol (vitamin E). The antioxidant activity of propofol has been demonstrated experimentally, both chemically and histologically, in the rat cerebrum, heart, intestinal mucosa, and joints.

Several attempts have been made within the last few years to determine the effect of antioxidant agents on the outcome of spermatogenesis and fertility during torsion, but to the best of our knowledge little has been published about this subject, especially with regard to the long-term preventive effects of adjunctive antioxidant therapies such as propofol on spermatogenesis. Additional clinical data are thus needed.

The aim of the present experimental study was to determine whether propofol protects spermatogenesis from ischemia–reperfusion injury in rats.

Materials and methods

This prospective experimental study was approved by the Urology and Nephrology Research Research Center, Hamadan University of Medical Sciences, Hamadan, Iran, and by the Chancellor of Research and Technology of Hamadan University of Medical Sciences and was conducted at an animal laboratory between December 2013 and November 2014.

The study was designed so as to minimize the number of rats required for the examination. Therefore, 40 mature, male albino Wistar rats weighing 200–235 g and aged 70–90 days were selected randomly from the animal laboratory and housed under standard conditions in individual cages and in a temperature- and light-controlled room with free access to sterile animal food and water. For the 12 hours immediately preceding the study, they were given only water. The experiments were performed after receiving ethical approval from the Animal Care and Ethics Committee of Hamadan University of Medical Sciences, in compliance with the principles of laboratory animal care (based on EU Directive 2010/63/EU for animal experiments).

Study protocol and groups

The rats were randomly divided into the following four groups of 10 animals each according to the planned treatment, prior to which they were anesthetized with an intraperitoneal injection of 50 mg/kg ketamine hydrochloride (50 mg/mL Ketamax, Rotexmedica, Trittau, Germany):

Group I (control group): required to determine the baseline values of histopathological parameters.

Group II (sham-operated): used to investigate the effect of surgical stress on spermatogenesis. The left testes were extracted through an ilioinguinal incision and then replaced by fixation to the scrotum without torsion.

Group III (torsion/detorsion [T/D]): the left testes were extracted through an ilioinguinal incision, and torsion was created by rotating them by 720º in a clockwise direction, which was maintained by fixing them to the scrotum with a Prolene (Supa Medical Devices Company, Tehran, Iran) suture through the tunica albuginea, as described by Turner et al. The suture was removed after 1 hour of torsion, and the left testes was untwisted and replaced into the scrotum for reperfusion (ie, detorsion). Orchiopexy was performed to prevent any new torsion during the assigned recovery period prior to final surgery.

Group IV (T/D plus propofol): the procedure was the same as for group III, but with the addition of an intraperitoneal injection of 50 mg/kg propofol (Propofol 1% Fresenius, Fresenius Kabi, Uppsala, Sweden) 30 minutes before detorsion. The propofol dose and administration method were adopted from the study by Ünsal et al.

In all groups, operations were performed under sterile conditions with the skin of the scrotal area shaved and prepared with 10% povidone–iodine solution before the operation. The scrotum was entered through a scrotal midline incision. The tunica vaginalis was opened, and the testes were delivered to the surgical field. After surgical intervention, the incision was closed using 4-0 silk sutures. After a brief recovery period from the anesthesia following the end of the experiment, the animals were allowed to consume food and water.

In all groups, ipsilateral orchiectomy was performed 30 days after the surgical procedure (or sham surgery) and reperfusion to determine mean testicular weights and enable histopathological examination of the testicular tissue using Johnsen’s mean testicular biopsy score (MTBS).
Histological analysis
All the rats were killed by cervical dislocation and the left testes was harvested 30 days after surgery (and reperfusion). All testes were weighed and evaluated with the aid of a light microscope (Nikon E200, Nikon Instruments Inc., Melville, NY, USA) by a pathologist who was blinded as to the treatment group from which they had been harvested. The testicular specimens were individually immersed in Bouin’s fixative, dehydrated in alcohol, and then embedded in paraffin. The tissue blocks were sectioned at a thickness of 4–5 µm, deparaffinized, and then stained with hematoxylin and eosin.24 Spermatogenesis was assessed histopathologically using Johnsen’s MTBS.25 About 100 seminiferous tubules were evaluated for each testis, and a score of 1–10 was assigned to each tubule according to the degree of cell maturation under Johnsen’s classification system, as follows: complete spermatogenesis, 10; germinal cell disorganization, 9; few spermatooza, 8; no spermatozoa, 7; few spermatids, 6; no spermatids, 5; few spermatocytes, 4; spermatagonia only, 3; Sertoli cells only, 2; and no cells in the tubules, 1. Finally, the average MTBS was calculated for all tubules assessed in each testis. The groups were compared with each other according to testis weight and Johnsen’s MTBS.

Statistical analysis
The data are reported as either mean ± SD values (for quantitative variables) or percentages (for categorical variables). The groups were compared using the Mann–Whitney U and Spearman correlations coefficient tests. Statistical software SPSS, version 14.0, for Windows (SPSS, Chicago, IL, USA) was used for all statistical analyses. The threshold for statistical significance was set at p<0.05.

Results
Forty male rats were evaluated in this prospective, randomized, experimental study, and all except three in group III (which were substituted) survived the follow-up period of 30 days after testicular T/D. The testicular weights in groups I, II, III, and IV were 1.65 ±0.32, 1.59 ±0.33, 1.11 ±0.56, and 1.08 ±0.50 g, respectively, and were significantly lower in the T/D groups (ie, III and IV) than in the non-T/D groups (I and II; p=0.009). The mean testicular weight did not differ significantly between groups III and IV (p=0.912), indicating that there was no improvement in testicular weight with propofol administration, and it also did not differ significantly between groups I and II (p=0.739). The testicular weights comparison between the four groups is shown in Table 1.

Similarly, the MTBSs were significantly lower in the T/D groups than in the control and sham-operated groups (p=0.000), but no positive effect was conferred by the administration of propofol (ie, group III vs group IV, p=0.315). Comparison of Johnsen’s MTBSs for the various groups is shown in Table 2. There was no significant correlation between mean testicular weight and Johnsen’s MTBS (p=0.050); on the other hand, in the presence of reducing Johnsen’s MTBS, testicular weight was not necessarily reduced.

Discussion
Spermatic cord torsion is a urological emergency, and failure to diagnose the condition or late treatment may cause permanent testicular damage.26 Although the mechanism underlying testicular tissue damage during torsion is not completely understood, studies conducted thus far have demonstrated that the primary pathophysiological event is ischemia–reperfusion injury1,4,7 resulting in the production of abundant amounts of nitrogen and ROS in the reperfusion period due to a considerable oxygen supply.1 ROS are usually eliminated by antioxidants, but in the case of excessive ROS production or failure of the cellular antioxidant defense system, exogenous antioxidants are potentially beneficial for neutralizing ROS.6 Several antioxidants or anti-inflammatory drugs, including propofol, have been investigated with a view to preventing ischemia–reperfusion injury in the testis.1,6,10,11,12 Propofol has exhibited potent antioxidant activity against lipid peroxidation, as demonstrated in the rat, possibly due to its phenolic hydroxyl group, which acts like α-tocopherol.13 As mentioned previously, evidence for this antioxidant activity in the rat
cerebrum, heart, intestinal mucosa and joints has already been published.9–10,14–21

Despite various attempts to determine the benefits of exogenous antioxidants with respect to the outcome of spermatogenesis following testicular torsion, there is a dearth of literature on the long-term preventive effects of adjunctive antioxidant therapies such as propofol on spermatogenesis. Therefore, additional clinical data are needed.

Taşkara et al27 investigated the long-term preventive effect of propofol on testicular ischemia–reperfusion injury in a rat model and found no apparent long-term therapeutic potential of this antioxidant in this context, despite the demonstrated short-term beneficial effects. Ünsal et al28 evaluated the attenuating effect of propofol on reperfusion injury after testicular T/D and concluded that the preference for propofol for anesthesia during the detorsion procedure may attenuate reperfusion injury. Finally, Yagmurdu et al29 performed two studies with an animal model of T/D to establish the mechanisms underlying the preventive effects of intravenous anesthetics and the preventive effects of propofol as an anesthetic agent on testicular ischemia–reperfusion injury.9 They concluded that as an anesthetic agent, propofol may attenuate germ-cell-specific apoptosis by decreasing nitric oxide biosynthesis and scavenging ROS and by inhibiting lipid peroxidation in this animal model.

Propofol was administered intraperitoneally as an antioxidant agent in male rats in the present prospective, randomized, experimental study in order to determine whether propofol protects spermatogenesis from ischemia–reperfusion injury. The data show that the ischemic injury produced by 1 hour of 720º torsion of the spermatic cord resulted in significant atrophy and histological testicular failure, which are the known results usually observed with ischemia-related injury, suggesting the presence of irreversible injury to tissue viability over the long term. The reduction in testis weight and the observed histopathological testicular stress in the T/D groups suggest that torsion exerts a direct toxic effect on spermatogenesis and Johnsen’s MTBSs. Nevertheless, propofol administration at the applied dose (50 mg/kg) and via the suggested route (intraperitoneal) cannot prevent ischemia–reperfusion-related injury to the testicular tissue; on the other hand, this method also did not result in significant rescue of testicular function by preserving the intact seminiferous tubular morphology in the testis.

Study limitations
This study was subject to some limitations that should be noted. The main limitation is that no biochemical measurements of chemical changes such as oxidative stress, oxidative damage, androgen levels, or immunological changes were made. In addition, the role of ketamine hydrochloride as a confounding factor may need to be evaluated in future studies. Finally, another different group (only propofol) for this study to evaluate the effect of propofol on testicular tissue and spermatogenesis was not considered.

Recommendations
The findings of the present study indicate that further investigations need to be performed, including assessments of the degree of vascular insufficiency, the use of pharmacological agents to evaluate the short- and long-term effects of oxidative stress, and the effects of torsion on long-term fertility.

Conclusion
The results of this study show that propofol has no significant long-term therapeutic benefits in the treatment of testicular ischemia–reperfusion injury caused by testis torsion. However, further evaluation of this issue is needed.

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Disclosure
All authors are employees of Hamadan University of Medical Sciences. The authors report no other conflicts of interest in this work.

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