Increase and regulation of synovial calcitonin gene-related peptide expression in patients with painful knee osteoarthritis

Shotaro Takano*  
Kentaro Uchida*  
Gen Inoue  
Atsushi Minatani  
Masayuki Miyagi  
Jun Aikawa  
Dai Iwase  
Kenji Onuma  
Manabu Mukai  
Masashi Takaso

Department of Orthopedic Surgery, Kitasato University School of Medicine, Sagamihara, Japan

*These authors contributed equally to this work

Background: Recent studies suggest that the vasodilatory neuropeptide calcitonin gene-related peptide (CGRP) is localized in the synovial tissue and may be involved in the pathology of hip and knee osteoarthritis (OA). However, the regulation and relationship between pain and CGRP expression levels in the synovial tissue of human OA patients are not fully understood.

Methods: Synovial tissues were harvested from 74 participants with radiographic knee OA (unilateral Kellgren/Lawrence grades 3–4) during total knee arthroplasty. CGRP-expressing cells in the resected tissue were identified by immunohistochemical analyses. To examine CGRP expression levels, CD14-positive (CD14+) (macrophage-rich cell fraction) and CD14-negative (CD14−; fibroblast-rich cell fraction) were isolated from the synovial tissue. To investigate the involvement of prostaglandin E2 (PGE2) in the regulation of CGRP expression, cultured CD14− and CD14+ cells were stimulated with PGE2. In addition, CGRP expression levels in the synovial tissue of OA patients with strong/severe (visual analog scale [VAS] ≥6) and mild/moderate pain (VAS <6) were compared.

Results: CGRP-positive cells were detected in the intimal lining layer and comprised both CD14− and CD14+ cells. CGRP expression in non-cultured CD14− fractions was significantly higher than that in CD14+ fractions. The expression levels of CGRP were significantly increased in cultured CD14− cell fractions treated with exogenous PGE2, compared to untreated CD14− cell fractions. In contrast, treatment with PGE2 did not increase CGRP regardless of whether or not CD14+ cells expressed CGRP. Furthermore, CGRP expression in the VAS ≥6 group was also significantly higher than that in the VAS <6 group.

Conclusion: These findings suggest that CGRP expression in the synovial fibroblasts is regulated by the COX-2/PGE2 pathway and that elevation of synovial CGRP levels may contribute to OA pain.

Keywords: calcitonin gene-related peptide, knee osteoarthritis, regulation, synovium, pain

Introduction

Patients with knee osteoarthritis (OA) typically present with joint pain, which is typically managed with nonsteroidal anti-inflammatory drugs and opioid analgesics. However, as these medications are associated with gastrointestinal and cardiorenal toxicity and are ineffective in certain patients, the development of specific and efficacious drugs for OA pain management represents an unmet clinical need.

Calcitonin gene-related peptide (CGRP) is a 37-amino-acid vasodilatory neuropeptide that binds to the calcitonin receptor-like receptor (CLR) and receptor activity-modifying protein 1 (RAMP1). Recent evidence suggests that CGRP is localized to the synovial tissue and is involved in the pathology of hip and knee arthritis.
Specifically, CGRP was detected in the synovial lining layer of patients with developmental dysplasia of the hip, and it stimulated inflammatory cytokine expression in cultured synoviocytes.\(^7\) CGRP expression in synoviocytes from OA patients was increased by prostaglandin E2 (PGE2).\(^6\) In addition to fibroblast-like synoviocytes, the synovial lining layer also contains macrophage-like cells, in which levels of CGRP expression have not been determined.

An association between OA pain and CGRP protein levels in several joint tissues from humans has been reported.\(^3,4\) Notably, CGRP-positive (CGRP+) nerve fibers are increased in the outer region of the menisci in patients with knee OA\(^7\) and in the joint capsule of those with painful hip OA.\(^8\) Moreover, CGRP expression is also increased in the synovial tissue of OA patients;\(^5,9\) however, the relationship between pain and CGRP expression in synovial tissue is not well understood in human OA.

Here, we investigated the localization and regulatory mechanisms of CGRP in the knee synovial tissue of patients with different degrees of OA pain.

**Materials and methods**

**Patients**

A total of 74 participants with radiographic knee OA (unilateral Kellgren/Lawrence grades 3 [28/74, 38%] and 4 [46/74, 62%]) underwent total knee arthroplasty at our institution. The study population comprised 15 men and 59 women aged 50–88 (mean ± standard error [SE] body mass index (BMI) of 26.2±0.5 kg/m\(^2\) (range 18.2–36.7) kg/m\(^2\)). Samples of synovial tissue were harvested from each operated knee during the total knee arthroplasty. A portion of each synovial tissue sample was immediately fixed synovial tissue samples were embedded in paraffin and sliced into 3-µm thick sections. The sections were deparaffinized with xylene for 1 h, hydrated in serial dilutions of ethanol (100%, 95%, and 70%) and then rinsed in distilled water. The sections were then treated for 6 h at 4°C with rabbit polyclonal primary antibody against CD14 (Abcam, Cambridge, UK) and mouse monoclonal primary antibody against CGRP (Abcam). The sections were further incubated for 1 h at room temperature with the secondary antibodies Alexa Fluor 488 goat anti-rabbit IgG and Alexa Fluor 594 goat anti-mouse IgG (Thermo Fisher Scientific, Waltham, MA, USA).

**Real-time (RT)-PCR analysis**

Total RNA extraction from 74 synovial tissue and cDNA synthesis were performed as described previously.\(^6\) The sequences of the PCR primer pairs used for RT-PCR analysis are listed in Table 2. The specificity of the amplified products was examined by melt curve analysis. Quantitative PCR was performed using a Real-Time PCR Detection System (CFX-96; Bio-Rad, Hercules, CA, USA) to determine the relative mRNA expression levels of CGRP in strong/severe (VAS≥6; n=30) and mild/moderate pain (VAS<6; n=44) groups. CGRP mRNA expression was normalized to the levels of GAPDH mRNA.

**Isolation of CD14+ and CD14− cells**

To investigate the involvement of PGE2 in the regulation of CGRP expression, synovial fibroblasts and macrophages were harvested from the synovial tissue collected from the knees of 10 OA patients. The synovial tissue was digested with 40 mL of 0.1% type I collagenase to release mononuclear cells, which were then suspended in 500 µL phosphate-buffered saline (PBS) containing biotinylated anti-CD14 antibody (Biolegend, San Diego, CA, USA). After incubation for 30 min at 4°C, the cells were washed once with PBS, mixed with streptavidin-labeled magnetic particles (BD IMag Streptavidin Particles Plus-DM; BD Biosciences, San Jose, CA, USA), and subjected to magnetic separation using an IMag

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**Table 1 Clinical characteristics of patients (VAS<6 and VAS≥6 groups)**

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>VAS&lt;6 (n=30)</th>
<th>VAS≥6 (n=44)</th>
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<tbody>
<tr>
<td>Age (years), mean ± standard error</td>
<td>75.5±1.2</td>
<td>72.8±1.3</td>
</tr>
<tr>
<td>Male/female (n)</td>
<td>9/21</td>
<td>6/38</td>
</tr>
<tr>
<td>BMI (kg/m(^2)), mean ± standard error</td>
<td>25.7±0.7</td>
<td>26.4±0.7</td>
</tr>
<tr>
<td>Kellgren/Lawrence grades 3/4 patients (n)</td>
<td>10/20</td>
<td>17/27</td>
</tr>
<tr>
<td>VAS (cm), mean ± standard error</td>
<td>3.6±0.3</td>
<td>8.4±0.2</td>
</tr>
</tbody>
</table>

**Abbreviations:** VAS, visual analog scale; BMI, body mass index.
separation system (BD Biosciences) to isolate labeled cells. After an 8-min incubation at room temperature, unbound (CD14–) cells were collected by the addition of warmed (37°C) α-minimum essential culture medium (α-MEM) to the cell suspension. The tube containing bound cells was removed from the magnetic support, and CD14+ cells were collected by the addition of 3 mL α-MEM. The CD14+ and CD14− cell fractions were centrifuged at 300 g for 10 min. CD14+ and CD14− cell fractions from five patients were directly used for quantitative PCR analysis of CGRP, whereas those from the other five patients were cultured in six-well plates containing α-MEM. After 7 days of culture, the cells were stimulated with PGE2 (10 μM) for 24 h. α-MEM containing no PGE2 was added to vehicle control cells and incubated for 24 h. The treated and control cells were harvested for RNA isolation and CGRP expression, which was performed by RT-PCR analysis.

Statistical analysis
The Mann–Whitney U test was used to compare the differences between CD14− and CD14+ cells, vehicle control and PGE2-treated cells, and CGRP expression in the strong/severe (VAS≥6) and mild/moderate pain (VAS<6) groups. A p-value of <0.05 was considered statistically significant for all analyses. All statistical analyses were performed using SPSS software (v. 19.0; SPSS, Chicago, IL, USA).

Results
Localization and expression of CGRP in synovial tissue of OA patients
To determine the cell type(s) that expresses CGRP in the synovial lining layer of knees in OA patients, immunohistochemical analysis of CGRP and CD14 was performed (Figure 1A–E). CD14+ cells (macrophages) were localized to the intimal lining layer (Figure 1B). Furthermore, CD14+ cells were also observed in the intimal lining layer and corresponded to both CD14+ and CD14− cells (fibroblasts; Figure 1C–E). However, RT-PCR analysis revealed that CGRP expression in CD14− fractions was significantly higher than that in CD14+ fractions (Figure 1F, p=0.020)

Effects of PGE2 on CGRP expression in cultured synovial fibroblasts and macrophages
CGRP expression in cultured cells isolated from synovial tissue was measured after treatment with or without exogenous PGE2. CD14 mRNA expression in CD14+ cells was significantly higher than that in CD14− cells (Figure 2A, p=0.016). CGRP expression increased significantly in CD14− cell fractions treated with PGE2, compared to vehicle control CD14− cells (Figure 2B, p=0.008). In contrast, treatment with PGE2 did not increase CGRP regardless of whether or not CD14+ cells expressed CGRP (Figure 2B, p=1.000).

CGRP expression in the VAS<6 and VAS≥6 groups
To investigate the relationship between synovial CGRP expression levels and pain, the OA patients were separated into strong/severe (VAS≥6) and mild/moderate pain (VAS<6) groups. There were no differences between the two groups in patient age, male/female ratio, BMI, or KL 3/4 ratio (Table 1). RT-PCR analysis indicated that CGRP expression in the synovial tissue of the VAS≥6 group was significantly higher than that in the VAS<6 group (Figure 3, p=0.035).

Discussion
In the synovial tissue of patients with knee OA, CD14− cells in the synovial lining layer highly expressed CGRP as compared to CD14+ cells. The treatment of CD14− cells with PGE2 stimulated CGRP expression. In addition, CGRP expression in the VAS≥6 group of OA patients was significantly higher than that in the VAS<6 group. Taken together, these findings suggest that synovial fibroblast-derived CGRP in the lining layer may be involved in mediating joint pain in OA.

Synovial tissue contains macrophage- and fibroblast-like cells in the lining layer. Previous studies reported that CGRP+ cells are present in the synovial lining layer of developmental dysplasia of the hip and OA patients; however, cell types that expressed CGRP were not determined. Here, double immunostaining of synovial tissue for CGRP and the synovial macrophage marker, CD14, revealed that CGRP+ cells corresponded to CD14− and CD14+ cells, although CGRP expression in CD14− cells was significantly higher than in CD14+ cells. This suggests that synovial fibroblasts are one of the main sources of CGRP production in synovial tissues.

CGRP expression is regulated by inflammatory cytokines, growth factors and PGE2 in several cell types, such as immune, epithelial, neural and synovial cells.
Figure 1. CGRP immunolocalization and mRNA expression in synovial tissue.

Notes: Synovial tissue stained with (A) DAPI (nuclei), (B) CD14 or (C) CGRP. (D) Merged image, (E) High-magnification merged image of (D). Scale bars =100 μm. (F) Real-time PCR analysis of CGRP expression in non-cultured CD14− and CD14+ fractions. All data are presented as mean ± standard error (n=5). *p<0.05.

Abbreviations: CGRP, calcitonin gene-related peptide; DAPI, 4′,6-diamidino-2-phenylindole; PCR, polymerase chain reaction.

Figure 2. Effects of PGE2 on CGRP expression in cultured synovial fibroblasts and macrophages.

Notes: Synovial fibroblasts and macrophages were stimulated in vitro with PGE2 (10 μM) or without PGE2 (vehicle) for 24 h prior to the extraction of total RNA for (A) CD14 and (B) CGRP expression analysis. All data are presented as mean ± standard error (n=5). *p<0.05.

Abbreviations: PGE2, prostaglandin E2; CGRP, calcitonin gene-related peptide.
Elevated synovial CGRP expression was associated with increased pain in OA patients with severe/strong pain. The findings from the present study suggest that modifying the regulation of CGRP expression in synovial tissue may be a promising therapeutic strategy for managing OA pain.

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**Disclosure**

The authors report no conflicts of interest regarding this work.

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