Intersession test–retest variability of 10-2 MAIA microperimetry in fixation-threatening glaucoma

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Purpose: To determine the intersession test–retest variability (TRV) of CenterVue Macular Integrity Assessment (MAIA) microperimeter in glaucoma patients with fixation-threatening field defects.

Methods: This is a prospective case–control study of 27 participants consisting of 13 patients with stable primary open-angle glaucoma and 14 control subjects including 5 healthy individuals and 9 retinal patients (5 with non-neovascular age-related macular degeneration and 4 with inherited retinal disease). Each participant underwent three microperimetry tests in one eye at 1-month intervals. Each test used an identical test strategy of 10-2 Cartesian grid and 4-2 staircase algorithm. We investigated TRV by calculating the coefficient of repeatability (CR) for mean sensitivity (MS) and point-wise sensitivity (PWS) for glaucomatous subjects and retinal and normal subjects. 95% confidence intervals (CIs) for CRs were calculated.

Results: There was no significant change in MS, and the median durations of microperimetry sessions were 9′26″, 8′52″, and 8′46″ across the three study visits. The intersession CRs for MS were 1.1, 2.5, and 1.8 dB, and the average CRs for PWS were 3.5, 7.4, and 8.6 dB for healthy controls and retinal and glaucoma patients, respectively. For test loci with 25–34 dB at baseline, CRs for PWS were 8.2 (95% CI: 7.5–8.9) and 4.3 (95% CI: 4.0–4.6) dB for glaucoma and control subjects, respectively.

Conclusion: We found differences in TRV of test loci depending on the baseline sensitivity value. Glaucoma patients had significantly worse TRV for loci that had sensitivity values within the normal range at baseline. The estimated CR has implications for sample size calculation in future glaucoma treatment trials using microperimetry as a clinical endpoint.

Keywords: microperimetry, MAIA perimeter, glaucoma, retinal dystrophy, optic neuropathy

Introduction

Glaucoma management relies critically on the ability of structural and functional assessments to differentiate disease stability from progression. Perimetry is the mainstay of functional assessment because visual acuity loss does not occur until end-stage disease. In some glaucoma patients, conventional perimetry may be unreliable for monitoring glaucomatous field progression because of reduced fixation stability or eccentric fixation in foveal-involving field defect from advanced glaucoma1 or retinal comorbidities such as diabetic maculopathy, retinal vein occlusion, age-related macular degeneration (AMD), or traction maculopathies.2 In such patients, fundus-controlled perimetry, or microperimetry, may offer a more suitable alternative tool for monitoring disease progression.

Microperimetry is an increasingly utilized form of static automated perimetry3 that tracks fundal landmarks to correct rapidly for shifts in gaze and enables manual centration of testing grid at the anatomic fovea. This allows accurate assessment of the central visual field even in the absence of stable and foveal fixation.3 While the...
use of microperimetry in glaucomatous cohorts is far from mainstream practice, it is beginning to make an entry in clinical research. With the increasing use of macular ganglion cell complex thickness to monitor glaucoma progression, microperimetry can also offer an opportunity for investigating and predicting patterns of disease progression based on structure–function correlation. Like any other psychophysical test, the establishment of test–retest variability (TRV) for microperimetry is essential prior to their adoption into clinical glaucoma practice. While TRV of conventional perimetry has been investigated extensively in glaucoma patients, there is limited information on the TRV of microperimetry in a glaucoma population. Previous studies have found differences in the TRV of microperimetry in normal healthy subjects, AMD, Stargardt disease, and X-linked retinoschisis. Therefore, it cannot be assumed that the TRV of microperimetry in glaucoma patients is equivalent to those reported for retinal diseases. Furthermore, prior to its use as a clinical outcome measure in those with glaucoma field defect threatening fixation (encroaching into 10° of eccentricity), it is important to have some information on test–retest parameters in this particular cohort.

Therefore, we conducted a pilot study to assess the difference in the intersession TRV of the CenterVue Macular Integrity Assessment (MAIA) between patients with primary open-angle glaucoma and control cohorts consisting of healthy subjects and patients with retinal dystrophy or macular degeneration.

Methods
Participants
This study was approved by the Human Research Ethics Committee of the Sir Charles Gairdner Hospital (protocol number: 2011-063), registered with the Therapeutic Goods Administration of the Australian Government (CTN: 263/2012) and adhered to the tenets of the Declaration of Helsinki. Written informed consent was obtained from all participants prior to enrolment.

Inclusion criteria for the three cohorts were treated open-angle glaucoma patients with stable visual field defects within the central 10°, patients with stable inherited retinal disease (IRD) or non-neovascular AMD serving as positive controls, and healthy subjects serving as negative controls. All glaucoma and retinal patients were recruited from the clinics at the Lions Eye Institute between September 2011 and September 2013. For glaucoma subjects, the definition of fixation-threatening defect was: a scotoma consisting of sensitivity reductions to probability <1% for all of three or more contiguous points within 10° of fixation, with one or more loci 3° from fixation and the others occupying loci at 9° from fixation tested on Humphrey 24-2 threshold standard automated perimetry tests. The minimum three-point defect was consistent over at least two consecutive field tests. The definition of a stable defect was three consecutive field tests or more following two baseline tests showing no change on the guided progression analysis over a period of 2 years or more. In addition, an experienced glaucoma specialist (WHM) reviewed all field test series to confirm clinical stability. Intraocular pressure (IOP) was not used as part of this definition. Clinical diagnoses of retinal degeneration were confirmed by an experienced retinal specialist (FKC). Normal subjects were recruited from staff or acquaintances of staff of the Lions Eye Institute, and retinal and optic nerve diseases were excluded based on history, ophthalmic examination (FKC), and fundus imaging. Although not representative of the general population or ophthalmic patients, this group is chosen to provide data that are representative of the minimum TRV and highest retinal sensitivity measurements for comparison with the other cohorts.

Exclusion criteria included ocular signs such as intraretinal cysts or subretinal deposits that suggested potential disease progression during the study period, significant media opacity, prior experience with microperimetry, or the inability to give informed consent. Participants with prior experience of performing nonfundus controlled visual field tests were not excluded since all glaucoma patients had previous experience with Humphrey field tests.

The participants were recruited as part of a larger study involving two microperimeters: the MP-1 and the CenterVue MAIA. Here, we will report the results from the 10-2 grid tests of patients with glaucoma, retinal degeneration, and normal subjects from the MAIA microperimeter as a case–control study.

Examination protocol
Participants were examined three times at three separate study visits, 1-month apart (Visits 1, 2, and 3). At each visit, best-corrected visual acuity assessment using the Early Treatment Diabetic Retinopathy Study (ETDRS) chart (Lighthouse International, New York, USA), MAIA microperimetry, and spectral-domain optical coherence tomography (SD-OCT) imaging was performed sequentially. At the baseline visit (Test 1), if both eyes were eligible for the study, one eye was chosen at random to be the study eye using a random number generator (www.random.org). Prior to any slit-lamp examination or imaging of the retina, the study eye was tested on both MAIA and MP-1 microperimeters consecutively. The order of
testing was assigned by a list generated by the online random number generator prior to commencement of the study for all study visits. Microperimetry testing is detailed in the Microperimetry Protocol section. Pupil dilation was not performed. Following completion of microperimetry tests, all participants underwent multimodal imaging using the Spectralis HRA + OCT device (Heidelberg Engineering, Heidelberg, Germany). Infrared reflectance fundus image, macular cube volume scans (covering 30°×25° area at scan separation of 120 µm), and optic disc nerve fiber layer circular scans were acquired. All scans were examined to ensure that there was no intraretinal cystic lesion that may change, and no disease progression had occurred during the three study visits.

**Microperimetry protocol**

Microperimetry was performed using the CenterVue MAIA (CenterVue, Padova, Italy) in a randomized order. Participants were provided with identical instructions on how to perform the assessment at each visit. Although we did not use a practice test prior to the first session, participants were given the opportunity to familiarize themselves with the response trigger. Testing was conducted in a dedicated quiet psychophysics dark room, and the lights were turned off after the nonstudy eye was patched for at least 5 min prior to commencement of testing, and the patient comfortably positioned at the device with chin on the chin rest and finger ready to press the response trigger.

The fixation target was a 2° diameter red ring broken into four segments: the dim white background had a luminance level of 1.27 cd/m²; the maximum stimulus intensity was 317.5 cd/m², producing a dynamic range of 0–36 dB; stimulus size was Goldmann III; stimulus duration was 200 ms; and testing protocol was 4-2 threshold strategy. The standard 10-2 grid was used, and it consisted of 68 test loci arranged in a Cartesian pattern covering the central 20°.

The MAIA microperimeter has a built-in whole-fundus registration software function that allows follow-up tests to be automatically registered to the baseline (reference) test, thus enabling accurate reassessment of retinal sensitivity at the same test loci examined during the baseline test. In this study, we reported the outcome measures that have been defined previously: mean sensitivity (MS; mean of all 68 loci) and point-wise sensitivity (PWS; individual score of each loci).

**Statistical analysis**

All left eye retinal sensitivity measurements were transposed to corresponding loci in the right eye allowing all data to be analyzed as if they were from the same side. A score of −1 dB was assigned to a retinal sensitivity measurement if the patient did not respond to the brightest stimulus (absolute scotoma). In contrast, a score of 0 dB was assigned to loci at which patient only responded to the brightest stimulus. The dynamic range was 37 dB. A P-value of <0.05 was considered statistically significant.

The Shapiro–Wilk test was used to determine if data were normally distributed. One-way ANOVA with repeated measures was used to ascertain if statistically significant changes in MS had taken place across three study visits. The Friedman test was used to determine if changes had taken place in visual acuity and test duration across three visits. We then calculated intersession coefficients of repeatability (CRs) for each outcome measure based on the formula as outlined by Bland and Altman:

\[
CR = 1.96 \times \sqrt{2 \times s_{w}^2} = 2.77 \times s_{w}
\]

where \(s_{w}^2\) is the mean within-subject variance that can be derived from the sum of squares about the subject mean divided by the degrees of freedom. The 95% confidence interval (CI) for the CR can be calculated based on the assumption that the square root of a chi-squared variable, \(s_{w}\), has an approximately normal distribution with the variance being −1/2. Standard error (SE) of \(s_{w}\) can then be estimated by the following formula based on the number of subjects (n) and number of repetitions (m):

\[
SE = \frac{s_{w}}{\sqrt{2n (m-1)}}
\]

95% CI of \(s_{w}\) = \(s_{w}\) ± 1.96 SE

\[
95\% \text{ CI of } CR = \frac{CR \pm CR \times 1.39}{\sqrt{n(m-1)}}
\]

Bland–Altman plots were used to assess test–retest characteristics across the range of magnitude of outcome measures, and the presence of systematic changes was assessed by calculating the rank correlation coefficient (Kendall τ-b). The average PWS of healthy subjects, retinal patients, and glaucoma patients were determined using a linear mixed-effects model. For this model, visit number was the fixed effect and stimuli points nested within subjects were the random effect. All statistical analyses were performed on the commercially available Statistical Package for Social
Results

Patient characteristics

A total of 27 participants meeting inclusion and exclusion criteria were recruited into the study. There were 13 participants with stable primary open-angle glaucoma and 14 controls. Five of these were healthy subjects, five had non-neovascular AMD, and four had IRD (two subjects with rod-cone dystrophy and two with Stargardt disease). There were 15 females and 12 males with a mean and median (range) age of 59 and 62 (21–86) years, respectively (Table 1). The mean (SD) deviation of 24-2 Humphrey Field Analysis for the glaucomatous cohort was −12.4 (5.4) dB. All participants completed three study visits.

Median (IQR) intervals between both Tests 1 and 2, and Tests 2 and 3 for the entire cohort, were 28 (28–33) and 28 (28–35) days, respectively. There was no significant difference in the intervals between the study visits (paired t-test, \(P=0.71\)). Review of SD-OCT macular cube and retinal nerve fiber layer scans showed no evidence of disease progression in any patient. There was no change in the median (IQR) visual acuities across three study visits for the entire cohort (Friedman test, \(P=0.12\)), and these were 84 (78–86), 83 (79–89), and 85 (80–90) ETDRS letters at Tests 1, 2, and 3, respectively.

Microperimetry outcome

The mean (IQR) duration for each microperimetry examination was 9′26″ (8′54″–10′33″), 8′52″ (8′16″–9′25″) and 8′46″ (8′30″–9′45″) for the three study visits. There was no statistically significant difference in testing duration (Friedman test, \(P=0.36\)). The average MS was 17.6 dB for all three study visits across all subjects. When analyzed by diagnosis, the average MS was 28.0, 27.8, and 28.1 dB for healthy subjects, 14.9, 15.8, and 15.7 dB for retinal patients, and 15.2, 15.2, and 15.0 dB for glaucoma patients, over the three study visits, respectively. A repeated-measures ANOVA with Greenhouse–Geisser correction demonstrated no statistically significant difference in MS over the three test visits for each of these three cohorts: healthy subjects (\(P=0.48\)), retinal patients (\(P=0.05\)), and glaucoma patients (\(P=0.84\)). Therefore, data from all three visits were included in the calculation of CRs.

Intersession TRV

The plot of within-subject SD versus mean for MS, over three test sessions (Figure 1) as outlined by Bland and Altman, demonstrated a moderate positive correlation for glaucomatous subjects (Kendall \(\tau-b; r=0.436, P=0.04\)) and a weak negative correlation for healthy subjects and retinal patients (Kendall \(\tau-b; r=-0.200, P=0.62\) and \(r=-0.111, P=0.68\)).

CRs for MS for normal subjects, retinal patients, and glaucoma patients were 1.1 (95% CI: 0.6–1.6), 2.5 (95% CI: 1.7–3.3), and 1.8 (95% CI: 1.3–2.3) dB, respectively. The average (SD) CRs for PWS (all 68 coefficients) for healthy controls, retinal patients, and glaucomatous patients were 3.5 (1.2), 7.4 (2.9), and 8.6 (4.4) dB, respectively.

We then analyzed the spread of within-subject SD for PWS (Figure 2A and B) and limits of agreement in PWS between second and third testing sessions (Figure 2C and D). The CRs for PWS were further analyzed by stratifying PWS into four categories of retinal thresholds based on first test session result: absolute scotoma (−1 dB), deep scotoma (0–14 dB), shallow scotoma (15–24 dB), and normal threshold (25 dB and above) (Table 2).

Table 1 Demographic and testing characteristics of three study cohorts

<table>
<thead>
<tr>
<th></th>
<th>Glaucoma patients</th>
<th>Control, retinal patients</th>
<th>Control, healthy subjects</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male/female (number of subjects)</td>
<td>5/8</td>
<td>4/5</td>
<td>3/2</td>
</tr>
<tr>
<td>Age (median years, range)</td>
<td>72 (39–83)</td>
<td>62 (30–86)</td>
<td>29 (21–33)</td>
</tr>
<tr>
<td>Visual acuity (median ETDRS letter score, range)</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Session 1</td>
<td>85 (74–93)</td>
<td>77 (10–85)</td>
<td>90 (83–93)</td>
</tr>
<tr>
<td>Session 2</td>
<td>85 (72–94)</td>
<td>82 (20–89)</td>
<td>90 (81–94)</td>
</tr>
<tr>
<td>Session 3</td>
<td>85 (75–95)</td>
<td>80 (19–90)</td>
<td>90 (84–94)</td>
</tr>
<tr>
<td>Testing duration (median minutes, and seconds range)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Session 1</td>
<td>9′31″ (7′2″–11′31″)</td>
<td>9′34″ (7′20″–11′37″)</td>
<td>8′29″ (8′14″–8′58″)</td>
</tr>
<tr>
<td>Session 2</td>
<td>8′49″ (6′46″–13′22″)</td>
<td>9′24″ (8′52″–28′5″)</td>
<td>8′43″ (8′16″–8′48″)</td>
</tr>
<tr>
<td>Session 3</td>
<td>8′34″ (6′56″–11′13″)</td>
<td>8′57″ (8′30″–17′33″)</td>
<td>8′46″ (8′25″–9′6″)</td>
</tr>
<tr>
<td>Mean sensitivity (average retinal sensitivity in dB, range)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Session 1</td>
<td>15.2 (4.8–23.3)</td>
<td>15.0 (−0.6–25.7)</td>
<td>28.0 (26.4–29.3)</td>
</tr>
<tr>
<td>Session 2</td>
<td>15.2 (4.5–21.3)</td>
<td>15.9 (1.8–25.1)</td>
<td>27.8 (26.2–29.0)</td>
</tr>
<tr>
<td>Session 3</td>
<td>15.0 (4.7–22.3)</td>
<td>15.8 (−0.5–24.8)</td>
<td>28.1 (27.0–29.5)</td>
</tr>
</tbody>
</table>

Abbreviation: ETDRS, Early Treatment for Diabetic Retinopathy Study.
Figure 1 A plot of within-subject standard deviation of the mean macular sensitivity against average of the three testing sessions for (A) glaucoma patients and (B) control subjects (including healthy and retinal patients showing weak relationship between variability and magnitude).

Figure 2 A plot of within-subject standard deviation of the point-wise sensitivity against average of the three testing sessions for (A) glaucoma patients and (B) control subjects (including healthy and retinal patients) showed increased variability when sensitivity dropped below 20 dB. Bland–Altman plot of differences against mean between testing sessions 2 and 3 for (C) glaucoma patients and (D) control subjects showed wider limits of agreement in glaucoma patients. Differences in point-wise sensitivity between testing sessions 1, 2, and 3 displayed as thresholds of second and third session against thresholds of first session show the increased spread of differences in (E) glaucoma and (F) control subjects as the baseline threshold decreases.
The variability of retinal threshold at second and third examinations for each level of retinal threshold obtained at the first examination is shown in Figure 2E and F. The spread of retest values increased significantly as retinal thresholds dropped below 25 dB. However, this spread was more marked in glaucoma patients, where the increase in spread was noticeable at retinal thresholds below 30 dB.

### Discussion

We demonstrated no learning effect in microperimetry over three testing sessions. The intersession CRs for MS were 1.1, 2.5, and 1.8 dB, and the average CRs for PWS were 3.5, 7.4, and 8.6 dB for healthy subjects, retinal patients, and glaucoma patients, respectively. Our results suggest that microperimetry reliability in glaucoma patients is worse than healthy control and may be worse than retinal patients in point-wise threshold values.

From our cohort of nine patients with non-neovascular AMD or IRD, we found a much higher CR for both the MS and PWS than that reported previously (Table 3). Notably, there is only one other study that had investigated intersession TRV using the MAIA microperimeter in a cohort of young, healthy normal subjects. However, a CR was not reported in that study. The intrasession CR value reported by Wu et al in a cohort of dry AMD patients for MS and PWS was 1.08 (95% CI: 0.83–1.33) dB and 3.7 (95% CI: 3.4–4.1) dB, respectively. The inclusion criteria for their cohort were drusen ≥125 µm, multiple drusen ≥63 µm with or without pigmented abnormalities, or noncentral geographic atrophy. The difference in severity of disease and testing grid (smaller than 10-2 coverage) and protocol (intrasession instead of intersession) may explain the large differences observed in the estimated CRs between ours and previous studies.

Microperimetry TRV in glaucomatous field defect has been reported recently. One study investigated TRV at the border of deep scotomas using the border of the optic nerve head in healthy individuals as a model of glaucomatous defect and found CR for PWS of ±12.99 dB. Another recent publication investigated TRV of MAIA microperimetry in a cohort of 30 individuals with primary open-angle glaucoma. However, the authors investigated novel parameters in the peripapillary region, and their results are not comparable to the traditional macular outcome measures presented in this study. TRV of conventional standard automated perimetry in glaucomatous subjects is large, particularly with worsening sensitivity. We observed a similar pattern in our cohorts. In both glaucoma and control subjects, we found increased within-subject SD below 20 dB of retinal sensitivity (Figure 2A and B). This concurs with previous findings and carries the implication that areas that are of greatest clinical importance are also the ones in which it is most difficult to determine true change.

To explore the effect of retinal sensitivity on CR of PWS, we stratified all test loci threshold results into four categories: absolute scotoma (~1 dB), deep scotoma (0–14 dB), shallow scotoma (15–24 dB), and normal threshold (25 dB or above). We found that the CR for the lower two categories were comparable between cohorts, as evidenced by the overlapping CIs. In contrast, the upper two categories had significant differences: 9.7 (95% CI: 8.9–10.5) versus 7.1 (95% CI: 6.5–7.8) dB and 8.2 (95% CI: 7.5–8.9) versus 4.3 (95% CI: 4.0–4.6) dB for glaucomatous versus retinal and normal subjects, respectively. The CR for normal threshold loci in retinal and normal subjects was almost half that of normal points in glaucomatous subjects. This observation is also demonstrated in Figure 2E and F, in which the spread of retest values is significantly larger in the glaucomatous cohort even when the baseline values were within the normal range (ie, >24 dB). The unexpected high TRV in regions of normal sensitivity in glaucomatous eyes may be driven by loci at the edge of an absolute glaucomatous defect. A slight misalignment between consecutive microperimetry tests may lead to a test loci falling into an absolute scotoma resulting in more than 25 dB fluctuation. In contrast, steep scotoma within regions of normal sensitivity is unusual in non-neovascular AMD or IRD because the region of atrophy (absolute scotoma) is usually surrounded by a wide penumbra of mild-to-moderate retinal dysfunction. Therefore, threshold profile in retinal degeneration is more forgiving to slight misalignment of testing grid in follow-up examinations.
Table 3 Summary of previously reported values of intersession and intrasession test–retest variability of the MAIA device

<table>
<thead>
<tr>
<th>Author</th>
<th>Cohort demographics</th>
<th>No of subjects</th>
<th>Testing protocol</th>
<th>Microperimetry protocol</th>
<th>Reported measures of repeatability</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wu et al</td>
<td>Dry AMD (69.1±10.5 yo) and normal controls (66.7±6.0 yo)</td>
<td>30 (AMD), 14 (controls)</td>
<td>3 tests at 1 session</td>
<td>MAIA; white-on-white 12° diameter, 37 loci</td>
<td>CR for mean sensitivity (dB): 1.08 (95% CI: 0.83–1.33) (dry AMD)</td>
</tr>
<tr>
<td>Wu et al</td>
<td>Dry AMD (69.6±8.0 yo)</td>
<td>71</td>
<td>2 tests in each eye at 1 session</td>
<td>MAIA; white-on-white 12° diameter, 37 loci</td>
<td>CR for point-wise sensitivity (dB): 4.37 (95% CI: 4.26–4.48)</td>
</tr>
<tr>
<td>Wu et al</td>
<td>Normal subjects (sampled at macula, peripapillary region, and ONH border); 38.4±10.9 yo</td>
<td>30</td>
<td>2 tests at 1 session</td>
<td>MAIA; white-on-white 14° diameter, 2 rows traversing the border of ONH, and 4 loci within the macular region; 4-2</td>
<td>CR for mean sensitivity (dB): 3.8 (macula)</td>
</tr>
<tr>
<td>Molina-Martin et al</td>
<td>Normal subjects; 27.0±8.5 yo</td>
<td>44</td>
<td>3 tests over 3 weeks</td>
<td>MAIA; white-on-white 10° diameter, 37 loci; 4-2</td>
<td>Passing-Bablok regression intercept: 3.66 (95% CI: -9.64 to 13.99)*</td>
</tr>
<tr>
<td>This study</td>
<td>Normal subjects (28.0±4.0 yo)</td>
<td>5</td>
<td>3 tests over 2 months</td>
<td>MAIA; white-on-white 20° grid, 68 loci; 4-2</td>
<td>CR for mean sensitivity (dB): 1.1 (95% CI: 0.6–1.5)</td>
</tr>
<tr>
<td>Primary open-angle glaucoma (67.0±11.0 yo)</td>
<td>9</td>
<td>3 tests over 2 months</td>
<td>MAIA; white-on-white 20° grid, 68 loci; 4-2</td>
<td>CR for mean sensitivity (dB): 2.5 (95% CI: 2.2–2.8)</td>
<td></td>
</tr>
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</table>

Note: *No coefficient of repeatability available.

Abbreviations: AMD, age-related macular degeneration; CR, coefficient of repeatability; dB, decibels; ONH, optic nerve head; SD, standard deviation; yo, years old.

References


Conclusion

This pilot study provided an estimated CR for MAIA microperimetry in glaucoma patients. The estimated CR has implications for sample size calculation in future glaucoma treatment trials using microperimetry as a clinical endpoint. The authors report no conflicts of interest in this work.

Disclosure

The authors report no conflicts of interest in this work.

Variability of MAIA perimetry in glaucoma

There are a number of limitations in this study. First, we used the 10-2 test grid. This may be too coarse a grid to study focal-threatening disease; however, it was chosen because glaucoma specialists are familiar with the grid and it allows large regions of structure–function correlation to be transferred to patients with both types of disease in whom ganglion cell microneurophysiological responses due to photoreceptor versus ganglion cell loss. The sample size was small and the cohorts were not age-matched, reflecting the difficulty in recruiting patients with either retinal degeneration or glaucoma were selected with ganglion cell complex thinning. Second, only patients with either advanced glaucoma or with ganglion cell complex thinning were examined in previous studies. Finally, our study involved only patients with either advanced glaucoma or with ganglion cell complex thinning.

We chose a study period of only 2 months as it was deemed too short a period for true progression to become a significant factor in assessing the TRV of the data. These limitations will need to be addressed by future studies using the pilot data from our study for sample size calculation.

The authors report no conflicts of interest in this work.


