Effect of menopause on hormonal receptors in ampullae of the fallopian tube with a special reference to the p53 signature

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Objectives: Age-related changes in the expression of hormonal receptors have not been well examined in the fallopian tube (FT). We herein report the effect of menopause on the hormone receptors in ampullae of the FTs (AFTs), in comparison with cortical inclusion cysts (CICs) of the ovary.

Methods: A total of 84 AFTs and 16 fimbriae of FTs, which were obtained from 26 premenopausal and 58 postmenopausal women; and 27 postmenopausal CICs were immunohistochemically studied for the expression of p53, Ki-67, estrogen receptor-alpha (ER- α), and progesterone receptor A (PRA). Apoptotic cells were identified using a TUNEL assay.

Results: Postmenopausal AFTs showed a significantly lower labeling index (LI) for Ki-67 ($P < 0.001$), apoptosis ($P = 0.03$), and PRA ($P < 0.001$) than premenopausal AFTs. No significant correlation with immunohistochemical markers was found in premenopausal AFTs, but the LI for PRA was positively correlated with that for Ki-67 ($P = 0.004$) and inversely with that for p53 ($P = 0.023$) in postmenopausal AFTs. The expression of immunohistochemical markers was closely correlated between ampullae and fimbriae of the FT. The p53 signature (p53S) was detected in five postmenopausal AFTs (mean age: 70.2 years) and was not detected in any CICs. The immunohistochemical profile of p53S was low expression of Ki-67, apoptosis, and PRA, and high expression of ER- α . The expression of PRA in CICs was significantly higher than that in AFTs ($P = 0.001$).

Conclusion: The expression of PRA was significantly lower in postmenopausal AFTs than in premenopausal AFTs, whereas the expression of PRA was well preserved in postmenopausal CICs.

Keywords: fallopian tube, progesterone receptor, estrogen receptor, p53 signature, apoptosis

Introduction

Although previous studies have investigated the expression patterns of estrogen receptor (ER) and progesterone receptor (PR) in human endometrium and documented both age-specific patterns of expression^{1,2} and hormone-dependent changes during the menstrual cycle,^{3,4} the expression of hormone receptors has not been well documented in the normal human fallopian tube (FT), especially the age-related expression of hormone receptors.

Recent morphologic and molecular genetic studies of women with a hereditary mutation in *BRCA1* and *BRCA2* have reported new findings regarding the carcinogenesis and histogenesis of pelvic serous carcinoma.⁵ It appears that a significant subset of pelvic serous carcinomas originates from the distal FT rather than the ovarian surface epithelium and/or ovarian cortical inclusion cyst (CIC).⁶ In addition, the p53

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signature (p53S) is associated with serous tubal intraepithelial carcinoma (STIC) and has been reported to be the earliest lesion in the development of tubal serous carcinoma.⁷ Tone et al reported that the gene expression profiles of BRCA1/2-mutated FT during the luteal phase more closely resembled those of high-grade serous carcinomas than those of BRCA1/2-mutated FT during the follicular phase.⁸ They further suggested that studies of the relative PR isoform expression in FT exhibiting p53 (p53S) and STICs should be performed to determine whether or not differential PR isoform expression may be involved in high-grade serous carcinoma development.⁹

In this study, we assessed the immunohistochemical profile of FTs and CICs along with the expression of ER- α , progesterone receptor A (PRA), Ki-67, p53, and apoptosis, and investigated the effect of menopause on hormonal receptors, proliferation, apoptosis, and presence of p53S in FTs in comparison with CICs of the ovary.

Materials and methods

Case selection

Eighty-four specimens of AFTs, obtained by conventional tubal sampling of macroscopically normal FTs, were retrieved from the database of the Pathology Department at the University of Occupational and Environmental Health Hospital between 2011 and 2012.

Among 26 premenopausal women, surgical specimens of AFTs were collected from 8 women with invasive cervical cancer, 7 with ectopic pregnancy, 5 with endometrial cancer, 3 with benign ovarian cyst, and 3 with ovarian cancer.

Among 58 postmenopausal women, surgical specimens of AFT were collected from 25 women with benign ovarian cyst, 18 with endometrial cancer, 6 with cervical cancer or dysplasia, 4 with ovarian cancer, 3 with benign uterine tumor, and 2 with peritoneal cancer. The postmenopausal women aged 60 years or older were included in the present study with a reference to age of menopause in medical records. No woman received hormones for the management of menopausal symptoms.

Sixteen of the 84 FTs, which were obtained from 8 premenopausal and 8 postmenopausal women, were able to be examined for both the ampullae and fimbriae. The CICs obtained from the simultaneously resected ovaries of 27 postmenopausal women were also included in this study.

The study was approved by the Review Board of the University Hospital of Occupational and Environmental Health on Ethical Issues (H27-185). The patients whose specimens were used in this study provided written, informed consent for the future use of the specimens for research.

Immunohistochemistry

For the immunohistochemical analyses, 4- μ m-thick sections were cut from formalin-fixed paraffin-embedded tissue blocks, deparaffinized in xylene, and rehydrated through sequential washes of alcohol and distilled water. Ki-67 and p53 were detected using ready-to-use monoclonal antibodies against Ki-67 and p53 (clones MIB-1 and DO-7, respectively; Dako Cytomation, Kyoto, Japan). ER- α and PRA were detected using monoclonal antibodies (clone 6F11, diluted 1:50, and clone 16, diluted 1:50, respectively; Novocastra, Fukuoka, Japan). ER- α and PRA are essential for the endometrium to proliferate, remodel, and shed in response to estrogen and progesterone.² The immunoreactivity of these antibodies was well established in the literature. The slides were heated in an autoclave at 120°C for 5 min in 0.01 M citrate buffer (pH 6.0) before immunostaining. The slides were incubated with the above-mentioned antibodies for 2 h at room temperature. Antibody binding was visualized using the EnVision+ Dual link system and diaminobenzidine as a chromogen (Dako Cytomation). The slides were counterstained with methyl green or hematoxylin and mounted.

Detection of apoptosis

3'-End-labeling of apoptotic cell DNA was performed using an Apop Tag in situ apoptosis detection kit (Millipore Corporation, Tokyo, Japan) in accordance with the manufacturer's instructions. Deparaffinized sections were pretreated with proteinase K for 15 min at room temperature and treated with methanol containing 3% H₂O₂ for 5 min, for endogenous peroxidase blockage. After applying 75 μ L/5 cm² of equilibration buffer for 30 min at room temperature, 55 μ L/5 cm² of working strength TdT enzyme was pipetted onto sections and incubated in a humidified chamber at 37°C for 1 h. After applying working strength stop/wash buffer for 10 min, 65 μ L/5 cm² anti-digoxigenin peroxidase conjugate was pipetted onto each slide, and the slides were incubated in a humidified chamber for 30 min at room temperature. After washing with phosphate-buffered saline, peroxidase was visualized by diaminobenzidine.

The interpretation of the immunohistochemical preparations was performed and the presence of apoptotic cells was confirmed with the help of the WinROOF image processing software program (Mitani Corp., Tokyo, Japan).¹

The p53 signature

p53S was defined as the presence of endometrial epithelial cells, with either glandular or surface growth patterns, that were morphologically benign in appearance but displayed 12 or more consecutive epithelial cell nuclei with strong p53 immunostaining.⁵

Table 1 Clinicopathological data of the ampullae of the fallopian tube

Variable	Ampullae of the fallopian tube		P-value
	Premenopausal	Postmenopausal	
Number of cases	26	58	
Age (years; mean $\pm$ SD)	37.3 $\pm$ 7.7	71.0 $\pm$ 7.7	<0.001
LI for p53 (%)	2 (0–28)	2 (0–12)	0.163
LI for Ki-67 (%)	6 (1–42)	2 (0–21)	<0.001
LI for apoptosis (%)	1 (0–3)	0 (0–2)	0.03
LI for ER- α (%)	83 (0–96)	88 (0–98)	0.1
LI for PRA (%)	98 (8–99)	12 (0–98)	<0.001

Abbreviations: ER, estrogen receptor; LI, labeling index; PRA, progesterone receptor A; SD, standard deviation.

Statistical analyses

Statistical analyses were carried out using the SPSS software program for Windows, version 22.0.0 (SPSS, Chicago, IL, USA). Data for the clinicopathological factors and immunohistochemical markers were evaluated using the chi-squared test or Mann–Whitney *U*-test. The mean age of the patients was assessed using Student's *t*-test. Spearman's rank correlation coefficient was used to determine the correlations between the immunohistochemical markers. Poisson regression analyses were used to examine the association of p53, ER- α , PRA, and Ki-67 with apoptosis. A value of $P < 0.05$ was considered statistically significant.

Results

Clinicopathological and immunohistochemical findings

The immunohistochemical data for premenopausal and postmenopausal AFTs are presented in Table 1. Although

premenopausal AFTs were collected in women with varied diseases, the labeling index (LI) for ER- α and PRA in premenopausal AFTs was consistently high regardless of the menstrual cycle or ectopic pregnancy (Figure 1). Postmenopausal AFTs showed significantly lower LI for Ki-67 ($P < 0.001$), apoptosis ($P = 0.03$), and PRA ($P < 0.001$) than premenopausal AFTs, whereas the LIs for p53 and ER- α showed no significant difference between premenopausal and postmenopausal AFTs. In the univariate analysis, the LI for apoptosis was inversely correlated with the menopausal status ($P = 0.021$) and significantly correlated with PRA ($P = 0.005$), whereas multivariate analyses showed that no variables were correlated with apoptosis (Table 2). No significant correlation among the immunohistochemical markers was found in premenopausal AFTs, whereas the LI for PRA was significantly correlated with those for Ki-67 ($P = 0.004$, $r = 0.373$) and p53 ($P = 0.023$, $r = -0.298$) in postmenopausal AFTs. An LI for PRA of 90% or more was found in 8 (13.8%) of 58 postmenopausal AFTs and in 24 of 26 (92.3%) premenopausal AFTs. Among 16 FTs examined for both ampullae and fimbriae, the expressions of p53, Ki-67, apoptosis, ER- α , and PRA (Figure 2) were significantly correlated between the ampullae and fimbriae ($r = 0.564$; $P = 0.023$; $r = 0.602$; $P = 0.016$; $r = 0.556$; $P = 0.025$; $r = 0.487$; $P = 0.011$; and $r = 0.956$; $P < 0.001$, respectively).

The immunohistochemical data of the p53S are presented in Table 3. The p53S was positive in 5 (12.7%) of 58 postmenopausal AFTs (Figure 3) and negative in 26 premenopausal AFTs and 16 fimbriae. The mean age of women with p53S positivity was 70.2 years. The immunohistochemical profile of p53S was not markedly different from that of postmenopausal AFTs with a background of p53S.

Figure 1 The ampullae of the fallopian tube in a 32-year-old woman with ectopic pregnancy (A). Moderate-to-strong expression of p53 is shown in the epithelial cells, but the staining pattern is not enough to be labeled p53 signature (B). The apoptotic body (C; arrow) and mild proliferative activity for Ki-67 (D) are shown. Positivity for estrogen receptor- α (E) and progesterone receptor A (F). All figures are presented at 40 $\times$ original magnification.

Table 2 Incidence rate ratios for the expression of apoptotic cells in the ampullae of the fallopian tube

Variable	Univariate			Multivariate		
	IRR	95% CI	P-value	IRR	95% CI	P-value
Menopause	0.51	0.29–0.89	0.021	0.93	0.41–2.08	0.851
p53	1.03	0.99–1.08	0.148	1.02	0.97–1.07	0.392
Ki-67	1.03	1.00–1.06	0.066	1.01	0.97–1.04	0.769
ER- α	1.01	0.99–1.03	0.473	1.01	0.99–1.03	0.475
PRA	1.01	1.00–1.02	0.005	1.01	1.00–1.02	0.068

Abbreviations: CI, confidence interval; ER, estrogen receptor; IRR, incidence rate ratio; PRA, progesterone receptor A.

The mean age $\pm$ standard deviation (SD) of women with CIC was 71.0 ± 7.7 years. Twenty-seven of the CIC specimens were immunohistochemically examined. The median and range of the LI for p53, Ki-67, apoptosis, ER- α , and PRA were 0% and 0%–5%, 2% and 0%–5%, 0% and 0%–1%, 70% and 0%–98%, and 92% and 0%–100%, respectively. Compared with AFTs, CICs had significantly lower LIs for p53 ($P < 0.001$), apoptosis ($P = 0.003$), and ER- α ($P < 0.001$) and higher LI for PRA ($P = 0.001$) (Figure 4). p53S was not detected in 27 CICs.

Discussion

Although we were unable to find any studies of age-related changes in the expression of hormone receptors in the FTs in the literature, the expression of PRA was significantly

decreased in postmenopausal AFTs compared with premenopausal AFTs, whereas the expression of ER- α was well preserved in both premenopausal and postmenopausal AFTs. Regarding the expression of Ki-67 in FTs, younger groups (45–50 years) reportedly showed higher proliferation than older groups (51–55 years), but this difference was not statistically significant.¹⁰ In the present study, postmenopausal AFTs (ranging in age from 60 to 86 years) showed significantly lower LIs for Ki-67 than premenopausal AFTs (ranging in age from 21 to 47 years). The expression of PRA was positively correlated with that of Ki-67 and inversely correlated with that of p53 in postmenopausal AFTs.

Although ER- α was reported not to be detected in the FT of women with ectopic pregnancy,¹¹ the LI for ER- α was consistently high in AFTs of women with ectopic pregnancy in the present study.

Ovarian high-grade serous carcinomas have been well known to show high expression of ER and low expression of PR.^{9,12} The expression of hormone receptors in postmenopausal AFTs had a similar feature as that of hormone receptors of ovarian high-grade serous carcinoma, while that of hormone receptors of CICs had an inverse feature. The strong expression of PR was reported to be a good prognostic indicator in ovarian high-grade serous carcinoma.¹² The increased expression of PR was reported in the CIC of patients with a hereditary predisposition compared with controls.¹³

Figure 2 Ampullae (A) and the fimbriated end (B) show positivity for progesterone receptor A in a 31-year-old woman with cervical carcinoma. Ampullae (C) and the fimbriated end (D) show negativity for progesterone receptor A in an 85-year-old woman with endometrial carcinoma. All figures are presented at 4 $\times$ original magnification.

Table 3 Immunohistochemical profile of the p53 signature in the ampullae of the fallopian tube

Case	Age (years)	Disease	Ki-67 (%)	ER- α (%)	PRA (%)	Apoptosis (%)
1	75	Endometrial cancer	0	100	0	0
2	80	Serous cyst	0	27	0	0
3	64	Mucinous cyst	0	100	2	0
4	77	Endometrial cancer	3	91	15	0
5	76	Endometrial cancer	0	100	0	0

Abbreviations: ER, estrogen receptor; PRA, progesterone receptor A.

Ovarian high-grade serous carcinoma is closely related to advanced age of patients, with a reported mean age of 63 years.⁶ In the nonprophylactic setting, Morrison et al¹⁴ and Gilks et al¹⁵ reported respective mean ages of 62.7 and 57 years among patients with STIC. Under non-neoplastic conditions of the FT, the incidence of STIC was also associated with advanced patient age.¹⁶ STIC is commonly located in the tubal fimbria and is associated with p53S.¹⁷ However, p53S was reported to be equally prevalent in FTs of BRCA mutation carriers and controls, with no particular association with age.^{18,19} p53S of AFTs was only found in elderly women (mean age; 70.2 years) in the present study. We previously examined the overexpression of p53 in normal endometrium,¹ endometrial polyps,²⁰ and the background endometrium of endometrial carcinomas.²¹ Our previous studies showed that endometrial p53S was closely related with advanced patient age, except in patients with Lynch syndrome. The postmenopausal endometrial glands have been consistently reported to be positive for ER- α and PRA.^{1,2} The incidence of p53S (12.7%) in postmenopausal AFTs was almost the same as that of p53S (11.1%) in postmenopausal endometrial

glands.¹ High proliferative activity with low apoptotic cell death was shown in the postmenopausal endometrial glands surrounded p53S,¹ but the immunohistochemical profile of fallopian tubal p53S was not markedly different from that of postmenopausal AFTs on a background of p53S.

A previous study of carcinoma of the FT showed that 92% of the tumors were situated within the tubal portion of the organ, most often in its distal two-thirds, and 8% of the tumors were confined to the fimbriae.²² However, the adoption of the sectioning and extensively examining the fimbriated end (SEE-FIM) protocol have led to increased awareness of tubal fimbriae as a source of high-grade serous carcinomas.²³ The immunohistochemical markers in the present study were unable to reveal a significant difference between the ampullae and fimbriae of the FTs. As such, differences between carcinoma of the ampullae and fimbriae of the FT should be examined in future studies.

Conclusion

The decreased expression of PRA was found in postmenopausal AFTs and was significantly associated with a low

Figure 3 Case 2: The ampullae of the fallopian tube in a 75-year-old woman with ovarian serous cystadenoma (A). p53 signature (p53S) is shown (B; arrow). p53S shows negativity for apoptosis (C), Ki-67 (D), and progesterone receptor A (F), and positivity for estrogen receptor-alpha (E). All figures are presented at 20 $\times$ original magnification.

Figure 4 Ovarian cortical inclusion cyst in the left normal looking ovary in a 68-year-old woman with right ovarian serous cystadenoma (A). The epithelial cells show negativity for p53 (B), apoptosis (C), and Ki-67 (D), and positivity for estrogen receptor-alpha (E) and progesterone receptor A (F). All figures are presented at 40× original magnification.

proliferative and apoptotic index but was not associated with changes in the expression of ER- α . The expression pattern of PRA and ER- α in postmenopausal AFTs is similar to that in ovarian high-grade serous carcinoma and inversely correlated with that in CICs.

Disclosure

The authors report no conflicts of interest in this work.

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