Rutin protects against neuronal damage in vitro and ameliorates doxorubicin-induced memory deficits in vivo in Wistar rats

Abstract: Doxorubicin (DOX) is the most widely used broad-spectrum anticancer agent, either alone or in combination, for most cancers including breast cancer. Long-term use of chemotherapeutic agents to treat breast cancer patients results in cognitive complications with a negative impact on survivors’ quality of life. The study objective was to evaluate rutin (RUT) for its neuroprotective effect against DOX in human neuroblastoma (IMR32) cells in vitro and study its potential to ameliorate DOX-induced cognitive dysfunction in Wistar rats. Cell viability assay (3-[4,5 dimethyl thiazol-2-yl]-2,5-diphenyl tetrazolium bromide), neurite growth assay, detection of apoptosis by (acridine orange/ethidium bromide) staining, intracellular reactive oxygen species (ROS) assay, and flowcytometric analysis were carried out to assess neuroprotective potential against DOX. An in vivo study was conducted for assessing protective effect of RUT against memory deficit associated with DOX-induced chemobrain using object recognition task (ORT). Locomotion was assessed using open field test. Serum biochemistry, acetylcholinesterase, oxidative stress markers in hippocampus, and frontal cortex were assessed. Histopathological analysis of major organ systems was also carried out. Prior exposure to RUT at 100 µM protected IMR32 cells from DOX (1 µM) neurotoxicity. DOX exposure resulted in increased cellular death, apoptosis, and intracellular ROS generation with inhibition of neurite growth in differentiated IMR32 cells, which was significantly ameliorated by RUT. Cognitive dysfunction was induced in Wistar rats by administering ten cycles of DOX (2.5 mg/kg, intra-peritoneal, once in 5 days), as we observed significant impairment of episodic memory in ORT. Coadministration with RUT (50 mg/kg, per os) significantly prevented memory deficits in vivo without any confounding influence on locomotor activity. RUT also offered protection against DOX-induced myelosuppression, cardiotoxicity, and nephrotoxicity. In conclusion, RUT may be a possible adjuvant therapeutic intervention to alleviate cognitive and other complications associated with DOX chemotherapy.

Keywords: breast cancer, chemobrain, cognitive deficit, doxorubicin, episodic memory, object recognition test

Introduction

Doxorubicin (DOX; adriamycin) is a highly effective broad-spectrum cytotoxic agent used in the treatment of most forms of neoplasia. Using chemotherapeutic agents to particularly manage various neoplastic diseases has opened novel prospects to improve survival rates in many cancers. The death rates due to breast cancer have gradually decreased in females since 1989. Furthermore, the 10- and 15-year survival rates (for all stages of breast cancer combined) are currently at 83% and 78%, respectively, in the United States. Despite this prolonged survival, chemotherapy-induced cognitive...
dysfunction (from now referred to as chemobrain/chemofog or mental fog) is a debilitating problem that negatively impacts day-to-day activities and quality of life (QOL) in survivors.

Chemobrain is a condition characterized by neurocognitive complications, which can often persistently be present for 5–10 years and, in some cases, even lifelong, following the cancer chemotherapy.

Cognitive deficits were reported to occur in 34%–70% of cancer patients following the chemotherapy. This phenomenon is particularly evident in breast cancer survivors because of the remarkable improvement in the survival rate, and hence the feasibility of longer follow-up. No treatment has been approved for this particular problem despite the large number of cancer patients reporting memory dysfunction following chemotherapy.

Hence, there exists a great need to develop interventions to combat the cognitive deficits accompanying chemobrain condition so as to improve the health-related QOL in cancer survivors.

Materials and methods

Chemicals and apparatus

DOX was purchased from Fresenius Kabi Oncology Ltd., Solan, India, while RUT, Trypsin-EDTA, 3-[4,5 dimethyl thiazol-2-yl]-2,5-diphenyl tetrazolium bromide (MTT), fetal bovine serum (FBS), retinoic acid, and acridine orange/ethidium bromide (AO/EB) were purchased from HiMedia Laboratories, Mumbai, India. Dichlorofluorescin diacetate and Dulbecco’s Modified Eagle’s Medium were procured from Sigma-Aldrich Co. LLC, St Louis, MO, USA. Sterile 96- and 6-well tissue culture plates and 25 and 75 cm² culture flasks, used in this study, were from Tarsons Products Pvt. Ltd., Kolkata, India.

Object recognition task (ORT) was carried out using 49 cm (length × width × height) square boxes. Behavior of animals was monitored with the help of a camera (model: Quickcam Pro9000, Logitech International S.A., Lausanne, Switzerland) that was mounted 150 cm above the behavioral observation arena. Locomotion and general activity of the animals was assessed using ANYmaze video tracking system (Version 4.82 m, Stoelting Co., Wood Dale, IL, USA).

Animals

Twelve-week old, healthy (n=36) female Wistar rats, weighing 180–230 g, were used in this study. The protocol was approved by the Institutional Animal Ethics Committee, Manipal Medical College, Manipal University, Manipal (approval number IAEC/KMC/17/2013) in association with Committee for the Purpose of Control and Supervision on Experimental Animals. Animals were maintained at Central Animal Research Facility of Manipal University, Manipal, as per the directions specified by Committee for the Purpose of Control and Supervision on Experimental Animals guidelines. A 12/12-hour light and dark cycle was maintained, with lights being turned on at 07 am. Standard rat pellet diet as well as potable water was provided ad libitum.
Cell culture and maintenance

Human neuroblastoma (IMR32) cells were procured from National Centre for Cell Science, Pune, India. Cells were maintained in a CO₂ incubator using Dulbecco’s Modified Eagle’s Medium supplemented with 10% FBS and suitable antibiotic in sterile cell culture flasks. Use of cell lines for in-vitro studies was approved by Doctoral Advisory Committee, Manipal University, Manipal. All the necessary precautions and care was taken for handling, maintenance and disposal of cell lines as per the protocol designed by the Institution.

In vitro cell viability and dose determination for neuroprotection studies

Initially we evaluated toxicity profile (IC₅₀) of RUT in IMR32 cells. Cell viability was assessed using MTT assay. Following 24 hours of seeding, cells were treated with RUT at different concentrations (50–500 µM), and cell viability was assessed after 24 hours of exposure. IC₅₀ of DOX was also assessed. Two concentrations that produced <10% cell death were selected for assessing neuroprotective activity of test flavonoid RUT against DOX-induced neurotoxic insult.

Neuroprotection studies

Neuronal viability using MTT assay

The neuroprotective potential of RUT was tested against DOX-induced neurotoxicity in IMR32 cells using in vitro cell viability assay, ie, MTT. Nontoxic concentrations of test compounds, ie, 50 and 100 µM were selected. Following 2-hour inhibition with RUT at both tested concentrations, toxicant DOX (1 µM) was added and incubated for a further 24 hours. Then, the intensity of purple color formed was calculated using a microplate reader (ELx800; BioTek Instruments Inc., Winooski, VT, USA) at 540 nM, reflecting the direct measurement of cell viability.

Cell cycle analysis by flowcytometry

One day after seeding, cells were incubated with RUT for 2 hours at 100 µM, then exposed to DOX at 1 µM, and further incubated for 24 hours. Then cells were fixed in 70% v/v ethanol for 4 hours and centrifuged, suspended in PBS containing propidium iodide with RNase, and kept in the dark for 20 minutes. Samples were subjected to analysis in a flow cytometer (BD Accuri™ C6; BD Biosciences, San Jose, CA, USA). The influence of test compounds, RUT, and DOX on cell population during the cell cycle progression was assessed.

Assessment of morphology and neurite length in differentiated IMR32 cells

After inducing differentiation into neurons with retinoic acid (10 µM), the cells were treated with RUT at 100 µM for 2 hours, exposed to DOX, and incubated for 24 hours. The cells were then examined using an inverted microscope (Eclipse TS100F; Nikon Instruments Inc., Melville, NY, USA). For measuring neurite length and cell body area, roughly 60–70 images were captured randomly by scanning each treatment well. The length of each neurite drawn was measured using ImageJ software supplemented with Neuron J plug-in provided by National Institutes of Health (Bethesda, MD, USA). Since neurite width is not equal in all the parts of neuron, the neurite width was calculated by dividing the cell body area with the neurite length.

Detection of apoptosis/necrosis in IMR32 cells by AO/EB staining

Differentiated neurons were incubated with DOX (1 µM), either alone or in combination with RUT (at 100 µM, 2 hours incubation prior to DOX exposure) in six well plates. Following incubation for 24 hours, wells were washed with PBS and fixed with 1 mL of ice-cold ethanol (100%) for 10 minutes at room temperature. Then, after washing with PBS, 1 mL AO/EB reagent stain was added to each well and incubated at 37°C for 10 minutes. Individual cells were observed under an inverted microscope with the use of fluorescence filters. Morphology of apoptotic and necrotic cells was recognized based on the staining pattern as described previously.

Quantitative estimation of apoptosis by AnnexinV assay

The Annexin V apoptosis assay was performed as per the manufacturer’s instruction manual. Briefly, 2×10⁴ differentiated cells/well, seeded in a six-well tissue culture plate, were treated with 1 µM DOX alone or a combination of DOX (1 µM) and RUT (50 and 100 µM) after 12 hours of seeding and then incubated for 48 hours. After 48 hours, the cells were trypsinized and centrifuged. After centrifugation, the cell pellets were resuspended in 100 µL of media containing 10% FBS. Approximately 100 µL Annexin V was added to the cell suspension and incubated for 20 minutes. The apoptotic profile of the cells was analyzed using Muse® Cell Analyzer (Model Number 0500-3115, Merck Millipore, Billerica, MA, USA).

Intracellular reactive oxygen species estimation

IMR32 cells in black 96-well plates were incubated with RUT at 50 and 100 µM for 2 hours and then subjected to DOX (1 µM) exposure. After 24-hour inhibition, culture supernatants were discarded and replaced with 100 µL of...
dichlorofluorescein diacetate (100 µM). Following 1-hour incubation with fluorescence dye, cells were washed with sterile Hank’s balanced salt solution. Fluorescence intensity was measured with the help of a fluorescence microplate reader (FLx800; BioTek Instruments Inc., Winooski, VT, USA). The reactive oxygen species (ROS) level was measured and calculated with respect to media control.34

In vivo chemobrain study
Experimental design
Four experimental groups (n=9 of each) were used in this study. Groups 1 and 4 were given normal saline, intraperitoneal, while groups 2 and group 3 were treated with DOX (2.5 mg/kg, intraperitoneal) once in 5 days over a period of 50 days. Animals of groups 2 and 3 were administered daily doses of sodium carboxymethyl cellulose (0.25% w/v in water for injection, intraperitoneal [ip]) and RUT (50 mg/kg, per os [po]), formulated as suspension in carboxymethyl cellulose, respectively. Dosing was begun from 1 week before first DOX cycle and continued thereafter until the end of study. Group 4 was administered RUT at 50 mg/kg, po, but without DOX, to identify the chronic per se effect of this flavonoid on cognitive function. The treatment protocol lasted over a period of 65 days as shown in Figure 1. The doses selected were based on the previous studies and also the earlier work conducted in our laboratory.35,55

ORT for episodic memory
Episodic memory was assessed in rodents using ORT. The test also involves the working memory component to some extent. Procedures adopted were based on earlier reports, with a few modifications.36,37 Briefly, the experiment was conducted over a period of 2 days between 09 am and 05 pm and comprised three phases, viz, habituation, familiarization, and choice trials. On the first day, animals were habituated to the square arenas for a period of 20 minutes. In pilot studies on female Wistar rats, we noticed that control animals were able to remember a familiar object up to an intertrial interval (ITI) of 2 hours.35 Therefore, on day 2, animals were subjected to object recognition test with this ITI of 2 hours. In the familiarization trial, animals were allowed to explore a pair of similar objects. The movement of rats toward the objects was recorded by an expert observer (blind to treatments) using a camera (LogitechPro9000, Logitech International S.A., Lausanne, Switzerland). Recognition trial was performed by replacing one of the familiar objects with a novel object, and the cumulative exploration time of each animal toward a familiar or novel object was noted for 3-minute trial duration. Recognition index and discriminative index that reflect the animal’s memory for objects were determined.38

Body weight monitoring
Body weights were recorded every 3 days throughout the study period. Average body weights were compared among the various experimental groups.

TNF-α levels
The level of TNF-α was estimated using ELISA kit (Novex™ Kit Number KRC3011, Thermo Fisher Scientific Inc., Waltham, MA, USA) in hippocampal and frontal cortex regions, as per the manufacturer’s instructions.

Open-field test for assessing locomotor activity
Rats were assessed for locomotion using an open-field (OF) test.39,40 Briefly, rats were individually placed in OF boxes for 15 minutes. Using ANYmaze video-tracking system, general activity, distance (cm) traveled, and mean velocity (cm/s) of animals during test duration were evaluated.

Hematological analysis
Complete blood profile was determined using automated veterinary blood cell counter (Model: PCE210 VET, ERMA Inc., Tokyo, Japan).

Figure 1 Chemobrain protocol adopted.
Note: Protocol for inducing chemobrain upon DOX administration to assess RUT for possible protective effect against DOX-induced chemobrain in Wistar rats.
Abbreviations: RUT, rutin; DOX, doxorubicin; ORT, object recognition test.
Organ index
Following blood sampling, animals were deeply anesthetized with ketamine, and whole-body perfusion was carried out with ice-cold saline. Kidney, liver, brain, and heart were collected. The weight of each organ per 100 g body weight of the animal was calculated as organ index and comparison was made among the different treatment groups.

Oxidative stress markers
The antioxidant defense systems, viz, superoxide dismutase (SOD), catalase, glutathione (GSH), and total thiols levels were estimated in hippocampus and frontal cortex regions of brain.\(^{41-45}\)

Acetylcholinesterase activity
Following hippocampal and frontal cortex isolation, samples were homogenized with phosphate buffer (pH 7.4), and supernatants were collected for estimation of acetyl cholinesterase.\(^{46}\)

Serum cortisol estimation
Cortisol levels in serum were assessed using rat cortisol ELISA kit (Elabscience Biotechnology Co. Ltd., Wuhan, People’s Republic of China), following the manufacturer’s procedure.

Serum biochemistry
Serum analysis was carried out for cholesterol, total protein, urea, creatinine, creatine kinase, aspartate transaminase, alanine transaminase, alkaline phosphatase, etc using fully automated autoanalyzer (Cobas C111; Roche Diagnostics India Pvt. Ltd., Basel, Switzerland), following manufacturer’s standard protocol.

Histopathology
Histopathological analysis was conducted for all major organs, viz, liver, heart, kidney, and cerebral cortex following the conventional tissue processing in gradient alcohol and xylene with the use of eosin/hematoxylin staining. Gross histopathological changes as a result of toxicant, ie, DOX and also the treatment, RUT was reported.

Statistical analysis
Statistical analysis was carried out using Prism Version 6.05 (fully functional demo; GraphPad Software Inc., La Jolla, CA, USA). All data are expressed as mean ± standard error of the mean of specified number of samples. Cell viability, % apoptotic cells, and intracellular ROS measures were analyzed by Kruskal–Wallis test, followed by Dunn’s post hoc test. For behavioral analysis, exploration time was compared by Student’s paired t-test (within the group and between the objects), whereas recognition index and discriminative index were analyzed by using Kruskal–Wallis test, followed by Dunn’s post hoc test. Locomotor activity, organ index, body weight, neurite length, acetyl cholinesterase, and hematological analyses were carried out by one-way analysis of variance, followed by Dunnett’s post hoc test. A value of \(P<0.05\) was considered as statistically significant.

Results

Cell viability and dose fixation for neuroprotection studies
During the preliminary cytotoxicity assessment of flavonoids or the toxicant in IMR32 cells, the IC\(_{50}\) for RUT, and its aglycone, quercetin was found to be 350 and 250 \(\mu\)M, respectively. Dose-dependent cytotoxic effect was observed for DOX in the concentration range (0.01–5 \(\mu\)M) tested and IC\(_{50}\) of DOX was found to be 1 \(\mu\)M. Approximately 80%–90% cell viability was observed for RUT and quercetin at 50 and 100 \(\mu\)M; therefore, the same concentrations have been used to test the neuroprotective ability of flavonoids in further studies evaluating protective effect against DOX.

Evaluation of neuroprotective effect of flavonoids in vitro against DOX
Neuronal cell viability by MTT assay
DOX (1 \(\mu\)M) produced 49.63% viability in IMR32 cells. However, prior incubation with flavonoids for 2 hours (RUT produced 64.89% and 73.93% viability at 50 and 100 \(\mu\)M, respectively) significantly \((P<0.001)\) prevented DOX-induced cell death in a dose-dependent manner, the most effective being 100 \(\mu\)M. Since 100 \(\mu\)M concentration showed comparatively more protection against DOX, we used this concentration, ie, 100 \(\mu\)M in later studies (Table 1).

<table>
<thead>
<tr>
<th>Serial number</th>
<th>Treatment</th>
<th>% viability</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>DMSO media control</td>
<td>100.0±1.914*</td>
</tr>
<tr>
<td>2</td>
<td>DOX control, 1 (\mu)M</td>
<td>49.63±1.44</td>
</tr>
<tr>
<td>3</td>
<td>DOX, 1 (\mu)M + RUT 50 (\mu)M</td>
<td>64.89±0.948*</td>
</tr>
<tr>
<td>4</td>
<td>DOX, 1 (\mu)M + RUT 100 (\mu)M</td>
<td>73.93±0.841*</td>
</tr>
</tbody>
</table>

Notes: Data represent mean ± SEM of three tests in triplicates for protective effect of RUT on cell viability against DOX-induced neurotoxicity in MTT assay on IMR32 cells, *P<0.001 compared to DOX control.

Abbreviations: DMSO, dimethyl sulfoxide; DOX, doxorubicin; MTT, 3-[4,5-dimethyl thiazol-2-yl]-2,5-diphenyl tetrazolium bromide; RUT, rutin; SEM, standard error of the mean.
Cell cycle analysis by flow cytometry

Treatment with DOX (1 µM) arrested the cell cycle progression in IMR32 cells at G2/M and S phases. Pretreatment with RUT partially prevented DOX-induced changes (Figure 2) with the cells predominately in G0/G1 and S phases.

Morphological changes and assessment of neurite length in differentiated IMR32 cells

Treatment with DOX induced morphological alterations in neuronal cells. The nucleus was condensed, and cell membrane was fragmented due to toxicant insult (Figure 3C). However, prior treatment with RUT resulted in the cells retaining their normal morphology (Figure 3D). DOX exposure significantly (P<0.001) inhibited the development of neurite outgrowth in differentiated IMR32 cells compared to media control. However, treatment with RUT at 100 µM produced prominent establishment of neurite and, hence, significantly (P<0.001) averted inhibitory effect of DOX on neurite growth in IMR32 cells. This indicates that RUT protected the differentiated neuronal cells in course of neurite
development against the neurotoxic insult produced by DOX. In contrast to neurite length, DOX treatment significantly increased the neurite width when compared to normal control. This increase in neurite width was attenuated by pretreatment with RUT. The neurite establishment, length, and width are illustrated in Figures 3 and 4.

Detection of apoptosis/necrosis by AO/EB staining
AO/EB staining showed that DOX produced marked apoptotic cell death in IMR32 cells with the finding of bright condensed nuclei and fragmented cellular structures (Figure 5). The percentage of cells with early apoptotic and late apoptotic morphology were noted. The percentage of apoptotic cells was significantly ($P<0.001$) high in DOX group when compared to the normal control (Table 2). RUT prior treatment reversed the DOX-induced elevation of apoptotic cell death and produced significant decline ($P<0.01$) in percentage of apoptotic cells. This shows that neuroprotective potential of flavonoids is through inhibition of apoptotic pathways.

Apoptotic detection by Annexin V staining
Treatment with DOX (1 µM) induced early and late apoptosis in IMR32 cells (Figure 6B). Pretreatment with RUT (50 and 100 µM) significantly decreased the number of apoptotic cells induced by DOX (Figure 6E), indicating the

Figure 4 Effect of DOX and RUT on (A) neurite length and (B) neurite width in IMR32 cells.
Notes: Illustration represents mean ± SEM of neurite length in micrometers for the protective effect of RUT against DOX-induced neurite inhibition in IMR32 cells, $^*P<0.001$ vs normal control, $^#P<0.001$ vs DOX control.
Abbreviations: DOX, doxorubicin; RUT, rutin; SEM, standard error of the mean.

Figure 5 Effect of treatments on apoptotic cell death in IMR32 cells.
Notes: Illustration represents the effect of RUT on DOX-induced apoptosis in AO/EB staining apoptosis assay; (A) normal control; (B) DOX control; (C) cells treated with RUT prior to DOX; (D) cells treated with rutin only. Arrows illustrate the represent images of IMR32 cells with normal (A, C and D) or apoptotic (B) morphology.
Abbreviations: DOX, doxorubicin; AO/EB, acridine orange/ethidium bromide; RUT, rutin.
neuroprotective effect of RUT. Furthermore, RUT prevented early, rather than late, apoptosis in IMR32 cell lines challenged with DOX (Figure 6C and D).

**Intracellular ROS estimation**

DOX produced significant \((P<0.001)\) elevation of intracellular ROS as compared to the normal media control. Prior treatment with RUT significantly \((P<0.001)\) inhibited the ROS generation by DOX. This shows the potential antioxidant role of this flavonoid and its ability to scavenge the intracellular ROS that was generated by DOX (Figure 7).

**Table 2** Percentage of apoptotic cells in AO/EB staining

<table>
<thead>
<tr>
<th>Serial number</th>
<th>Treatment</th>
<th>% apoptotic cells</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Normal control</td>
<td>16.54±1.06</td>
</tr>
<tr>
<td>2</td>
<td>DOX control, 1 µM</td>
<td>54.78±3.56*</td>
</tr>
<tr>
<td>3</td>
<td>DOX, 1 µM + RUT, 100 µM</td>
<td>31.22±3.76*</td>
</tr>
<tr>
<td>4</td>
<td>RUT, 100 µM</td>
<td>12.56±2.76**</td>
</tr>
</tbody>
</table>

**Notes:** Data represent mean ± SEM of three tests in triplicate for the effect of RUT on cellular apoptosis against DOX-induced neurotoxicity in differentiated IMR32 cells. \(*P<0.001\) compared to normal control; \(#P<0.01\) and \(##P<0.001\) compared to DOX control.

**Abbreviations:** DOX, doxorubicin; RUT, rutin; AO/EB, acridine orange/ethidium bromide; SEM, standard error of the mean.

**OF test for locomotor activity**

It was found that neither DOX nor RUT had any influence on locomotor activity. Hence, there was no confounding influence of DOX or RUT in the assessment of episodic memory using ORT. No significant differences for either distance traveled or mean velocity were noted among the four treatment groups, which proves the validity of the ORT in assessing the episodic memory deficits associated with DOX chemotherapy (Figure 11).

**Hematological analysis**

Treatment with DOX over 50 days resulted in a significant \((P<0.01)\) reduction in red blood cells (RBC) and hemoglobin compared to vehicle control animals. This myelosuppressive effect of DOX was significantly \((P<0.01)\) reversed by RUT coadministration at the dose of 50 mg/kg, po. White blood cells (WBC) counts were comparatively reduced in DOX control group with respect to vehicle. However, the differences were not statistically significant. Rats treated with only RUT without any DOX treatment showed improvement of RBC and hemoglobin when compared to the normal vehicle control, which supports RUT’s protective effect on DOX-induced hematological changes (Table 3).

**Organ index**

No significant difference was observed among the treatment groups for organ index of any of the organs studied, ie, heart, liver, brain, kidney, etc (data not shown).

**Oxidative stress status**

Catalase, GSH, total thiols, and SOD levels were significantly \((P<0.01)\) lower in frontal cortex and hippocampal regions for the DOX control group as compared to the normal control. However, these alterations were significantly \((P<0.01)\) prevented by coadministration of RUT at a dose.
Figure 6 Quantitative analysis of the effect of treatments on apoptotic patterns in iMr32 cells.

Notes: Illustration represents the effect of RUT on DOX-induced apoptosis in Annexin V staining; (A) normal cells; (B) DOX treated; (C) cells treated with RUT (50 µM) prior to DOX; (D) cells treated with RUT (100 µM) prior to DOX; (E) percentage apoptosis in cells due to treatments where values are represented as mean ± SEM.

*P<0.001 compared to normal control; †P<0.01 compared to DOX control.

Abbreviations: apop, apoptosis; DOX, doxorubicin; RUT, rutin; SEM, standard error of the mean.
Hippocampal and frontal cortex acetylcholinesterase activity
No significant difference in acetylcholinesterase activity was noted among the treatment groups, either in frontal cortex or hippocampus. DOX treatment did not produce any inhibition or activation of this enzyme activity. This shows that the pathophysiology of chemobrain is different from Alzheimer’s disease–associated dementia (Table 5).

Serum cortisol levels
Changes in serum cortisol levels were found to be insignificant among the treated groups, showing that the test treatments did not influence cognitive assessment with depression-like states (Data not shown).

Serum biochemistry
It was noted that the creatine kinase, urea, triglyceride, and total cholesterol levels were significantly elevated in DOX control group compared to vehicle control. However, RUT coadministration prevented the changes observed with DOX treatment. Other parameters, viz, glucose, AST, ALT, ALP, total bilirubin, creatinine, total protein, and albumin were found to be unaltered among different groups (Table 6).

Histopathology
Histopathological analysis revealed that heart, liver, kidney, and cerebral cortex samples of the animals treated with DOX showed abnormal architecture with degenerative changes as
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Rutin ameliorates chemobrain through neuroprotection

... compared to vehicle control samples. Treatment with DOX along with RUT at 50 mg/kg, po, protected pathological changes observed with DOX control samples. This indicates that RUT is able to reverse the pathological abnormalities induced by DOX to the major organ systems and preserves their structural integrity and function (Figure 12).

Discussion

Oncologists often receive complaints from breast cancer survivors following the standard-/high-dose chemotherapy on cognitive complications for diverse components of memory, viz, episodic, working memories, difficulty in multitasking, attention, planning, etc. Increasing evidence shows that chemotherapeutic agents, because of their nonspecific effects, are highly toxic to the rapidly proliferating neuronal cells like adult hippocampal neuronal stem cells. These agents also cause excessive activation of microglial cells that can ultimately lead to neuroinflammation and death.

The first report on chemotherapy-related cognitive dysfunction was made in the 1980s, which revealed that patients undergoing chemotherapy for various cancers like leukemia, Hodgkin’s disease, digestive and respiratory malignancies, etc, performed significantly low on various neuropsychological cognitive tests.47 This study also identified the fact that permeation of chemotherapeutic agents into the brain was not a prerequisite for inducing chemofog condition that resulted from a noncentral nervous system–directed chemotherapy. Although most of the cytotoxic agents do not cross blood–brain barrier effectively (eg, DOX), they still produce chemobrain-associated cognitive dysfunction through their peripheral cytotoxic effects in animals.48

Increased levels of oxidative free radicals like ROS, RNS, and peroxynitrite (ONOO⁻) in the periphery that can easily cross blood–brain barrier are deleterious to normal cell population as a result of protein and lipid oxidation products such as protein carbonyls, 3-nitrotyrosine and malondialdehyde, 4-hydroxy nonenal, etc.49 Furthermore, in rodent studies, DOX was found to increase the susceptibility of brain mitochondria to calcium-induced permeability transition pore opening and oxidative stress predisposing neuronal cells to degeneration.50 Hence, we have selected to study DOX-induced chemobrain and to test RUT as a possible intervention.

To date, no novel chemical entities are found to be effective to alleviate the symptoms of chemofog. There is a great need for animal studies to explore cognitive impairment objectively following the cytostatic treatment and also to identify possible underlying mechanism. Hence, in this study, we demonstrated neuroprotective potential of a flavonoid glycoside, ie, RUT against DOX toxicity in neuronal cells.

Figure 9 Effect of treatments on body weight of animals.

Notes: Illustration represents time course of change in body weight and the effect of RUT on DOX-induced body weight change (n=9).

Abbreviations: DOX, doxorubicin; RUT, rutin.

Figure 10 Effect of treatments on TNF-α levels in brain regions.

Notes: Illustration represents mean ± SEM of TNF-α level in (A) hippocampus and (B) frontal cortex regions, *P<0.001 compared to vehicle control, #P<0.001 compared to DOX control (n=6). RUT significantly prevented the elevated levels of TNF-α in both hippocampus and frontal cortex regions in brain.

Abbreviations: TNF-α, tumor necrosis factor alpha; DOX, doxorubicin; SEM, standard error of the mean; RUT, rutin.
This was followed by in vivo evaluation of RUT in an established model of chronic DOX-induced chemobrain condition accompanying memory deficits in female Wistar rats.

In clinical oncology, the chemotherapy regimen often comprises a combination of different cytotoxic agents to treat various cancers. However, it is equally essential to assess the individual chemotherapeutic agents for their potential contribution to induce a chemobrain-like condition. DOX is the most widely used cytotoxic drug for many forms of cancer. Further, it is a known fact that despite its use globally for many forms of cancers, not much attention has been paid to DOX-induced neurotoxicity and its role in inducing cognitive impairment. Hence, we focused mainly on DOX-induced neurotoxicity and also the associated cognitive impairment in vivo.

We hypothesize that natural flavonoid, RUT, through its pleiotropic actions, may combat chemotherapy-related cognitive dysfunction. Furthermore, we tried to identify the mechanism underlying the protective potential of RUT against DOX-induced chemobrain through in vitro neuroprotection studies in IMR32 neuronal cells. IMR32, the human neuroblastoma cell line, is widely used in vitro system to assess the neuroprotective ability (with regard to neuronal damage, morphological changes, and neuritogenic effects) of novel test compounds.

In vitro studies revealed that DOX produced neuronal death in IMR32 cells, while prior treatment with RUT resulted in significant protection against DOX-induced neurotoxicity in a dose-dependent manner, as described in MTT assay. Furthermore, it was evident that the neuroprotective effect of RUT was through reducing the generation of intracellular ROS and apoptotic bodies induced by DOX treatment.

With the aforementioned promising neuroprotective effects, the flavonoid, RUT, was further evaluated in vivo for its potential to ameliorate chemobrain (DOX)-associated episodic-like memory deficits. We assessed RUT in vivo for its chemobrain preventing potential in a well-established DOX-induced chemofog animal model, which is associated with episodic memory deficits.

Memory of autobiographical events in relation to times and places is termed as episodic memory and is most useful form of memory to accomplish the day-to-day activities in

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**Table 3: Hematological profiling**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Vehicle control</th>
<th>DOX control</th>
<th>RUT + DOX</th>
<th>RUT</th>
</tr>
</thead>
<tbody>
<tr>
<td>RBC (\times 10^6 cells/µL)</td>
<td>8.04±0.146</td>
<td>5.89±0.219*</td>
<td>9.07±0.42*</td>
<td>9.28±0.124*</td>
</tr>
<tr>
<td>Hb % (g/dL)</td>
<td>12.96±0.172</td>
<td>9.81±0.158*</td>
<td>13.82±0.385*</td>
<td>14.14±0.283*</td>
</tr>
<tr>
<td>WBC (\times 10^9 cells/µL)</td>
<td>8.66±0.682</td>
<td>7.21±0.48</td>
<td>7.32±0.676</td>
<td>8.17±0.516</td>
</tr>
<tr>
<td>Granulocytes (\times 10^9 cells/µL)</td>
<td>1.28±0.098</td>
<td>1.04±0.073</td>
<td>1.22±0.145</td>
<td>1.3±0.278</td>
</tr>
<tr>
<td>Lymphocytes (\times 10^9 cells/µL)</td>
<td>7.04±0.549</td>
<td>6.62±0.458</td>
<td>7.33±0.267</td>
<td>7.85±0.289</td>
</tr>
<tr>
<td>Monocytes (\times 10^9 cells/µL)</td>
<td>1.1±0.103</td>
<td>1.05±0.05</td>
<td>1.11±0.115</td>
<td>0.88±0.068</td>
</tr>
<tr>
<td>Platelets (\times 10^3 cells/µL)</td>
<td>548.11±23.545</td>
<td>521.11±22.497</td>
<td>523.88±31.023</td>
<td>581.55±17.84</td>
</tr>
</tbody>
</table>

**Notes:** Data represent mean ± SEM of various hematological parameters. *P<0.01 compared to vehicle control, †P<0.01 compared to DOX control (n=6).

**Abbreviations:** DOX, doxorubicin; RUT, rutin; RBC, red blood cells; Hb, hemoglobin; WBC, white blood cells; SEM, standard error of the mean.
one’s life.56 Prefrontal cortex is mainly involved in encoding and retrieval of episodic memory.57,58 Moreover, the anatomical structures that are crucial for episodic memory include amygdala, brainstem, and hippocampus. In animal species like rodents, episodic component is generally assessed using ORT where animals encounter different episodes in which they remember different objects presented to them.

In ORT, rats treated with DOX explored both familiar and novel objects without any discrimination during choice trial, which indicates that the rats were unable to recognize either familiar or novel objects. However, the animals with saline treatment were able to discriminate the novel object from the familiar one. Treatment with RUT alone or in combination with DOX produced significant increase in recognition and discriminative indices compared to DOX control group. This showed that RUT was able to prevent the cognitive deficits associated with DOX chemotherapy. The locomotor activity was also assessed for any inhibitory or stimulatory effects influencing treatments. In our study, we did not find any significant effects on locomotion among the treatment groups that support the validity of assessing episodic memory using ORT.

Body weight recordings displayed moderate reduction in body weight for the group treated with DOX alone as compared to saline control. RUT treatment was able to reverse the DOX-induced decline in body weight and was comparable to that of vehicle group. Hematological profiling indicated that chronic DOX treatment resulted in significant reduction of RBC and hemoglobin levels with moderate effect on WBC and platelets compared to vehicle. However, chronic RUT treatment (alone or in combination with DOX) produced significant myeloprotective effects against DOX. Organ index difference for major organs was found to be insignificant among the treated groups.

Acetylcholinesterase activity in hippocampus and frontal cortex was almost similar across the treatment groups and no significant difference was noticed. This indicates that the pathological changes underlying chemobrain are different from those of Alzheimer’s disease. In Alzheimer’s disease, inhibition of acetylcholinesterase seemed to be an effective but only a symptomatic therapeutic strategy.

TNF-α levels have been significantly elevated in DOX control group in hippocampus and frontal cortex, which indicate an indirect neurotoxic mechanism of DOX treatment through periphery. However, this increase was significantly prevented by RUT cotreatment, which proves the potential anti-inflammatory effect of the flavonoid, RUT to alleviate chemobrain condition.

Histopathological analysis of major organs revealed chronic DOX-produced gross structural abnormalities in major organs such as liver, heart, kidney, and brain. RUT pretreatment averted these abnormalities. This shows that RUT was able to reverse the DOX-induced structural, and thereby the, functional abnormalities of major organ systems to some extent.

Growing evidence shows that adult hippocampal neurogenesis is a continuous process throughout the life span and is essential to form new recognition memories. Dentate gyrus, the hippocampal subregion, is the principal and active site of neurogenesis. Even when a small quantity of cytotoxic drugs

Table 4 Effect of treatments on oxidative stress markers in brain

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Hippocampal markers</th>
<th>Frontal cortex markers</th>
<th>Total thiol (µg of protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Catalase (units/mg of protein)</td>
<td>SOD (units/mg of protein)</td>
<td>GSH (µg of protein)</td>
</tr>
<tr>
<td>Vehicle control</td>
<td>1.90±0.11</td>
<td>12.45±1.04</td>
<td>4.77±0.71</td>
</tr>
<tr>
<td>DOX control</td>
<td>0.94±1.3</td>
<td>7.35±0.54</td>
<td>2.11±0.31</td>
</tr>
<tr>
<td>RUT + DOX</td>
<td>1.89±0.32</td>
<td>12.18±1.04</td>
<td>4.52±0.52</td>
</tr>
<tr>
<td>RUT</td>
<td>2.01±0.11</td>
<td>13.08±1.12</td>
<td>4.90±0.42</td>
</tr>
</tbody>
</table>

Notes: Data represent mean ± SEM of oxidative stress markers in hippocampal and frontal cortex regions in brain; *P<0.01 compared to vehicle control, **P<0.01 compared to DOX control (n=6).

Abbreviations: DOX, doxorubicin; RUT, rutin; SOD, superoxide dismutase; GSH, glutathione; SEM, standard error of the mean.

Table 5 Acetylcholinesterase activity in brain

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Acetylcholinesterase activity (µM of acetylthiocholine hydrolyzed/min/mg of protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Hippocampus</td>
</tr>
<tr>
<td>Vehicle control</td>
<td>8.275±0.403</td>
</tr>
<tr>
<td>DOX control</td>
<td>8.646±0.514</td>
</tr>
<tr>
<td>RUT + DOX</td>
<td>7.973±0.467</td>
</tr>
<tr>
<td>RUT</td>
<td>7.196±0.711</td>
</tr>
</tbody>
</table>

Notes: Data represent mean ± SEM of acetylcholinesterase enzyme activity in hippocampal and frontal cortex regions in brain (n=6). No significant difference observed between groups.

Abbreviations: DOX, doxorubicin; RUT, rutin; SEM, standard error of the mean.
Table 6 Effect of treatments on biochemical parameters

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Vehicle control</th>
<th>DOX control</th>
<th>RUT + DOX</th>
<th>RUT</th>
</tr>
</thead>
<tbody>
<tr>
<td>AST (U/L)</td>
<td>128.72±8.44</td>
<td>122.53±4.85</td>
<td>116.50±8.26</td>
<td>117.68±8.15</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>43.60±3.03</td>
<td>45.47±3.19</td>
<td>44.75±3.53</td>
<td>43.98±2.38</td>
</tr>
<tr>
<td>ALP (U/L)</td>
<td>103.53±5.80</td>
<td>107.98±7.55</td>
<td>95.44±6.16</td>
<td>98.13±8.29</td>
</tr>
<tr>
<td>CK (U/L)</td>
<td>376.05±38.37</td>
<td>801.83±167.91</td>
<td>398.73±52.92</td>
<td>421.18±37.80</td>
</tr>
<tr>
<td>Glucose (mg/dL)</td>
<td>73.46±6.92</td>
<td>81.65±2.92</td>
<td>73.67±5.22</td>
<td>80.28±4.20</td>
</tr>
<tr>
<td>Urea (mg/dL)</td>
<td>36.66±5.06</td>
<td>96.96±8.87</td>
<td>41.21±7.82</td>
<td>33.10±4.32</td>
</tr>
<tr>
<td>Total cholesterol (mg/dL)</td>
<td>55.12±3.18</td>
<td>86.40±3.77</td>
<td>61.54±2.17</td>
<td>58.44±3.29</td>
</tr>
<tr>
<td>Triglyceride (mg/dL)</td>
<td>61.98±4.67</td>
<td>147.58±7.53</td>
<td>79.13±5.46</td>
<td>49.73±4.41</td>
</tr>
<tr>
<td>Total bilirubin (mg/dL)</td>
<td>0.10±0.00</td>
<td>0.10±0.00</td>
<td>0.08±0.02</td>
<td>0.1±0.02</td>
</tr>
<tr>
<td>Creatinine (µmol/L)</td>
<td>31.39±2.17</td>
<td>31.22±1.88</td>
<td>30.06±0.55</td>
<td>30.53±2.01</td>
</tr>
<tr>
<td>Total protein (g/dL)</td>
<td>7.02±0.41</td>
<td>7.87±0.17</td>
<td>6.55±0.26</td>
<td>6.43±0.24</td>
</tr>
<tr>
<td>Albumin (mg/dL)</td>
<td>4.33±0.30</td>
<td>4.52±0.23</td>
<td>3.9±0.37</td>
<td>4.01±0.15</td>
</tr>
</tbody>
</table>

Notes: Data represent mean ± SEM of various biochemical parameters (n=6). *P<0.05, **P<0.01 compared to vehicle control; *P<0.05, **P<0.01, ***P<0.001 compared to DOX control.

Abbreviations: AST, aspartate transaminase; ALT, alanine transaminase; ALP, alkaline phosphatase; CK, creatinine kinase; DOX, doxorubicin; RUT, rutin; SEM, standard error of the mean.

Although RUT showed protective effects in this study in structural and functional assessments, not knowing the exact mechanism by which this happened is a limitation of this study. Most probable mechanisms include neuroprotection, combating the oxidative stress, blood-forming effect, and prevention of neuroinflammation (TNF-α). Further studies are warranted to assess the effect of RUT in models entering brain, it can be highly toxic to this adult hippocampal neurogenesis pool of neuronal stem cells, which can result in cognitive complications and chemofog.

Figure 12 Histopathological analysis of major organ systems.

Notes: (A) Cerebral cortex: (a) healthy vehicle control showing normal histological signs; (b) DOX control showing irregular and degenerated morphological features along with marked gliosis; (c) rats treated with DOX and RUT (50 mg/kg, po) reflecting the minor morphological changes with reduced gliosis as compared to DOX control; (d) rats treated with only RUT showing the histological features as that of healthy control; (B) heart: (a) healthy control displaying general histological architecture of normal myocardium; (b) DOX control with numerous vacuoles and degeneration of myocardial structure; (c) rats treated with DOX and RUT (50 mg/kg, po) reflecting slight histological changes as compared to normal control; (d) rats treated with RUT only displaying healthy myocardial architecture as that of normal control; (C) liver: (a) vehicle control exhibiting histological architecture of healthy hepatocyte parenchyma and central vein with no abnormalities; (b) rats treated with DOX showing disruption of hepatocyte parenchyma with central vein dilation and necrosis; (c) RUT- and DOX-treated animals displaying minor changes of hepatocyte parenchymal and central vein histological architecture; (d) RUT- only treated group exhibiting liver histology as that of normal healthy control; (D) kidney: (a) healthy control rats showing normal histological morphology of kidney with complete renal corpuscle and proximal tubules; (b) DOX control rats showing abnormal architecture with necrosis of renal corpuscles, and proximal tubules with medullary congestion; (c) rats treated with DOX and RUT displaying healthy histological features as that of normal control; (d) RUT- only treated rats exhibiting kidney morphology as that of healthy control. Arrows in DOX control animal samples represent the histological abnormality profile to the major organ systems assessed. (All the images were captured at 40x magnification.)

Abbreviations: DOX, doxorubicin; po, per os; RUT, rutin.
of other cognitive components and also the combination chemotherapy-induced cognitive dysfunction.

From these results, it is evident that RUT mitigates DOX-induced cognitive dysfunction, myelosuppression, and oxidative stress. This may be attributed to its antioxidant and anti-inflammatory activity. The cytotoxic action of DOX is mediated through the disruption of the redox potential in cancer cells, which leads to an environment of oxidative stress. Hence, it is imperative to verify whether RUT suppresses the side effects associated with DOX in normal cells without affecting its cytotoxic potential in cancer cells. There are reports of use of antioxidant agents as anticancer drugs. It is well established that RUT modulates Wnt, JAK-STAT, and EGF signaling, and AP-1, NF-κB, and Akt in cancer cells, which are implicated in its anticancer property.59 In this regard, further studies are underway to evaluate the efficacy of RUT in the prevention of DOX-induced toxic effects in mammary carcinoma rats.

Conclusion
DOX on chronic treatment produced episodic memory deficits in Wistar rats, and the flavonoid glycoside RUT is able to prevent these deficits associated with DOX chemotherapy through its neuroprotective effect when coadministered. Hence, RUT may be a promising and potential adjuvant therapeutic intervention to alleviate the cognitive deficits associated with DOX-induced chemobrain in vivo as well as other treatment-related side effects.

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Disclosure
The authors report no conflicts of interest in this work.

References


