Long noncoding RNA LINC01296 is associated with poor prognosis in prostate cancer and promotes cancer-cell proliferation and metastasis

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Background/purpose: Long noncoding RNAs (lncRNAs) have emerged as important regulators and biomarkers of tumor development and progression. This study investigated the clinical significance, biological functions, and underlying mechanisms of long intergenic non-protein-coding RNA 1296 (LINC01296) in prostate cancer.

Materials and methods: LINC01296 expression in prostate cancer tissues and cell lines was detected by quantitative real-time polymerase chain reaction (qRT-PCR). The correlation between LINC01296 expression and clinicopathologic characteristics of prostate cancer was analyzed using Kaplan–Meier analysis and the Cox proportional-hazard model. Small interfering RNA was used to suppress LINC01296, and knockdown efficiency was examined by qRT-PCR. Cell Counting Kit 8 assay, colony-formation assay, migration and invasion assays, and Western blot assay were used to explore the role of LINC01296 in tumor progression further.

Results: LINC01296-expression level was higher in prostate cancer tissues and prostate cancer cells than in adjacent nontumor tissues and immortalized normal prostate stromal WPMY1 cells. LINC01296 expression was correlated with preoperative prostate specific antigen (P=0.002), lymph-node metastasis (P=0.035), Gleason score (P<0.001), and tumor stage (P=0.036). Patients with higher LINC01296 expression displayed advanced clinical features and shorter biochemical recurrence-free survival time than those with lower LINC01296 expression. Multivariate analysis showed that LINC01296 expression was an independent predictor of biochemical recurrence-free survival in prostate cancer. Additionally, LINC01296 knockdown inhibited prostate cancer-cell proliferation, migration, and invasion, demonstrated in an in vitro study involving regulation of PI3K–Akt–mTOR signaling and epithelial–mesenchymal transition.

Conclusion: The results demonstrated that LINC01296 is a novel molecule involved in prostate cancer development and progression, and thus is a potential biomarker of prognosis.

Keywords: long noncoding RNA, prostate cancer, prognosis, proliferation, migration, invasion

Introduction

Prostate cancer is one of the most common malignant tumors in males worldwide, particularly in developed Western countries.1,2 The latest data indicate that the incidence and mortality of prostate cancer are estimated to reach up to 180,890 new cases and 85,920 new deaths by the year 2016 in the US.3 In addition, the incidence and mortality of prostate cancer among Asian men has increased rapidly during the past decade.4 In recent years, the incidence of prostate cancer in China has also increased rapidly, and this phenomenon is associated with lifestyle changes; moreover, increased detection of prostate cancer is attributed to prostate-specific antigen screening and improved detection of prostate cancer at an early stage.
biopsy techniques. Early-stage prostate cancer patients often show good prognosis after comprehensive treatment. Nevertheless, recurrence, metastasis, and development of castration-resistant prostate cancer remain the leading causes of mortality. The detailed molecular mechanisms underlying prostate cancer carcinogenesis remain to be fully elucidated. Therefore, studies on prostate cancer remain extremely important.

Recent evidence has indicated that noncoding RNAs (ncRNAs) play a key role in cancer. Based on their sizes, ncRNAs can be classified into two major groups: small ncRNAs (eg, microRNAs) and long ncRNAs (lncRNAs). lncRNAs are a class of transcripts that are longer than 200 nucleotides and do not contain an open reading frame. lncRNAs have been proved to be involved in the development and progression of prostate cancer; however, research on the involvement of lncRNA in prostate cancer is still in its preliminary stage.

Based on the relationship between lncRNA and prostate cancer, lncRNA can be roughly divided into three categories: carcinogenicity lncRNA, tumor-suppressor lncRNA, and lncRNA with unknown functions. PCA3, also known as carcinogenicity lncRNA, is the first lncRNA to be identified in prostate cancer, and has been used in different applications, including diagnosis, treatment, and prediction of prostate cancer. Additionally, MALAT1, PCAT1, and SCHLAP1 demonstrate oncogenic functions in prostate cancer. PCGEM1 is an androgen-induced prostate-specific lncRNA that is highly expressed in early stages of prostate cancer; PCGEM1 was also detected in African-American prostate cancer patients or in prostate cancer patients with a family history of prostate cancer. PCGEM1 is associated with the targeted androgen receptor, which contributes to the development of castration-resistant prostate cancer. In addition, MEG3 is a tumor suppressor. MEG3 can promote apoptosis and inhibit proliferation by stabilizing p53 expression and by selectively activating the expression of the p53 target gene. IncRNA H19 plays a tumor-suppressive role in metastatic prostate cancer by repressing the effects of TGFβ. However, newer lncRNAs are being investigated through high-throughput technologies. Moreover, the functional role and underlying molecular mechanisms of the lncRNAs in the development and progression of prostate cancer remain unclear.

With the rapid development of new-generation sequencing, RNA-sequencing technology has been widely used to analyze the transcriptomes of various organisms; this technology is useful in the discovery of new therapeutic targets and prognostic indicators. Our previous microarray data have revealed the novel intergenic lncRNA LINC01296 to be significantly upregulated (P<0.05, fold-change >2). LINC01296 is located at chromosome 14q11.2.

In this study, we analyzed the expression of the novel lncRNA LINC01296 (ENSG00000244306) in prostate cancer tissue samples and investigated its potential biomedical functions. Our results demonstrated that LINC01296 is a potential new molecular biomarker or a new basis for prostate cancer treatment.

Materials and methods

Ethics statement

Written informed consent was obtained from all participants involved in this study. This research was approved by the institutional review board of the First Affiliated Hospital of Nanjing Medical University (Nanjing, China).

Patient samples and microarray analysis

Prostate cancer tissues and paired adjacent normal tissues were obtained from 73 patients who had undergone surgical resection of prostate cancer from February 2009 to April 2014. After resection, all tissue samples were immediately frozen and stored in liquid nitrogen until further analysis. The diagnosis of prostate cancer was confirmed by histopathology. Three sample pairs were used for microarray analysis, and other pairs were used for additional studies. A total of 8,277 lncRNA probes were used in microarray assays. Differentially expressed lncRNAs were identified by comparing the expression levels in tumor samples and in adjacent normal samples.

Cell lines and culture conditions

Human prostate cancer cell lines (22Rv1 and LNCaP) and a normal myofibroblast stromal cell line (WPMY1) were purchased from the Institute of Biochemistry and Cell Biology of the Chinese Academy of Sciences (Shanghai, China). These cells were cultured in Roswell Park Memorial Institute 1640 medium or Dulbecco’s Modified Eagle’s Medium (Thermo Fisher Scientific, Waltham, MA, USA) supplemented with 10% fetal bovine serum, 100 mg/mL streptomycin, and 100 U/mL penicillin (Thermo Fisher Scientific) in a humidified atmosphere at 37°C with 5% CO₂.

RNA isolation and quantitative real-time polymerase chain reaction

Total RNA was extracted from tissues or cultured cell lines using Trizol reagent (Thermo Fisher Scientific) according
to the manufacturer’s protocols. For reverse-transcription polymerase chain reaction (PCR), 1 μg of total RNA was reverse-transcribed to complementary DNA in a final volume of 10 μL by using random primers (10 μM) and a reverse-transcription kit (Takara, Kyoto, Japan). Reverse transcription was performed at 37°C for 15 minutes and then at 85°C for 5 seconds and 4°C for 5 minutes according to the manufacturer’s instructions. Quantitative real-time PCR (qRT-PCR) was performed on an ABI StepOnePlus (Thermo Fisher Scientific) using a standard protocol from the manufacturer’s instructions; the reaction was performed in 10 μL reaction volume containing 5 μL of SYBR Premix (2x), 0.4 μL of PCR forward primer (10 μM), 0.4 μL of PCR reverse primer (10 μM), 0.2 μL of ROX Reference Dye II (50x), 3 μL of double-distilled water, and 1 μL of complementary DNA of the sample. qRT-PCR was performed under the following conditions: 95°C for 10 minutes, 95°C for 30 seconds, 95°C for 5 seconds, 61°C for 30 seconds in cycles for a total of 50 cycles, and a final extension step at 72°C for 5 minutes. ΔCt values were normalized to β-actin. The relative quantitative value was obtained using the 2−ΔΔCt method. Relative quantitative values greater than 2 were defined as high expression, and less than 0.5 as low expression. Each experiment was performed in triplicate and repeated three times. The primers were: LINC01296, forward 5′-AAGTGCCACGCTCCTAC-3′, reverse 5′-CGGCCAAGT TCTTACCAC-3′; β-actin, forward 5′-ACTGGAACGGTGAAGGTGAC-3′, reverse 5′-AGAGAAGTGGGTGCTT-3′.

Small interfering RNA transfection
Small interfering RNA (siRNA) that targets LINC01296 RNA and unspecific control was purchased from GenePharma (Shanghai, China). The target sequences for siRNA-LINC01296 were: siRNA-LINC01296-1 (S1), 5′-GGCCGGAUAUUCCCUATTTT-3′; siRNA-LINC01296-2 (S2), 5′-CUGAACAUAUUCGGUGGT-3′; and siRNA-LINC01296-3 (S3), 5′-GCAGCUGGGAUUUGAAGU-3′. The experiments showed that S1 and S2 demonstrated the highest inhibition efficiency and thus were used in the subsequent experiments. Synthetic sequence-unspecific siRNA was used as negative control (NC) siRNA. Prior to transfection with siRNA-LINC01296 and NC siRNA using Lipofectamine 2000 (Thermo Fisher Scientific) according to the manufacturer’s instructions, the cells were cultured in six-well plates containing complete growth media until cell density reached 60%. The cells were harvested after 48 hours for qRT-PCR and Western blot analyses.

Cell-proliferation assays
A CCK-8 assay (Dojindo, Kumamoto, Japan) was used to analyze the proliferation of the siRNA-transfected cell lines. After siRNA transfection for 48 hours, 2,000 cells per well were incubated in 96-well plates, each well containing 200 μL of complete culture medium. After 6–8 hours of culture and 24, 48, 72, and 96 hours, the cells were treated with 10 μL of CCK-8, which was added into the complete culture medium. The cells were incubated at 37°C for 4 hours. Absorbance was measured at 450 nm. All experiments were performed in triplicate.

Colony-formation assay
For the colony-formation assay, cells were trypsinized into single-cell suspensions after transfection. A total of 1,000 cells were plated into each median plate and maintained in complete culture medium, which was replaced every 3–5 days. After 14 days, colonies were fixed with 4% paraformaldehyde and stained with 0.1% crystal violet (Beyotime, Haimen, China) for 20 minutes. Finally, visible colonies (>50 cells/colony) were manually counted after being washed twice with double-distilled H2O. Triplicate plates were measured in each treatment group.

Cell-migration and -invasion assays
Cell-invasion and -migration assays were performed in a transwell chamber coated with or without Matrigel (BD Biosciences, San Jose, CA, USA) 48 hours after siRNA transfection. Cells (2.5×10⁶) were suspended in 200 μL of serum-free medium and seeded in the upper chamber of a transwell. Culture medium (500 μL) containing 10% fetal bovine serum was placed into the bottom chamber. After incubation for 72 hours in a humidified atmosphere of 5% CO₂ at 37°C, the cells on the upper surface of the transwell chambers were removed using cotton swabs. Cells on the bottom surface of the transwell chambers were fixed with polyoxymethylene and stained with 0.1% crystal violet for 20 minutes. The invaded cells were counted in five randomly selected fields under microscopy, and the average value was calculated. Each experiment was conducted in triplicate. Matrigel invasion assays were performed with the same methods.

Protein extraction and Western blot assay
Transfected cells were washed twice with cold phosphate-buffered saline and lysed using the mammalian protein-cleavage
reagent radioimmunoprecipitation assay (Beyotime) supplemented with protease inhibitors (Hoffman-La Roche, Basel, Switzerland) and phenylmethylsulfonyl fluoride (Hoffman-La Roche) at 4°C for 30 minutes. Cellular debris were subsequently removed by centrifugation at 12,000 rpm for 10 minutes. The supernatants were mixed with quarter-volume 5× sodium dodecyl sulfate sample buffer and boiled for 10 minutes. Protein samples were separated by 8%–10% sodium dodecyl sulfate polyacrylamide-gel electrophoresis and then transferred onto polyvinylidene difluoride (PVDF) membranes (Bio-Rad Laboratories, Hercules, CA, USA). The PVDF membranes were soaked in 5% w/v skim milk at room temperature for 2 hours and then incubated overnight with primary antibodies at 4°C. The PVDF membranes were subsequently incubated with horseradish peroxidase-conjugated goat antirabbit secondary antibodies (Cell Signaling Technology, Danvers, MA, USA) for 1 hour at room temperature. An enhanced-chemiluminescence chromogenic substrate was used to visualize the bands, and the intensity of the bands was quantified by Image Lab software (Quantity One; Bio-Rad). GAPDH antibody (Cell Signaling Technology) was used as control. Rabbit monoclonal antibodies of PI3K, Akt, mTOR, vimentin, E-catenin, N-cadherin, and MMP9 were purchased from Cell Signaling Technology.

Statistical analysis
All results are expressed as mean ± standard deviation, and statistical analyses were performed using SPSS version 19.0 (IBM, Armonk, NY, USA). Student’s t-test and χ² tests were performed, and P<0.05 indicated statistical significance. The Kaplan–Meier method was applied for survival analysis, while related data were analyzed using Cox proportional-hazard regression model.

Results
LINC01296 expression in prostate cancer tissues and cell lines
lncRNA-expression profiles of three pairs of prostate cancer samples were screened through microarray data analysis (Figure 1A). qRT-PCR was used to examine the expression of
LINC01296 in 70 pairs of prostate cancer tissues and adjacent normal tissues. The qRT-PCR result was consistent with the microarray data. Additionally, LINC01296 expression was significantly upregulated in prostate cancer tissues compared with paired adjacent normal tissues (Figure 1B). LINC01296 expression in two prostate cancer cell lines (22Rv1 and LNCaP) and in a normal human myofibroblast stromal cell line (WPMY1) was further detected by qRT-PCR. The result revealed that LINC01296 was overexpressed in 22Rv1 and LNCaP, but not in WPMY1 (Figure 1C).

Correlations between LINC01296 expression and clinicopathologic features in prostate cancer patients

Based on LINC01296 messenger RNA expression, patients were classified into two groups: a high LINC01296-expression group (n=54) and a low LINC01296-expression group (n=16). Clinicopathologic features of these groups were compared (Table 1). Statistical results demonstrated that the high LINC01296-expression group was significantly correlated with high preoperative prostate-specific antigen (P=0.002), lymph-node metastasis (P=0.035), Gleason score (P<0.001), and tumor stage (P=0.036) compared with the low LINC01296-expression group. LINC01296-expression level was not correlated with age (P=0.569), prostate volume (P=0.794), or multiple lesions (P=0.171).

Table 1 The association between LINC01296 expression and clinicopathological features of prostate cancer

<table>
<thead>
<tr>
<th>Age (years)</th>
<th>Total (%)</th>
<th>LINC01296 expression</th>
<th>P-value</th>
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<tr>
<td>&lt;65</td>
<td>14 (20)</td>
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<td>10 (71.4)</td>
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<td>≥65</td>
<td>56 (80)</td>
<td>12 (21.4)</td>
<td>44 (78.6)</td>
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<td>Prostate volume (cm³)</td>
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<tr>
<td>&lt;50</td>
<td>37 (52.9)</td>
<td>8 (21.6)</td>
<td>29 (78.4)</td>
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<tr>
<td>≥50</td>
<td>33 (47.1)</td>
<td>8 (24.2)</td>
<td>25 (75.8)</td>
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<td>Preoperative PSA (ng/mL)</td>
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<tr>
<td>&lt;10</td>
<td>29 (41.4)</td>
<td>12 (41.4)</td>
<td>17 (48.6)</td>
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<tr>
<td>≥10</td>
<td>41 (58.6)</td>
<td>4 (9.8)</td>
<td>37 (90.2)</td>
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<td>Lymph-node metastasis</td>
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<td>Absence</td>
<td>32 (45.7)</td>
<td>11 (34.4)</td>
<td>21 (65.6)</td>
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<tr>
<td>&lt;7</td>
<td>23 (32.9)</td>
<td>12 (52.2)</td>
<td>11 (47.8)</td>
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<tr>
<td>≥7</td>
<td>47 (67.1)</td>
<td>4 (8.5)</td>
<td>43 (91.5)</td>
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<td>T2</td>
<td>28 (40)</td>
<td>10 (35.7)</td>
<td>18 (64.3)</td>
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<td>T3–T4</td>
<td>42 (60)</td>
<td>6 (14.3)</td>
<td>36 (85.7)</td>
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<tr>
<td>Presence</td>
<td>43 (61.4)</td>
<td>9 (20.9)</td>
<td>34 (79.1)</td>
</tr>
<tr>
<td>Absence</td>
<td>27 (38.6)</td>
<td>7 (25.9)</td>
<td>20 (74.1)</td>
</tr>
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Association between LINC01296 expression and prognosis of prostate cancer

We further analyzed the correlation of LINC01296 expression with the prognosis of patients with prostate cancer after radical prostatectomy. Kaplan–Meier analysis results showed that prostate cancer patients with high LINC01296 expression displayed significantly shorter biochemical recurrence-free survival time than those with low LINC01296 expression (P=0.01, Figure 2). As shown in Table 2, LINC01296-expression level and Gleason score were significantly correlated with biochemical recurrence-free survival rate of prostate cancer patients (P<0.05). Univariate and multivariate analyses showed that the relative expression of LINC01296 and Gleason score were independent prognostic markers for biochemical recurrence-free survival of prostate cancer patients (Table 2). These data collectively indicated that high LINC01296 expression was an independent risk factor for prostate cancer patients.

Knockdown of LINC01296 inhibits prostate cancer-cell proliferation

To verify the biological functions of LINC01296 in prostate cancer cell lines, we downregulated LINC01296 expression by transfecting siLINC01296 or an NC siRNA into 22Rv1 and LNCaP cells. qRT-PCR results showed that siLINC01296 demonstrated the highest inhibition efficiency,
Table 2 Cox proportional-hazard model of biochemical recurrence-free survival in prostate cancer

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<thead>
<tr>
<th></th>
<th>Univariate analysis</th>
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<th>Multivariate analysis</th>
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<td></td>
<td>HR</td>
<td>95% CI</td>
<td>P-value</td>
<td>HR</td>
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<tr>
<td>Age (years)</td>
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<tr>
<td>≥65 vs &lt;65</td>
<td>1.18</td>
<td>0.545–2.552</td>
<td>0.675</td>
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<td>Prostate volume (cm³)</td>
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<tr>
<td>≥50 vs &lt;50</td>
<td>0.911</td>
<td>0.476–1.746</td>
<td>0.78</td>
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<tr>
<td>Preoperative PSA (ng/mL)</td>
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<tr>
<td>≥10 vs &lt;10</td>
<td>1.879</td>
<td>0.906–3.898</td>
<td>0.09</td>
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<tr>
<td>Lymph-node metastasis</td>
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<td>Presence vs absence</td>
<td>1.319</td>
<td>0.687–2.535</td>
<td>0.406</td>
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<tr>
<td>Gleason score</td>
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<tr>
<td>≥7 vs &lt;7</td>
<td>3.429</td>
<td>1.426–8.247</td>
<td>0.491</td>
<td>2.965</td>
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<td>Tumor stage</td>
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<tr>
<td>T3–T4 vs T2</td>
<td>1.272</td>
<td>0.642–2.519</td>
<td>0.514</td>
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<td>Multiple lesions</td>
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<tr>
<td>Presence vs absence</td>
<td>0.799</td>
<td>0.407–1.569</td>
<td>0.006</td>
<td>0.002</td>
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<tr>
<td>LINC01296 expression</td>
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<tr>
<td>High vs low</td>
<td>6.584</td>
<td>1.951–22.224</td>
<td>0.044</td>
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</table>

Abbreviations: HR, hazard ratio; CI, confidence interval; PSA, prostate specific antigen.

Cell colony-formation assays were used to investigate further the effect of LINC01296 on proliferation of 22Rv1 and LNCaP cells. The results showed that the colony-formation numbers of 22Rv1 and LNCaP cells transfected with siLINC01296 were obviously lower than those transfected with NC siRNA (Figure 3C and D, P<0.05). All of the results were consistent with those of cell-proliferation assays.
Western blot analysis was used to investigate the effect of LINC01296 knockdown on the PI3K–Akt–mTOR signaling pathway. Figure 3E–G shows that LINC01296 knockdown significantly reduced the protein-expression levels of phosphorylated PI3k (p-PI3k), phosphorylated Akt (Ser473) (p-Akt), and phosphorylated mTOR (p-mTOR) (P<0.05). These results suggested that LINC01296 is involved in the promotion of prostate cancer-cell proliferation and seems to be mediated by regulation of PI3K–Akt–mTOR pathway.

Knockdown of LINC01296 inhibited cell migration and invasion

We explored the effect of LINC01296 knockdown on prostate cancer-cell migration/invasion. Figure 4A and B shows that knockdown of LINC01296 inhibited the migration capacity of 22Rv1 and LNCaP cells compared with cells in the NC groups (P<0.05). Moreover, transwell invasion assays were performed to assess the effect of LINC01296 on the invasiveness of prostate cancer cells. The results showed that the invasion ability of siLINC01296-transfected prostate cancer cells was reduced compared with control cells (Figure 4C and D, P<0.05). These data collectively imply that LINC01296 is involved in prostate cancer-cell migration and invasion in vitro.

Considering that LINC01296 promoted prostate cancer-cell migration and invasion, we further investigated the effects of LINC01296 on epithelial–mesenchymal transition (EMT) markers and MMP9, a gene that is usually upregulated during
EMT in cancer cells. The result of Western blot analysis demonstrated that LINC01296 knockdown increased the protein expression of E-cadherin and reduced the protein expression of N-cadherin and vimentin. Moreover, MMP9 protein expression was significantly reduced (Figure 4E–G, *P<0.05).

**Discussion**

IncRNAs have recently emerged as critical elements in cell biology, as they contribute to a wide variety of cellular behaviors and functions. In cancer, IncRNAs are associated with the development and progression of tumors, contributing to essential cancer-cell functions, such as proliferation, invasion, and metastasis. Similar to other cancer types, prostate cancer is a complicated biological process characterized by myriad molecular abnormalities. IncRNAs in prostate cancer are nominated as critical actors in disease pathogenesis. Therefore, identification of cancer-associated IncRNAs and investigation of their clinical significance and functions may
reveal the missing piece of the well-known oncogenic and tumor-suppressor network puzzle.

According to the microarray data, we future-validated and confirmed the result. In our study, we retrospectively analyzed the expression of LINC01296 in 70 prostate cancer patients and found that LINC01296 was overexpressed in prostate cancer, consistent with the microarray data. Studies have also revealed that LINC01296 is upregulated in 22Rv1 and LNCaP cell lines but not in WPMY1, which is a normal prostate myofibroblast stromal cell line. Specifically, we investigated the relationship between the expression levels of LINC01296 and the clinicopathologic features of prostate cancer patients. Our results demonstrated that prostate cancer patients with high LINC01296 expression displayed poor biochemical recurrence-free survival. More importantly, univariate and multivariate analyses demonstrated that LINC01296 should be an independent prognostic factor for prostate cancer patients, and that it plays an important role in the development and progression of prostate cancer. A previous study showed that colorectal cancer patients with high expression of LINC01296 displayed significantly worse overall survival than those with low LINC01296 expression. However, the biomedical functions of LINC01296 in prostate cancer remain unclear.

We performed in vitro experiments to highlight further the function of LINC01296. We focused on the identification and characterization of LINC01296 in prostate cancer and its role in proliferation, migration, and invasion. siRNA-mediated knockdown of LINC01296 significantly reduced the proliferation and metastasis capability of 22Rv1 and LNCaP cells compared with the control group, suggesting that LINC01296 knockdown can suppress the development of prostate cancer. Our results suggested that high LINC01296 expression increases the malignant phenotypes of prostate cancer cells. We summarized the characteristics of LINC01296 relative to those of other previously reported lncRNAs: 1) LINC01296 expression was closely correlated with Gleason score and tumor stage of prostate cancer; 2) higher expression of LINC01296 resulted in worse biochemical recurrence-free survival; and 3) LINC01296 knockdown can significantly inhibit the growth and metastasis of prostate cancer cells. These findings suggested that LINC01296 can be used as a novel biomarker and therapeutic target in prostate cancer.

To identify the potential molecular mechanism through which LINC01296 contributes to cell proliferation of prostate cancer cells, we investigated the PI3K–Akt–mTOR signaling pathway. This is one of the three main signaling pathways, and plays an important role in the development and progression of cancer. Our previous research found that in prostate cancer, the IncRNA LOC400891 regulated cell proliferation through the PI3K–Akt–mTOR signaling pathway. In addition, CRNDE, an IncRNA, promotes glioma cell growth via the mTOR-signaling pathway. These results showed that siRNA-mediated LINC01296 silencing significantly reduced the PI3K–Akt–mTOR signaling pathway’s relevant protein-expression level compared with NC. Therefore, we hypothesized that LINC01296 regulates the PI3K–Akt–mTOR signaling pathway via epigenetic modification, thereby promoting tumorigenesis. EMT is an important step in metastasis, during which epithelial cells are transformed into mesenchymal cells under specific physiological and pathological conditions. MMP9 is frequently observed in prostate cancer progression, and the upregulated expression of MMP9 is usually correlated with the occurrence of metastasis or local progression of cancer. Studies have found that IncRNA promotes cancer metastasis by regulating the EMT process. In this study, we investigated the role of LINC01296 in the EMT process in prostate cancer, and the results indicated that LINC01296 knockdown significantly suppressed the EMT process by reducing N-cadherin-, vimentin-, and MMP9-expression levels and by increasing E-cadherin expression compared with the control group. We mechanistically demonstrated that LINC01296 is possibly an inducing factor of EMT, making it an important factor in prostate cancer metastasis. IncRNAs usually bind with protein targets to perform their function in tumorigenesis. Therefore, the binding partner of LINC01296 should be further explored.

**Conclusion**

We demonstrated that increased LINC01296 expression is a common event underlying prostate cancer, suggesting that LINC01296 acts as an independent negative prognosis factor for prostate cancer patients. The effects of LINC01296 on proliferation, migration, and invasion suggest that it promotes tumorigenesis in prostate cancer. Increased LINC01296 expression promotes prostate cancer growth and metastasis by activating the PI3K–Akt–mTOR signaling pathway and EMT, and thus is a promising prognostic factor and therapeutic target to treat prostate cancer.

**Acknowledgment**

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**Disclosure**

The authors report no conflicts of interest in this work.
References


