Focal adhesion kinases crucially regulate TGFβ-induced migration and invasion of bladder cancer cells via Src kinase and E-cadherin

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Abstract: Focal adhesion kinase (FAK) is a non-receptor protein-tyrosine kinase that is triggered off by special extracellular signals such as some growth factors and integrins. FAK is found in cell–matrix attachment sites and implicated in cell migration, invasion, movement, gene expression, survival and apoptosis. In this study, we aimed to investigate whether FAK plays a role in invasion and migration of bladder cancer cells. Using an FAK-specific small interfering RNA (siRNA) and an FAK inhibitor PF-228, we found that inhibition of FAK tyrosine phosphorylation or knockdown of FAK suppressed invasion and migration of bladder cancer cells. Src is an important mediator of FAK-regulated migratory and invasive activity. Tyrosine phosphorylation of Src and FAK is mutually dependent and plays a key role in transforming growth factor beta (TGFβ)-induced invasion and migration. E-cadherin acts downstream of FAK and is a critical negative regulator in FAK-regulated invasion and migration of bladder cancer cells. These findings imply that FAK is involved in oncogenic signaling of invasion and migration, which can be a novel therapeutic target to treat patients with bladder cancer.

Keywords: focal adhesion kinase, transforming growth factor beta, migration/invasion, bladder cancer, Src kinase, E-cadherin

Introduction

Bladder cancer is the eleventh most frequent malignancy, which accounts for 3.1% of all new cancer cases worldwide. Annually, ~429,793 people suffer from this disease and ~165,084 people die from this disease.¹ Moreover, bladder cancer is the most common malignancy in the urinary system and the seventh most common cancer in men all over the world. About 75% of patients who were diagnosed with bladder cancer that was limited to the mucosa or sub-mucosa, were grouped as non-muscle-invasive bladder cancer. However, ~25% of patients were diagnosed with muscle-invasive bladder cancer, which implied that cancer cells had invaded into bladder detrusor.² Moreover, 10%–20% of initially diagnosed non-muscle-invasive bladder tumors develop into muscle-invasive cancer after organ-preserving treatment, in 5 years.³ Approximately 25% of patients with muscle-invasive bladder cancer present with cancer cell metastasis to lymph nodes when radical cystectomy was performed, and it was thought that undetected metastases occurred in one-third of patients suffering from muscle-invasive bladder cancer at the time of the first diagnosis.⁴ Consequently, these facts underlined a critical need for the study on how to inhibit the metastasis and invasion of bladder cancer.

Focal adhesion kinase (FAK), a 125 kDa protein, is one of a small and young family of structurally distinct protein tyrosine kinases without receptors such as FAK...
and PYK2, which can be triggered off by special extracellular signals such as some growth factors and integrins. The \textit{pik2} gene, which encodes the messenger RNA (mRNA) of FAK, locates at 8q24.3, a site commonly amplified in several cancers such as serous ovarian cancer\(^6\) and gastric cancer.\(^7\) Therefore, the role of FAK in the malignant progression was assessed in several cancers.

Up till now, unusual increases in FAK expression were found in multiple types of cancers,\(^8\) such as lung cancer,\(^9\) ovarian cancer,\(^10\) breast cancer\(^11\) and cervical cancer.\(^12\) It is known that FAK is a scaffolding protein with several functional domains that links the input of transmembrane signals from growth factor receptors or integrins to several effector proteins within cells such as PI3K–AKT, Src and JNK.\(^13\) Therefore, FAK is important for signal transduction within the tumor microenvironment.\(^14\) FAK proteins regulate several processes of cellular biology, which are important for the occurrence and progression of cancer, including cell survival and apoptosis, as well as adhesion, invasion and migration.\(^15\)

Recent reports showed that FAK is overexpressed in bladder cancer,\(^6\) one of the most common tumors of the urinary system. However, no study up till now is available regarding whether FAK is related to invasion and metastasis of bladder cancer. In this study, we explored the role of FAK in the invasion and migration of bladder cancer cells.

Materials and methods
Preparation and transfection of small interfering RNAs (siRNAs)
As described previously,\(^16\) siRNAs were designed against Src (S: 5′ ACU CGC CUU CUU AGA GUU U [dT][dT] 3′, AS: 5′ AAA CUC UAA GAA GGG GAG GAG U [dT][dT] 3′), FAK (S: 5′ UAA UAC UCG CUC CAU UGC ACC [dT][dT] 3′, AS: 5′ GGU GCA AUG GAG CGA GUA UUA [dT][dT] 3′) and E-cadherin (S: 5′ GGG UUA AGG ACA ACA GCA A [dT][dT] 3′, AS: 5′ UUG CUG UUG UGC AGA ACC C [dT][dT] 3′). The duplexes of the siRNAs were chemically synthesized (GeneChem, Shanghai, China) and used for transfecting bladder cancer cells by Lipofectamine\(^{TM}\) 2000 Transfection Reagent (Thermo Fisher Scientific, Waltham, MA, USA).

Cell culture
T24 and 5637 cells, two cell lines of human bladder cancer, were obtained from American Type Culture Collection. These cells were maintained in Dulbecco’s Modified Eagle’s Medium (GIBCO, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (FBS) at 37°C under a humidified atmosphere containing 5% CO\(_2\).

Quantitative polymerase chain reaction (PCR)
Total RNAs were prepared from the cells using TRIzol\(^{®}\) reagent (Thermo Fisher Scientific) in accordance with the manufacturer’s instructions. Primers were synthesized (Sangon, Shanghai, China) for quantitative PCR, which was performed with SYBR Green dye in an ABI7300 Real-time PCR System (Applied Biosystems, Foster City, CA, USA). The expression levels of target genes mRNA were relatively quantified with the 2\(^{-}\Delta\Delta t\) method.\(^17\)

Transwell assays
Transwell cell migration assays were performed in 24-well Transwell plates (Costar, Cambridge, MA, USA) according to the slightly modified method of Pelletier et al.\(^18\) The lower chambers of the Transwell plates were filled with medium with 20% FBS, and the upper chambers of the Transwell plates were used for culturing cells. The upper and lower chambers were separated by membranes with a pore size of 8 μm. After 24 hours of incubation, the upper surface of the Transwell membranes was lightly scraped to clean non-migrated cells, and the membrane was stained with crystal violet to count the migrated cells in the lower surface. Transwell cell invasion assays were performed in a similar way, but the upper chambers were coated with Matrigel.

Western blotting
Western blotting was carried out as described elsewhere.\(^19\) The treated cells were washed and collected and then resuspended in lysis buffer for further protein extraction. For each sample, 50 mg of protein was loaded onto gel electrophoresis (10% SDS-PAGE). ImageJ software (NIH, USA) was used to calculate the cumulative gray levels of each Western blotting band for further relative quantified analysis. The following antibodies were used as primary antibodies: β-actin (sc-1616-R) and GAPDH (sc-59540) from Santa Cruz (Santa Cruz, CA, USA) and E-cadherin (3195), pFAK (3283), FAK (3285), pSRC (2101) and Src (2109) from Cell Signaling (Beverly, MA, USA). Enhanced chemiluminescence (ECL; Pierce, Rockford, IL, USA) was used to visualize the bands.

Statistical analyses
Each experiment was done at least thrice. SPSS 12.0 (SPSS, USA) was used for analyzing the data. Student’s \(t\) test and one-way analysis of variance (ANOVA) were performed.
for statistical analyses of the data. For all statistical results, \( P \)-value <0.05 was considered statistically significant difference.

Results
Reducing FAK impairs invasion and migration of bladder cancer cells
To identify if reducing FAK impaired invasion and migration of bladder cancer cells, a specific siRNA duplex was used for knocking down FAK in bladder cancer cells, which was selected by quantitative PCR from several candidates. The results of Western blotting showed powerful interference effect of the selected siRNA duplex against the expression of FAK protein in bladder cancer T24 and 5637 cells as shown in Figure 1A and B. Further, the results of Transwell experiments showed that the selected siRNA duplex against FAK attenuated migratory and invasive ability of bladder cancer T24 cells (Figure 1C).

Transforming growth factor beta (TGF\(\beta\)) induces FAK phosphorylation and enhances invasive and migratory potential of bladder cancer cells
TGF\(\beta\) is regarded as one of the important metastasis-related growth factors. We further determined the effects of TGF\(\beta\) on FAK phosphorylation, as well as cell invasion and migration. The results of Western blotting showed that TGF\(\beta\) (5 ng/mL) did not change the expression of general FAK but significantly increased the expression of tyrosine-phosphorylated FAK in bladder cancer T24 and 5637 cells (Figure 2A and B). Src, an FAK-related kinase, was also tyrosine phosphorylated by TGF\(\beta\) (Figure 2A and B). Moreover, TGF\(\beta\) significantly

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**Figure 1** FAK was knocked down by a selected siRNA in bladder cancer cells.
**Notes:** T24 and 5637 cells were transfected with a selected siRNA. (A and B) Expression of FAK protein was measured by Western blotting. (C) Cell invasion and migration were measured by Transwell assays. Scale bar, 200 \( \mu \)m.

**Abbreviations:** FAK, focal adhesion kinase; siRNA, small interfering RNA; TGF\(\beta\), transforming growth factor beta; conRNA, control RNA.
decreased the expression of E-cadherin, a metastasis-related protein (Figure 2A). The results of Transwell experiments showed that TGFβ significantly increased cell migration and invasion of bladder cancer T24 cells (Figure 2C).

### Inhibition of FAK phosphorylation reduces TGFβ-induced migration and invasion of bladder cancer cells

To further explore if inhibition of FAK phosphorylation regulated TGFβ-induced migration and invasion of bladder cancer cells, we employed PF-228 (Sigma, USA), a small-molecule selective inhibitor of FAK, to reduce tyrosine phosphorylation level of FAK. The results of Western blotting showed that PF-228 (10 μM) significantly decreased TGFβ-stimulated tyrosine phosphorylation of not only Src but also FAK (Figure 3A and B). The results of Transwell experiments showed that PF-228 reduced TGFβ-induced cell migration and invasion of T24 cells (Figure 3C).

Src is an important mediator of FAK-regulated invasive and migratory activity of bladder cancer cells

To further study if suppression of Src phosphorylation regulated TGFβ-induced migration and invasion of bladder cancer cells, PP2 (Sigma, USA), a selective Src family tyrosine kinase inhibitor, was used for reducing tyrosine phosphorylation level of Src. The results of Western blotting showed that PP2 (10 μM) significantly cut down TGFβ-stimulated tyrosine phosphorylation levels of not only FAK but also Src (Figure 4A and B), which was similar to PF-228. The results of Transwell experiments showed that PP2 reduced TGFβ-induced cell migration and invasion of T24 cells (Figure 4C). To further study the relationship of tyrosine phosphorylation of Src and that of FAK, a specific siRNA duplex against Src was employed to knock down Src in bladder cancer cells. The results of Western blotting showed that the siRNA not only knocked down Src but also reduced...
tyrosine-phosphorylated levels of FAK (Figure 4D and E). The results of Transwell experiments demonstrated that the siRNA significantly reduced TGFβ-induced cell migration and invasion of T24 cells (Figure 4F).

**E-cadherin regulates invasive and migratory capability of bladder cancer cells as a downstream signal of FAK**

The loss of E-cadherin was one of the most important risk factors for metastasis of several carcinomas. Therefore, we further studied whether E-cadherin regulated the invasive and migratory capability of bladder cancer cells as a downstream signal of FAK. Western blotting analysis showed that E-cadherin was significantly upregulated when FAK was repressed by PF-228 or specific siRNA in bladder cancer T24 and 5637 cells (Figure 5A and B). On the other hand, the expression of not only tyrosine-phosphorylated but also general FAK was not regulated via the knockdown of the E-cadherin by specific siRNA (Figure 5C and D). The results of Transwell experiments demonstrated that the inhibiting effect of PF-228 on TGFβ-stimulated cell invasion and migration was reversed by the knockdown of the E-cadherin in T24 cells (Figure 5E and F).

**Discussion**

In 1992, FAK was first found by Schaller et al. and identified as a 125 kDa substrate of v-Src in chicken embryo fibroblasts. They found that FAK was a major phosphotyrosine-containing protein and tyrosine phosphorylated in the cells that were transformed by v-Src. The protein was named as focal adhesion kinase for its location to focal adhesions. Subsequently, it was found by Schaller et al. in 1994 that integrin-linked FAK exhibited tyrosine-phosphorylation and that this phosphorylation regulated cell migration and invasion. Therefore, FAK is a crucial molecule in the regulation of cellular motility and invasion.

Figure 3 Suppression of FAK phosphorylation reduces TGFβ-induced migration and invasion of bladder cancer cells.

Notes: T24 and 5637 cells were treated with 5 ng/mL TGFβ and 10 μM PF-228. (A and B) Expression of Src, pSrc, FAK and pFAK was examined by Western blotting. (C) Cell invasion and migration were measured by Transwell assays. Scale bar, 200 μm.

Abbreviations: FAK, focal adhesion kinase; TGFβ, transforming growth factor beta.
phosphorylation depending on extracellular matrix and physically associated to Src through their SH2 domains. Moreover, they also found that Tyr397 was a major site for FAK auto-phosphorylation. These findings suggested an elementary model that described the mechanisms of FAK activation and the mutual regulation of FAK and Src. Currently, it was found that FAK regulates cell migration, invasion, movement, gene expression, survival and apoptosis. The goal of this study was to explore whether suppression of FAK impaired migration and invasion of bladder cancer cells.

RNA interference (RNAi) is a powerful tool for silencing specific genes by degrading the target mRNA. Several studies have shown that expression of FAK was downregulated by some specific siRNAs targeting mRNA

![Image](https://www.dovepress.com/)

**Figure 4 (Continued)**
Figure 4 The role of Src in FAK-regulated migratory and invasive activity of bladder cancer cells.

Notes: T24 and 5637 cells were treated with 10 μM PP2 or siRNA against Src. (A, B, D and E) Expression of Src, pSrc, FAK and pFAK was measured by Western blotting. (C and F) Cell invasion and migration were examined by Transwell assays. Scale bar, 200 μm.

Abbreviations: FAK, focal adhesion kinase; siRNA, small interfering RNA; TGFβ, transforming growth factor beta; conRNA, control RNA.

Figure 5 (Continued)
of ptk2 gene. Hence, RNAi was employed to explore the role of suppressing FAK on the migration and invasion of bladder cancer cells. In the further experiments, the results of knockdown demonstrated that siRNAs against ptk2 gene significantly inhibited both the protein of FAK and the mRNA of ptk2 gene.

FAK is a focal adhesion-related protein and has been shown to be associated with migration and invasion since its initial discovery in 1992.20,21 Up to now, FAK was implicated as an important regulator of invasion and migration of several cancers such as colon cancer,28 ovarian cancer,29 lung cancer,30 tongue squamous31 and thyroid cancer.32 Our results showed that inhibiting FAK expression via RNAi significantly attenuated invasion and migration of bladder cancer cells.

It is known that FAK is a vital mediator of TGFβ signaling.33 Walsh et al34 suggested that TGFβ stimulated FAK synthesis as well as FAK phosphorylation through p38- and Smad-dependent pathways. However, most researchers found that TGFβ regulated FAK mainly via FAK phosphorylation.35-37 Our results showed that TGFβ stimulated FAK phosphorylation rather than FAK synthesis. Moreover, TGFβ enhanced the migration and invasion of bladder cancer cells. Because PF-228 is a highly selective small molecular inhibitor of FAK and can inhibit FAK phosphorylation at Tyr397,38 we employed PF-228 as a tool to inhibit FAK phosphorylation in TGFβ-stimulated bladder cancer cells. The experimental results showed that PF-228 was able to inhibit TGFβ-stimulated tyrosine phosphorylation of Src and FAK and suppress invasion and migration of bladder cancer cells.

Src was identified as an FAK-related protein since its initial discovery in 1992.20,21 In effect, input signaling from growth factor receptors and integrins leads to FAK autophosphorylation at Tyr397 where Src links to FAK subsequently. Subsequently, FAK proteins are phosphorylated at Y576 and Y577, and then, FAK–Src complexes are formed and play a key role in mediating the signaling from growth factor receptors and integrins to downstream molecules such as ERKs and paxillin.39,40 So, Src protein is one of the most important regulators in FAK-associated signal transduction. PP2 is an inhibitor of Src family tyrosine kinases and shows >10,000-fold selection over JAK2 and ZAP-70.41 PP2 is able to block Tyr416 phosphorylation of Src. We employed PP2 as a tool to inhibit Src phosphorylation in TGFβ-stimulated bladder cancer cells. The experimental results showed that PP2 was able to inhibit tyrosine-phosphorylated levels of not only Src but also FAK in bladder cancer cells stimulated by TGFβ. PP2 also inhibited invasion and migration of TGFβ-stimulated bladder cancer cells. Moreover, knockdown of Src not only decreased the protein expression of Src but also inhibited tyrosine phosphorylation of FAK. It was no surprise that knockdown of Src suppressed invasion and migration of TGFβ-stimulated bladder cancer cells.

E-cadherin is an invasion and tumor suppressor protein, which is a prototype of the “classical” type 1 cadherins and one of the most important components of adherens junctions known as intercellular adhesions.42 The loss of E-cadherin regulated FAK signaling through α5-integrin upregulation in trophoblast cells.43 Our results showed that not only FAK inhibitor PF-228 but also siRNA against FAK were

**Figure 5** The role of E-cadherin in FAK-regulated migratory and invasive activity of bladder cancer cells.

**Notes:** T24 and 5637 cells were treated with sirna against e-cadherin or FaK. (A–D) Expression of e-cadherin, FaK and pFaK was measured by Western blotting. (E and F) Cell invasion and migration were measured by Transwell assays. Scale bar, 200 μm.

**Abbreviations:** FAK, focal adhesion kinase; sirna, small interfering RNA; TGFβ, transforming growth factor beta; e-cad, e-cadherin; conRNA, control RNA.
able to increase E-cadherin expression in TGFβ-stimulated bladder cancer cells. However, knockdown of E-cadherin did not change general and tyrosine-phosphorylated FAK. Moreover, the results of Transwell experiments showed that FAK inhibitor PF-228 suppressed TGFβ-stimulated invasion and migration of bladder cancer cells, which was able to be restored via knockdown of E-cadherin.

We demonstrated that FAK was a vital regulatory molecule of invasion and migration of bladder cancer cells. Knockdown of FAK inhibited invasion and migration of bladder cancer cells. TGFβ induced FAK phosphorylation and enhanced migratory and invasive potential. Moreover, suppression of FAK phosphorylation reduced TGFβ-induced invasion and migration. Src was one of the most important mediators of FAK-related migratory and invasive activity. Tyrosine phosphorylation of FAK and Src was mutually dependent and played a key role in TGFβ-induced invasion and migration. E-cadherin acted downstream of FAK and was a critical negative regulator in FAK-regulated invasion and migration of bladder cancer cells. These results suggested a mechanism for invasion and migration involving FAK-related oncogenic signal as well as a novel therapeutic target and strategy for treating patients who develop bladder cancer.

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Disclosure
The authors report no conflicts of interest in this work.

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