

Neutralizing activities against seasonal influenza viruses in human intravenous immunoglobulin

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Abstract: Influenza viruses A/H1N1, A/H3N2, and B are known seasonal viruses that undergo annual mutation. Intravenous immunoglobulin (IVIG) contains anti-seasonal influenza virus globulins. Although the virus-neutralizing (VN) titer is an indicator of protective antibodies, changes in this titer over extended time periods have yet to be examined. In this study, variations in hemagglutination inhibition (HI) and VN titers against seasonal influenza viruses in IVIG lots over extended time periods were examined. In addition, the importance of monitoring the reactivity of IVIG against seasonal influenza viruses with varying antigenicity was evaluated. A/H1N1, A/H3N2, and B influenza virus strains and IVIG lots manufactured from 1999 to 2014 were examined. The HI titer was measured by standard methods. The VN titer was measured using a micro-focus method. IVIG exhibited significant HI and VN titers against all investigated strains. Our results suggest that the donor population maintains both specific and cross-reactive antibodies against seasonal influenza viruses, except in cases of pandemic viruses, despite major antigen changes. The titers against seasonal influenza vaccine strains, including past strains, were stable over short time periods but increased slowly over time.

Keywords: IVIG, influenza, seasonal, neutralization, vaccine

Introduction

Influenza viruses A/H1N1, A/H3N2, and B are known seasonal viruses that undergo annual mutation. Therefore, to control/prevent influenza, vaccine programs against virus strains are determined every season by the World Health Organization and implemented by the health authorities of various countries. Intravenous immunoglobulin (IVIG) possesses significant neutralizing activities against influenza viruses.¹⁻⁴ IVIG is manufactured from pooled plasma that is derived from >10,000 healthy donors. Therefore, IVIG contains various antibodies against numerous human pathogens. The anti-seasonal influenza virus activity of IVIG seems to be an accumulation of host reactions against previous vaccinations and natural infections. Consequently, IVIG maintains a high neutralizing titer reflecting the epidemiological status of the donor population at a given time period and region.

The virus-neutralizing (VN) titer is considered a significant indicator of IVIG activity. However, anti-influenza virus activity is usually determined by the hemagglutination inhibition (HI) titer. This assay evaluates the inhibition of viral particles bound to sialic acid. In contrast, the VN assay evaluates the inhibition of viral attachment and penetration into the cell and the release of progeny virus particles. Although VN is considered a more significant functional marker than HI, the fluctuation of VN

activity in IVIG lots over extended time periods has yet to be carefully evaluated. Here, we determined the fluctuation in HI and VN titers in IVIG lots manufactured from 2009 to 2014 by examining their activities against influenza A/H1N1, A/H3N2, and B vaccine strains. In addition, we evaluated the fluctuation in HI and VN titers in several IVIG lots following prolonged storage (1999 to 2009) to evaluate how they responded to virus antigenic mutations in adults over the long term.

Materials and methods

The influenza A/H1N1 viruses were A/Yamagata/32/1989 (A/Yam), A/New Caledonia/20/1999 (A/NC), A/Brisbane/59/2007 (A/Bri), and A/California/7/2009 (A/Cal). Two clinical isolates, A/Osaka/18/2008 and A/Osaka/16/2008, were also used. The influenza A/H3N2 viruses were A/Uruguay/716/2007 (A/Uru), A/Victoria/210/2009 (A/Vic210), A/Victoria/361/2009 (A/Vic361), A/Texas/50/2012 (A/Tex), and A/New York/39/2012 (A/NY). The influenza B viruses were B/Florida/4/2006 (B/Flo), B/Brisbane/60/2008 (B/Bri), B/Wisconsin/01/2010 (B/Wis), and B/Massachusetts/02/2012 (B/Mas). The seasonal influenza vaccine strains from 2006 to 2015 are summarized in Table 1.

Three IVIGs, Venoglobulin IH 5% I.V. (V-IH; Japan Blood Products Organization, Tokyo, Japan; formerly the Benesis Corporation, Osaka, Japan [manufactured from 1999 to 2014]), Nisseki Polyglobin-N 5% I.V. (Poly-N; formerly the Japanese Red Cross Society, Tokyo, Japan [manufactured from 2013 to 2014]), and Poly-N 10% I.V. (formerly the Japanese Red Cross Society [manufactured from 2013 to 2014]), were used in this study. Experiments using these IVIGs were approved by the ethical committee of the Japan Blood Products Organization.

The VN assays were performed at two facilities (Facilities A and B) using identical protocols. In brief, viruses were

propagated in Madin–Darby canine kidney (MDCK) cells and the allantoic cavity of 10-day-old embryonated chicken eggs. Culture media and allantoic fluid were aliquoted and stored at -80°C prior to use. Virus infectivity was titrated by focus-forming units/mL in MDCK cells using the peroxidase–anti-peroxidase staining technique (micro-focus method).^{2,4,5} The results are expressed as the reciprocal of the lowest dilution demonstrating >50% neutralization. The HI assays were performed at a commercial test company (Facility C). The results are expressed as the reciprocal of the highest two-fold serial IVIG dilution demonstrating HI.

Results

The HI and VN titers of IVIG lots against the A/H1N1 influenza vaccine strains used from seasons 2008/09 to 2014/15 (A/Bri [to 2009/10] and A/Cal [from 2009/10]) are shown in Figure 1. The titers were stable for A/Bri but decreased with time when the virus was changed to A/Cal. Interestingly, the titers against A/Cal in lots manufactured from late 2010 to 2012, derived from source plasma collected before 2009, showed weak reactivity. This result suggested that the donor population had cross-reactive globulin against pandemic A/Cal. The titers increased in lots manufactured from 2012 to 2013, derived from source plasma collected around 2009. These results seem to indicate a major antigen change, such as an antigenic shift in the 2009 pandemic, and that the donor population quickly developed the anti-A/Cal-specific globulin by viral infection and/or vaccination. The HI and VN titers were correlated.

The HI and VN titers against the A/H3N2 influenza vaccine strains from seasons 2008/09 to 2014/15 (A/Uru [2008/09–2009/10], A/Vic210 [2010/11–2011/12], A/Vic361 [2012/13], A/Tex [2013/14], and A/NY [2014/15]) are shown in Figure 2. These results indicate that the antibody titers were stable over short time periods but slowly increased over extended periods. This suggests that the A/H3N2 virus did

Table 1 Vaccine strains from 2005/06 to 2014/15 seasons in Japan

Seasons	A H1N1		A H3N2	B (lineage)
	Before 2009 pandemic	After 2009 pandemic		
2005/06	A/New Caledonia/20/1999	–	A/New York/55/2004	B/Shanghai/361/2002 (Yamagata)
2006/07	A/New Caledonia/20/1999	–	A/Hiroshima/52/2005	B/Malaysia/2506/2004 (Victoria)
2007/08	A/Solomon Islands/3/2006	–	A/Hiroshima/52/2005	B/Malaysia/2506/2004 (Victoria)
2008/09	A/Brisbane/59/2007	–	A/Uruguay/716/2007	B/Florida/4/2006 (Yamagata)
2009/10	A/Brisbane/59/2007	–	A/Uruguay/716/2007	B/Brisbane/60/2008 (Victoria)
2009/10	–	A/California/7/2009	–	–
2010/11	–	A/California/7/2009	A/Victoria/210/2009	B/Brisbane/60/2008 (Victoria)
2011/12	–	A/California/7/2009	A/Victoria/210/2009	B/Brisbane/60/2008 (Victoria)
2012/13	–	A/California/7/2009	A/Victoria/361/2011	B/Wisconsin/01/2010 (Yamagata)
2013/14	–	A/California/7/2009	A/Texas/50/2012	B/Massachusetts/02/2012 (Yamagata)
2014/15	–	A/California/7/2009	A/New York/39/2012	B/Massachusetts/02/2012 (Yamagata)

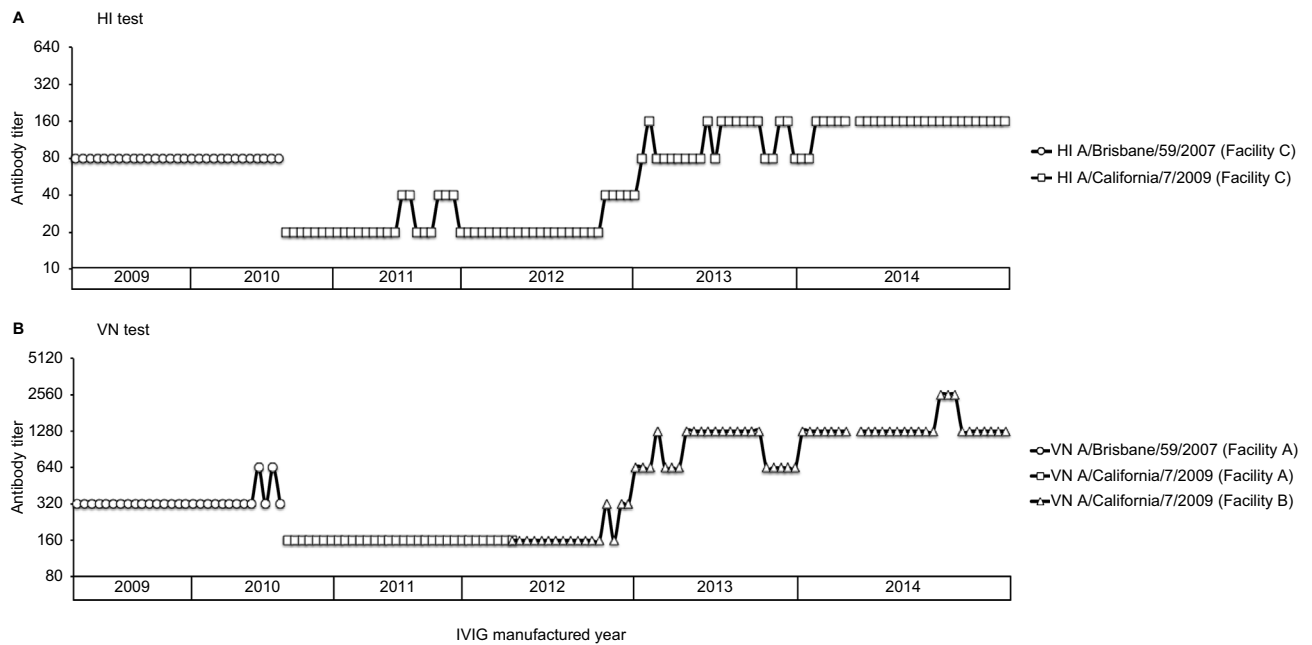


Figure 1 Changes in antibody titers against A/H1N1 influenza viruses in V-IH lots manufactured from 2009 to 2014.

Notes: (A) HI titers. Open circle: A/Brisbane/59/2007 (until 2010); open square: A/California/7/2009 (from 2010). Both tests were performed in Facility C. (B) VN titers. Open circle: A/Brisbane/59/2007 in Facility A until 2010; open square: A/California/7/2009 in Facility A between 2010 and 2012; open triangle: A/California/7/2009 in Facility B from 2012.

Abbreviations: V-IH, Venoglobulin IH; HI, hemagglutination inhibition; VN, virus neutralizing; IVIG, intravenous immunoglobulin.

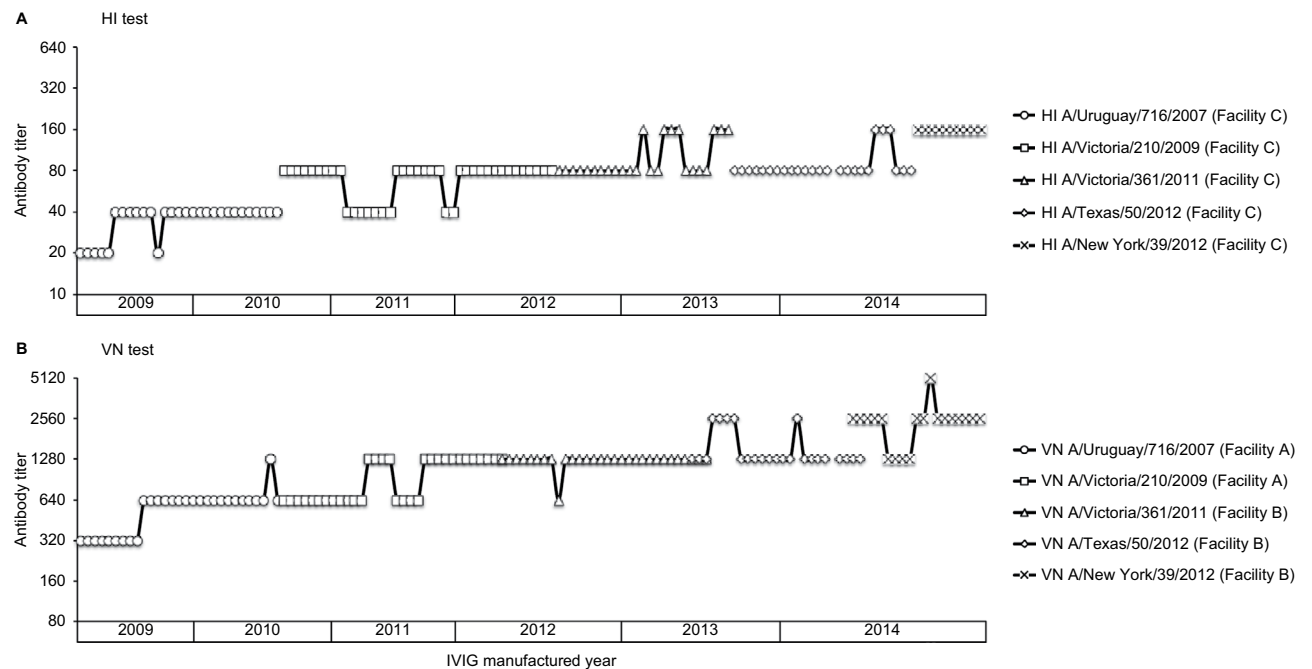


Figure 2 Changes in antibody titers against A/H3N2 influenza viruses in V-IH lots manufactured from 2009 to 2014.

Notes: (A) HI titers. Open circle: A/Uruguay/716/2007 until 2010; open square: A/Victoria/210/2009 between 2010 and 2012; open triangle: A/Victoria/361/2011 between 2012 and 2013; open diamond: A/Texas/50/2012 between 2013 and 2014; cross: A/New York/39/2012 from 2014. All tests were performed in Facility C. (B) Virus-neutralizing (VN) titers. Open circle: A/Uruguay/716/2007 in Facility A until 2010; open square: A/Victoria/210/2009 in Facility A between 2010 and 2012; open triangle: A/Victoria/361/2011 in Facility B between 2012 and 2013; open diamond: A/Texas/50/2012 in Facility B between 2013 and 2014; cross: A/New York/39/2012 in Facility B from 2014.

Abbreviations: V-IH, Venoglobulin IH; HI, hemagglutination inhibition; VN, virus neutralizing; IVIG, intravenous immunoglobulin.

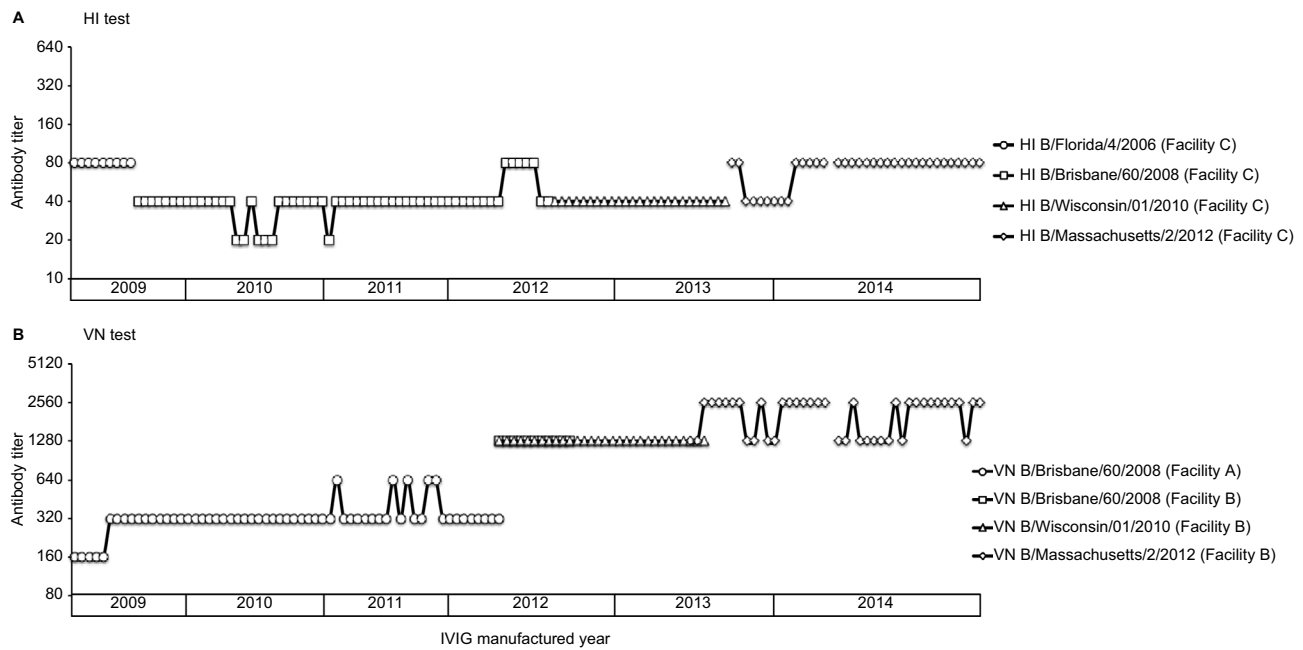


Figure 3 Changes in antibody titers against B influenza viruses in V-IH lots manufactured from 2009 to 2014.
Notes: (A) HI titers. Open circle: B/Florida/4/2006 until 2009; open square: B/Brisbane/60/2008 between 2009 and 2012; open triangle: B/Wisconsin/01/2010 between 2012 and 2013; open diamond: B/Massachusetts/2/2012 from 2013. All tests were performed in Facility C. (B) VN titers. Open circle: B/Brisbane/60/2008 in Facility A until 2012; open square: B/Brisbane/60/2008 in Facility B in 2012; open triangle: B/Wisconsin/01/2010 in Facility B between 2012 and 2013; open diamond: B/Massachusetts/2/2012 in Facility B from 2013.

Abbreviations: V-IH, Venoglobulin IH; HI, hemagglutination inhibition; VN, virus neutralizing; IVIG, intravenous immunoglobulin.

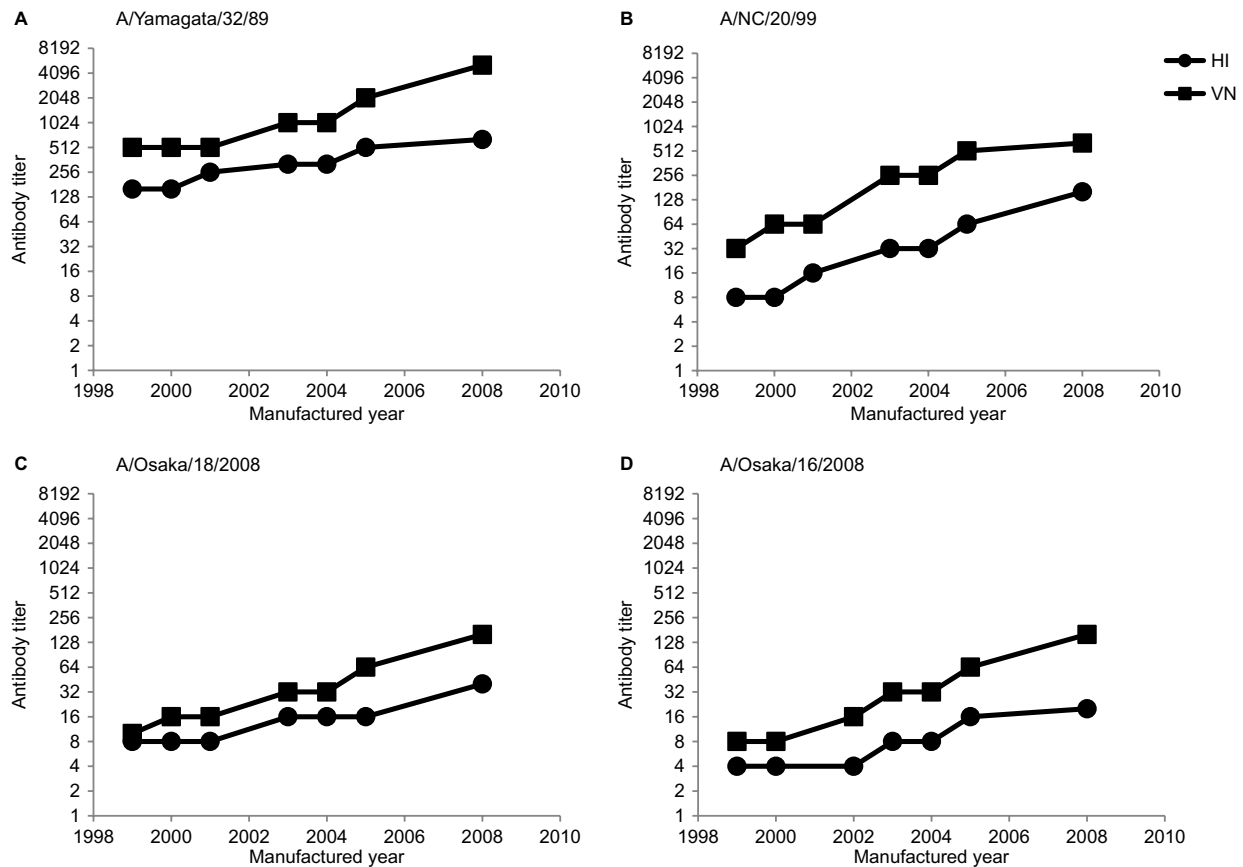


Figure 4 Changes in HI and VN titers against H1N1 influenza viruses isolated from 1989 to 2008 in representative V-IH lots manufactured from 1999 to 2008.
Notes: Antibody titers against vaccine strains (A and B). Antibody titers against clinical isolates (C and D). Closed circle: HI titer; closed square: VN titer.

Abbreviations: HI, hemagglutination inhibition; VN, virus neutralizing; V-IH, Venoglobulin IH.

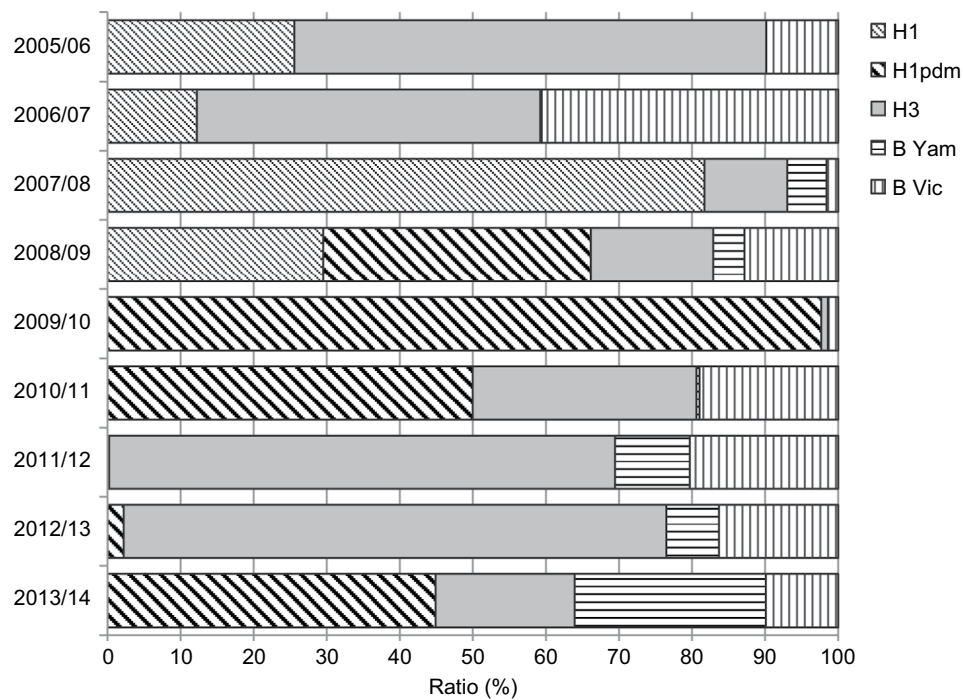


Figure 5 Influenza virus types isolated from 2005/2006 to 2014/2015 seasons in Japan.

Notes: Thin oblique line pattern: A/H1N1 (H1); thick oblique line pattern: A/H1N1 2009 pandemic (H1pdm); gray: A/H3N2 (H3); horizontal line pattern: B/Yamagata-lineage (B Yam); vertical line pattern: B/Victoria-lineage (B Vic).

not undergo a major antigen change. The HI and VN titers were again correlated.

The HI and VN titers against the influenza B vaccine strains from seasons 2008/09 to 2014/2015 (B/Flo [2008/09], B/Bri [2009/10–2011/12], B/Wis [2012/13], and B/Mas [2013/14–2014/15]) are shown in Figure 3. Interestingly, the VN titers against B/Bri measured by Facilities A and B showed differing results. In contrast, the VN titers against B/Bri (Victoria-group) and B/Wis (Yamagata-group) were similar. These results indicate that the titers were stable over long time periods. Note that the titers may indicate different results measured by different facilities.

These results indicate that, although the virus antigen underwent a change, the status of the donor population was not significantly altered. Taken together, this suggests that HI and VN titers can exhibit several patterns (stable, slowly increasing, and quickly increasing) when the target virus is changed; a significant antigenic shift has a greater impact than no major antigenic shift. The HI and VN titers are closely associated, though their mechanisms are different.

Variations in the HI and VN titers against A/H1N1 influenza viruses isolated from 1989 to 2008 in representative IVIG lots manufactured from 1999 to 2008 were also evaluated. The HI and VN titers against the viruses increased slowly with increasing year of manufacture (Figure 4A–D). In addition,

the IVIG lots exhibited higher HI and VN titers against older than recent isolates. The types of influenza viruses isolated from 2005/06 to 2014/15 are summarized from infectious agent surveillance reports (Figure 5).⁶ The proportion of isolated virus types varied in each season. Hence, the increase in HI and VN titers over extended time periods seems to be independent of the antigenicity of the epidemic viruses.

V-IH and Poly-N are manufactured using different process conditions. Thus, differences in the HI and VN titers of the three IVIG products were evaluated. The mean HI and VN titers are summarized in Table 2. The results indicate almost the same titer between V-IH and Poly-N (5% immunoglobulin G [IgG] for both). In addition, the results with Poly-N 5 and 10% indicated that the titers were proportional to concentration. The dilution methods of these tests were 2× serial dilutions. Therefore, values within 2× of the titers are non-significant differences. These results suggest that the titers are not affected by the manufacturing process conditions, although they may be affected by virus strains.

Discussion

In this study, the HI and VN titers of IVIG against influenza A/H1N1, A/H3N2, and B vaccine strains were evaluated. IVIG contains various antibodies against numerous human pathogens. Importantly, the individual donor plasma used for

Table 2 Antibody titers of IVIG products

Seasons	Virus strain	HI titers			VN titers		
		V-IH (IgG: 5%)	Poly-N (IgG: 5%)	Poly-N (IgG: 10%)	V-IH (IgG: 5%)	Poly-N (IgG: 5%)	Poly-N (IgG: 10%)
2012/13	A H1N1						
	A/California/7/2009	147 ± 100 (n = 27)	NE	NE	450 ± 402 (n = 27)	NE	NE
2013/14	A/California/7/2009	255 ± 81 (n = 22)	224 ± 105 (n = 10)	432 ± 227 (n = 10)	1135 ± 275 (n = 22)	864 ± 454 (n = 10)	3136 ± 2100 (n = 10)
2014/15	A/California/7/2009	320 ± 0 (n = 19)	267 ± 80 (n = 9)	512 ± 175 (n = 5)	1482 ± 480 (n = 19)	1138 ± 282 (n = 9)	2048 ± 701 (n = 5)
2012/13	A H3N2						
	A/Victoria/361/2011	178 ± 51 (n = 27)	NE	NE	1256 ± 123 (n = 27)	NE	NE
2013/14	A/Texas/50/2012	178 ± 60 (n = 22)	144 ± 34 (n = 10)	240 ± 84 (n = 10)	1571 ± 549 (n = 22)	1664 ± 810 (n = 10)	3456 ± 1484 (n = 10)
2014/15	A/New York/39/2012	152 ± 25 (n = 19)	116 ± 42 (n = 9)	224 ± 88 (n = 5)	2425 ± 842 (n = 19)	1489 ± 675 (n = 9)	3584 ± 1402 (n = 5)
2012/13	B						
	B/Wisconsin/01/2010	314 ± 175 (n = 27)	NE	NE	1280 ± 0 (n = 27)	NE	NE
2013/14	B/Massachusetts/02/2012	465 ± 163 (n = 22)	480 ± 169 (n = 10)	640 ± 0 (n = 10)	2095 ± 630 (n = 22)	2048 ± 661 (n = 10)	4352 ± 1237 (n = 10)
2014/15	B/Massachusetts/02/2012	589 ± 120 (n = 19)	640 ± 0 (n = 9)	896 ± 351 (n = 5)	2088 ± 634 (n = 19)	1422 ± 427 (n = 9)	4608 ± 1145 (n = 5)

Notes: Data are expressed as mean ± SD. V-IH and Poly-N are manufactured by different processes. Also, Poly-N comes in two different IgG concentrations: 5% (50 mg/mL) and 10% (100 mg/mL).

Abbreviations: IVIG, intravenous immunoglobulin; HI, hemagglutination inhibition; VN, virus neutralizing; V-IH, Venoglobulin IH; IgG, immunoglobulin G; Poly-N, Nisseki Polyglobin-N; NE, not examined; SD, standard deviation.

the IVIG lots manufactured from 2009 to 2014 examined in this study was collected several years prior to manufacture. Therefore, the HI and VN titers against vaccine strains exhibit time-lag effects, and the time-lag periods are not constant. With the lots used in this study, the lead time of V-IH was >3 years. Thus, the HI and VN titers of IVIG lots reflect the status of the donor population during the donation period.

Previously, we examined the HI and VN activities of IVIG lots against A/H1N1 2009 pandemic and A/H2N2 1957 pandemic.^{2,4} Interestingly, the results showed that IVIG possessed HI and VN activities against these viruses, although most donors were not exposed to either pandemic. These results suggest that the donor population plasma maintained cross-reactive globulins against the pandemic A/H1N1 and A/H2N2 viruses. Prior to this study, the HI and VN titers against seasonal influenza viruses over extended time periods had not been examined. Importantly, variations in the HI and VN titers before and after a major antigenic change, such as a pandemic caused by a novel virus, had not been evaluated.

The results of this study showed that all IVIG lots contained HI and VN activities against seasonal influenza vaccine strains. Overall, HI and VN activities were stable over short time periods and slowly increased over extended periods,

with little variance following changes in the vaccine strain. In contrast, in specific cases such as the major antigenic change that occurred during the 2009 pandemic, the HI and VN activities decreased, but subsequently increased following the spread of the virus to the donor population. These results suggest that IVIG contains stable levels of specific and cross-reactive antibodies against seasonal influenza viruses. In addition, the HI and VN titers varied slowly over extended periods, except when the vaccine strain was altered or a major antigenic change occurred in a pandemic.

Variances in HI and VN titers against influenza viruses isolated from 1989 to 2009 in representative IVIG lots manufactured from 1999 to 2008 indicate that these titers increased with year of manufacture. This result suggests that the level of HI and VN activity of IVIG reflects the effects of vaccination programs and natural infection of donor populations. Furthermore, monitoring HI and VN titers over extended time periods is important, especially when the vaccine strain is changed or an antigenic shift occurs, such as in the 2009 pandemic. Further studies are required to elucidate the mechanism(s) underlying the maintenance of antibody titers against influenza viruses over extended time periods and how they are affected by antigenic shift and drift.

IVIGs contain neutralizing antibodies against various influenza viruses and are thus expected to have a therapeutic effect. Unfortunately, knowledge of the therapeutic effect and the mechanism of IVIG against influenza has been limited, although some findings have been reported.^{7–10} Evaluation of the therapeutic effect of IVIG against influenza using an animal model has also been reported.^{11,12} IVIGs derived from plasma collected prior to 2009 showed a therapeutic effect on mouse-adapted A/NC; however, the effect against the 2009 pandemic virus was lower than against A/NC.¹¹ IVIG containing a high titer of antibodies against the 2009 pandemic virus elicited 100% survival in contrast with the 50% survival using IVIGs derived from plasma collected prior to 2009.¹² The mechanism underlying this effect requires further study because the efficacy of IVIG against influenza has yet to be verified.

Conclusion

IVIG exhibited significant HI and VN titers against all influenza strains investigated in this study. The HI and VN titers against seasonal influenza vaccine strains, including past strains, were stable over short time periods and increased slowly over extended periods. Our results suggest that the donor population maintains both specific and cross-reactive antibodies against seasonal influenza viruses over extended periods, except in cases of pandemic viruses with a major antigen change. Moreover, our results suggest that new antibodies were acquired and added to globulin, including a few cross-reactive antibodies, when antigenic shift occurred. New antibodies were also acquired against mutated viruses and added to globulin, maintaining reactivity to past viruses when antigenic drift occurred. Monitoring of VN titers following major antigenic changes, such as an antigenic shift, will be valuable for evaluating the function of IVIG against pandemic influenza strains.

Acknowledgments

This study was conducted in part as a collaborative research project between Osaka University, Rakuno Gakuen University, The Research Foundation for Microbial Diseases of Osaka University, and the Japan Blood Products Organization (formerly the Benesis Corporation). The authors thank Dr. Kaoru Sakai (Japan Blood Products Organization) for valuable discussions and technical support. We would also like to thank Editage (www.editage.jp) for English language editing.

Disclosure

H. Onodera, T. Urayama, and M. Yunoki are employees of the Japan Blood Products Organization. T. Urayama and M. Yunoki are also former employees of the Benesis Corporation. H. Onodera is a former employee of the Japanese Red Cross Society. K. Maeda, R. Kubota-Koketsu, K. Hirota, Y. Okuno, and K. Ikuta are employees of The Research Foundation for Microbial Diseases of Osaka University. K. Ikuta was also a Professor at Osaka University. K. Takahashi is currently affiliated to the Department of Clinical Laboratory, International University of Health and Welfare Hospital. The authors report no other conflicts of interest.

References

- Pichyangkul S, Jongkaewwattana A, Thitithanyanont A, et al. Cross-reactive antibodies against avian influenza virus A (H5N1). *Emerg Infect Dis*. 2009;15(9):1537–1539.
- Yunoki M, Kubota-Koketsu R, Urayama T, et al. Significant neutralizing activity of human immunoglobulin preparations against pandemic 2009 H1N1. *Br J Haematol*. 2009;148(6):948–963.
- Hong DK, Tremoulet AH, Burns JC, Lewis DB. Cross-reactive neutralizing antibody against pandemic 2009 H1N1 influenza A virus in intravenous immunoglobulin preparations. *Pediatr Infect Dis J*. 2011;30(1):67–69.
- Kubota-Koketsu R, Yunoki M, Okuno Y, Ikuta K. Significant neutralizing activities against H2N2 influenza A viruses in human intravenous immunoglobulin lots manufactured from 1993 to 2010. *Biologics*. 2012;6:245–247.
- Okuno Y, Tanaka K, Baba K, Maeda A, Kunita N, Ueda S. Rapid focus reduction neutralization test of influenza A and B viruses in microtiter system. *J Clin Microbiol*. 1990;28:1308–1313.
- NIID [webpage on the Internet]. *Infectious Agents Surveillance Report*. Available from: <http://www.nih.go.jp/niid/ja/typhi-m/iasr-reference/230-iasr-data/5492-iasr-table-v-p.html>. Accessed December 14, 2016.
- Gokturk B, Pekcan S, Guner SN, et al. Efficacy of intravenous immunoglobulin treatment in immunocompromised children with H1N1 influenza: a clinical observation. *Clin Respir J*. 2016;10(2):223–230.
- Davey RT Jr, Markowitz N, Beigel J, et al. INSIGHT FLU005: an Anti-Influenza Virus Hyperimmune Intravenous Immunoglobulin Pilot Study. *J Infect Dis*. 2016;213(4):574–578.
- Donovan S, Bearman GM. Use of intravenous immunoglobulin in critically ill patients. *Curr Infect Dis Rep*. 2014;16(12):447.
- Hung IF, To KK, Lee CK, et al. Hyperimmune IV immunoglobulin treatment: a multicenter double-blind randomized controlled trial for patients with severe 2009 influenza A(H1N1) infection. *Chest*. 2013;144(2):464–473.
- Hagiwara K, Kawami S, Kato-Mori Y, et al. Protective role of human intravenous immunoglobulin from influenza A virus infection in mice. *Open Hematol J*. 2012;6:8–11.
- Hohenadl C, Wodal W, Kerschbaum A, et al. Hyperimmune intravenous immunoglobulin containing high titers of pandemic H1N1 hemagglutinin and neuraminidase antibodies provides dose-dependent protection against lethal virus challenge in SCID mice. *Virol J*. 2014;11:70.

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